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Total number of authors: 
15

Published in: 
New Phytologist

Link to article, DOI: 
10.1111/nph.14018

Publication date: 
2016

Document Version 
Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA): 
The decision to germinate is regulated by divergent molecular networks in spores and seeds

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Summary

• Dispersal is a key step in land plant life cycles, usually via formation of spores or seeds. Regulation of spore- or seed-germination allows control over the timing of transition from one generation to the next, enabling plant dispersal. A combination of environmental and genetic factors determines when seed germination occurs. Endogenous hormones mediate this decision in response to the environment. Less is known about how spore germination is controlled in earlier-evolving nonseed plants.

• Here, we present an in-depth analysis of the environmental and hormonal regulation of spore germination in the model bryophyte Physcomitrella patens (Aphanoregma patens).

• Our data suggest that the environmental signals regulating germination are conserved, but also that downstream hormone integration pathways mediating these responses in seeds were acquired after the evolution of the bryophyte lineage. Moreover, the role of abscisic acid and diterpenes (gibberellins) in germination assumed much greater importance as land plant evolution progressed.

• We conclude that the endogenous hormone signalling networks mediating germination in response to the environment may have evolved independently in spores and seeds. This paves the way for future research about how the mechanisms of plant dispersal on land evolved.

Introduction

Transition from one generation to the next in land plants is mediated by the formation of desiccation-resistant dispersal units (Finch-Savage & Leubner-Metzger, 2006). Within the spermatophyte lineage, these dispersal units are multicellular seeds, whereas in bryophytes and nonseed vascular plants (lycophytes and ferns) the functionally equivalent dispersal units are unicellular spores (Linkies et al., 2010). How and when germination is initiated in a seed or spore is critical for plant and species reproduction, movement and survival.

Hormonal and environmental factors are both well-established as key players in the regulation of seed germination (Holdsworth et al., 2008a). The regulation of seed germination is highly complex, and involves integration of environmental signals by hormones within the seed and within different seed compartments and cell types (Yamaguchi et al., 2001; Holdsworth et al., 2008b; Linkies et al., 2010; Dekkers et al., 2013). Much is still unknown about exactly how seed germination is controlled at a cellular level (Nonogaki et al., 2010; Basel et al., 2014).

Spores, unlike seeds, are haploid and are derived from the sporophyte stage of the plant life cycle via meiosis (reviewed in Rubinstein et al., 2010). Despite the different developmental origins of spores and seeds, previous work suggests that at least some aspects of germination regulation may be conserved between the two types of dispersal unit, as outlined later. Because spores are unicellular structures, understanding spore germination provides us with a simplified system for the study of the cellular and hormonal basis of germination regulation, and how this has evolved.

Environmental regulation of seed germination in both monocots and dicots is controlled in part by a phytochrome-mediated reversible system, with red (R) light promoting germination, and even brief exposure to far-red (FR) light inhibiting R light-induced germination (Borthwick et al., 1952; Shinomura et al., 1994; Hennig et al., 2002), although this trait has been bred out of some commercial cereal crops (Barrero et al., 2012). A similar R–FR reversible system regulates spore germination in several ferns (Mohr et al., 1964; Raghavan, 1973; Wayne & Hepler, 1984; Scheuerlein et al., 1989; Tsuboi et al., 2012). In the earliest-evolving land plant lineage, bryophytes, complete inhibition of spore germination by FR light, and reversal of this inhibition by R light via phytochromes, has been demonstrated (Possart & Hilbrunner, 2013). Phytochrome regulation of spore germination is likely to be extremely ancient, as it appears to exist outside the land plant lineage, also, including in spores of the Charophycean algae Spirogyra and Chara, and in fungi (Calpouzos &
ABA is central to dormancy establishment and maintenance in Arabidopsis (Finkelstein et al., 2008). Freshly harvested Arabidopsis seeds contain high levels of ABA and show primary dormancy. ABA is also implicated in the imposition of secondary dormancy by, for example, high temperatures, via de novo synthesis of this hormone (Finch-Savage & Leubner-Metzger, 2006; Toh et al., 2008, 2012). The majority of bryophyte species' spores have not been reported to show primary dormancy (McLetchie, 1999; Glime, 2015). Whether spores can have secondary dormancy imposed on them is currently not well-characterized (Glime, 2015).

ABA also protects plants against abiotic stresses such as desiccation and freezing (Lee & Luan, 2012; Dekkers et al., 2015). Formation of both mature seeds and spores involves desiccation, and aspects of ABA signalling during abiotic stress responses in bryophytes are conserved with angiosperms (Knight et al., 1995; Cuming et al., 2007; Khandelwal et al., 2010). ABA-mediated stress tolerance in bryophytes occurs at least in part via accumulation of soluble sugars including sucrose (Burch & Wilkinson, 2002; Nagao et al., 2006; Oldenhof et al., 2006; Bhyan et al., 2012; Erxleben et al., 2012). Notably, soluble sugars (sucrose, glucose) can inhibit germination in Arabidopsis seeds (Dekkers et al., 2004; Li et al., 2012).

Several additional hormones regulate seed germination. Strigolactones (SLs) produced by host plants are potent promoters of parasitic plant seed germination (Bouwmeester et al., 2003; Yoneyama et al., 2010). In Arabidopsis, strigolactone signalling pathway mutants show reduced seed germination (Stanga et al., 2013) and strigolactone can overcome the secondary dormancy imposed by exposing Arabidopsis seeds to high temperatures (Toh et al., 2012). Ethylene also promotes seed germination via multiple routes (Stewart & Freebairn, 1969; Logan & Stewart, 1991; Linkies & Leubner-Metzger, 2012), whereas an inhibitory role for cytokinin in germination is implied in Arabidopsis (Riefler et al., 2006). A role for auxin in seed germination under normal conditions has not been demonstrated (Holdsworth et al., 2008a; Park et al., 2011); neither has the effect of these hormones on moss spore germination been investigated systematically (Chopra & Kumra, 1988; Glime, 2015).

Here, we sought for the first time to define comprehensively how hormones and environmental processes regulate spore germination in the bryophyte lineage by using Physcomitrella as a model system.

Materials and Methods

Physcomitrella culture and spore generation

Physcomitrella patens (Gransden wild-type (WT) strain, and Ppcps/ks and ced8 mutants) was cultured and sporation induced as in Moody et al. (2012). Sporophytes were harvested after maturation (dark brown sporophytes with a slightly ‘sparkly’ appearance) using sterile forceps under a SMZ645 dissecting microscope (Nikon, Tokyo, Japan) and air-dried in sterile tubes for c. 1 wk before storage at room temperature.

Spore germination assays

Spores from a minimum of three sporophytes (all of the same age) of a particular genotype were used within each assay. For
larger assays, three sporophytes’ worth of spores were used for every 10 Petri dishes (9 cm diameter). For assays comparing WT with a mutant, spores of each genotype were chosen to be of the same age, harvested at the same time.

Sporophytes were bleached in groups of two to three in 1 ml 25% Parozone™ (Jeyes Group, Thetford, UK) for 10 min and then washed three times in 1 ml sterile distilled water (10 min each) in a sterile flow cabinet. The sporophytes were then crushed in 100–200 μl of sterile distilled water to allow plating of 500 μl of spore solution per Petri dish. Spores were plated on cellophane-overlaid BCD supplemented with 5 mM CaCl₂ and 5 mM ammonium tartrate, or on water agarose for the dormancy experiments in Fig. 1. Cellophane discs (A.A. Packaging Ltd, Preston, UK) were autoclaved for 15 min at 121°C, before use.

For hormone/chemical treatments, the treatment in the relevant solvent (or a solvent-only control) was added to 1 ml of sterile water, which was then added to BCD medium just before plate-pouring to achieve the desired final concentration in the plates. Within each assay, all solvent-containing plates were matched so that all contained exactly the same volume of solvent, even if the hormone concentration varied. The solvents used were methanol for diterpenoids and norflurazon, acetone for GR24 and water for ethylene.

Plated spores were air-dried in a laminar flow hood, sealed with micropore tape, and placed at 22°C in long-day conditions. Spores were counted daily under the 94 objective of a Leica compound microscope with a 910 eyepiece; total magnification 940. A minimum of 200 spores per plate was counted to define the number of germinated and ungerminated spores. Data were expressed as percentage germination, that is: (germinated spores/total spores counted) × 100. A spore was defined as having germinated as soon as the very first deformation of the spore coat by the emerging protonemal filament was observed.

Two to three technical replicates were contained within each assay, and each assay was repeated a minimum of three times on different dates, using different batches of spores and medium, to provide biological replicates. Data from one representative assay is shown in each figure.

RNA preparation

RNA was prepared from dry spores (c. 250 harvested sporophytes), imbibed spores (c. 250 sporophytes bleached then soaked in liquid BCD for 18 h), germinating spores (spores from c. 250 bleached sporophytes plated at high density and left for 7 d until c. 50% germination was seen) and 100 mg vegetative (protonemal or leafy) tissue using the Bioline Isolate II Plant RNA preparation kit according to the manufacturer’s instructions (London, UK). Average RNA yield was c. 300 ng μl⁻¹, with OD260/280 > 2 and average OD 260/230 c. 1.8.

RT-PCR

Reverse transcription polymerase chain reaction (RT-PCR) was carried out on 20 ng RNA from each sample using the Bioline MyTaq™ one step RT-PCR kit according to the manufacturer’s instructions. Primer sequences are detailed in Supporting Information Table S1.

Generation of Physcomitrella patens lines with disrupted PpCPS/KS functionality

The moss line pCL755#29 is described in Zhan et al. (2015), generated using the methods described in (Bach et al., 2014). Briefly, a cassette containing p35S-nptII-CaMVter, expressing Neomycin Phosphotransferase II (conferring resistance to G418), was excised from pMBL6 (http://www.biology.wustl.edu/moss/pmb6.jpg) using XhoI and inserted into the XhoI site in pDONR201:CPS/KS, generating the knock-out construct pDONR201:CPS/KS-nptII. Following transformation into P. patens of this construct, one line – pCL755#29 – with disrupted PpCPS/KS functionality was used for further studies.

Fig. 1 Physcomitrella patens spore germination stages. (a) Dry spore. (b) Imbibed spore. (c) Spore coat cracking. (d) Protrusion of one protonemal (chloronemal) filament. (e) Protrusion of two protonemal filaments. (f) Established protonemal colony. Bars, 100 μm.
A second line, pBK3, was generated by disrupting \textit{PpCPS/KS} via targeted gene replacement using the pBK3 vector first described in Pan \textit{et al.} (2015) and utilizing the method described in King \textit{et al.} (2016), which contains a p35S-aph4-CaMVter cassette flanked by 5' and 3'-genomic sequence of \textit{PpCPS/KS} on its 5' and 3' end, respectively. Therefore, genomic sequence of \textit{PpCPS/KS} was replaced with p35S-aph4-CaMVter, which gives hygromycin resistance. The two lines were genotyped by PCR with the combination of primers that specifically bind to genomic DNA or selection marker cassette to distinguish knock-out mosses from WT (Fig. S1).

**GC-MS analysis of diterpenoids**

All GC-MS analyses were performed on a Shimadzu GCMS-QP2010 plus (GC-2010) with a CTC auto sampler AOC-5000, with cooled trays, agitation oven, and needle bake-out.

GC-MS analysis utilizing solid-phase microextraction fibers was previously published (Drew \textit{et al.}, 2012; Andersen \textit{et al.}, 2015). Briefly, the injection port temperature was set to 230°C, with a sampling time of 1 min. The flow control mode was pressure control with a total flow of 2.3 ml min⁻¹, with H₂ as carrier gas, and a purge flow of 1.0 ml min⁻¹. The column was a 30 m HP-5MS column. The oven temperature program was 35°C for 3 min, rising by 10°C per min to 230°C and a hold for 3 min.

The MS settings were: Ion source temperature 260°C, interface temperature 280°C and the scan range from m/z 50 to m/z 350 with 70 eV electrical ionization.

All data were analysed using the Shimadzu software Lab Solutions, GCMS S OLUTIONS v.2.70, using the libraries provided by NIST (NIST 08) and WILEY (WILEY 8.0). Obtained spectra were compared with the spectra in the mass spectral libraries. Compounds were identified comparing the data with library information of MS and retention indices (I). All reference Is were taken from Adams (2007).

**Results**

\textit{Physcomitrella} spores are nondormant and do not require cold-stratification or after-ripening treatment for germination.

We define a \textit{Physcomitrella} imbibed spore as having ‘germinated’ as soon as the first visible deformation of the spore coat occurs and the spore no longer appears spherical. This marks the first protrusion of protonemal filament(s), which grow with the subsequent formation of further filaments (Fig. 1a–f). Spores typically begin to germinate 2–7 d after imbibition/plating. We found that there was no correlation between spore age (time of dry storage post-harvest) and speed of germination (Fig. 2a), suggesting that
after-ripening does not occur as it does in seeds. Moreover, a period of chilling, which breaks dormancy in seeds, did not affect spore germination (Fig. 2b,c). This suggests an absence of primary dormancy or after-ripening in Physcomitrella spores.

Far-red inhibition of Physcomitrella spore germination is not rescued by application of diterpenes or inhibition of ABA synthesis

Although Physcomitrella spores appear not to have primary dormancy, we investigated whether their germination could be inhibited by environmental signals, as occurs in seeds (imposition of secondary dormancy in otherwise germination-competent seeds). In both seeds and Physcomitrella spores, germination can be fully inhibited by a pulse of FR light (Seo et al., 2009; Possart & Hiltbrunner, 2013). In eudicot seeds, FR-inhibition of germination can be fully rescued by treatment with bioactive GAs or by inhibition of ABA biosynthesis (Ikuma & Thimann, 1960; Schopfer et al., 2001; Oh et al., 2006; Seo et al., 2006; Lee et al., 2012).

In order to investigate whether the interface between light- and diterpene/ABA-signalling in Physcomitrella spores is conserved with the regulation seen in Arabidopsis seeds, we FR-treated spores in the presence of a diterpenoid known to be bioactive in moss (Hayashi et al., 2010), the fern antheridiogen GA9-methyl ester, and the carotenoid biosynthesis inhibitor norflurazon, which blocks ABA biosynthesis in flowering plants (Chamovitz et al., 1991). We found that neither GA9-methyl ester nor norflurazon were able to rescue the germination-inhibitory effects of a pulse of FR light on Physcomitrella spores, even when control spores had germinated to 100% (Fig. 3a). This suggests that diterpenes and ABA do not facilitate spores’ responses to the environmental signal of light at certain wavelengths, and thus the hormonal control of Physcomitrella spores and dicot seeds is not conserved.

Physcomitrella spore germination can be inhibited by high temperatures, but this cannot be rescued by ABA-inhibition, or application of diterpenes or strigolactones

In order to further explore the role of diterpenes and ABA in the control of environmentally regulated germination, we inhibited spore germination using another environmental trigger: high temperature (thermoinhibition). Substantial and reversible thermoinhibition of seed germination (imposition of secondary dormancy) is seen at 32°C in Arabidopsis (Tamura et al., 2006; Toh et al., 2008), and this can be rescued by GA3, norflurazon or strigolactone application (Toh et al., 2012).

Incubation of spores at 35°C (but not 32 or 34°C; data not shown) caused complete inhibition of germination that was fully reversible upon return to normal growth conditions (22°C) (Fig. 3b). Thermoinhibition of Physcomitrella spores at 35°C could not be alleviated at all by GA9-methyl ester, norflurazon or the synthetic strigolactone GR24 (Fig. 3c). Although high temperatures can inhibit germination in Physcomitrella spores, as in Arabidopsis seeds, the hormones mediating this response in Arabidopsis are not the same as in Physcomitrella, as was also seen with the FR light response.

Diterpenoids can promote Physcomitrella spore germination

Our previous work suggested conservation of some hormone function in spore and seed germination, via an ABA-ARABIDILLO/PHYSCODILLO signalling module (Moody et al., 2016). Thus, we further explored the effects of diterpene hormones in Physcomitrella, to compare their effects with those of GAs in seeds. We examined Physcomitrella mutants in the gene encoding the first enzyme in the putative moss gibberellin biosynthesis pathway, *ent-COPALYL DIPHOSPHATE SYNTHASE/ent-KAURENE SYNTHASE* (*CPS/KS*), which makes no ent-kauren and hence no bioactive diterpenoids/gibberellins (*Ppcps/ks*; Hayashi et al., 2010; Fig. S1), similar to the mutants used by Hayashi et al. (2010). Two different Ppcps/ks mutant alleles showed a reduced germination speed compared with WT: they attained a lower percentage of germination at any given time on the upwards slope of the graph, although they eventually attained 100% germination (Figs 4a, S1a), and this phenotype could be rescued by application of two diterpenoids known to be bioactive in moss (Hayashi et al., 2010): the fern antheridiogen GA9-methyl ester or ent-kaurene (Fig. 4b,c). This result indicates that bioactive diterpenoid hormones in Physcomitrella have a positive effect on spore germination.

In order to further investigate this possibility, we examined the effect of diterpenes on WT Physcomitrella spore germination. GA9-methyl ester and ent-kaurene both enhanced spore germination (Figs 4b,c, S1b), whereas GA3 did not (Fig. S1c). Conversely, the diterpenoid hormones that promote moss spore germination cannot fully rescue the germination defect of the Arabidopsis *gal-1* mutant (Fig. S2). Together, these data: (i) show that diterpenoid hormones are not absolutely required for spore germination, unlike in seeds, corroborating Hayashi et al. (2010); (ii) indicate that diterpenoid hormones increase Physcomitrella spore germination speed, thus have a positive effect on germination; and (iii) lend support to the notion that bryophyte bioactive diterpenoid hormones differ from those in seed plants.

ABA reduces Physcomitrella spore germination acting synergistically with sucrose

We have shown previously that ABA inhibits Physcomitrella spore germination in a dose-dependent manner (Moody et al., 2016). Physcomitrella spores require approximately five-fold higher concentrations of ABA for strong inhibition of germination than Arabidopsis (Finkelstein, 1994).

In order to examine the effect of inhibiting ABA biosynthesis on spore germination, we treated WT spores with norflurazon. We saw a small but reproducible promotion of germination (Fig. 5a; see also Fig. 3a). To investigate a potential link between diterpenoid hormones and ABA-regulation of spore germination, we first tested whether the* cpls/ks* mutant phenotype can be rescued via inhibition of ABA biosynthesis in moss spores.
Norflurazon enhanced the germination of *cps/ks* mutant spores (Fig. 5b). Moreover, exogenously applied diterpenoid hormones could reverse the inhibitory effect of ABA on spore germination (Fig. 5c). This suggests that the balance of ABA and diterpenoid hormone levels may be important for regulating spore germination, but not with the same prominent role that these hormones have assumed in seed germination.

As ABA-mediated stress responses in *Physcomitrella* vegetative tissue involve accumulation of soluble sugars (Burch & Wilkinson, 2002; Nagao *et al.*, 2006; Oldenhof *et al.*, 2006; Bhyan *et al.*, 2012; Erxleben *et al.*, 2012), we tested the effect of low concentrations of sucrose (0.1–1%) on spore germination. Sucrose inhibited spore germination in a dose-dependent manner (Fig. 5d) and acted synergistically with ABA (Fig. 5e). Together these data show that ABA at relatively high concentrations reduces moss spore germination, as it does (although not as strongly as in) in seed germination, and suggest that *Physcomitrella* ABA-mediated inhibition of spore germination could share downstream mechanisms with ABA-mediated desiccation and freezing tolerance responses in the *Physcomitrella* gametophyte.

Fig. 3 Effects of environmental signals on *Physcomitrella* spore germination. (a) A pulse of far-red (FR) light inhibits spore germination and this cannot be rescued by norflurazon or GA₉-methyl ester (GA, gibberellin). Spores plated on control medium or GA₉-methyl ester (GA₉-ME) or norflurazon were treated with FR light (‘No FR’ and ‘FR’, respectively) and immediately placed in the dark for 7 d. The spores’ germination percentage was measured 7 d after treatment. All spores were then moved to white light for a further 7 d (‘No FR > White’ and ‘FR > White’), under which conditions all spores germinated to completion, showing that the effect of FR light is completely reversible. A Kruskal–Wallis test indicates differences between FR and white light-treated samples, *P* < 0.05. Error bars, ± SEM. (b) A temperature of 35°C inhibits spore germination and this is completely reversible when spores are returned to 22°C. (c) High-temperature inhibition of spore germination cannot be rescued by norflurazon, GA₉-methyl ester or the synthetic strigolactone analogue GR24 (‘strigolactone’ on the graph). Spores were germinated for 7 d at either 22°C or 35°C. The 35°C-treated spores were then transferred to 22°C for another 7 d (‘35°C > 22°C’). A Kruskal–Wallis test indicated differences between 35°C and 22°C spores, *P* < 0.05. Error bars, ± SEM.
Diterpenes promote Physcomitrella spore germination. (a) Comparison of wild-type (WT) and copalyl-diphosphate synthase/kaurene synthase (cps/ks) mutant (Zhan et al., 2015) spore germination. (b) Effect of exogenous GA9-methyl ester on WT and cps/ks mutant spores (GA, gibberellin). (c) Effect of exogenous ent-kaurene (entK) on cps/ks mutant spores. Error bars, ± SEM.

Ent-kaurene and ABA biosynthesis and signalling genes are expressed in spores

Our data suggest that diterpenoid hormones and ABA have subtle effects on spore germination compared with the absolute requirement for these hormones in regulating seed germination. To extend these findings, we asked whether the putative homologues of genes encoding the proteins responsible for biosynthesis and signal transduction of diterpenes and ABA are expressed in spores or during spore germination. We extracted RNA from dry spores, imbibed spores, germinating spores, protonemal filaments and leafy gametophytes. We performed semi-quantitative RT-PCR to detect expression of the Physcomitrella homologues of the ent-kaurene biosynthesis genes CPS/KS and CYP701A3 (ent-KO) (Hayashi et al., 2006; Miyazaki et al., 2011), the first and second enzymes (respectively) in the putative moss diterpenoid hormone biosynthesis pathway (Hayashi et al., 2006). The CPS/KS transcript was detectable during spore germination and was absent from dry spores, whereas the ent-KO transcript was detectable largely in dry spores (Fig. 6a), suggesting that spatial and temporal regulation of different stages of diterpene biosynthesis occurs during the Physcomitrella life cycle.

We also examined the expression of the putative gibberellin receptors GLP1 and GLP2 (Yasumura et al., 2007), and the two PpGAMYB transcription factors PpGAMYB1 and PpGAMYB2 (Aya et al., 2011). The putative gibberellin receptor PpGLP1, like CYP701A3 (ent-KO), showed its highest expression in dry spores, decreasing upon imbibition and undetectable once germination occurs (Fig. 6b). The second putative gibberellin receptor, PpGLP2, was expressed more strongly than PpGLP1, but again showed strong expression in dry spores, decreasing markedly during imbibition (Fig. 6b). PpGLP2 expression also was detected in germinating spores and, later, in leafy tissue (Fig. 5b). PpGAMYB1 was expressed in all tissues tested, whereas PpGAMYB2, similarly to the PpGLPs and ent-KO, was detected in dry spores but decreased upon imbibition (Fig. 6b). PpGAMYB2 was absent from germinating spores but present in protonemal and gametophyte tissues (Fig. 6b), corroborating Aya et al. (2011).

We also assessed the expression of the putative genes encoding the final two (cytosolic, ABA-specific) steps in the ABA biosynthesis pathway, namely two putative Physcomitrella ABA DEFICIENT2 (ABA2) homologues and two putative Physcomitrella ABCSCISIC ALDEHYDE OXIDASE3 (AAO3) homologues (Hanada et al., 2011). All genes showed expression in dry spores, germinating spores and leafy tissue, with one PpABA2 and one PpAAO3 also present in imbibed spores and one PpAAO3 also present in protonema (Fig. 7a). We also tested the expression of putative ABA signalling genes. The four putative PYRABACTIN-RESISTANCE 1/PYRABACTIN RESISTANCE 1-RELATED/REGULATORY COMPONENT OF ABA RECEPTOR (PYR/PYL/RCAR) ABA receptors (Takezawa et al., 2011) were expressed in all tissues tested (Fig. 7b), as were the two putative Class II SnRK phosphatases that were detectable in this assay (Fig. 7c) (out of the total six SnRKS in Physcomitrella, Takezawa et al., 2011). Out of the two Physcomitrella ABI-INSENSITIVE1 (ABI1) protein phosphatases (Komatsu et al., 2013), PpABI1a was expressed ubiquitously (Fig. 7d), whereas PpABI1b was highly expressed in dry spores and leafy tissue (Fig. 7d). Out of the three ABA-regulated transcription factors Physcomitrella ABA INSENSITIVE3A, -3B and -3C (PpABI3A, -3B and -3C) (Khandelwal et al., 2010), PpABI3A and PpABI3C were expressed in all tissues tested, whereas PpABI3B was largely absent from imbibed spores but present in other tissues (Fig. 7e).

In summary, all putative ABA biosynthesis genes were expressed in dry spores, with lower levels during imbibition, as is
Fig. 5 Abscisic acid (ABA) and sucrose synergistically inhibit Physcomitrella spore germination. (a) Effect of norflurazon on wild-type (WT) spore germination. Five micromolar norflurazon has no further effect (data not shown). (b) Effect of 2 µM norflurazon on cps spores. (c) GA₉-methyl ester can rescue the inhibition of germination by ABA (GA, gibberellin). (d) Dose-dependent inhibition of germination by sucrose. Sucrose was dissolved in BCD medium. (e) Synergistic inhibitory effect of ABA and sucrose on spore germination: an intermediate concentration of both ABA and sucrose was used; the 0.5% sucrose control also contains matched solvent. Error bars, ± SEM.
Physcomitrella

Fig. 6 Expression of diterpene biosynthesis and putative response genes in Physcomitrella tissues. (a) Reverse transcription (RT)-PCR of the Physcomitrella diterpene biosynthesis gene PpCPS/KS and putative diterpene biosynthesis gene PpentKO in Physcomitrella tissues compared to a PpTUBULIN control. (b) RT-PCR of the putative Physcomitrella diterpene response genes PpGLP1 and PpGLP2 and PpGAMYB1 and PpGAMYB2 in Physcomitrella tissues compared to a PpTUBULIN control. Dry, Dry spores; Imb, imbibed spores; Ger, germinating spores; Prot, protonema; Leaf, Leafy gametophores; W, water control.

Strigolactones inhibit Physcomitrella spore germination

In order to extend our findings around the hormonal control of spore germination, we examined the effect of other hormones known to affect seed germination. Strigolactones promote seed germination in a variety of plants (Akiyama & Hayashi, 2006) and have been suggested to affect Physcomitrella spore germination (Proust et al., 2011). We tested whether strigolactones affected spore germination by comparing the germination of WT spores with those of the Physcomitrella ccd8 mutant, which cannot synthesize SLs (Proust et al., 2011). The ccd8 mutant showed increased germination (Fig. 8a), corroborating an unpublished observation by Proust et al. (2011). The ccd8 mutant’s increased germination could be reduced to levels closer to that of WT spores by exogenous application of GR24 (Fig. 8b). Moreover, exogenous GR24 reduced the germination speed of WT Physcomitrella spores at concentrations of 0.1 μM (Fig. 8b). This indicates that in Physcomitrella, unlike in Arabidopsis and parasitic plants, strigolactones have an inhibitory role in the germination process.

The ethylene precursor ACC inhibits Physcomitrella spore germination

Like strigolactone, ethylene has been shown to promote seed germination in Arabidopsis and other dicots, acting antagonistically to ABA. We showed that application of 50 μM–1 mM 1-aminocyclopropane-1-carboxylic acid (ACC), the ethylene precursor that is cleaved by ACC oxidase to release ethylene, inhibited Physcomitrella spore germination in a dose-dependent manner.
Fig. 8 Strigolactones and an ethylene precursor reduce Physcomitrella spore germination rate. (a) Ppccd8 strigolactone biosynthesis mutants (Proust et al., 2011) germinate faster than their corresponding wild-type (WT). (b) Ppccd8 mutant germination can be inhibited by exogenously applied synthetic strigolactone GR24, which also inhibits WT spore germination. Day 5 data shown; a Kruskal–Wallis test indicates significant differences between samples, \( P < 0.05 \) as indicated by the letters on the graph. (c) The ethylene precursor 1-aminocyclopropane-1-carboxylic acid (ACC) inhibits WT spore germination rate. (d) The ethylene inhibitor aminoisobutyric acid (AIB) can rescue the effects of ACC on spore germination. Error bars ± SEM.

(Fig. 8c), which is the converse of its effects in seeds (where experiments commonly use a dose of 1 mM ACC), but similar to strigolactone’s effects on spores. Moreover, the ethylene biosynthesis inhibitor aminoisobutyric acid, which competitively inhibits ACC oxidase, can overcome the effects of ACC (Fig. 8d).

Discussion

Environmental regulation of germination by light and high temperature is not integrated by the same hormones in Physcomitrella spores compared with seeds

We have shown that unlike most seeds, Physcomitrella spores grown in laboratory conditions do not show primary dormancy and after-ripening. A lack of primary dormancy has been reported in several moss and liverwort species (McLetchie, 1999; Glime, 2015).

We have also shown that, in a similar way to seeds, germination-competent Physcomitrella spores can be prevented from germinating using environmental cues. This is in line with evidence that conditional (secondary) dormancy exists in Sphagnum allowing formation of a spore bank (Sundberg & Rydin, 2000) and in Physcomitrium sphaericum (Furness & Hall, 1981). The interface of environmental signals with known hormonal networks is not conserved between Physcomitrella spores and flowering plant seeds.

Seed germination and spore germination can both be reversibly inhibited by a pulse of far-red (FR) light (e.g. Seo et al.,...
Research

et al. compared with seeds: live, and move around, on land. Our experiments suggest a

gests that this is an evolutionarily early adaptation for plants to

respond to environmental cues that block germination, and sug-

This shows that both spores and nondormant seeds

can be rescued (Chopra & Kumra, 1988), although this was

not shown to be reversible (Chopra & Kumra, 1988).

Unlike in seeds (Ikuma & Thimann, 1960; Schopfer et al.,

2001; Oh et al., 2006; Seo et al., 2006; Nelson et al., 2009), FR-
inhibition of Physcomitrella spore germination cannot be rescued

by addition of diterpenes or inhibition of abscisic acid (ABA)
synthesis. Furthermore, unlike in seeds (Toh et al., 2012), addition

dieterpenes, norflurazon or strigolactones (SLs) cannot res-
cue the germination of spores inhibited by high temperature.

Thus, a rescue mechanism for FR- and thermo-inhibition in

Physcomitrella spores is divergent compared with seeds, and

remains to be elucidated.

In seeds, environmentally regulated inhibition of germination

impinges on ABA and gibberellin (GA) synthesis and metabolism

(Seo et al., 2006; Toh et al., 2012). Crosstalk between light and

GA/ABA signalling in seeds occurs at multiple levels, including

via effects on hormone metabolism (and subsequent hormone

levels), via transcriptional changes in signalling genes (reviewed in

Piskurewicz et al., 2009), and via interactions between transcrip-
tion factors from different pathways (e.g. Richter et al., 2010;

Casal, 2013; Tang et al., 2013). For example, FR light

blocks germination in dicots by repressing the expression of GA-

oxidase (Toyomasu et al., 1998; Yamauchi et al., 2004), which

catalyses a step in gibberellin biosynthesis that does not exist in

Physcomitrella (Stewart & Freebairn, 1969; Hayashi et al., 2010;

Zhan et al., 2015). Moreover, Physcomitrella seems not to have

clear orthologues of FH13/FAR1 or ABI5 (Rensing et al., 2008)

(which integrate ABA and light signalling in Arabidopsis; Tang

et al., 2013) and has divergent DELLA proteins that have not

been shown to transduce gibberellin signalling (Yasumura et al.,

2007). Thus, the ’wiring’ of the interface between environmental

and hormonal regulation of seed germination evolved after the

divergence of the bryophyte lineage. Our work suggests that 

a novel trigger for light-induced germination exists in moss. Fur-

thermore, divergent molecular networks mediate conserved

developmental responses to environmental stimuli in spores and

seeds to enable plant movement on land.

Diterpenes and ABA affect germination in Physcomitrella

spores but appear to have a modulatory role, in contrast to

the critical role of GA and ABA in seeds

We showed, on the one hand, that certain diterpenoid hor-

mones have a positive effect on germination in Physcomitrella

spores and, on the other, that ABA has a negative effect on

spore germination. Furthermore, Physcomitrella ent-KO, ABA

synthesis genes and putative diterpene- and ABA-signal-trans-
duction pathway genes were expressed in spores. Corroborating

previous work (Hayashi et al., 2010; Zhan et al., 2015), we

saw that the bioactive diterpenes in Physcomitrella are those at

an early step in the biosynthesis pathway (ent-kaurene), or

those that show activity in ferns and also spore-bearing plants

(GA₃-methyl ester), rather than those active in seed plants. We

found that diterpenoids are not required for Physcomitrella

spore germination (corroborating Hayashi et al., 2010), but

that they do influence germination in a positive way. Although

Hayashi et al. (2010) did not report a germination phenotype

for their cps mutant, their mutant spores were compared with

wild-type (WT) only at a single time point, so no measure of

germination rate was made and thus relatively subtle differ-

ences were probably overlooked. Physcomitrella GAMYB pro-
teins, homologues of which are regulated by gibberellin

signalling in flowering plants, are required for correct spore

coat formation (Aya et al., 2011).

Mutant analysis in Arabidopsis has demonstrated that bioactive
gibberellins and gibberellin signalling are absolutely required for

seed germination to occur (Koornneef & van der Veen, 1980),

whereas Arabidopsis seed germination is completely inhibited by

concentrations of ABA as low as 5 μM (Finkelstein, 1994). The

effects of gibberellins and ABA in Physcomitrella spores were not

as extreme: the cps mutant has a slower germination rate than

WT, but cps mutant spores can eventually germinate to the same

level as WT controls, demonstrating no loss of germination

potential. The concentration of ABA required for strong inhibi-
tion of Physcomitrella spore germination was also five- to 10-fold

higher than for Arabidopsis seeds, although different spore

batches vary in their ABA sensitivity, as is the case for Arabidopsis

seeds (Finkelstein, 1994), as might be expected in an ephemeral

species (Glime, 2013).

The effect of norflurazon on Physcomitrella spores was subtle,

which may be due to the lack of primary dormancy in spores,

which are essentially ’ready to germinate’. The extent to which

norflurazon reduces ABA levels in Physcomitrella is not known.

This suggests that diterpenes and ABA perform a modulatory

role during Physcomitrella spore germination, and gained a more

prominent and complex role in multicellular seeds after co-

option into the sporophyte during the evolution of seed plants

(e.g. Piskurewicz et al., 2009). Our data comparing the behaviour

of physcodillo mutant spores and arabadillo mutant seeds on ABA

suggest that these Armadillo-related protein homologues may

represent a conserved node in an ancient regulatory network

(Moody et al., 2016).

The function of ABA in Physcomitrella previously has been

studied in gametophytic vegetative tissues, where a role in stress

responses (such as drought) has been demonstrated, as in the

flowering plant sporophyte (Knight et al., 1995; Cuming et al.,

2007; Khandelwal et al., 2010; Takezawa et al., 2011). Several

studies have demonstrated a role for sugars (including sucrose)

acting synergistically with ABA during moss abiotic stress resis-
tance (Burch & Wilkinson, 2002; Nagao et al., 2006; Oldenhof

et al., 2006; Bhyan et al., 2012; Erdleben et al., 2012). We
demonstrated that a similar synergism may exist during spore germination. The level of sucrose that can decrease spore germination rate (0.1–1% in this study) is 10- to 100-fold lower than that used to cause osmotic stress in moss (10%: Garrocho-Villegas & Arredondo-Peter, 2008). Evidence exists for a regulatory role of sugars during seed germination: an inhibitory effect of low levels of sucrose (0.5–3%) on Arabidopsis seed germination up to 5 d has been observed (Chen et al., 2006; Li et al., 2012). This effect requires functional ABA biosynthesis, because aba2 mutant seeds are sucrose-insensitive (Li et al., 2012). However, Finkelstein & Lynch (2000) showed that 1–2% sucrose could counteract the effects of 3 μM ABA on Arabidopsis seed germination after 7 d, suggesting that different interactions between sucrose and ABA may occur at different times.

A novel role for SLs and ethylene in inhibiting spore germination in Physcomitrella

We showed that SLs have a negative effect on Physcomitrella spore germination. The role of SLs in seed plant germination is a positive one: they act as signals to promote seed germination either between plants (via root exudates) or within one plant (e.g. during thermoinhibition) (Bouwmeester et al., 2003; Yoneyama et al., 2010; Toh et al., 2012; Stanga et al., 2013). This occurs via regulation of gibberellin biosynthesis (Nelson et al., 2009), lending weight to the hypothesis that SLs have a different mechanism of action in Physcomitrella germination. SLs are well known as regulators of shoot branching in seed plants (Waldie et al., 2014). This branching function is conserved in the Physcomitrella gametophyte (Proust et al., 2011). In addition, SLs in moss promote ‘self-awareness’ and delimit colony spread, acting as quorum-sensing molecules (Proust et al., 2011), suggesting that the ‘between-plant’ communication function of SLs arose early in land plant evolution. Perhaps in bryophytes the inhibitory role of SLs in spore germination arose as a quorum-sensing function; thus, when spores are released from the plant, the formation of one colony from a spore could prevent the germination of a second colony-forming spore close by, and hence aid colony establishment without competition for resources.

Interestingly, ethylene also has a positive role in seed germination and the ethylene precursor ACC has a negative role in spore germination: as ethylene is also a small, gaseous and easily diffusible hormone, its presence in a developing moss gametophyte could also signal to neighbouring spores and prevent their germination. Ethylene affects seed germination via crosstalk with ABA signalling/synthesis, via synergism with GA signalling and via direct effects on cell separation of the endosperm in a number of plant species (Linkies & Leubner-Metzger, 2012). This suggests that ethylene may have been co-opted separately into spores and seeds to perform different roles.

Comparing the regulation of germination in Physcomitrella spores and nondormant seeds

The regulation of equivalent developmental processes between gametophyte and sporophyte seems to show an amazing degree of similarity in comparative studies on key model organisms. For example, Physcomitrella thizoids are developmentally equivalent to Arabidopsis root hairs (Menand et al., 2007), and GAMYB functions in spore and reproductive organ development are equivalent (Aya et al., 2011), whereas the liverwort Marchantia polymorpha shows circadian regulation of the vegetative-to-reproductive transition, as in the seed plant sporophyte (Kubota et al., 2014). We have shown that spores and seeds respond to the same environmental cues to generate the same developmental output, but via different mechanisms.

Our results suggest evolution of novel hormonal regulation of germination between Physcomitrella spores and nondormant/after-ripened seeds, with ABA and GA assuming much greater importance in seed plants, multilevel crosstalk between environmental and hormone pathways evolving in seeds, and some hormones being co-opted into different roles in spores and seeds. Similar network rewiring has been identified between root hairs and rhizoids, between RHD SIX-LIKE transcription factors and auxin signalling (Jang et al., 2011; Pires et al., 2013).

One possibility is that the multicellular nature of seeds may have led to a requirement for complex, coordinated hormonal regulation of the different tissues during germination. Determining the molecular nature of the signalling pathways that regulate spore germination is now a key target for future research. Alternatively, as spores appear not to show dormancy, there may be no requirement for a complex hormonal regime regulating dispersal. It is possible that a ‘bet-hedging’ strategy controlling germination rates due to life history may exist in Physcomitrella spores as in Arabidopsis (Springthorpe & Penfield, 2015). The absence of observed dormancy in moss spores also raises the question of how these dispersal units function to colonize new environments.

Acknowledgements

We thank Peter Hedden for providing extensive supplies of Gibberellin A3-methyl ester and ent-kaurene, and Catherine Rameau for providing ccd8 spores. We thank Daniel Gibbs and Petra Stamm for providing critical comments on the manuscript, Ottoline Leyser and Binne Zwanenburg for providing GR24, Steve Penfield for use of the Far-Red LED array and Andy Cuming for initial discussions around bryophyte spore germination. We thank Heinho Düring, Ralf Reski and Janice Glim for discussions about spore dormancy. E.F.V. was supported by a NERC Doctoral Training Scholarship. Y.S. and S.J.B. were supported by Leverhulme Trust Grant F/00094/BA. L.A.M. was supported by a Biotechnology and Biological Sciences Research Council (BBSRC) Doctoral Training Scholarship. D.H., A.W., S.N., A.C., B.B. and D.M. performed research using University of Birmingham Funds. H.B. was supported by a grant from the Danish Council of Independent Research – Technology and Production. B.C.K. and H.T.S. were supported by SpotLight, a grant from the Danish Council for Strategic Research. G.W.B. was supported by BBSRC grants BB/L0102321/1 and BB/J017604/1 and J.C.C. was supported by a Royal Society-Leverhulme Trust Fellowship (2013-14).
Author contributions

References
Research


Takezawa D, Komatsu K, Sakata Y. 2004. Additional Supporting Information may be found online in the Supporting Information tab for this article.


Supporting Information

Additional Supporting Information may be found online in the Supporting Information tab for this article.

**Fig. S1** Moss bioactive gibberellins promote *Physcomitrella* spore germination.

**Fig. S2** Gibberellins that are bioactive in *Physcomitrella* cannot rescue the *Arabidopsis ga1-3* mutant seed germination phenotype and substitute for GAs.

**Table S1** Primers used for RT-PCR analysis

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