



Anthelmintic activity of forage chicory (*Cichorium intybus*) and field efficacy of ivermectin against gastrointestinal nematodes in Danish cattle

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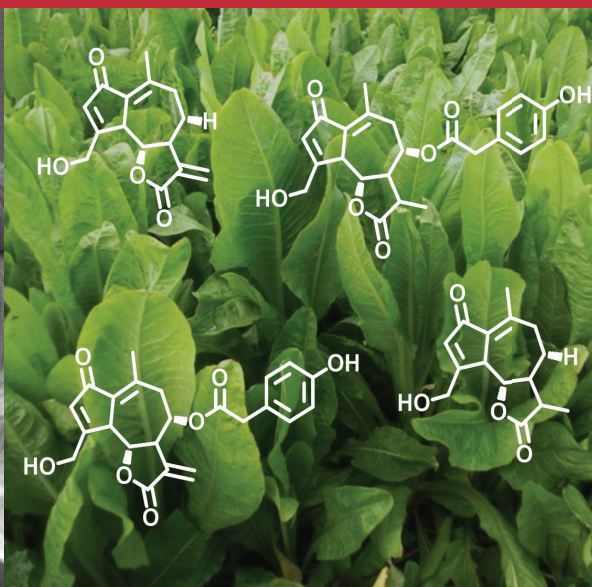
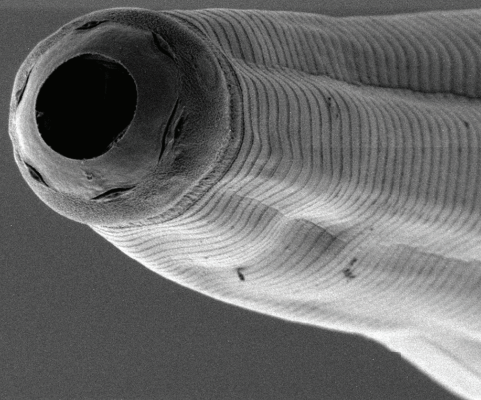
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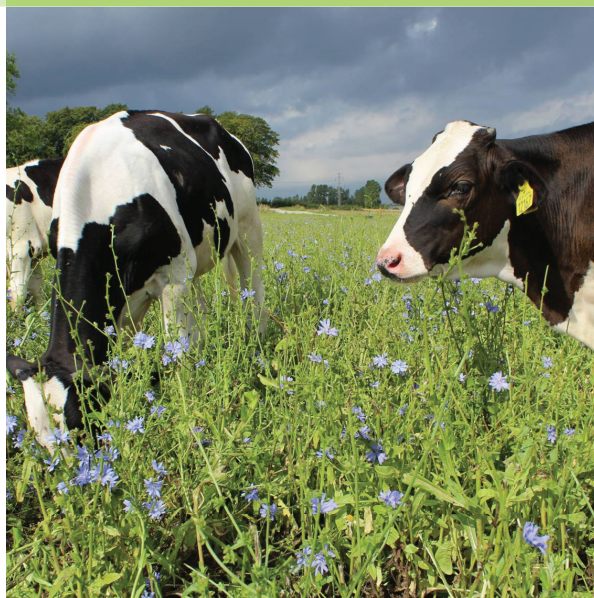
Anthelmintic activity of forage chicory (*Cichorium intybus*) and field efficacy of ivermectin against gastrointestinal nematodes in Danish cattle

PhD thesis



Miguel Peña-Espinoza

2016



**Anthelmintic activity of forage chicory
(*Cichorium intybus*) and field efficacy of
ivermectin against gastrointestinal
nematodes in Danish cattle**

Miguel Angel Peña-Espinoza

PhD thesis

Section for Bacteriology, Pathology and Parasitology
National Veterinary Institute
Technical University of Denmark
Copenhagen
2016

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Preface

The research work presented in this PhD thesis was conducted between October 2012 and December 2015 in the Section for Bacteriology, Pathology and Parasitology and Section for Immunology and Vaccinology at the National Veterinary Institute, Technical University of Denmark, and in the Section for Parasitology and Aquatic Diseases, Department of Veterinary Disease Biology, University of Copenhagen. The PhD studies were funded by the EMIDA ERA-NET project “Coping with anthelmintic resistance in ruminants” (CARES; 3405-11-0430/32) and CONICYT Chile (Becas Chile, segunda convocatoria 2009).

The thesis is organized as a succinct review of gastrointestinal nematodes in cattle, anthelmintic resistance and the use of bioactive plants as an alternative helminth control approach, with focus on studies with forage chicory. Next, a brief description of the performed investigations is presented, including the experimental designs and methods, a summary of the major findings and conclusions, and final remarks on future research. The thesis culminates with the following scientific articles describing in detail the research work conducted during the PhD studies:

Manuscript I. Efficacy of ivermectin against gastrointestinal nematodes of cattle in Denmark evaluated by different methods for analysis of faecal egg count reduction. Submitted to International Journal for Parasitology: Drugs and Drug Resistance.

Manuscript II. Anthelmintic effects of forage chicory (*Cichorium intybus*) against gastrointestinal nematode parasites in experimentally infected cattle. Parasitology 2016, 143, 1279–1293.

Manuscript III. Sesquiterpene lactone containing extracts from two cultivars of forage chicory (*Cichorium intybus*) show distinctive chemical profiles and *in vitro* activity against *Ostertagia ostertagi*. International Journal for Parasitology: Drugs and Drug Resistance 2015, 5, 191–200.

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I wish to sincerely thank all the farmers that accepted to participate in our field studies, for all the very good talks about cows and parasites and your warm hospitality. Special thanks to Esben Møller-Xu, whose passion for organic cattle farming inspired me so much at the start of the project and also introduced me to *cikorie* Spadona. *Tusind tak!*

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My years in Copenhagen have been an amazing experience of learning and discovery thanks to the company, inspiration and the big help of all my friends in Denmark, and now around the world. I feel really grateful to have met you all and to have shared so many brilliant moments in the dark winter days and the sunny summer nights. My wholehearted thanks to my sisters and brothers in the road, Paula, Giovanna, Miriam, Noelia, Teresa, Eva, Marcela, Alejandro, Olivier, Sundar, César, Leo, Laue, Sebastian, Kristian, Myriam, José and all the big CPH family. *Tusind tusind tak!!!*

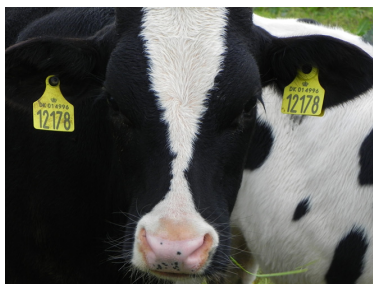
To my families in Chile and Austria, who inspired and filled me with energy, love and support throughout my PhD studies. *A mi Mamá, Papá y Hermana, Marisol, Miguel y Natalia: los amo mucho y muchas gracias por todo su amor, alegría, apoyo incondicional y fuerzas durante todos estos años lejos de Chile. En cada camino que se abre, como siempre, donde yo voy ustedes van conmigo! An Gerda, Karl, Susanne und Bernhard: Danke, für den herzlichen Empfang und die Aufnahme in die Familie seit dem ersten Tag, für eure wunderbare Unterstützung, Großzügigkeit und Freundschaft während der letzten Jahre. Dankeschön!*

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Ich liebe dich so sehr! 

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Miguel Peña-Espinoza

Abstract

Parasitism with gastrointestinal (GI) nematodes is ubiquitous in grazing cattle and its control is critical to sustain satisfactory levels of animal health, welfare and productivity. Currently, conventional parasite control strategies relying exclusively on anthelmintic drugs are severely threatened by the development of anthelmintic resistance (AR) in bovine nematodes in several countries. In Denmark, cattle farmers and their veterinarians depend largely on macrocyclic lactones (ML), mainly ivermectin (IVM), for nematode control. However, no studies have evaluated the field efficacy of IVM against GI nematodes in Danish cattle. Alternative control approaches to minimize selection for AR and that can complement the use of the still effective anthelmintics in integrated control strategies are urgently needed. A promising complementary control strategy is to include bioactive forages with anti-parasitic effects in the animals' diets. Forage chicory (*Cichorium intybus*) has demonstrated anthelmintic effects against sheep nematodes, but comparable evidence in cattle is absent. The direct anthelmintic activity of forage chicory has been associated with its content of plant secondary metabolites (PSM), particularly sesquiterpene lactones (SL). Still, no studies have investigated the content of SL in chicory used as anthelmintic forage and no evidence exists of a direct effect of chicory SL against parasitic nematode stages. Thus, the main objective of the present investigations was to contribute towards the integrated control of bovine GI nematodes by evaluating the field efficacy of ivermectin in naturally-infected Danish cattle and by exploring the direct anthelmintic activity of forage chicory and its SL against bovine nematodes *in vivo* and *in vitro*.

Faecal egg count reduction tests (FECRT) were performed to evaluate the field efficacy of IVM against natural infections with GI nematodes (Manuscript I). Six cattle farms with a history of macrocyclic lactones use were included. At day of treatment (Day 0), 20 naturally infected grazing calves per farm were stratified by initial faecal egg counts (FEC) and randomly allocated to a treatment group (IVM; n=10) or an untreated control group (CTL; n=10). Calves in the IVM group were treated with 0.2 mg IVM kg⁻¹ body weight (s.c.). Individual FEC were obtained at Day 0 and 14 days post-treatment and pooled larval cultures for species-specific detection of *Ostertagia ostertagi* and *Cooperia oncophora* by real-time PCR. Treatment efficacies were analysed as FEC reduction percentages (FECR%) using five different methods of calculations: the recommended WAAVP method (with and without CTL) and two Bayesian methods; Bayescount (with and without CTL) and EggCounts. A simulation study was performed to evaluate the performance of the three methods (without CTL) to correctly identify the FECR% of simulated populations with values as observed in three investigated farms. In the FECRT, reduced IVM efficacy was confirmed in three farms by all methods based on the FEC of treated animals only and in one farm according to the two methods including the FEC from treated and untreated cattle. Post-treatment, *C. oncophora* was detected in treated groups of all herds and *O.*

ostertagi was identified in one farm with confirmed reduced IVM efficacy. In the simulation study, high FEC aggregation markedly affected the performance of the methods of calculation. Furthermore, there is an urgent need for standardised international guidelines for the design and evaluation of FECRT in cattle. This is the first report of reduced efficacy of an anthelmintic in Danish cattle. However, due to the low number of enrolled farms in the FECRT, this study may not represent the general situation in Danish cattle. In addition, the presence of IVM-resistant nematode strains suggested by the FECRT in the investigated farms should preferably be confirmed by a controlled efficacy test. Yet, evidence of IVM inefficacy to reduce FEC in Danish cattle suggests that farmers and their advisors should closely monitor the efficacy of the anthelmintic treatments and include alternative control strategies to reduce their dependence on anthelmintics.

Two independent trials studied the direct anthelmintic effects of forage chicory-rich diets against GI nematodes in experimentally infected cattle (Manuscript II). In Experiment 1, 15 stabled, nematode-naïve Jersey-bull calves (body weight [BW]: 92 ± 21 kg) were randomly allocated, after BW stratification, into chicory (CHI₁, n=9) or control (CTL₁, n=6) groups and fed *ad libitum* with forage chicory (cultivar [cv.] Spadona) silage or ryegrass/clover hay, respectively. Animals were supplemented with concentrate to balance protein/energy intakes between groups throughout the trial. After 16 days of adaptation to diets, all calves were infected with 10,000 *O. ostertagi* and 66,000 *C. oncophora* L3 (Day 0 post infection [p.i.]). In Experiment 2, 20 parasite-naïve Holstein bull-calves (153 ± 24 kg) were stratified by BW and randomly assigned to pastures of either pure forage chicory (cv. Spadona; CHI₂, n=10) or ryegrass/clover (CTL₂, n=10). After 7 days, all calves were infected with 20,000 *O. ostertagi* L3 (Day 0 p.i.) and moved regularly to avoid re-infections from pasture. Due to poor regrowth of the chicory pasture, the CHI₂ group was supplemented with chicory silage. Faecal egg counts per g of faecal dry matter (FECDM) were performed in both trials. Condensed tannins (CT) and SL were analysed in all the experimental feeds. At Day 40 p.i. (Experiment 1) and Day 35 p.i. (Experiment 2) calves were slaughtered for worm counts. In Experiment 1, cumulative FECDM was not significantly different between groups, but lower numbers of *O. ostertagi* L3 were detected in pooled larval cultures of CHI₁ calves from Day 26 p.i. At slaughter, *O. ostertagi* counts were significantly reduced in CHI₁ by 60% (geometric mean reduction; $P < 0.01$), whereas *C. oncophora* burdens were unaffected ($P = 0.12$). In Experiment 2, FECDM were markedly lowered in CHI₂ from Day 22 p.i onwards ($P < 0.01$) and *O. ostertagi* adult burdens were significantly reduced in CHI₂ by 66% ($P < 0.001$). Condensed tannins were not detected in any of the experimental feeds and SL were only identified in forage chicory (fresh and silage) in both experiments, at levels ranging from 12.3 to 22.5 g SL kg⁻¹ dry matter. Forage chicory-rich diets ($\geq 70\%$ of chicory dry matter in the diet) led to a significant reduction in adult worm burdens and FECDM of *O. ostertagi* in calves, while no activity was observed against *C. oncophora*. Chicory (fresh and silage) was readily consumed by calves and the anthelmintic effects were preserved despite ensiling.

To confirm the relationship between the observed *in vivo* anthelmintic effects of forage chicory and its content of SL, *in vitro* studies were performed to test the direct anti-parasitic activity of SL-containing chicory extracts against free-living and parasitic stages of *O. ostertagi* (Manuscript III). Total (free and unbound) SL were extracted from leaves of chicory cv. Spadona and cv. Puna II. *O. ostertagi* eggs isolated from mono-infected calves were hatched and L1 were used in a larval feeding inhibition assay (LFIA), while cultured L3 were used in a larval exsheathment inhibition assay (LEIA). Adult worms were immediately recovered after slaughter and used for motility inhibition assays (AMIA). Analyses by high-performance liquid chromatography of Spadona and Puna II extracts revealed the presence of lactucin (LAC), 8-deoxylactucin (8-DOL) and lactucopicrin (LCP). In addition, the 11, 13-dihydro (DI) derivatives of the mentioned SL were also detected. Between 60 – 65% of the tested extracts was composed of SL, while the remaining (unknown) compounds were not determined. Comparable levels of total and individual SL and unknown compounds were detected in both extracts; however, Spadona-extract contained significantly higher concentrations of DI-8-DOL ($P=0.01$), while Puna II-extract had increased levels of DI-LAC ($P<0.0001$). In the LFIA, both extracts reduced larval feeding at increasing concentrations, but Spadona extract showed higher potency confirmed by significantly lower EC_{50} ($P<0.0001$). In the LEIA, neither of the two extracts interfered with the exsheathment of L3 ($P>0.05$). In the AMIA, both extracts induced a dose-dependent worm paralysis effect but Spadona extract exerted a more rapid and lethal activity on adult worms than Puna II extract, with significantly lower EC_{50} ($P<0.0001$). Sesquiterpene lactone-containing extracts from forage chicory demonstrated a direct and potent anti-parasitic activity against *O. ostertagi* L1 and exerted a rapid and lethal paralysis of *O. ostertagi* adults, and these molecules may be the responsible active compounds for the anthelmintic activity of chicory. Differences in the anti-parasitic activity between extracts from two chicory cultivars seem related with their distinct content of SL and could help to further identify the most active compounds.

In conclusion, reduced IVM efficacy was confirmed in three of six Danish cattle farms and emphasizes the need to perform larger surveys, monitor the efficacy of prescribed anthelmintic treatments and to include alternative control methods to reduce the selection pressure for AR. Forage chicory-rich diets show promise as selective anti-*Ostertagia* feed for cattle and may help to reduce infection levels and the need for anthelmintic treatments in grazing cattle. However, further studies are needed to investigate the on-farm use of forage chicory as part of integrated parasite control strategies for cattle. Sesquiterpene lactone-containing chicory extracts demonstrated direct and dose-dependent anthelmintic effects against *O. ostertagi in vitro* and distinct potencies observed between chicory cultivars may be related to differences in their individual SL.

Sammen drag

Gastrointestinale nematoder (GIN), også kaldet løbetarmorm, er almindeligt forekommende hos græssende kvæg, og det er afgørende for opretholdelsen af sundhed, velfærd og produktivitet i kvægbruget at kontrollere disse parasitter. Flere steder i verden udgør udviklingen af ormemiddelresistens (OR) igennem de senere år imidlertid en alvorlig trussel mod den gængse parasitkontrol, der oftest udelukkende er baseret på anvendelse af ormemidler. I Danmark kontrolleres GIN hos kvæg overvejende ved anvendelse af ormemidler tilhørende gruppen af makrocykliske laktoner (ML), især ivermectin (IVM), men hidtil har ingen studier undersøgt effekten af disse medikamenter i danske kvægbesætninger. Alternative kontrolmetoder er nødvendige med henblik på at mindske selektionen for OR. Anvendelse af bioaktive planter med antiparasitær effekt i foderet har vist lovende resultater som et supplement til traditionelle kontrolforanstaltninger. Fodring med cikorie (*Cichorium intybus*) har således vist god effekt mod GIN hos får, men tilsvarende resultater overfor kvægets GIN findes ikke. Cikoriens anthelmintiske aktivitet er blevet sat i forbindelse med dens indhold af bioaktive plantemetabolitter, især stofferne sesquiterpene-laktoner (SL). Hovedformålet med de nærværende studier var derfor at bidrage mod en integreret kontrol af GIN hos kvæg i) ved at evaluere effekten af IVM i danske besætninger; og ii) ved at undersøge den anti-parasitære effekt af cikorieekstrakter og cikorie dels *in vitro* dels *in vivo* overfor kvægets GIN.

Fækale ægreduktionstests blev gennemført i 6 danske kvægbesætninger med henblik på at vurdere IVMs effekt overfor naturlige infektioner med GIN. Førsteårsgræssende kalve i hver besætning blev behandlet ved injektion med den anbefalende dosis IVM (CHI₁; n=10) eller de indgik i forsøget som ubehandlede kontroller (CTL₁; n=10). Den fækale ægudskillelse blev vurderet ved behandling samt 14 dage senere, og samtidig blev larvekulturer opsat for hver behandlingsgruppe med henblik på artsidentifikation af 3. stadielarver (L3) v.h.a. «real-time quantitative PCR». Reduktionsprocenten blev beregnet ud fra fem forskellige metoder, bl.a. gældende retningslinier fra World Association for the Advancement of Veterinary Parasitology (WAAVP, med eller uden CTL) og to Bayesianske metoderne: EggCounts (uden CTL) and Bayescount (med eller uden CTL). Effekten af IVM var lavere end forventet i alle 6 besætninger, og nedsat effekt, defineret som en reduktionsprocent under 95% og en nedre grænse for 95% konfidensintervallet under 90%, blev bekræftet i 3 besætninger ved anvendelse af de tre metoder uden inklusion af ubehandlede kontroller. Efter behandling viste larvekulturerne, at tyndtarmsormen, *Cooperia oncophora* var til stede i alle de behandlede grupper, mens løbeormen, *Ostertagia ostertagi* blev fundet i en af besætningerne med bekræftet nedsat effekt af IVM.

Den direkte antiparasittære effekt af fodercikorie blev undersøgt i to uafhængige forsøg hhv. et opstaldnings- og et markforsøg. I Eksperiment 1 blev 15 nematode-naïve Jersey kalve (92 ± 21 kg) tilfældigt fordelt (stratificeret efter vægt) mellem en cikorie- (CHI₁, n=9) eller kontrol- (CTL₁, n=6) gruppe, som blev fodret *ad libitum* med henholdsvis cikorie-ensilage af sorten Spadona eller med hø af kløvergræs. Med henblik på afbalancering af protein- og energiindtagelsen mellem grupperne blev foderet suppleret med kalvepiller (kraftfoder) igennem hele forsøgsperioden. Efter en adaptationsperiode på 16 dage blev kalvene inficeret med 10.000 *O. ostertagi* og 66.000 *C. oncophora* L3-larver (Dag 0 post infektion [p.i.]). I Eksperiment 2 blev 20 nematode-naïve kalve af racen Dansk Holstein (153 ± 24 kg) stratificeret efter vægt og tilfældigt allokeret til marker med ren fodercikorie af sorten Spadona (CHI₂; n=10) eller kløvergræs (CTL₂; n=10). Alle kalvene blev eksperimentelt inficerede med 20.000 *O. ostertagi* L3-larver efter 7 dages adaptation (Dag 0 p.i.). Forsøgsgruppen (CHI₂) blev fodret supplerende med cikorie-ensilage p.g.a. ringe plantevækst i forsøgsperioden. Under begge eksperimenter blev der foretaget fækale ægtællinger pr. g tørstof (faecal egg counts per g of faecal dry matter [FECDM]), og koncentrationen af kondenserede tanniner (CT) samt SL i foderet blev analyseret. Ormebyrden i de eksperimentelt inficerede kalve blev optalt efter slagtning Dag 40 p.i. (Eksperiment 1) og Dag 35 p.i. (Eksperiment 2). Det kumulative antal æg pr. g tørstof var ikke signifikant forskellig mellem grupperne i Eksperiment 1, men fra Dag 26 p.i. blev der påvist lavere antal *O. ostertagi* i larvekulturerne fra CHI₁ kalvene. Ved slagtning var antallet *O. ostertagi* signifikant reduceret med 60% i CHI₁ gruppen (gennemsnitlig geometrisk reduktion; $P < 0.01$), mens antallet af *C. oncophora* var uændret i forhold til kontrolgruppen ($P = 0.12$). I Experiment 2, var FECDM markant lavere hos CHI₂ kalvene fra Dag 22 p.i. ($P < 0.01$) og antallet af *O. ostertagi* var signifikant reduceret med 66% ($P < 0.001$). Kondenserede tanniner kunne ikke påvises i foderet i hverken det ene eller det andet forsøg, og SL i koncentrationer mellem 12.3 og 22.5 g SL pr. kg fodertørstof blev udelukkende identificeret i cikorie (både frisk og ensileret). Fodring med cikorie i høje koncentrationer (≥ 70 % cikorie tørstof på diæt) medførte en signifikant reduktion i både ægudskillelsen og antallet af *O. ostertagi* hos kalvene, mens der ikke blev fundet nogen anthelmintisk effekt overfor *C. oncophora*. Kalvene optog villigt cikorien (frisk såvel som ensileret), og den anthelmintiske effect blev bevaret trods ensilering.

Med henblik på undersøgelse af sammenhængen mellem indholdet af SL i cikorien og de observerede anthelmintiske effekter *in vivo* blev der gennemført *in vitro* studier for at teste den direkte antiparasittære effect af SL-holdige ekstrakter overfor fritlevende og parasitære stadier of *O. ostertagi*. Sesquiterpene-laktoner blev ekstraheret fra cikoriesorterne Spadona og Puna II og testet i «larval feeding inhibition assay» (LFIA) vha. L1-larver isoleret fra kalve mono-inficerede med *O. ostertagi*. Tillige blev den anthelmintiske effekt overfor L3 testet ved «larval exsheathment inhibition assay» (LEIA) og adulte nematoder testet ved «motility inhibition assay» (AMIA) på *O. ostertagi* isoleret umiddelbart efter slagtning. Analyse af cikorieekstrakterne vha. “High-performance

væskekromatografi” (HPLC) afslørede forekomst af lactucin (LAC), 8-deoxylactucin (8-DOL) and lactucopicrin (LCP) samt disses 11, 13-dihydro (DI) derivater. Sesquiterpenlaktoner udgjorde mellem 60 og 65 % af de testede ekstrakter, mens de resterende, ukendte forbindelser ikke kunne bestemmes. Der blev fundet sammenlignelige koncentrationer af SL (total og individuelle) og “ukendte” forbindelser i ekstrakter fra begge cikoriesorter. Spadona-ekstrakt indeholdt signifikant højere koncentrationer af DI-8-DOL ($P=0.01$), mens Puna II havde højere koncentrationer af DI-LAC ($P<0.0001$). Der var en omvendt korrelation mellem stigende koncentrationer af ekstrakter fra begge cikoriesorter og fødeoptaget hos *O. ostertagi* L1-larver i LFIA-testen, men Spadona ekstrakt var mere potent end Puna II reflekteret i en signifikant lavere EC_{50} -værdi ($P<0.0001$). Ingen af ekstrakterne påvirkede L3-larverne i LEIA-testen ($P>0.05$), mens begge cikoriesorter inducerede en dosisafhængig paralyse af adulte orm i AMIA; Spadona ekstrakt udviste dog en hurtigere og mere dødelig aktivitet end Puna II, hvilket blev afspejlet i en signifikant lavere EC_{50} -værdi ($P<0.0001$). Ekstrakter af fodercikorie indeholdende SL havde således en direkte og potent antiparasitær effekt overfor *O. ostertagi* L1-larver og var i stand til at paralisere adulte *O. ostertagi* hurtigt og effektivt. Det er derfor sandsynligt, at disse molekyler er afgørende for cikoriens antiparasitære aktivitet. De markante forskelle i potens mellem de to cikoriesorter var tilsyneladende korreleret med forskelle i SL profilerne.

Studierne gennemført i forbindelse med nærværende ph.d-afhandling viste nedsat effekt af IVM i 3 af 6 danske kvægbesætninger, hvilket understreger behovet for løbende overvågning af effekten af de ormemedler, som anvendes i danske besætninger samt for alternative kontrolmetoder, der kan reducere udviklingen af OR. Fodring med cikorie viste lovende resultater overfor *O. ostertagi*, og kan medvirke til at nedbringe infektionsniveauet samt behovet for behandling med ormemedler, men yderligere undersøgelser er nødvendige med henblik på implementering i praksis. Ekstrakter af cikorie indeholdende SL demonstrerede en direkte og dosisafhængig anthelmintisk effekt overfor *O. ostertagi in vitro*, og vores resultater tyder på, at forskellene i effekten mellem cikoriesorterne kan være relaterede til forskelle i deres indhold af SL.

Resumen

El parasitismo con nematodos gastrointestinales (GI) es frecuente en el ganado bovino a pastoreo y su control es crítico para sostener niveles satisfactorios de salud, bienestar y productividad animal. En la actualidad, las estrategias de control parasitario convencionales dependientes exclusivamente del uso de fármacos antihelmínticos están seriamente amenazadas por el desarrollo de resistencia antihelmíntica (AR) en nematodos bovinos en distintos países. En Dinamarca, los ganaderos y sus asesores veterinarios basan mayoritariamente el control parasitario del ganado bovino en el uso de lactonas macrocíclicas (ML), principalmente ivermectina (IVM). Sin embargo, ningún estudio a la fecha ha evaluado la eficacia en terreno de IVM contra nematodos GI en rebaños bovinos daneses. Más aún, enfoques alternativos de control que minimicen la selección de AR, y que se puedan complementar con el uso de los fármacos antihelmínticos aún efectivos en estrategias de control integrado, son urgentemente necesarios. Una potencial estrategia de control complementaria es la introducción de forrajes bioactivos con actividad antiparasitaria en la dieta de los animales. La achicoria forrajera (*Cichorium intybus*) ha demostrado efectos antihelmínticos contra nematodos del ovino, pero hasta el momento no existe evidencia comparada en bovinos. Los efectos antiparasitarios directos de la achicoria forrajera han sido asociados con la presencia de metabolitos secundarios en la planta, particularmente lactonas sesquiterpénicas (SL). No obstante, ningún estudio a la fecha ha investigado el contenido de SL en achicoria usada como forraje antihelmíntico y no existe evidencia de un efecto directo de SL de achicoria contra estadios parasitarios de nematodos. Por lo tanto, el principal objetivo de las presentes investigaciones fue contribuir al control integrado de nematodos GI del bovino a través del estudio de la eficacia en terreno de IVM contra infecciones parasitarias en ganado bovino danés y la investigación de la actividad antihelmíntica de achicoria forrajera y sus SL contra nematodos del bovino *in vivo* e *in vitro*.

Tests de la reducción de la oviposición (“Faecal egg count reduction tests”/FECRT) fueron realizadas para evaluar la eficacia en terreno de IVM contra infecciones naturales con nematodos GI (Manuscrito I). Seis predios bovinos con uso previo de ML fueron incluidos en los estudios. En el día del tratamiento (Día 0), 20 terneros a pastoreo naturalmente infectados en cada predio fueron estratificados en base al conteo de huevos de nematodos en las heces (“Faecal egg counts”/FEC) y asignados al azar a un grupo tratado (IVM; n=10) o a un grupo control no tratado (CTL; n=10) con niveles similares de FEC. Terneros en el grupo IVM fueron tratados con 0.2 mg IVM kg⁻¹ peso vivo (s.c.). Muestras fecales fueron obtenidas el Día 0 y 14 días post-tratamiento para realizar FEC individuales y la preparación de cultivos de larvas por grupo para la detección de *Ostertagia ostertagi* y *Cooperia oncophora* por medio de PCR en tiempo real. La eficacia de los tratamientos fue analizada como el porcentaje de reducción de FEC (FECR%) usando cinco métodos de

cálculo distintos: el método sugerido por la WAAVP (con y sin grupo CTL) y dos métodos bayesianos; Bayescount (con y sin grupo CTL) y EggCounts. Además, se llevó a cabo un estudio de simulación para evaluar el rendimiento de los tres métodos (sin grupo CTL) para identificar correctamente FECR% de poblaciones de nematodos simuladas con valores observados en los rebaños investigados. En los FECRT, una eficacia reducida de IVM fue confirmada en tres predios por todos los métodos incluyendo el FEC de animales tratados solamente y en un predio por los dos métodos incluyendo el FEC de animales tratados y no tratados. Post-tratamiento, se detectó la presencia de *C. oncophora* en grupos tratados de todos los predios y *O. ostertagi* en un predio con una confirmada eficacia reducida de IVM. En el estudio de simulación, una alta agregación del FEC afectó el rendimiento de todos los métodos de cálculo. Guías armonizadas internacionales para el diseño y evaluación de FECRT en ganado bovino son urgentemente necesarias. Este es el primer estudio que reporta la eficacia reducida de un antihelmíntico en ganado bovino danés. Sin embargo, y debido al bajo número de predios investigados, este estudio no es representativo de la situación general en Dinamarca. Además, cepas de nematodos resistentes a IVM indicadas en el FECRT deben ser confirmadas por medio de un test de eficacia controlada, que incluye el tratamiento de animales y el conteo de nematodos post-mortem. Sin embargo, la evidencia de una eficacia reducida de IVM en ganado bovino danés sugiere que los productores y sus asesores técnicos deben monitorear atentamente la eficacia de los tratamientos antihelmínticos utilizados e incluir estrategias de control parasitario alternativas para reducir la dependencia a los antihelmínticos.

Dos ensayos independientes se llevaron a cabo para estudiar el efecto antiparasitario de dietas ricas en achicoria forrajera contra nematodos GI en bovinos infectados experimentalmente (Manuscrito II). En el Experimento 1, 15 terneros Jersey machos estabulados sin contacto previo con nematodos (peso vivo 92 ± 21 kg) fueron asignados al azar, luego de randomización en base a peso vivo, a un grupo achicoria (CHI₁, n=9) o grupo control (CTL₁, n=6) de pesos similares y alimentados *ad libitum* con ensilaje de achicoria forrajera (cultivo [cv.] Spadona) o heno de ballica/trébol, respectivamente. Los animales fueron suplementados con concentrado para equilibrar los consumos de proteína/energía entre los grupos a lo largo del ensayo. Luego de 16 días de adaptación a las dietas, todos los terneros fueron infectados con 10,000 L3 de *O. ostertagi* y 66,000 L3 *C. oncophora* (Día 0 post-infección [p.i.]). En el Experimento 2, 20 terneros Holstein machos sin contacto previo con nematodos (153 ± 24 kg) fueron estratificados en base a peso vivo y asignados al azar a praderas con achicoria forrajera pura (cv. Spadona; CHI₂, n=10) o con ballica/trébol (CTL₂, n=10). Luego de 7 días, todos los terneros fueron infectados con 20,000 L3 de *O. ostertagi* (Día 0 p.i.) y se rotaron regularmente para prevenir reinfecciones en pradera. Debido al pobre rebrote de la pradera de achicoria, el grupo CHI₂ fue suplementado con ensilaje de achicoria. Conteo de huevos en las heces por g de materia seca fecal (FECDM) fueron realizados en ambos ensayos. Taninos condensados (CT) y lactonas sesquiterpénicas (SL) fueron analizados en todos los forrajes y concentrados. Al Día 40 p.i. (Experimento 1) y Día

35 p.i. (Experimento 2) los terneros fueron eutanasiados para conteo de nematodos post-mortem. En el Experimento 1, no se detectaron diferencias significativas en el FECDM acumulativo entre grupos, pero se identificó un número reducido de L3 de *O. ostertagi* en los cultivos de larvas del grupo CHI₁ desde el Día 26 p.i. Post-mortem, la carga parasitaria de *O. ostertagi* fue significativamente reducida in CHI₁ en un 60% (reducción media geométrica; $P < 0.01$), mientras que la carga de *C. oncophora* no fue afectada ($P = 0.12$). En el Experimento 2, el FECDM fue marcadamente reducido en CHI₂ desde el Día 22 p.i. ($P < 0.01$) y la carga de *O. ostertagi* adultos fue significativamente reducida en CHI₂ en un 66% ($P < 0.001$). No se detectaron CT en ninguno de los alimentos experimentales y SL fueron identificadas sólo en achicoria forrajera (pradera y ensilaje) en ambos experimentos, a niveles entre 12.3 y 22.5 g SL kg⁻¹ materia seca. Dietas ricas en achicoria forrajera ($\geq 70\%$ de achicoria materia seca en la dieta) indujeron una reducción significativa en la carga de nematodos adultos y FECDM de *O. ostertagi* en terneros, mientras que no se observó actividad contra *C. oncophora*. Achicoria (fresca y ensilaje) fue consumida sin problemas por los terneros y los efectos antihelmínticos se conservaron a pesar del ensilado.

Para confirmar la relación entre los efectos antihelmínticos *in vivo* de achicoria forrajera y su contenido de lactonas sesquiterpénicas (SL), se realizaron estudios *in vitro* para evaluar la actividad antiparasitaria directa de extractos de achicoria conteniendo SL contra estadios de vida libre y parasitarios de *O. ostertagi* (Manuscrito III). Se extrajeron SL totales de hojas de achicoria cv. Spadona y cv. Puna II. Huevos de *O. ostertagi* aislados de terneros infectados fueron eclosionados y las L1 usadas para el ensayo de inhibición de la alimentación larvaria ("larval feeding inhibition assay"/LFIA), mientras que L3 cultivadas se usaron para el ensayo de inhibición del desenvaine larvario ("larval exsheathment inhibition assay"/LEIA). Estadios adultos fueron aislados inmediatamente post-mortem y usados en el ensayo de inhibición de la motilidad ("adult motility inhibition assay"/AMIA). Análisis químicos con cromatografía líquida de alta eficacia de los extractos de Spadona y Puna II revelaron la presencia de las SL lactucin (LAC), 8-deoxylactucin (8-DOL) y lactucopicrin (LCP). Además, los 11, 13-dihydro (DI) derivados de las mencionadas SL fueron detectados. Entre el 60 – 65% del contenido de los extractos testeados correspondió a SL, mientras que los restantes compuestos (desconocidos) no fueron determinados. Niveles similares de SL totales e individuales y de compuestos desconocidos fueron detectados en ambos extractos; sin embargo, extractos de Spadona contenían concentraciones significativamente altas de DI-8-DOL ($P = 0.01$), mientras que extractos de Puna II tenían altos niveles de DI-LAC ($P < 0.0001$). En el LFIA, concentraciones crecientes de ambos extractos inhibieron la alimentación de las L1, pero el extracto de Spadona demostró una mayor potencia que fue confirmada por valores de EC₅₀ significativamente menores ($P < 0.0001$). En el LEIA, ninguno de los dos extractos impidió el desenvaine de las L3 ($P > 0.05$). En el AMIA, ambos extractos causaron una parálisis dosis-dependiente en nematodos, pero el extracto de Spadona ejerció una actividad más rápida y letal en los nematodos adultos que el extracto de Puna II, con valores de EC₅₀ significativamente

menores ($P < 0.0001$). Extractos de achicoria forrajera conteniendo SL demostraron potentes y directos efectos antiparasitarios contra L1 de *O. ostertagi* y causaron una rápida y letal parálisis en estadios adultos de *O. ostertagi*. Las SL son probablemente los compuestos activos responsables por la actividad antihelmíntica de achicoria. Las diferencias en los efectos antiparasitarios entre extractos de dos cultivos de achicoria parecen estar relacionadas con su contenido particular de SL y podría ayudar a identificar los compuestos más activos.

En conclusión, se confirmó una eficacia reducida de IVM en tres de seis predios bovinos en Dinamarca, lo que recalca la necesidad de realizar estudios con un mayor número de rebaños, el monitoreo de la eficacia de los tratamientos antihelmínticos prescritos y el incluir métodos alternativos de control parasitario para reducir la selección de AR. Dietas ricas en achicoria forrajera tienen el potencial de ser usadas como forraje anti-*Ostertagia* en ganado bovino, lo que podría ayudar a reducir los niveles de infección parasitaria y la necesidad de efectuar tratamientos antihelmínticos. Sin embargo, se necesitan de estudios adicionales que investiguen el uso de achicoria forrajera en un contexto productivo en predios bovinos como parte de estrategias de control integrado de nematodos GI. Extractos de achicoria forrajera conteniendo SL demostraron efectos antihelmínticos directos y dosis-dependientes contra *O. ostertagi in vitro*, y las diferencias en potencias antiparasitarias entre cultivos de achicoria puede estar relacionada con su contenido particular de SL individuales.

Abbreviations

AMIA	adult motility inhibition assay
AR	anthelmintic resistance
BW	body weight
CP	crude protein
CT	condensed tannins
CET	controlled efficacy test
CI	confidence interval
cv.	cultivar
DI-8-DOL	11,13-dihydro-8-deoxylactucin
DM	dry matter
DMSO	dimethyl sulfoxide
EM	electron microscopy
EPG	nematode eggs per g of faeces
FEC	faecal egg counts
FECDM	faecal egg counts adjusted for faecal dry matter
FECR	faecal egg count reduction
FECRT	faecal egg count reduction test
FSG	first-season grazing
GI	gastrointestinal
GIN	gastrointestinal nematodes
ha	hectare
HPLC-MS	high-performance liquid chromatography with mass spectrometry
IVM	ivermectin
ITS-2	second internal transcribed spacer
L1/L2/L3	first/second/third-stage nematode larvae
LEIA	larval exsheathment inhibition assay
LFIA	larval feeding inhibition assay
MCMC	Markov chain Monte Carlo
ML	macrocyclic lactones
MOX	moxidectin
NDF	neutral detergent fibre
PBS	phosphate-buffer solution
p.i.	post-infection
PSM	plant secondary metabolites
s.c.	subcutaneous
SEM	scanning electron microscopy
SL	sesquiterpene lactones
TEM	transmission electron microscopy
TST	targeted selective treatment
VetStat	Danish system for surveillance of the veterinary use of drugs for production animals
WAAVP	World Association for the Advancement of Veterinary Parasitology

Introduction

Cattle farming is a human activity of vast socio-economic significance, contributing to societies in terms of food production and as source of income, agricultural diversification and asset savings for rural communities (FAO, 2002; Herrero et al., 2009; Swanepoel et al., 2010). However, the sustainability of cattle farming is facing multiple and remarkable challenges in the first half of the 21st century. The foreseen growing demand for animal products by 2050, mostly in low and middle-income economies, is projected to be met by higher yields from an increasingly constrained array of natural resources (Thornton, 2010). Concurrently, the widely recognized environmental impact of cattle farming, mainly caused by greenhouse gas emissions, land and water use, and the food-feed competition resulting from the use of human-edible crops for livestock feeding, are expected to augment following the increasing pace of food production (Herrero et al., 2011; Makkar, 2016).

Generally speaking, the main strategies suggested to address these concomitant challenges are: to increase the efficiency of livestock systems aiming to maximize productivity, in order to reduce the environmental impacts per unit of product (Thornton, 2010; FAO, 2014), and to reduce the consumption of animal products, hence leading to shrinking of livestock numbers and their environmental impacts (McMichael et al., 2007; Foley et al., 2011). A complementary approach is to increase the feeding of ruminants with feed components that do not directly compete with human consumption (“not food-competing feedstuffs”), like grasslands, crop residues or by-products from food production (Eisler et al., 2014). Although the production from roughage-fed cattle systems is expected to be lower than from concentrate-fed cattle, thus resulting in a lower carbon footprint for the latter, the lower production efficiency from the first approach is offset by the benefit of reducing the production of human-edible crops for livestock in arable lands (Schader et al., 2015). As a result, it has been suggested that a global food system with pasture-based ruminant products can provide sufficient food for the expected increase in global demand and to reduce livestock environmental impacts (Schader et al., 2015), although with reductions in the consumption of animal products particularly from monogastric animals (usually fed with food-competing feeds). Furthermore, well-managed grasslands can also act as “carbon sinks” (FAO, 2010; Smith, 2014) and access to pasture has been related with improved animal health and welfare parameters (Burow et al., 2011; 2013).

Certainly, the success of current and future pasture-based cattle systems will depend on permanent advancements in animal breeding, nutrition and health. This PhD thesis tries to contribute to this wider goal through the study of novel strategies for the control of gastrointestinal nematodes in cattle, which remain the most prevalent infections in grazing livestock in Denmark and worldwide.

Chapter 1: Background

1.1 Gastrointestinal (GI) nematodes of cattle: *Ostertagia ostertagi* and *Cooperia oncophora*

Parasitic infections with GI nematodes are ubiquitous in grazing cattle of all ages, inducing clinical and subclinical disease with negative consequences for animal health, welfare and productivity. Worldwide, the most common GI nematodes infecting cattle are from the genus *Haemonchus*, *Ostertagia*, *Trichostrongylus*, *Cooperia* and *Nematodirus* (Armour, 1989; Taylor *et al.*, 2016). In Denmark, as in other temperate regions, the most prevalent and economically important GI nematodes are *Ostertagia ostertagi* and *Cooperia oncophora* (Nansen *et al.*, 1993; Monrad, 2010), both belonging to the order *Strongylida*, family *Trichostrongylidae* (Anderson, 2000).

1.1.1 Life cycle

Ostertagia ostertagi (Stiles, 1982) infects the abomasum of cattle while *C. oncophora* (Railliet, 1898) parasitise the small intestine, with both species having a direct life cycle (Fig. 1.1). In *O. ostertagi*, eggs at the morula stage are passed with the faeces to the pasture. Inside the dung pat, eggs develop into L1, which then moult to L2 until reaching the infective L3 stage that retains the cuticle (sheath) of the L2. The development from egg to L3 varies according to the temperature in the dung pat, and under optimal conditions (25°C) can occur within 7 days (Pandey, 1972). Ensheathed L3 actively migrate outside the dung pat or are translocated by rain splash into the surrounding pasture (Grønvold and Høgh-Schmidt, 1989). When cattle consume the infective larvae with the contaminated herbage, the L3 undergo exsheathment in the rumen and migrates to the abomasum, infecting the abomasal gastric glands. Inside the glands, *O. ostertagi* develop into the L4 and L5 stages, and approximately 18 days post-infection, the L5 emerge from the gastric glands and the worms moult into the adult stage, which can be found on the abomasal mucosa. After mating, female worms start laying eggs around 21 days post-infection (pre-patent period), which are passed to the pasture with the faeces. Some infective L3 ingested by cattle during the end of the grazing season (autumn) do not develop into adult worms in three weeks as described above; instead they remain “arrested” as early L4 (hypobiotic larvae) in the gastric glands (Michel, 1974). Arrested L4 usually resume their development in late winter or early spring, and the synchronized emergence of a large number of L5 into the abomasal lumen can lead the clinical condition known as “Ostertagiosis Type II” (Entrocasso *et al.*, 1986).

The life cycle of *C. oncophora* is essentially similar to that described for *O. ostertagi*, apart from the fact that infective L3, which undergo exsheathment in the abomasum, migrate to the small intestine where the nematodes develop into L4, L5 and adult worms, with the first females laying eggs as soon as 15 days post-infection (Frankena, 1987). Arrested development has also been described in *C. oncophora* (Michel et al., 1978)

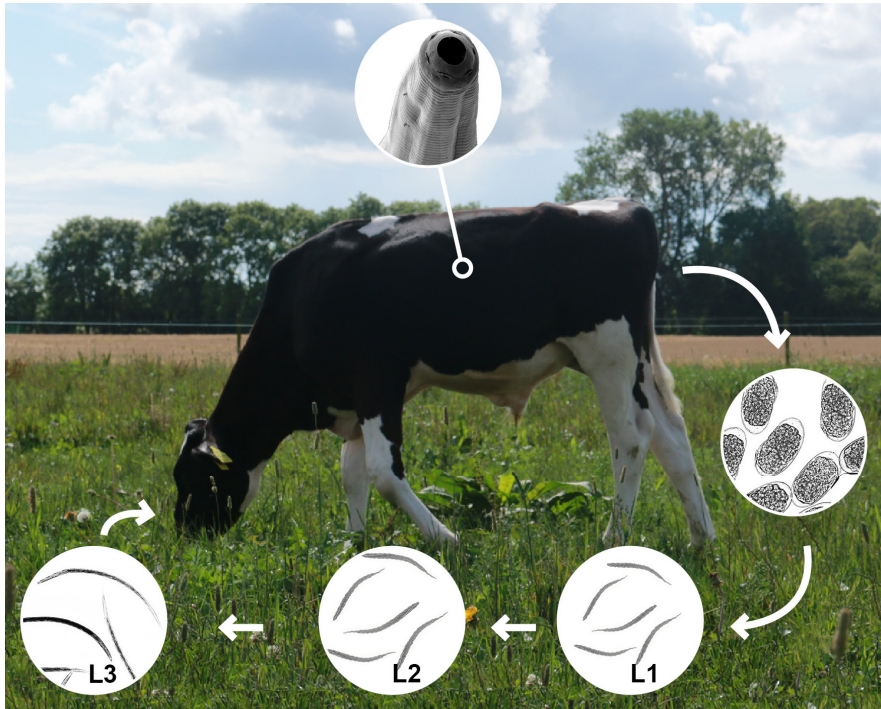


Fig. 1.1. Life cycle of *Ostertagia ostertagi*. Development from egg to L3 occurs inside the dung pat and the L3 must migrate onto the herbage to be ingested by the host and continue its development until adult stage.

1.1.2 Pathophysiology

1.1.2.1 *Ostertagia ostertagi*

Ostertagia ostertagi is commonly regarded as the most pathogenic cattle nematode in temperate areas due to the severe damage that can induce in the abomasum. The pathogenicity of *O. ostertagi* is mainly attributed to the emergence of L5 from the gastric glands and the presence of adult worms in the abomasal lumen, which can induce severe epithelial damage (resulting in protein loss to the lumen), loss of parietal cells and hyperplasia of mucous-secreting cells, inhibition of acid secretion (thus increasing abomasal pH and allowing survival of anaerobic bacteria) and increased serum gastrin and pepsinogen levels (McKellar et al., 1986; Taylor et al., 1989; Fox, 1997; Simpson, 2000). In chronic infections, macroscopic lesions in the abomasal mucosa can be observed, such as oedema, haemorrhages and formation of small nodules. Fox et al. (1989) reported that calves trickle infected with 10,000 *O. ostertagi* L3 daily during 46 days had significantly lower nitrogen digestibility and rate of passage of digesta and a marked reduction in voluntary feed intake. In subsequent studies, reduced feeding intake has been further confirmed as the main factor responsible for the reduced weight gain in infected calves (Fox, 1993; Fox, 1997; Forbes et al., 2000)

1.1.2.2 *Cooperia oncophora*

Cooperia oncophora has been considered a mild pathogen in cattle and a small number of studies have investigated its pathological effects. Coop et al. (1979) observed that calves daily infected with up to 20,000 *C. oncophora* L3 during 20 weeks had only a slight hypoalbuminemia and minor pathological damage in the small intestine, with a limited compression of the intestinal villi only in the areas with presence of worms, without penetration of the mucosa. In the same study, only a small reduction in weight gains were observed with no differences in the DM intake between infected or non-infected groups. In contrast, Armour et al. (1987) reported that calves daily infected with 10,000 *C. oncophora* L3 over six weeks had atrophied and thickened intestinal villi with excessive mucus production, loss of plasma protein into the intestinal lumen, reduced feed intake and weight loss; however, 12 weeks post-infection the authors observed a significant regeneration of the intestinal mucosa, associated with the expulsion of most of the worms, although loss of plasma protein to the intestine continued.

1.1.2.3 Interactions of *O. ostertagi* and *C. oncophora*

Parkins et al. (1990) reported that mixed infections with *O. ostertagi* and *C. oncophora*, which are the norm in the field, can induce larger pathological effects than mono-infections, leading to diarrhoea, weight loss, anorexia and hypoalbuminemia. However, the authors only studied the course of mixed infections, without comparing with mono-infections under the same conditions. Satrija and Nansen (1993) described watery diarrhoea in calves co-infected with *O. ostertagi* and *C. oncophora*, but did not observe clinical signs in calves mono-infected with *O. ostertagi* and no differences were detected in serum pepsinogen, albumin or weight gains between groups. In the same study, *C. oncophora* did not affect the establishment or the course of *O. ostertagi* infections.

1.1.3 Impact of GI nematodes on animal health and productivity

Clinical parasitism with GI nematodes, particularly in susceptible cattle, is usually manifested by poor body condition, anorexia, lower weight gain or weight loss, dull hair coat, (watery) diarrhoea, FEC>200 EPG and serum pepsinogen >5 unit of tyrosine (Eysker and Ploeger, 2000; Vercruyse and Claerebout, 2001). Although clinical disease can still be observed, it is widely recognized that the major impact of GI nematodes under a context of widespread use of (effective) anthelmintic drugs is at the subclinical level, inducing production losses without obvious clinical symptoms (Corwin, 1997; Charlier et al., 2014). Subclinical parasitism with GI nematodes has been reported to induce reduced growth rate (calves), loss of carcass yield and quality (heifers), decreased pregnancy rates (cows and heifers), reduce milk yield (cows) and longer calving to conception interval (primiparous and multiparous cows) (Gibbs, 1992; Hawkins, 1993; Shaw et al., 1998; Vercruyse and Claerebout, 2001; Charlier et al., 2009). Growth rates of FSG calves treated chemoprophylactically with anthelmintics have been reported to increase by 150/315 g day compared with untreated calves suffering subclinical/clinical infections (Shaw et al., 1998), while in treated cows an average increase in milk yield of ~1 kg/day has been observed (Charlier et al., 2009). Also, it has been reported that reduced growth rates as a result of GI parasitism during the first grazing season can persist in heifers during their second grazing season, thus suggesting a lasting impact of helminth infections on animal production (Larsson et al., 2011).

1.1.4 Immune response

Cattle is able to develop acquired immunity against GI nematode infections, but the rate and extent of this response varies largely towards different worm species (Armour, 1989; Claerebout and Vercruyse, 2000). Under field conditions, a high level of immunity to *C. oncophora* is observed in FSG calves after around 4 months of exposure, illustrated by a marked reduction in FEC and adult worms at winter housing (Armour, 1989). Under experimental conditions, challenge with 100,000 *C. oncophora* L3 resulted in the expulsion of adult worms from most animals after 35-42 days post-infection (Kanobana et al., 2001). Yearling and adult cattle exposed to *C. oncophora* during previous grazing seasons have none or very low infection levels of this species (Armour, 1989), although *C. oncophora* adult stages can still be detected (yet very rarely) in cows (Borgsteede et al., 2000). In contrast, immunity towards *O. ostertagi* develops slower (Gasbarre, 1997) and calves at housing after their first grazing season can still have a high worm burden (Armour, 1989). Yearling cattle are still susceptible to new infections in their second grazing season, and a marked decrease in *O. ostertagi* burdens and/or reduce establishment of newly acquired L3 do not occur before the animals are >18-24 months-old (Williams et al., 1993; Gasbarre, 1997). However, a reduced egg output related with development of acquired immunity in young cattle exposed to *O. ostertagi* in the previous grazing season is clear (Grønvold et al., 1992; Gasbarre et al., 2001). Interestingly, it is known that the development of immunity to *O. ostertagi* in yearling cattle is inversely correlated to the efficacy (and intensity) of anthelmintic treatments performed during the first grazing season, which limit the exposure to infections and the resulting build-up of immunity (Vercruyse et al., 1994). Adult cattle can still harbour small numbers of *O. ostertagi* adults (Agneessens et al., 2000; Claerebout and Vercruyse, 2000) and have a very low faecal egg excretion, although enough to preserve the contamination of the pastures with this species (Armour, 1989).

1.1.5 Epidemiology

Similarly as in other North European countries, the epidemiology of GI nematodes of cattle in Denmark is determined by the presence of overwintered L3 on pastures as a source of infection for cattle turned out (~early May) on that fields (Nansen et al., 1978; 1987). Infective L3 ingested by FSG (naïve) calves develop into adult worms that generally reach a peak of faecal egg excretion around 2 months post-turn out, resulting in the accumulation of newly developed L3 on pasture from mid-summer onwards (Fig. 1.2). Under Danish conditions, development of L3 in the field is slow during the first half of the grazing season (<15°C) but rapidly increases as the temperature rises, leading to a “synchronized” larval development of infective larvae from mid-summer (Monrad, 2010). By late summer/early autumn, a high concentration of L3 is expected on the pastures which can lead to severe clinical disease of grazing animals, condition known as “Ostertagiosis Type I” (Henriksen et al., 1976; Nansen, 1987). Higher stocking rates exacerbates the level

of infections as animals are forced to graze more close to the faecal pats, where most of the infective L3 are present (Nansen et al., 1988). Susceptible calves are infected mainly with *C. oncophora* during the first-half of the grazing season, while contamination of pastures with *O. ostertagi* L3 increases and peaks from mid-summer until early autumn (Henriksen et al., 1976). However, unusual climatic conditions can significantly change the described pattern of pasture infectivity (Nansen et al., 1989). During very dry Danish summers in the mid-1970's, faecal pats were not disintegrated and infective L3 developed inside the dung did not migrate onto the grass (thus pasture infectivity remained low in late summer). The following spring, the same dung pats acted as reservoirs of a high number of overwintered larvae that were translocated onto the herbage, leading to clinical disease already on early spring/summer, termed "early-season Ostertagiosis Type I" (Nansen et al., 1989).

The infection dynamics described above imply that susceptible cattle are kept on the same permanent pasture the entire grazing season. Certainly, farming practices that reduce the contact with a high number of infective larvae may effectively prevent significant production losses and clinical parasitism. Differences in parasite exposure are greatly influenced by production systems and climatic factors, as recently described for *O. ostertagi* in dairy cows from different European countries (Bennema et al., 2010). Moreover, the influence of climate change on both the worm biology and livestock management is likely to further alter the epidemiology of pasture-borne parasitism in cattle (Morgan and Wall, 2009), to which the control strategies will need to adapt.

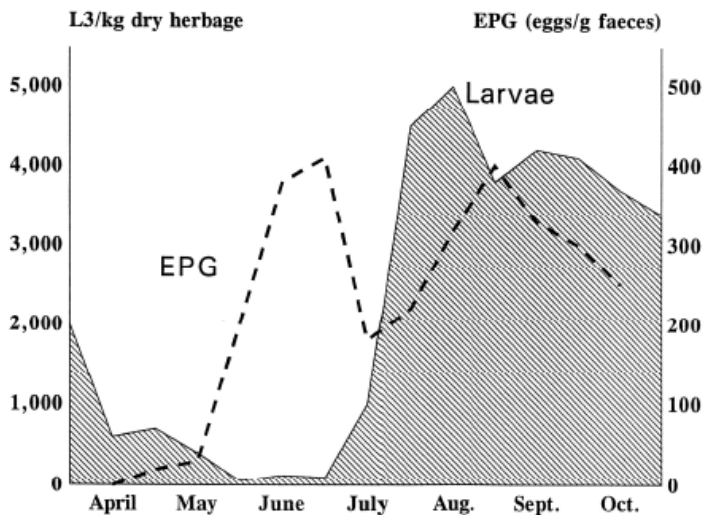


Fig. 1.2. Number of infective L3 on pasture (L3/kg dry herbage; hatched area) on a permanent pasture grazed by FSG calves in Denmark with turn-out mid-April. Faecal egg counts (EPG; dotted line) are presented as group means for each time point (adapted from Monrad, 2010).

1.2. Control of GI nematodes of cattle with anthelmintics

Since the release of the first broad-spectrum benzimidazoles in the 1960's and the advent of ivermectin and the macrocyclic lactones (ML) in the 1980's (Egerton et al., 1981), the use of synthetic anthelmintics has become the basis for the control of GI nematodes in intensively grazed cattle (Geurden et al., 2015; Sutherland and Bullen, 2015). Apart from the low toxicity of these drugs in treated animals and their initial high anti-parasitic efficacy when released, farmers also rapidly adopted anthelmintics because they could be used in different production systems without requiring major changes in husbandry practices (Nansen, 1993). In Denmark, drugs within three broad-spectrum anthelmintic classes with distinct modes of action are currently registered for use in cattle: *i*) benzimidazoles (e.g. fenbendazole, albendazole), *ii*) imidazothiazoles/tetrahydro-pyrimidines (e.g. levamisole) and *iii*) ML (e.g. ivermectin, moxidectin, eprinomectin, etc.). Additionally, two novel broad-spectrum anthelmintic groups have been described: the amino-acetonitrile derivatives (ADD; Kaminsky et al. (2008) and the spiroindoles (Little et al., 2011). The ADD have a high efficacy against GI nematodes of ruminants and a commercial product from this group, monepantel, has been registered in the EU for use in sheep (EMA, 2009). In Denmark, as in other European countries, the ML are the most widely used anti-parasitics in cattle, mainly due to their high efficacy against adult and larval stages of parasitic nematodes and their persistent efficacy against re-infections, but also because of their effect against ectoparasites and availability as topical (pour-ons) formulations (Vercruyssen and Rew, 2002).

1.2.1 Ivermectin: mechanisms of action and expected *in vivo* efficacy in cattle

Ivermectin (IVM) and other ML selectively increase the permeability of excitable membranes in the nervous system of parasitic nematodes (Gill and Lacey, 1998) by binding irreversibly to glutamate-gated chloride ion channels (and in higher concentrations also to γ -aminobutyric acid [GABA] and glycine receptors), interfering with the muscle contractions that regulate worm feeding, reproduction and movement (Cully et al., 1994; Kotze et al., 2014). However, not all cattle nematodes are equally susceptible to IVM. When IVM was released, clinical trials in cattle revealed that *C. oncophora* was the “dose limiting” species for this drug, requiring a minimum dosage of 0.2 mg IVM kg BW⁻¹ (s.c. injection) to eliminate adults and L4 stages by $\geq 97\%$ (Egerton et al., 1981). In contrast, *O. ostertagi* adults and L4 were eliminated by $>99\%$ with a dose of 0.05 mg IVM kg⁻¹ BW s.c. (Egerton et al., 1981). Later research demonstrated that IVM can be detected at higher active concentrations in the abomasal mucosa, in comparison with the small intestine (Lifschitz et al., 2000). As a result, the recommended dose for injectable IVM was established to 0.2 mg kg BW⁻¹, but even at this dosage, small percentages of exposed worms are able to survive each treatment, thereby contributing to the selection of resistant nematode populations.

1.2.2 Anthelmintic resistance in GI nematodes of cattle

Anthelmintic resistance (AR) has been defined as the capacity of a worm population (or individual worms within the population) to tolerate doses of an anthelmintic that would otherwise kill a normal population from the same species, and to transmit this “resistant” fitness to their progeny (Prichard et al., 1980). The accumulation of resistant genes in a nematode population is therefore an evolutionary process which depends on: *i*) the genetic diversity of the parasite populations under selection for AR, *ii*) the selection pressure (i.e. applications of anthelmintics) and *iii*) time (Prichard, 2002). Therefore, it can be expected that parasite populations exposed to anthelmintic drugs will evolve gradually from fully susceptible to fully resistant, and at different speeds under different circumstances (Kaplan et al., 2007).

Anthelmintic resistance is nowadays a widespread problem for the control of GI nematodes of small ruminants (Kaplan and Vidyashankar, 2012). In bovine nematodes, AR was originally sporadically identified (Jackson et al., 1987; Vermuntt et al., 1996; Stafford and Coles, 1999). However, since the 2000's, cases of AR in GI nematodes of cattle have been continuously reported in surveys worldwide, sometimes even involving multi-drug resistant populations (e.g.: Mejia et al., 2003; Waghorn et al., 2006; Soutello et al., 2007; Sutherland and Leathwick, 2011; Gasbarre, 2014; Geurden et al., 2015). In the majority of these studies, AR has been declared to ML, and in temperate regions mainly involving *C. oncophora*, although cases of AR in *O. ostertagi* developed in the field have also been confirmed (Edmonds *et al.* 2010; Gasbarre, 2014). In Scandinavia, the presence of AR in cattle nematodes has been only investigated in Sweden and clear indications were detected of reduced field efficacies of injectable and pour-on ML (Demeler et al., 2009; Areskog et al., 2013).

It is evident that, at the moment, the development and impact of AR in cattle has not reached the alarming levels observed in sheep and goat farming (Kaplan and Vidyashankar, 2012). In addition, the impact of an inefficacious treatment is likely to be more obvious in small ruminants (particularly if infected with *Haemonchus contortus*) than in cattle, where a reduced drug efficacy may only translates in subclinical production losses. Nevertheless, it has been suggested that some differences in the husbandry of cattle have limited the progression of AR in bovine helminths. One explanation is the higher level of the cattle nematode population *in refugia*, i.e. the proportion of the worm population not exposed to anthelmintics which is outside the host as free-living stages or in untreated animals (Coles, 2002). In cattle nematodes, worms *in refugia* are expected to be present mainly as free-living stages in the dung and on pasture, which may constitute > 99% of the entire parasite population in a farm (Nansen, 1993). In addition, adult cattle are not frequently treated with anthelmintics, in contrast with adult sheep, and this may have mitigated a rapid selection pressure (Prichard, 2002). Nevertheless, an intensification of treatments in adult

cattle, due to ease of administration and development of drug formulations without milk withdrawal to maximise production, may increase the selection for AR by decreasing the *in refugia* population (Stafford and Coles, 1999; Coles, 2002; Sutherland and Leathwick, 2011). Furthermore, topical (pour-ons) anthelmintics are widely used in cattle, and whether the varying drug concentrations observed in animals treated with these formulations can increase the selection for AR needs to be evaluated (Sargison et al., 2009; Sutherland and Bullen, 2015). Considering the current situation of AR in cattle nematodes, it seems opportune to reduce the selection pressure on the still effective anthelmintics before drug-resistance becomes a more extensive problem (Prichard, 2002; Taylor et al., 2010). A first step towards this goal is the precise detection of the drug efficacy on-farm.

1.2.3 Field detection of anthelmintic resistance in GI nematodes of cattle

In the absence of quantitative molecular techniques for detection of AR to ML and the high cost of the controlled efficacy test (the gold standard method to diagnose AR), at present the only readily available technique to diagnose AR on farms is the faecal egg count reduction test (FECRT). Various *in vitro* assays have also been described (Demeler et al., 2010; 2012), although they are currently not widely available.

The FECRT evaluates the efficacy of an anthelmintic by measuring the reduction in FEC in a treated group before and after treatment or between a treated and untreated group (Taylor et al., 2002). Current guidelines for the design and analysis of the FECRT in cattle are derived from recommendations developed for the detection of AR in sheep nematodes (WAAVP; Coles et al., 1992). Since then, several possible biases have been highlighted concerning the use of FECRT in cattle, mainly the high degree of density-dependence of cattle nematodes (thus reducing the correlation between FEC and actual worm burden), the lower faecal egg excretion commonly detected in cattle as compared with sheep, and the highly aggregated (over-dispersed) distribution of FEC between animals (Coles, 2002; Coles et al., 2006; Demeler et al., 2010; El-Abdellati et al., 2010; Sutherland and Leathwick, 2011). In addition, the inclusion of untreated control animals in the FECRT has been subject of much scientific debate (Dobson et al., 2012; Calvete and Uriarte, 2013; Lyndal-Murphy et al., 2014), and although untreated controls have been included in FECRT in cattle (Waghorn et al., 2006; Lyndal-Murphy et al., 2010), no field studies have compared the validity of such designs. Another problem is unreliable detection of AR when the drug efficacy is still high (80 – 95%), which prevents implementation of measures at an early stage to avoid the progression of AR into treatment failure. To reduce the uncertainty of the FECRT, much information on the true drug efficacy can be gained when studying the 95% confidence interval of a treatment (Levecke et al., 2012). Robust statistical analyses by Bayesian modelling using Markov chain Monte Carlo (MCMC) methods have been advocated to cope with low FEC data and to increase the accuracy in detecting uncertainty intervals (Denwood et al., 2010; Torgerson et al., 2014). These methods are being

increasingly used for monitoring AR in cattle nematodes (Neves et al., 2014; O'Shaughnessy et al., 2014; Geurden et al., 2015; Ramos et al., 2016). However, the influence of varying levels of aggregation observed in cattle FEC data on the performance of such Bayesian methods has not been evaluated. In addition, sensitive and species-specific tests to detect the nematode species surviving treatment are critical for a specific diagnosis of AR and are urgently required for cattle (Coles, 2002; Sutherland and Leathwick, 2011).

1.3 Alternative parasite control strategies

In parallel to the development of AR in worm populations, there has been an increasing public concern on the prophylactic use of veterinary drugs in food animals and the potential residues in the environment, as well as a wish to reduce the input of synthetic chemicals in organic and sustainable farming systems (Waller and Thamsborg, 2004; Hoste et al., 2014). All these factors have motivated the search for alternative and sustainable control methods that can complement or substitute the sole use of anthelmintics in farms (Thamsborg et al., 1999; Waller, 2006). Various alternative parasite control approaches to anthelmintics have been explored in ruminants (Hoste and Torres-Acosta, 2011). A common objective of these strategies is to reduce the parasite burden in the animals below a threshold of clinical or production impact, instead of achieve a high efficacy like anthelmintics, and to combine different methods as part of integrated parasite control programs (Thamsborg et al., 1999; Waller and Thamsborg, 2004; Ketzis et al., 2006). In general, an integrated parasite control program combines: *a*) strategies that reduced or eliminate the parasite burden in the host, *b*) strategies that increase the immunity and/or the resilience of the host to a parasite infection and *c*) strategies that lower the infection rate of the host (Hoste and Torres-Acosta, 2011). In cattle, some alternative or complementary approaches to synthetic anthelmintics for the control of GI nematodes have been investigated, like: pasture management, selection of resistant breeds, biological control and development of vaccines (Vercruyse and Dorny, 1999; Waller, 1999; Rinaldi and Geldhof, 2012; Vlamincx et al., 2015). However, one of the most promising novel approaches for nematode control, and not yet widely explored in cattle, is the use of bioactive plants to modulate the parasite biology.

1.4 Bioactive plants

In the Nordic countries, as in many other areas of the world, there is a long tradition of using plants with anthelmintic properties to treat parasitised animals (Waller et al., 2001). In less favoured regions of the world plants or plant extracts are normally used as dewormers, particularly where the access to synthetic anthelmintics is limited or costly. However, the believed anthelmintic activity of these plants is generally based on anecdotal evidence without scientific validation (Githiori et al., 2006). Yet, during the last decades, and largely driven by the development of AR in parasitic nematodes, extensive research has been conducted to confirm and validate the anthelmintic activity of several bioactive plants (Athanasiadou et al., 2007; Hoste et al., 2015). Most of these studies have explored the potential use of forages with anthelmintic activity as part of the animal's diets, as a nutraceutical approach (Waller and Thamsborg, 2004; Hoste et al., 2014). A nutraceutical, in the context of veterinary science, has been defined as any livestock feed which combines nutritional value with beneficial effects on animal health, and is usually offered to animals during several days or weeks (Hoste et al., 2015). Under this approach, the available scientific evidence indicates that bioactive plants may control parasitic infections by exerting direct anthelmintic effects in the host and/or indirectly by enhancing the immune response of the host to nematode infections through an improved nutrition (Hoste et al., 2012). The direct anthelmintic effects of plant nutraceuticals have been attributed to the activity of certain phytochemicals or plant secondary metabolites (PSM) (Barry et al., 2001; Athanasiadou and Kyriazakis, 2007). Condensed tannins (CT) have been the most widely investigated PSM for anthelmintic effects and promising results have been obtained using tannin-rich plants of the family *Fabaceae* (legumes), mainly against GI nematodes in small ruminants (reviewed by Hoste et al., 2006; 2012). Plants with PSM other than CT have also been examined for anthelmintic effects, like papaya, fig and pineapple containing cysteine proteinases (Stepek et al., 2004) and forage chicory containing sesquiterpene lactones (further discussed in Section 1.5).

To date, very few *in vivo* studies have examined the use of bioactive forages against GI nematodes of cattle, which may be attributed to less incentives due to limited development/diagnosis of AR in cattle nematodes and a stronger immunity of adult cattle to parasitic nematodes (compared to small ruminants), as well as higher cost of cattle as experimental animals (Sandoval-Castro et al., 2012). Nonetheless, evidence of direct anthelmintic activity of condensed tannins against free-living and parasitic stages of cattle nematodes has been confirmed *in vitro* (Novobilský et al., 2011; 2013; Desrues et al., 2016). Sievers and Nannig (2006) reported that calves with access to plantain (67.3% plantain DM in the field) during 7 h daily had significantly lower FEC of *Ostertagia* spp. and *Trichostrongylus* spp. in comparison with animals grazing ryegrass/clover and exposed to similar larval challenge from pasture. Xhomfulana et al. (2009) reported that Nguni and crossbreed cattle naturally infected with GI nematodes and supplemented with *Acacia*

karroo leaf meal (rich in CT) had lower FEC and worm burdens of *Haemonchus contortus* and *Oesophagostomum colombianum*, in comparison with non-supplemented controls. In the same study, a third group of cattle supplemented with sunflower cake (energy content not indicated) had also lower worm burdens compared to controls, thus, the anthelmintic effects of *A. karroo* leaf meal may have been a result of an improved nutrition due to supplementation rather than a direct anthelmintic activity of CT (Xhomfulana et al., 2009). Recently, Shepley et al. (2015) described that dairy heifers, naturally infected with GI nematodes and supplemented with mixed birdsfoot trefoil (32% birdsfoot trefoil DM in the mix) during 45 min daily, had significantly lower FEC compared with heifers grazing the same pasture but supplemented with alfalfa silage. Undoubtedly, substantial research is needed to further explore known and novel anthelmintic forages in cattle, with the aim to include them in integrated parasite control approaches and reduced the reliance on the still-effective anthelmintics.

1.4.1 Methods for evaluation of anthelmintic activity

Prior to on-farm use, the confirmation of the anthelmintic effects of bioactive forages and their PSM as well as their potential toxic effects must be evaluated in controlled studies (Githiori et al., 2006). Several *in vitro* assays and recommendations for *in vivo* studies have been proposed to investigate these issues, some of which are briefly described here.

1.4.1.1 *In vitro* assays

In vitro studies involve the incubation of GI nematodes with the plant extract or PSM of interest, aiming to test a direct effect on the parasite integrity or other biological processes, while excluding any host-related factors (Jackson and Hoste, 2010). *In vitro* studies can be performed as a preliminary screening of the best plant candidates for further *in vivo* trials and/or after *in vivo* studies to confirm the direct anthelmintic effects of plant compounds (Hoste et al., 2015). *In vitro* assays allow the testing of a wide range of plant compound/extract concentrations and using different nematode life stages. However, PSM concentrations tested *in vitro* may not correspond to the levels reaching the site of infection in the host, thus extrapolation of results from *in vitro* to *in vivo* conditions is complicated (Hoste et al., 2015). Detailed guidelines of *in vitro* assays have been described (Jackson and Hoste, 2010).

Several *in vitro* assays have been proposed to test the direct anthelmintic effects of plant compounds against free-living stages, most of them originally developed to investigate the anti-parasitic activity of synthetic anthelmintics (Coles et al., 1992; Demeler et al., 2010). *In vitro* assays with free-living stages are the most commonly used methods to test the anthelmintic activity of PSM and those relevant for the present PhD studies are summarised

here. The egg hatch assay (EHA) measures the ability of the compound of interest to inhibit the hatching of nematode eggs to L1, incubated under optimal conditions for hatching. The larval feeding inhibition assay (LFIA) investigates the inhibitory effects of a tested compound on the feeding behaviour of L1 exposed to fluorescein-labelled *Escherichia coli*; unaffected larvae are distinguishable by the presence of the labelled bacteria in their gut observed under fluorescence microscopy. The larval development assay (LDA) investigates the effect of a compound to inhibit the development of nematode eggs into infective L3. The larval motility inhibition assay (LMIA) measures the ability of a compound to paralyse L3 and inhibit their migration through a nylon mesh. And the larval exsheathment inhibition assay (LEIA) test the effect of a plant compound to delay the artificially-induced exsheathment of L3.

In contrast with free-living stages occurring in the environment, parasitic stages in the host have different biochemical characteristics and detoxification mechanisms, thus the same compound can exert different effects and affect distinct targets in free-living or parasitic stages (O'Grady and Kotze, 2004; Hoste et al., 2015). Moreover, parasitic (immature and adult) stages are the expected targets of dietary PSM in the host, and therefore it is of interest to confirm a direct activity of plant compound against them. The adult motility inhibition assay (AMIA) tests the effect of PSM to paralyse live adult worms, recovered from animals immediately post-mortem, and incubated in culture medium for up to 48 – 96 h (depending on the nematode species). However, the isolation of live adult nematodes requires the slaughter of infected animals, involving considerable resources that limit its routine performance. Alternative methods have been proposed to test the direct anthelmintic effect of plant compounds using the adults of the free-living nematode *Caenorhabditis elegans* (Katiki et al., 2011; Kumarasingha et al., 2014). Recently, an image-based whole-organism assay using *H. contortus* L3 and developing L4 has been explored for the low cost testing of compounds with potential anthelmintic activity (Preston et al., 2015).

1.4.1.2 Controlled *in vivo* studies

In vivo studies are essential to confirm the anthelmintic potential of bioactive forages in infected animals before on-farm use, as well to detect potential toxic effects in the animals. Based on the available evidence from CT-rich plants, it is expected that the reduction in worm burdens induced by bioactive forages is lower than following treatment with (efficacious) synthetic anthelmintics (Ketzi et al., 2006). Nevertheless, it is critical to establish the reduction levels of parasitism achieved by bioactive plants, particularly a dose-response, before on-farm studies can be designed (Githiori et al., 2006; Hoste et al., 2015). At the moment, no standardized guidelines exist for the evaluation of anthelmintic forages in controlled *in vivo* trials, although some recommendations have been proposed (Hoste et al., 2015). When testing the direct anthelmintic effect and the role of PSM it is of particular importance to exclude confounding factors such as improved nutrition (resulting from

high-protein/energy diets) which is known to enhance the immune response to helminth infections (Mansour *et al.* 1992; Van Houtert and Sykes, 1996; Coop and Kyriazakis, 2001). To overcome this, nutrient-balanced diets should be offered to the tested and control groups, particularly regarding protein and energy levels (Hoste *et al.*, 2015).

1.5 Chicory

Chicory (*Cichorium intybus* L., *Asteraceae*) is a perennial, deep-rooting herb that can be found as a wild plant on natural grasslands or in roadsides (identifiable by its distinctive bright blue flowers; Fig 1.3) and as cultivated varieties in most temperate areas of the world, including Denmark (Barry, 1998; Li and Kemp, 2005; Nielsen *et al.*, 2009; Wang and Cui, 2011). In Europe and other regions, chicory has traditionally been used for human consumption and as a medicinal plant (Lucchin *et al.*, 2008; Hitova and Melzig, 2014). Nowadays, cultivated chicory can be classified in four types, based on its applications: *i*) “industrial” or “root” chicory, for the production of inulin and as a coffee substitute; *ii*) “Brussels” or “witloof” chicory, for production of etiolated leaves (“chicons” or “*julesalat*” in Danish); *iii*) “leaf” chicory for fresh or cooked consumption and *iv*) “forage” chicory for animal feeding (Street *et al.*, 2013). Selection of forage cultivars for livestock nutrition in the 1980’s culminated in the release of the first commercial forage variety, “Grassland Puna” in New Zealand (Rumball, 1986). Since then, several forage varieties have been developed (Fig. 1.3)(Rumball *et al.*, 2003a; 2003b; Li and Kemp, 2005; Foster *et al.*, 2006).



Fig. 1.3. Chicory flowers (left) and forage chicory cv. Spadona (middle) and cv. Puna II (right)

Cultivars of forage chicory are predominantly active in warm seasons, with plant growth occurring at $>10^{\circ}\text{C}$ (if enough precipitation) and dormant in winter. In general, forage chicory cultivars are rich in minerals (e.g. Zn, B, Mn), have variable crude protein levels ($\sim 100 - 250$ g CP kg DM⁻¹), higher organic matter digestibility and higher rumen fractional disappearance rate than grasses, low NDF levels and high voluntary feed intakes (reviewed by Li and Kemp, 2005). Inclusion of forage chicory is increasingly advocated to enhance the protein, mineral and vitamin content of pastures and to sustain forage

production during warm and dry periods when ryegrass growth is low (Sanderson et al., 2003; Høgh-Jensen et al., 2006; Pirhofer-Walzl et al., 2011; Langworthy et al., 2015; Roca-Fernández et al., 2016). Similar or higher weight gains have been reported in cattle grazing pure forage chicory, compared with animals grazed on ryegrass (Fraser et al., 1988; Clark et al., 1990; Parish et al., 2012), and comparable positive effects on animal growth have been observed in sheep and deer (Scales et al., 1995; Hoskin et al., 1999b; Kidane et al., 2014). Chicory-based diets have also been reported to sustain milk production in cattle (Vaugh et al., 1998; Chapman et al., 2008; Muir et al., 2014) and in sheep (Di Grigoli et al., 2012). However, the rapid growth of low-quality reproductive stems in late spring/early summer (from the second year onwards) is an agronomical shortcoming of forage chicory, while its relatively low persistency (of around 3 years) and its dormant state during winter adds further limitations to its use (Clark et al., 1990; Sanderson et al., 2003; Li and Kemp, 2005; Lee et al., 2015; Muir et al., 2015).

1.5.1 Effects of forage chicory on GI nematodes of ruminants

1.5.1.1 *In vivo* anthelmintic effects of forage chicory

Since the mid-1990's, several *in vivo* studies have explored the potential anthelmintic effects of dietary forage chicory on parasitic nematodes, mainly in sheep (Scales et al., 1995; Marley et al., 2003b; Athanasiadou et al. 2005; Tzamaloukas et al., 2005; Tzamaloukas et al., 2006; Athanasiadou et al., 2007; Heckendorn et al., 2007; Nielsen et al., 2009; Kidane et al., 2010; Miller et al., 2011) but also in deer (Hoskin et al., 1999a) and cattle (Marley et al., 2014). The main parasitological findings of these *in vivo* studies, as well as the chicory cultivar used and the percentage of chicory DM in the diets, are summarised in Table 1.1. None of these studies investigated the content of SL in forage chicory, and when CT were screened they were detected at levels <0.5% CT in DM. Scales et al. (1995) reported a reduced FEC and *Teladorsagia* worm burden in lambs grazing forage chicory. They also recovered a lower number of infective L3 from chicory swards, as compared with ryegrass pastures. These authors described three hypothetical explanations for the reduced parasitism observed in lambs grazing chicory: *i*) a direct dietary modulation of chicory on the worms, *ii*) an increased animal resilience to the infection (due to a higher nutritional quality) and/or *iii*) to an indirect reduction in pasture-borne infections (Scales et al., 1995). From the subsequent *in vivo* studies with chicory, particularly short feeding trials with experimental infections in lambs and preventing reinfection from pasture, it was strongly suggested that feeding forage chicory could directly affect abomasal worms, but not intestinal nematodes (Athanasiadou et al. 2005; Tzamaloukas et al., 2005; Heckendorn et al., 2007). Tzamaloukas et al. (2006) reported that lambs, trickle infected with *T. circumcincta* while grazing chicory, then moved to a ryegrass pasture and challenged with *T. circumcincta* L3, had lower late L4 and adult worm counts than lambs only grazing ryegrass. The authors concluded that chicory enhanced the immune response against *T.*

circumcincta, evidenced by higher levels of mucosal mast cells and globule leucocytes in chicory lambs, and they attributed these findings to the high protein content in the chicory pasture, and not to a direct anthelmintic effect.

Marley et al. (2014) reported the only *in vivo* study to date exploring the anthelmintic effects of forage chicory in grazing cattle. In their study, beef steers were naturally infected with GI nematodes and grazed a mixed chicory/ryegrass pasture (~24% chicory DM in the field) for 18 weeks which resulted in similar FEC, proportion of *O. ostertagi* or *Cooperia* spp. L3 in larval cultures, serum pepsinogen and *O. ostertagi*-antibodies levels compared to infected controls grazing ryegrass. Post-mortem worm counts were not conducted. Evidence from *in vivo* studies in sheep suggests that increased levels of chicory ($\geq 50\%$ chicory DM in the diet) may be necessary to affect GI parasitism in the host.

Table 1.1. Summary of published *in vivo* studies investigating the anthelmintic effects of forage chicory in ruminants.

Species	Chicory cv.	Sown	Chicory DM in diet (%)	Study design	Effects in chicory fed animals	Reference
Sheep	Grasslands Puna	pure	>85%	Grazing, exp. + nat. infections with mixed GI nematodes	Lower FEC and <i>Teladorsagia</i> worm counts (P<0.05)	Scales et al., 1995
Deer	Grasslands Puna	pure	>56%	Grazing, nat. infections with mixed GI nematodes and lungworms	No effect on FEC or lungworm L1 counts. Lower clinical signs associated with parasitism, requiring less anthelmintic treatment	Hoskin et al., 1999a
Sheep	Grasslands Puna	pure	n.i.	Grazing, nat. infections with mixed GI nematodes	No effect on FEC or intestinal worm counts. 19-41% lower abomasal adult counts (P<0.001) and 32-60% lower abomasal L4 (P<0.01)	Marley et al., 2003b
Sheep	n.i.	pure	80%	Grazing, exp. infections with <i>Trichostrongylus colubriformis</i>	No effect on FEC, adult or immature worm counts	Athanasiadou et al., 2005
Sheep	n.i.	pure	87%	Grazing, exp. infections with <i>Teladorsagia circumcincta</i>	No effect on FEC, female per capita fecundity or immature worm counts. Reduction in adult male worms (P<0.01) and lower total adult counts by 43% (P=0.14)	Tzamaloukas et al., 2005
Sheep	n.i.	pure	91%	Lambs trickle infected with <i>T. circumcincta</i> while grazing chicory. After anthelmintic treatment, lambs were challenged with <i>T. circumcincta</i> L3 and grazed on ryegrass	No effect on FEC. Reduced numbers of late L4 and adult worms (P<0.05)	Tzamaloukas et al., 2006

(continuation Table 1.1)

Sheep	Grasslands Puna	pure	~80%	Stabled lambs fed fresh-cut chicory. Exp. infections with <i>Haemonchus contortus</i> and <i>Cooperia curticei</i>	Reduced total egg output and FECDM of <i>H. contortus</i> ($P<0.01$). No effect on FECDM of <i>C. curticei</i> , on total FECDM or on adult worm counts	Heckendorn et al., 2007
Sheep	Grasslands Puna	pure	~80%	Grazing, nat. infections with mixed GI nematodes in ewes and lambs. Ewes were treated or not with anthelmintics	Lower FEC in lambs from untreated ewes on chicory ($P<0.001$). No effect on FEC of undrenched ewes or on adult worm burden in lambs	Athanasiadou et al., 2007
Sheep	Grasslands Puna	mixed	2-51% in the field; <6% intake	Grazing, nat. infections with mixed GI nematodes	No effect on FEC	Nielsen et al., 2009
Sheep	Puna II	pure	50%	Grazing, nat. infections with <i>T. circumcincta</i>	Reduced FEC ($P<0.001$) and lower L3 recovered from chicory swards ($P=0.07$)	Kidane et al., 2010
Sheep	Oasis	pure	n.i.	Grazing, nat. infections with mixed GI nematodes	Reduced FEC counts ($P<0.05$), lower FAMACHA scores requiring less anthelmintic treatments	Miller et al., 2011
Cattle	Puna II	mixed	~24%	Grazing, nat. infections with GI nematodes	No effect on FEC, proportion of <i>O. ostertagi</i> or <i>Cooperia</i> spp. L3 in larval cultures, serum pepsinogen or <i>O. ostertagi</i> -antibodies	Marley et al., 2014

pure = monoculture; mixed = sown as mix sward; n.i. = not indicated; exp. = experimental; nat. = natural

1.5.1.2 Effect of forage chicory on the development and survival of free-living nematode stages

Studies have also described a lower recovery of infective L3 in chicory pastures, potentially attributed to a reduced development of free-living stages. Moss and Vlassoff (1993) reported a lower recovery of total L3 from a pure chicory sward, compared with a ryegrass field, seeded with strongyle and *Nematodirus* spp. eggs. The authors attributed their findings to the markedly lower DM in the chicory field (40% less DM, compared with ryegrass) which could have changed the micro-climate in the sward and limited the development and survival of L3 (Moss and Vlassoff, 1993). Reduced herbage mass and plant cover has also been suggested to limit the development and survival of larval stages from sheep nematode (Thamsborg et al., 1996). Niezen et al. (1998) described a lower total number of L3 and reduced larval survival, particularly of *T. circumcincta*, recovered from a chicory sward seeded with GI nematode eggs, as compared with a ryegrass field. No information of the DM in the chicory field was provided. Marley et al. (2006) reported a significantly lower recovery of L3 (per kg forage DM) from chicory seeded with *C. curticei* eggs and cultivated under greenhouse conditions (as compared with ryegrass) and a significantly lower *T. circumcincta* L3 (per kg forage DM) from chicory swards (compared to ryegrass pastures). Kidane et al. (2010) recovered a lower number of L3 from a chicory sward grazed by naturally infected lambs, although not significantly different from the larval recovery from a ryegrass/clover field. These studies strongly suggest a potentially lower infection pressure on animals grazing chicory swards. Experiments have also explored a potentially reduced development of free-living nematode stages isolated from animals fed with forage chicory. Schreurs et al. (2002) reported no differences in the *in vitro* egg hatching or larval development of GI nematodes isolated from faeces of deer grazing chicory. In contrast, Marley et al. (2003a) observed a significantly lower development of *H. contortus* and *C. curticei* eggs into L3 in cultured faeces from lambs fed with forage chicory, in comparison with the larval development in faeces from animals grazing ryegrass/clover. A decreased larval development of *T. colubriformis* has also been reported in faeces from animals fed CT-rich diets (Niezen et al., 2002).

The available literature summarised here suggests that forage chicory can negatively affect infections with GI nematodes through: a) direct anthelmintic effects on parasites in the host, b) indirect effects through improved nutrition and enhancement of immunity, and c) a reduced larval intake from pasture due to a lower development and survival of infective nematode stages in chicory swards and in faeces of chicory-fed animals. The direct anthelmintic effects of chicory, which are the main subject of this PhD thesis, are believed to derive from its content of bioactive compounds or plant secondary metabolites (PSM).

1.5.2 Bioactive compounds in forage chicory

As a member of the *Asteraceae*, chicory is known to produce biologically-active sesquiterpene lactones (SL), which are also responsible for its bitter taste (Rees and Harborne, 1985; Peters and van Amerongen, 1997; Kisiel and Zielin, 2001; Zidorn, 2008). Several other phytochemicals have been described in chicory, such as chicoric, chlorogenic and caffeic acids (all derivatives of hydroxycinnamic acids), coumarins (e.g. chicoriin, aesculin) and flavonoids (e.g. several quercetin and kaempferol derivatives) (Rees and Harborne, 1985; Heimler *et al.*, 2009; Street *et al.*, 2013; Sinkovič *et al.*, 2015). Chicory also has very low levels of condensed tannins (CT), normally <0.5% CT in DM (Jackson *et al.*, 1996; Tzamaloukas *et al.*, 2005; Heckendorn *et al.*, 2007; Lombardi *et al.*, 2015). Chicory contains sugars (fructose, glucose and sucrose) in both roots and leaves and highly soluble fructans (e.g. inulin), primarily in the roots and with only traces detectable in the leaves (Ernst *et al.*, 1995). Forage chicory also has high levels of pectins (80 – 90 g kg DM⁻¹; Liu *et al.*, 2013). Inulin fructans have been indicated to stimulate the human immune system, reducing the risk of GI diseases (Roberfroid, 2005), while the inclusion of chicory leaves and roots as dietary fibre can modulate the gut microbiota in pigs and chickens (Ivarsson *et al.*, 2012; Liu *et al.*, 2012). Chicory roots have been reported to significantly reduce the numbers of *Ascaris suum* in pigs (Jensen *et al.*, 2011), while pigs fed with pure inulin had reduced worm burdens of *Oesophagostomum dentatum* (Petkevicius *et al.*, 2003). These anthelmintic effects have been linked with the rapid fermentation of inulin in the large intestine and the consequent increased production of short-chain fatty acids that are known to directly affect the worms (Petkevicius *et al.*, 2004). No similar parasitological studies with inulin have been conducted in ruminants, although the use of fructan-rich diets as prebiotics is increasingly being investigated in cattle (Samanta *et al.*, 2013).

1.5.3 Sesquiterpene lactones

Sesquiterpene lactones are a group of extremely diverse natural terpenoids, including around 5,000 different molecules, mainly produced by plants of the family *Asteraceae* (Seaman, 1982; Schmidt, 2006). These compounds have a wide range of biological functions but are mainly associated with plant defence (Rees and Harborne, 1985; Gershenzon and Dudareva, 2007). Natural SL are classified in several groups, including: germacranolides, eudesmanolides, elemanolides, heliangolides, cadinanolides, eremophilanolides, xanthanolides, guaianolides and pseudoguaianolides, among others (Schmidt, 2006; Amorim *et al.*, 2013). In the *Cichorieae* tribe of the *Asteraceae*, around 360 distinct SL molecules have been reported, all part of the guaianolides (~243 molecules), eudesmanolides (~73 molecules) and germacranolides (~44 molecules) groups (Zidorn, 2008). In chicory, three guaianolide SL and their 11, 13-dihydro-derivatives have been described: lactucin, 8-deoxylactucin, lactucopicrin, 11, 13-dihydrolactucin, 11, 13-dihydro-8-deoxylactucin and 11, 13-dihydrolactucopicrin (Fig. 1.4) (Rees and Harborne, 1985; Price

et al., 1990; Foster et al., 2011; Ferioli and D'Antuono, 2012; Wulfkuehler et al., 2014; Ferioli et al., 2015). Sesquiterpene lactones are present in chicory leaves and roots as free molecules or as glycosides bound to carbohydrates (Ferioli et al., 2015).

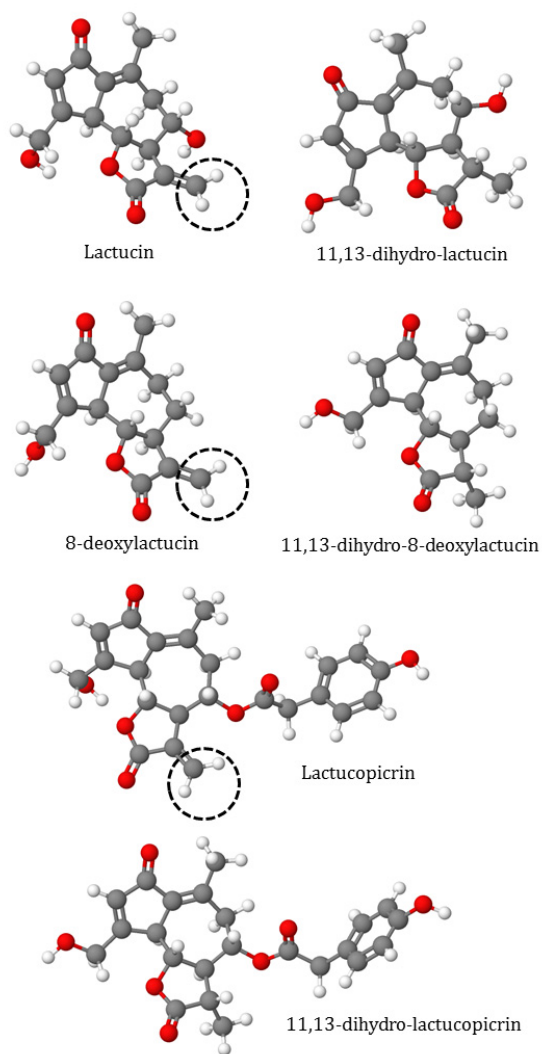


Fig. 1.4. Sesquiterpene lactones described in chicory leaves and roots. Dotted circles indicate the α -methylene (CH_2) group (absent in 11,13-dihydro-derivatives). Atoms: Grey = C; Red = O; White = H (structures designed in MolView.org).

1.5.3.1 Biological activities of sesquiterpene lactones

Numerous studies have demonstrated substantial biological activities exerted by SL, including antibiotic, antiprotozoan, antioxidant and anticancerigenous properties (Bischoff et al., 2004; Cavin et al., 2005; Schmidt, 2006; Barrera et al., 2013; Chadwick et al., 2013). As an example, artemisinin (and its derivatives), a SL originally isolated from sweet wormwood (*Artemisia annua*), is currently one of the major antimalarial drugs used worldwide (Meshnick, 2002; Gershenzon and Dudareva, 2007). Guaianolide and other SL are known to exert potent cytotoxic activities, mainly related to the presence of an α -methylene (CH_2) group attached to the γ -lactone in the SL molecule (Fig. 1.4) (Simonsen et al., 2013). This α -methylene group reacts with sulfhydryl (thiol) groups in any cysteine and cysteine-containing peptide, enzyme or other protein by a Michael-type addition, leading to an alkylation of cellular macromolecules and disruption of cellular functions (e.g. impairing cell signalling, cell replication and mitochondrial respiration) (Schmidt, 2006; Simonsen et al., 2013). As an example, SL were reported to reduce the intracellular concentration of free glutathione in *Leishmania mexicana mexicana*, which led to a toxic intracellular accumulation of reactive oxygen species (ROS) and blocked cell proliferation (Barrera et al., 2013). However, dihydro-derivatives of SL (also present in chicory) lack this α -methylene group but still can exert a strong biological activity (Lee et al., 1977; Schmidt, 2006; Ghantous et al., 2010), indicating that there is not only one biological mechanism of action for all SL.

1.5.3.2 Toxicity of sesquiterpene lactones

The unspecificity of the α -methylene–cysteine reaction induced by most SL, which is responsible for the broad biological activities of these molecules, can potentially induce toxic effects in humans and animals exposed (Amorim et al., 2013). As an example in veterinary medicine, SL have been described as the toxic compounds behind the noxious effects to animals of *Helenium* spp. and *Hymenoxys* spp. plants, which contain pseudoguaianolides, helenanolides and secohelenanolides (Burrows and Tyrll, 2004). However, the long tradition of growing and consumption of chicory by humans and the increasing use of chicory for animal feeding suggest that chicory and its guaianolide SL are non-toxic at the concentrations present in human and animal diets. To the knowledge of the author, only one study has investigated the toxicity of chicory in mammals; Schmidt et al. (2007) evaluated the toxicity of chicory root extracts containing 60% of guaianolide SL in rats dosed orally for 28 days (top dose 1000 mg extract BW kg^{-1} day $^{-1}$). The authors did not detect any clinical, post-mortem or histopathological signs of treatment-related effects in animals from any dosage group. However, the same chicory extract induced an inhibition in colony numbers of *Salmonella typhimurium* and *E. coli* when incubated *in vitro* at doses of 5 mg/plate (Schmidt et al., 2007). These results suggest a marked difference in

susceptibility to dietary SL from chicory in mammals and other organisms, and similar effects may be induced in GI nematodes.

1.5.3.3 Sesquiterpene lactones as anthelmintic compounds in forage chicory

The role of SL in the anthelmintic activity of forage chicory was first suggested by Schreurs et al. (2002), who reported a reduced *in vitro* motility of lungworms L1 when incubated in rumen and abomasal fluid from deer grazing chicory. The authors speculated that SL present in rumen and abomasal fluid of chicory-grazed deer were involved in the lower motility of lungworm L1; however, no detection of SL was performed. Molan et al. (2003) further explored the anthelmintic effects of SL from chicory and reported direct inhibitory effects of crude SL-extracts from chicory roots on the motility of lungworm L1 and L3 and of GI nematode L3 from deer. Still, chemical confirmation of SL in the tested extracts was not performed. The strongest evidence of the anthelmintic effect of SL from chicory was reported by Foster et al. (2011), who observed a dose-dependent inhibition in the egg hatching in *H. contortus* by extracts containing free SL isolated from forage chicory cv. Grasslands Puna and cv. Forage Feast. The study also characterised the three main SL of chicory in the tested extracts (LAC, 8-DOL and LCP) and detected an increased anthelmintic effect of the Grasslands Puna-extract, which was linked with its higher content of 8-DOL (Foster et al., 2011). Further, the authors suggested that dihydro-derivatives of the three main SL could have been present in their extracts, although their detection was not performed and the contribution of these compounds to the anthelmintic activity of forage chicory is unknown (Foster et al., 2011). Moreover, no studies have examined the direct effect of SL or SL-containing extracts from forage chicory against parasitic nematode stages, which are the expected primary targets of dietary SL in the host, or towards key mechanisms in the nematode life cycle that may clarify the *in vivo* anthelmintic effects of forage chicory.

Chapter 2. The present investigations

2.1 Objectives:

The overall objective of the present investigations was to contribute towards the integrated control of bovine GI nematodes by exploring the efficacy of the most common drug used for anthelmintic treatment in Danish cattle and the use of forage chicory as a potential complementary parasite control strategy.

The specific objectives were:

- I. To assess the efficacy of ivermectin against GI nematodes in naturally infected Danish cattle by the FECRT and examining different statistical approaches for estimation of drug efficacy, and to report the general use of anthelmintics in selected farms (Manuscript I).
- II. To investigate the anthelmintic effects of forage chicory-rich diets against experimental infections with GI nematodes in stabled and grazing calves, in relation with the PSM content in chicory (Manuscript II).
- III. To study the direct effects of sesquiterpene lactone-containing extracts isolated from two forage chicory cultivars on key biological processes of free-living and parasitic stages of the cattle nematode *Ostertagia ostertagi* (Manuscript III).

2.2 Materials and methods

A brief summary of the materials and methods used in the present investigations (including unpublished pilot studies) is outlined in this section and further details can be found in the attached Manuscripts I – III.

2.2.1. Field efficacy of ivermectin and prescription of anthelmintics in Danish cattle

2.2.1.1 Efficacy of ivermectin against GI nematodes evaluated by different methods for analysis of faecal egg count reduction data (Manuscript I)

The field efficacy of ivermectin against GI nematodes in Danish cattle was investigated by the FECRT. Cattle farms (~50) were contacted during spring 2013 and 2014 through local veterinarians across Denmark. A total of 19 farmers accepted the invitation and 20 FSG calves were sampled in each farm. Animals were not treated with anthelmintics 8 weeks prior to sampling and had ≥ 4 weeks of access to pastures. Due to very low FEC in most of the properties only six farms were selected for the final study. All six farms were visited during June – September of 2013/2014 to conduct the FECRT (Fig. 2.1). In each farm, 10 FSG calves were treated with recommended dose of injectable IVM (IVM group) while 10 calves remained untreated as controls (CTL group). Faecal samples were collected from all animals at day of treatment (Day 0) and 14 days post-treatment (Day 14) and transported to the laboratory for FEC analysis. Pooled larval cultures per group were prepared on Day 0 and 14 for the species-specific diagnosis of *O. ostertagi* and *C. oncophora* L3 by real-time PCR (Höglund et al., 2013).

Estimation of drug efficacy was performed by calculating the arithmetic mean FEC reduction percentage (FECR%) post-IVM treatment according to WAAVP guidelines (Coles et al. 1992). Furthermore, we used two Bayesian modelling approaches to evaluate the efficacy of IVM treatments; the “Bayescount” method modified from Denwood et al. (2010) and Geurden et al. (2015) and the “EggCounts” method described by Torgerson et al. (2014). Both MCMC methods model the observed pre-and post-treatment FEC data as separate gamma-Poisson (negative binomial) distributions; the difference of the means between these two distributions is the FECR%. These methods also account for the FEC aggregation within animal groups and the Poisson errors of the egg counting process. However, the major difference between these Bayesian methods is in the prior distributions used in the modelling. The EggCounts method defines a single prior to the dispersion parameter (over-dispersion), and as a consequence, assumes that no variation in drug efficacy exists between animals in the same treated group. In contrast, the Bayescount relaxes this assumption and allows such variation in the modelling. Treatment efficacies

estimated by the WAAVP and Bayescount methods were calculated using the FEC variation in the IVM and CTL groups (with CTL) or the FEC variation only in the IVM group (without CTL) between Day 0 and 14, whereas the analysis by the EggCounts only included the FEC data from the IVM group at Day 0 and 14.

The efficacy of IVM treatments by each method was interpreted based on the mean FECR% and lower 95% CI as recommended by Coles et al. (1992) as well as the upper 95% CI as suggested by Lyndal-Murphy et al. (2014), from which we categorised three conditions:

- i)* Efficacious, when mean FECR% and upper CI $\geq 95\%$ and lower CI $\geq 90\%$;
- ii)* Reduced efficacy, when mean FECR% and upper CI was $< 95\%$ and lower CI $< 90\%$;
- iii)* Inconclusive, when none of the above conditions were met.

In addition, a simulation study was performed to compare the performance of the WAAVP method and the two Bayesian approaches (without CTL) to estimate drug efficacy in datasets based on the observed field data from the FECRT. Datasets were simulated based on treatment groups of 10 animals, with simulated FECR = 85% or 97% and varying levels of FEC aggregation values representing those observed in the FECRT studies. A total of 6,000 datasets were generated and analysed by all methods of calculation. The performance of the methods was investigated by: i) studying the probability of the simulated FECR% being included in the 95% CI provided by each method (i.e. coverage probability of the 95% CI), and ii) the classification probability that the methods would correctly detect a reduced efficacy (for a simulated FECR = 85%), efficacious treatment (for a simulated FECR = 97%) or inconclusive results using the interpretation described above. Further, detailed information about prescriptions of anthelmintics in the six selected farms between 2002 and 2012 was retrieved from the Danish system for surveillance of the veterinary use of drugs for production animals, VetStat, as described in Section 2.2.1.2.



Fig. 2.1. First-season grazing calves included in faecal egg count reduction tests performed in Danish cattle farms during 2013 and 2014.

2.2.1.2. Prescription of anthelmintics in Danish cattle: preliminary study

In parallel to the study of the field efficacy of IVM in the selected farms, we briefly examined the prescription of anthelmintics for the full Danish cattle population over the period between 2010 and 2014. Data were retrieved from the Danish system for surveillance of the veterinary use of drugs for production animals, VetStat¹. The selected drugs with their active compounds, administration routes and indications are presented in Table 2.1 and correspond to all anthelmintics registered for use in cattle in Denmark during the period. For each prescription, data retrieved from VetStat included: *a*) the total amount of drug prescribed, *b*) farm identity, *c*) the target body system for which the treatment was prescribed (i.e. GI, respiratory, integumentary or other) and *d*) the targeted age group. In VetStat, cattle are divided into three age groups: *i*) calves < 12 months-old (heifer and bull calves); *ii*) young cattle ≥ 12 months-old (heifers until first calving and steers until slaughter) and *iii*) adult cattle (cows after first calving). The exact number of treatments performed based on each prescription is not recorded in VetStat. Thus, we used the total amount of a prescribed drug, the recommended dose of a given anthelmintic (considering its formulation) and a defined BW for each cattle age group to estimate the number of “defined doses” (DD) for each prescription. The defined BWs for the age groups were estimated from national databases on Danish cattle (Danmap, 2012) and were defined as: 200 kg BW for calves, 450 kg BW for young cattle and 620 kg BW for adult cattle. Due to the defined BW and the likely variation in the actual amount of active compound used for different animals following each prescription, the calculated DD is only a proxy of the number of treated animals. In addition, it was assumed that whenever an anthelmintic was prescribed and sold to a farmer, the drug was used within a month, which may not always have been the case. However, this potential bias is assumed to be consistent across the different drugs and age groups. Data on the total number of cattle and cattle farms in Denmark were extracted from Statistics Denmark (www.statbank.dk, visited on 15 December 2015).

¹ <https://www.foedevarestyrelsen.dk/Leksikon/Sider/VetStat.aspx#>

Table 2.1. Anthelmintics registered for use in cattle in Denmark between 2010 and 2014. The prescription data were retrieved from the Danish system for surveillance of the veterinary use of drugs for production animals, VetStat.

Active compounds	Administration routes	Indicated for parasites of: ^a
Albendazole (ALB)	oral	GI (including liverflukes); RES
Doramectin (DOR)	injectable	GI; RES; IG
Eprinomectin (EPR)	pour-on	GI; RES; IG
Fenbendazole (FEB)	oral	GI; RES
Ivermectin (IVM)	injectable; pour-on	GI; RES; IG
Ivermectin + Clorsulon (IVM + CLR) ^b	injectable	GI (including liverflukes); RES; IG
Ivermectin + Closantel (IVM + CLO)	pour-on	GI (including liverflukes); RES; IG
Levamisole (LEV)	injectable	GI; RES
Moxidectin (MOX)	injectable; oral; pour-on	GI; RES; IG
Oxfenbendazole (OXF)	bolus	GI; RES
Triclabendazole (TRI) ^c	oral	Liverflukes only

^aIndications are based on the efficacy claims of each drug against particular parasite species and summarised to match the prescription registration of the VetStat system by target body system: GI= gastrointestinal system; RES=respiratory system; IG = integumentary system. ^bIntroduced in Denmark in 2011.

^cNot labelled for veterinary use in DK and only available for treatment in individual farms following laboratory diagnosis and dispensation from the general rules issued by the veterinary authorities.

2.2.2. *In vivo* studies of the anthelmintic effects of forage chicory against GI nematodes in experimentally infected cattle (Manuscript II)

Two independent and consecutive experiments were conducted to investigate the direct anthelmintic effects of forage chicory against GI nematodes in cattle. Calves were experimentally infected with laboratory parasite strains in two trials (indoor = Experiment 1 and outdoor = Experiment 2) with similar study design. Re-infections from pasture (Experiment 2) were prevented, thus excluding any potential effect of a lower parasite challenge from chicory pastures as previously described (Moss and Vlassoff, 1993; Marley et al., 2003). In Experiment 1, protein and energy intakes were balanced between feeding groups to relate the possible anthelmintic activity of chicory to its content of PSM and to exclude any indirect effect due an improved nutritional status. Moreover, short infection periods (≤ 42 days) were designed to avoid any potential development of natural immunity during the studies. The two major differences between the trials were:

i) Experiment 1 was performed during October – December 2013 as an indoor trial with calves fed either chicory silage or ryegrass/clover hay during 56 days (Fig 2.2). Throughout Experiment 1, the fodder consumption of calves was monitored to balance protein and energy intakes between groups using concentrates. Experiment 2 in contrast was conducted during August – September 2014 as a grazing trial with calves allocated to a pure chicory sward or to ryegrass/clover pasture for 42 days (Fig. 2.3). In Experiment 2 calves were not supplemented with concentrates and the protein/energy intakes between groups were not controlled.

ii) Experiment 1 tested the anthelmintic effects of chicory silage against mixed-infections with *O. ostertagi* and *C. oncophora* and calves were infected 16 days after the start of the feeding trial. Experiment 2 was conducted to primarily confirm results from Experiment 1 by investigating the effect of fresh chicory and chicory silage on mono-infections with *O. ostertagi* and the animals were infected 7 days after the beginning of the grazing trial.



Fig. 2.2. Experiment 1: calves fed with chicory silage (n=9, left) and control calves fed with ryegrass/clover hay (n=6, right). Calves were penned in subgroups of three animals throughout the study (3 chicory subgroups and 2 control subgroups)



Fig 2.3. Experiment 2: calves grazing a second-year pure chicory sward progressing into reproductive (flowering) stage (n=10, left) and control calves grazing a ryegrass/clover field (n=10, right).

2.2.3. *In vitro* studies of the anthelmintic activity of plant secondary metabolites of forage chicory

2.2.3.1. Studies of the direct anthelmintic effects of purified forage chicory extracts against *Cooperia oncophora*: preliminary investigations

Previous to the commencement of *in vivo* and *in vitro* studies with forage chicory described in Manuscripts II and III, preliminary investigations studied the direct anthelmintic effect of purified extracts from chicory leaves (cv. Spadona) against free-living and parasitic stages of *C. oncophora*. Chicory leaves were collected from an organic dairy farm in Årre, Denmark, on December 2013. Purified chicory extracts were extracted following the method described by Foster et al. (2011), which is a modification of the method reported by Tamaki et al. (1995) to extract and purify free SL from plant material. Briefly, dried leaf tissue was extracted in a Soxhlet apparatus using methanol. The resulting extract was fractionated using solid-phase extraction cartridges to enhance isolation of PSM, such as SL. The obtained eluate was dried, dissolved in 100% dimethyl sulfoxide (DMSO) and serially diluted to obtain concentrations ranging from 30 to 5000 µg dry extract mL⁻¹. Two calves mono-infected with *C. oncophora* (Section 2.2.4) were used for collection of nematode eggs from faeces and for isolation of live adult worms post-mortem. Isolated eggs and adults were used in an EHA or AMIA, respectively. Eggs or adult worms were incubated at the described concentrations of chicory extract (final DMSO concentration of 2% in Mili-Q-H₂O) or 2% DMSO in Mili-Q-H₂O (negative controls).

2.2.3.2. Studies of the direct anthelmintic effects of SL-containing extracts from forage chicory against *Ostertagia ostertagi* (Manuscript III)

Following Experiment 1 (indoor trial), the potential role of SL in the anthelmintic activity of forage chicory against *O. ostertagi* was investigated using SL-extracts from chicory leaves. At the moment of the experiments, only two of the three guaianolide SL from chicory were available as pure standards (lactucin and lactucopicrin) but their high costs made them inaccessible in high enough quantities for *in vitro* studies. Therefore, these pure SL were only used as pure standards for HPLC-MS analyses. In addition, none of the SL dihydro-derivatives from chicory were commercially available. Hence, purified SL-extracts were prepared for the *in vitro* studies. Sesquiterpene lactones were extracted from leaves of chicory Spadona and Puna II (Fig. 2.4) following the procedure described by Ferioli and D'Antuono (2012) with the modifications detailed in Manuscript III. This method selectively fractionates total (free and unbound) SL from phenols and other plant compounds. The resulting dried extracts were not readily dissolved in PBS but were successfully dissolved with 100% DMSO. Stock solutions of both purified extracts in DMSO were prepared for the *in vitro* assays. Purified extracts dissolved in 100% methanol were used for characterization of SL by HPLC-MS. *Ostertagia ostertagi* L1 or L3 were used for

LFIA or LEIA, respectively, as described by Jackson and Hoste (2010) and Novobilský et al. (2011), with the modifications described in Manuscript III. In the LFIA, final concentrations of Spadona and Puna II extracts ranged from 10 to 500 µg dry extract mL⁻¹. In the LEIA, final concentrations ranged from 250 to 1000 µg dry extract mL⁻¹. Adult (male and female) worms immediately collected post-mortem were used in two independent adult motility inhibition assays (AMIA), following the methods described by Paolini et al. (2004) and Jackson and Hoste (2010), with the modifications described in Manuscript III. In the AMIA, adult worms were exposed to Spadona and Puna II extracts tested at final concentrations ranging from 10 to 1000 µg dry extract mL⁻¹. In addition, adult worms exposed to 1% DMSO (negative controls) and to the highest concentration of Spadona and Puna II extracts for 24 h were selected for scanning electron microscopy (SEM) investigations.

2.2.4 Parasite material and animals

First-season grazing calves in the field trials acquired natural infections with GI nematodes in the six investigated farms (Manuscript I). For the *in vivo* studies with forage chicory (Manuscript II) anthelmintic-susceptible strains from two sources were used: In Experiment 1, experimental infections were performed with *O. ostertagi* and *C. oncophora* strains kindly provided by Prof. Dr. Janina Demeler (Freie Universität Berlin, Germany; “Berlin strains”). While in Experiment 2, experimental infections were conducted with an IVM-susceptible *O. ostertagi* strain obtained from Ridgeway Research, UK (Batch number: OOSG10; “Ridgeway strain”), which was also used for *in vitro* studies (Manuscript III).

Heifer and bull calves (Manuscript I) or only bull calves (Manuscript II – III) between 3 – 12 months-old were used. First-season grazing calves included in the field trials in Danish cattle farms (n= 120; Manuscript I) were Danish Holstein, Danish Holstein crossbreeds and Danish Jersey. Bull nematode-naïve calves (reared indoor since birth) enrolled in the *in vivo* trials of forage chicory (Manuscript II) were Danish Jersey (n=15; Experiment 1) and Danish Holstein (n=20; Experiment 2). Danish Jersey bull-calves (n=4) were used as donor calves for propagation of infective strains of *O. ostertagi* and *C. oncophora* (“Berlin strains”) for Experiment 1 (Manuscript II); two calves were infected with pure *C. oncophora*, while two animals had a mixed infection composed of 30% *O. ostertagi* and 70% *C. oncophora* (according to morphological identification and qPCR). Danish Jersey bull-calves (n=2) were infected with pure *O. ostertagi* (“Ridgeway strain”) and acted as donor calves for propagation of infective L3 for Experiment 2 (Manuscript II) and as a source of nematode eggs for culture of L1 and L3 and of live adult worms for *in vitro* studies (Manuscript III).

2.2.5 Experimental feeds for *in vivo* studies (Manuscript II) and plant material for *in vitro* studies (Manuscript III)

Two chicory fields were cultivated for our investigations; a chicory cv. Spadona field cultivated as a pure sward (7.8 kg seeds ha⁻¹) and a chicory cv. Puna II pasture mixed with timothy (*Phleum pratense*, 6 kg chicory seeds + 6 kg timothy seeds ha⁻¹). Both chicory fields were sown on early May 2013 at the Experimental Farm of the University of Copenhagen, Tåstrup, Denmark. The chicory Spadona field was cultivated to provide forage for *in vivo* studies (Manuscript II) and the Puna II field was cultivated as a back-up in case of poor establishment of the chicory Spadona. Leaves from both cultivars were collected on 23 July 2013 for extraction of SL for *in vitro* studies (Manuscript III). At the time of collection chicory plants were at the vegetative stage (Fig. 2.4).



Fig. 2.4. Chicory cv. Spadona (left) and cv. Puna II (right) leaves at the time of collection for extraction of sesquiterpene lactones (Manuscript III).

When the chicory Spadona field was harvested on September 2013 to prepare chicory silage for *in vivo* Experiment 1 (Manuscript II) approximately 45% of the DM in the field corresponded to unsown species, mainly chamomile (*Matricaria recutita*), with some mugwort (*Artemisia vulgaris*) and shepherd's purse (*Capsella bursa-pastoris*) (Fig. 2.5). For Experiment 1 (Manuscript II), ryegrass/clover (*Lolium perenne*/*Trifolium repens*) hay was used as control feed and all animals were supplemented with a commercial concentrate for calves (Grønmix®, Danish Agro, Denmark) to balance protein and energy intakes between the groups.



Fig. 2.5. Chicory cv. Spadona field two weeks before the cut for ensiling (left), wrapped chicory leaves at ensiling on September 2013 (middle) and chicory silage offered to calves in Experiment 1 (right).

For Experiment 2 (Manuscript II), the chicory Spadona field was allowed to regrow from September 2013 until May 2014, when it was cut for ensiling to provide potentially supplementary feed for calves allocated to the chicory group during the grazing trial. At the start of Experiment 2, chicory plants were progressing into their reproductive (flowering) stage and calves quickly consumed the chicory leaves and the upper parts of stems and flowers, which, in addition to poor leaf regrowth during the study, resulted in a low amount of chicory on the pasture by the second half of the trial (Fig. 2.6, left). Consequently, chicory calves were supplemented *ad libitum* with chicory silage prepared in May 2014 (Fig. 2.6, right) during the last 19 days of the trial. As a control pasture for Experiment 2, a 0.8 ha ryegrass/clover field, left ungrazed during the 2013 and mowed on mid-June 2014, was used. Calves allocated to control pastures were not supplemented during Experiment 2. All experimental feeds used in both *in vivo* studies (Manuscript II) were screened for CT using the acetone-butanol-HCl method reported by Grabber *et al.* (2013) and for SL following the method described by Ferioli and D’Antuono (2012) with the modifications detailed in Manuscript III.

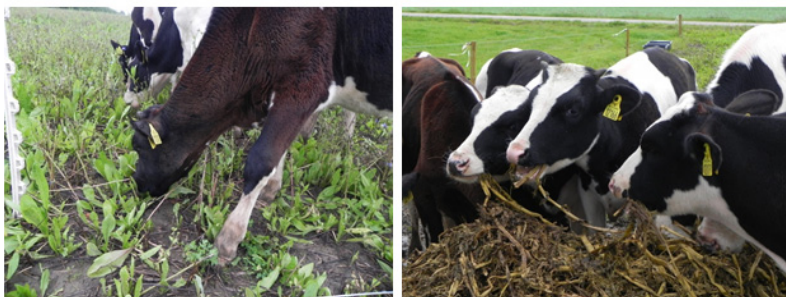


Fig. 2.6. Experiment 2; Calves grazing forage chicory (left) and supplemented with chicory silage in the field (right).

2.2.6 Parasitological analyses

Faecal egg counts in naturally-infected calves (Manuscript I) and experimentally infected animals (Manuscript II) were analysed using a modified McMaster technique with a sensitivity of 5 EPG (Henriksen and Aagard, 1975). Faecal larval cultures were prepared according to Roepstorff and Nansen (1998) and used to isolate infective L3 from naturally-infected cattle (Manuscript I) and from experimentally infected animals (Manuscript II and III). All L3 recovered from naturally-infected cattle before and after IVM treatment (Manuscript I) were analysed by real-time qPCR for species-specific detection and quantification of *O. ostertagi* and *C. oncophora* (Höglund et al., 2013).

In the *in vivo* chicory trials (Manuscript II), adult nematodes were recovered immediately after slaughter for worm counting from the abomasum and small intestine (Experiment 1) or only abomasum (Experiment 2) of experimental calves. Organs were opened into individual buckets, thoroughly washed with warm saline solution (0.9% NaCl, 38°C) and subsamples of diluted digesta were passed through a 25 µm sieve for collection of adult nematodes. Live adult worms from donor calves for *in vitro* assays (AMIA, Manuscript III) were recovered immediately after slaughter using the agar-migration method described by Slotved et al. (1996), in which worms actively migrate from a digesta-agar mixture, adhered to cloths, into warm saline (Fig. 2.7). Migrated live worms were collected using a 20 µm sieve, washed and immediately used in the AMIA.

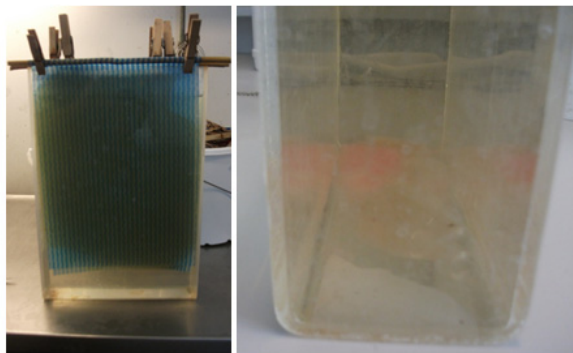


Fig. 2.7. Agar-migration method for isolation of live worms: digesta-agar mixture adhered to three cloths and placed in a container with warm saline (left) and *Ostertagia ostertagi* adult collected in the bottom of the container (red pellets) after 3 h incubation at 37°C (right).

Chapter 3: Manuscripts and preliminary studies

– results and discussion

The main results of the present investigations and their significance are briefly summarised in this chapter and the detailed descriptions of our findings can be found in the attached Manuscripts I – III, with the exception of the preliminary studies presented here.

3.1. Field efficacy of anthelmintics in Danish cattle

3.1.1 Prescription of anthelmintics in Danish cattle: preliminary results

Between 2010 and 2014 (5 years), an estimated 534,858 defined doses with anthelmintics were prescribed for the entire Danish cattle population, with at least one prescription registered in 6,142 different farms (corresponding to 48% of all Danish cattle farms at the time of the investigations). The estimated number of defined doses per active compound, year and age cattle group is presented in Fig. 3.1. Considering the total Danish cattle population for each age group between 2010 and 2014, the estimated defined doses presented in Fig. 3.1 corresponded to the treatment of approximately 4.8% of all calves, 8.4% of all young cattle and 3.3% of all adult cattle once per year in Denmark. Considering all the investigated drugs (in all formulations), treatments with ML accounted for 85% of all prescriptions, and of these 72.1% were performed with IVM alone or in combination with CLR or CLO. Topical anthelmintics were the predominant formulations used for the treatment of calves (71.3%), young cattle (73.7%) and adult cattle (80.2%). Based on the number of estimated defined doses for each indication (“target system”), calves and young cattle were primarily treated against infections of the GI system (45.3% of all treatments), followed by infections of the respiratory system (32.2%) and the integumentary system (20.1%). In comparison, adult cattle were predominantly treated against infestations of the integumentary (39.1% of all treatments), followed by infections of the GI (36.3%) and respiratory systems (20.1%). Whether the commonly-used topical treatments are still efficacious against GI in Danish cattle need to be assessed, but poor efficacy of pour-ons has been previously reported under field conditions in Northern Europe (Sargison et al., 2009; Areskog et al., 2013). Moreover, the selection for AR in nematode populations exposed to variable and sub-therapeutic drug concentrations in cattle treated with pour-on ML has been suggested (Sargison et al., 2009; Sutherland and Bullen, 2015) but this remains to be confirmed.

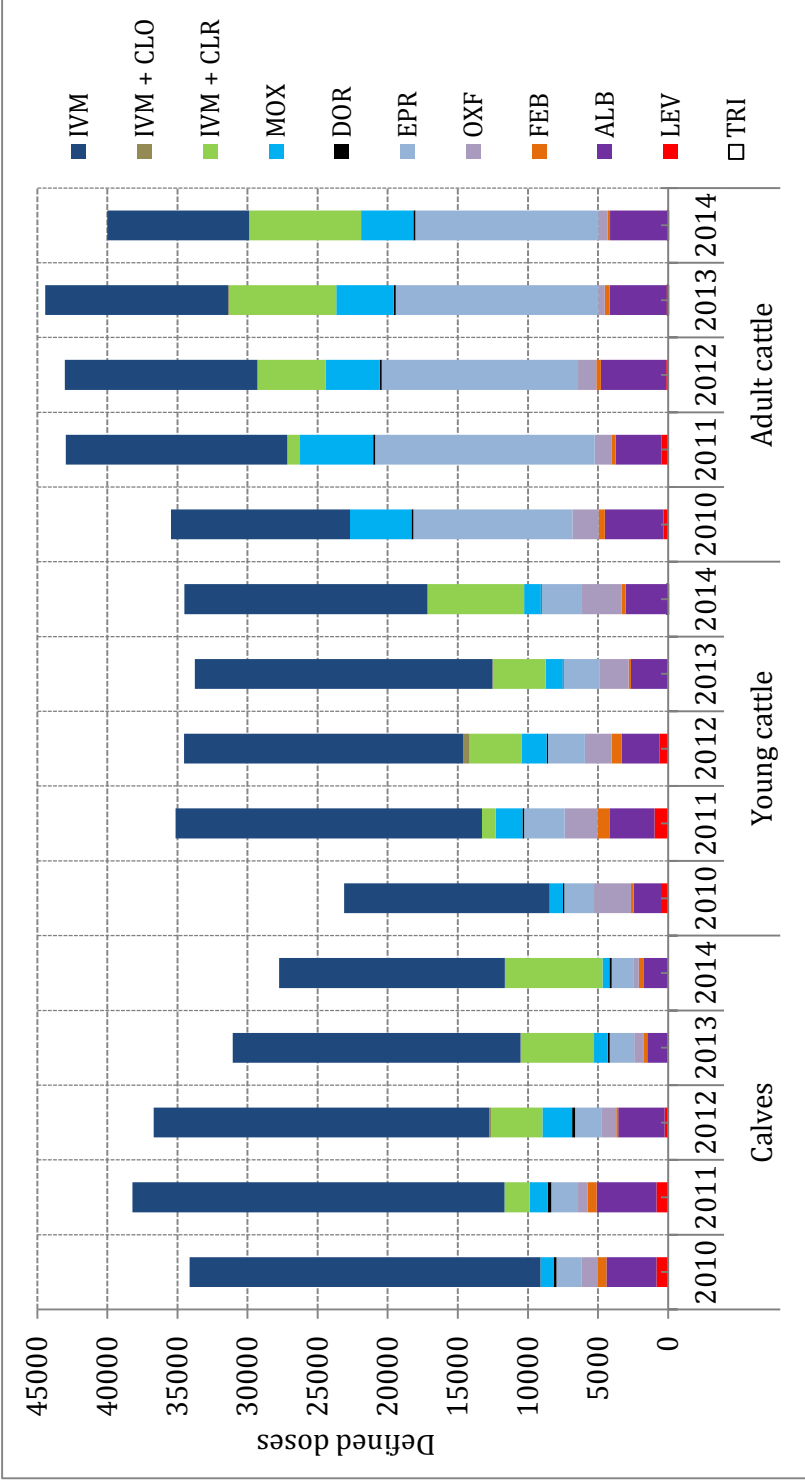


Fig. 3.1. Defined doses of anthelmintics prescribed for all indications in Danish cattle between 2010 and 2014. Data was extracted from the Danish system for surveillance of the veterinary use of drugs for production animals, VetStat. Details on the calculation of defined doses are presented in Section 2.2.1.2. ALB = albendazole; CLR= clorsulon; CLO = closantel; DOR = doramectin; EPR = eprinomectin; FEB = fenbendazole; IVM = ivermectin; LEV = levamisole; MOX = moxidectin; OXF = oxfendazole; TRI = triclabendazole.

3.1.2 Reduced efficacy of ivermectin against GI nematodes in Danish cattle and comparison of methods to analyse FECR data (Manuscript I)

In the initial screening, faecal samples from a total of 380 FSG calves in 19 farms were analysed for FEC during mid-June and early September in 2013 and 2014. Strongyle eggs were detected in all properties, but very low egg excretion levels (mean FEC \leq 44 epg) were seen in most farms (63%). As a result, six farms with mean FEC \geq 75 EPG were selected for FECRT.

In the FECRT, the WAAVP and the Bayescount methods including the FEC of IVM and CTL groups (with CTL) declared reduced treatment efficacy in farm #2, while reduced efficacy was declared in farm #1 only by the WAAVP method. Further, the WAAVP method (with CTL) declared the treatment to be efficacious in farm #5, while IVM treatments in the remaining farms were inconclusive as determined by both methods (with CTL). In comparison, the WAAVP, Bayescount and EggCounts analysing FEC data only from IVM groups (no CTL) unanimously declared reduced IVM efficacy in farms #1, #2 and #4. Additionally in farm #1, five CTL calves with high FEC at Day 14 were treated with injectable moxidectin (0.2 mg kg⁻¹ BW s.c., Cydectin® inj. 10 mg/mL) and FEC at 21 days post-treatment revealed a mean FECR = 89% (95% CI = 82 – 93) by the EggCounts method (data not shown). All methods (no CTL) declared inconclusive results in farm #5 and in two farms the methods were in disagreement: in farm #6 only the two Bayesian methods declared reduced efficacy, while in farm #3 reduced IVM efficacy was only detected by the EggCounts method. Post-treatment, qPCR analyses of L3 from pooled larval cultures revealed that *C. oncophora* was the predominant species surviving injectable IVM in all farms detected, which is in agreement with previous European reports (Demeler et al., 2009; Geurden et al., 2015). The more pathogenic *O. ostertagi* was detected post-treatment in IVM groups from two farms. Yet, reduced efficacy by all methods (no CTL) was only detected in one of these farms (#4) and therefore the potential presence of an *O. ostertagi*-strain less susceptible to IVM in this herd should be further investigated.

The interpretation of FECRT results was affected by the method of calculation used to estimate treatment efficacy. Methods including the FEC of untreated animals estimated a higher drug efficacy when the FEC of control calves increased between Day 0 and 14 (farms #3, #4, #5 and #6) compared with methods not including FEC of CTL groups. Nevertheless, similar drug efficacies were detected by all methods when the FEC of CTL animals slightly decreased between samplings (farms #1 and #2). Inclusion of untreated controls in a FECRT has been suggested to allow the detection of such fluctuations in FEC not related with the treatment, which could affect the interpretation of the test (Coles et al., 1992; Lyndal-Murphy et al., 2014). Yet, dynamics of FEC excretion in untreated cattle are not directly comparable with the egg excretion in treated animals due to density-dependent control of female fecundity and poor correlation between worm burdens and FEC. The FEC

of untreated animals may for example be reduced in response to new infections, and FEC of treated animals may increase the egg excretion of females due to lower worm burden and competition (Michel, 1967; 1969; Dobson et al., 2012). Moreover, when ≤ 20 infected animals are available for FECRT in a farm (as observed in most farms in our study) the inclusion of untreated controls will reduce the size of the treatment group and consequently increase uncertainty of the estimated efficacy (Denwood et al., 2010). Therefore, our results suggest that under Danish conditions inclusion of untreated cattle in the FECRT is not recommended. Instead, it may be advisable to include more animals in the treatment group ($n \geq 15$), which is likely to increase the certainty of the estimated treatment efficacy (Denwood et al., 2010; Levecke et al., 2012).

Similarly to the observations at the national level, the study of the anti-parasitic prescriptions in the six farms between 2002 and 2012 showed that 98% of all treatments were ML, mostly IVM pour-on products, regardless of age group. However, we did not aim to directly correlate anthelmintic use and development of AR to IVM given the extensive use of this drug in Danish cattle since the 1990's. Nevertheless, the widespread selection of IVM as the drug of choice in most of the farms where a reduced efficacy was confirmed in the FECRT needs close monitoring to prevent further deterioration of the situation and to identify drugs that are still effective in these farms. Evidence from research on small ruminant nematodes suggest that treatment of selected animals in a group (TST approach), leaving some (or most) of the animals untreated to guarantee a *refugia* of unselected worms, is perhaps the most important strategy to slow the development of AR in nematode populations. Still, it is critical that the anthelmintic used in a TST approach is highly efficacious (Knox et al., 2012). This is also particularly relevant under Danish regulations of prescription-only use of anthelmintics, in which therapeutic treatments should have a high efficacy to cure clinically-affected animals.

In the simulation study, highly aggregated FEC data were poorly analysed by the WAAVP and EggCounts methods, which were unable to optimally include the simulated FECR% in the 95% CI provided by the methods. This was further reflected in the higher frequency of “false negatives” (i.e. declaring efficacious treatment when the simulated FECR = 85%) provided by the EggCounts and the WAAVP methods with highly aggregated simulated FEC data, suggesting that drug efficacy estimates calculated by these two methods with over-dispersed FECR data should be interpreted with caution. The poor coverage probability of 95% CI by the EggCounts derives from the distribution priors, which exclude the possibility of extremely high over-dispersion. In contrast, at low and moderate FEC aggregations, the EggCounts provided the higher rate of “true positive” (i.e. declaring reduced efficacy for simulated FECR = 85) and “true negative” (i.e. declaring efficacious treatment for simulate FECR = 97%) results for the simulated sample size values, followed by the WAAVP and the Bayescount. The Bayescount method, on the other hand, provided no false negative results despite varying FEC aggregation levels, but it provided the highest

rate of inconclusive results in comparison with the other two methods. This is a limitation of the Bayescount as a sample size of 10 animals was too small to provide conclusive results and to correctly distinguish between a FECR of 85% and 97% in the presence of extreme over-dispersion. Given the difficulties associated with the EggCounts method when FEC are highly over-dispersed, and the Bayescount method when the number of samples is low, we suggest that the use of both Bayesian methods requires the selection of appropriate prior distributions assigned in the models (to accurately include in the model the [over] dispersion parameters of the data) and an increased sample size. Nonetheless, there is an urgent need for standardised international guidelines for the design and evaluation of FECRT in cattle that can be used in different farming contexts and considering the characteristics of FEC excretion in cattle, e.g. low FEC and high aggregation. Furthermore, molecular methods for detection and quantification of resistant alleles are undoubtedly needed to detect AR before it reaches clinical levels and become apparent in tests like the FECRT.

Notwithstanding, reasons other than AR may result in a lower than expected efficacy in the FECRT, such as under dosing (e.g. due to inaccurate estimation of BW) and/or altered drug pharmacokinetics and pharmacodynamics (e.g. due to nutritionally related variations in fat body reserves that may affect the persistent efficacy of ML, erratic absorption of the drug from the site of s.c. injection and/or due to interactions with other co-administered drugs) (González Canga et al., 2008; El-Abdellati et al., 2010; Areskog et al., 2012; De Graef et al., 2013; Areskog et al., 2014). All these factors may impact the correct estimation of drug efficacy and detection of AR, particularly in the dose-limiting species *C. oncophora*. At the moment, the controlled efficacy test (CET) is the gold standard method to confirm drug-resistant nematode strains initially reported by the FECRT. In general, a good correlation exists in cattle between CET and FECRT when $FECR \geq 98\%$ or $< 80\%$, i.e. when the treatment is highly effective or when a high level of resistance is likely to be present, respectively (Mejía et al., 2003; Neves et al., 2014). However, when the FECR is 80 – 95% (as in the present study), the confirmation of AR in the CET has proved more challenging. For example, *C. oncophora* populations declared resistant to the recommended dose of injectable IVM by FECRT in two Swedish cattle farms (using girth tape for weight estimation and with $FECR\% \text{ [upper CI]} = 78\% \text{ [97\%]}$ and $79\% \text{ [98\%]}$ in each farm; Demeler et al., 2009) were declared IVM-susceptible when tested in calves under controlled conditions and weighed using scales (Areskog et al., 2014). Due to the absence of weight scales in all of our investigated farms, the BW of the animals was estimated based on girth measurements. Therefore, the reduced efficacy declared by FECRT in the present study may not be related to phenotypic resistance but could have originated from under dosing or changes in the pharmacokinetics of the drug under field conditions, and therefore our findings require confirmation by CET or at least a repeated FECRT. Nevertheless, the evidence of reduced IVM efficacy in three farms and the widespread use of ML in Denmark emphasises the importance of increased surveillance of anthelmintic efficacy in Danish

cattle and to develop novel control strategies that can lower the reliance on anthelmintics and the selection for AR in cattle nematodes.

3.2 Forage chicory selectively reduces *O. ostertagi* worm burdens in two independent experiments, while *C. oncophora* is unaffected (Manuscript II)

The anthelmintic effects of forage chicory against GI nematodes of cattle were investigated in two independent experiments. In Experiment 1, calves infected with *O. ostertagi* and *C. oncophora* and fed with chicory silage ($\geq 70\%$ chicory in the DM intake) had largely a similar faecal egg excretion pattern throughout the trial compared with infected control calves fed a standard balanced protein/energy diet. At Day 19 p.i., *O. ostertagi* and *C. oncophora* L3 were present at a similar ratio in pooled larval cultures from chicory and control groups; however, on Day 26 and 33 p.i., a markedly lower number of *O. ostertagi* L3 was observed in larval cultures from the chicory-fed calves. Post-mortem nematode recovery revealed that calves fed with chicory silage had significantly lower worm burdens of *O. ostertagi* in the abomasum (arithmetic/geometric mean reduction of 52/60%; $P < 0.01$) in comparison with control animals. In contrast, adult counts of *C. oncophora* in the small intestine were not statistically different between groups ($P = 0.12$). Chicory and control calves had comparable protein and energy intakes throughout the trial and similar daily weight gain until Day 19 p.i. (chicory = 383 [± 116] vs. control = 325 [± 236] g day⁻¹). From Day 20 p.i. until slaughter, chicory and control groups had mean growth rates of 811 [± 59] and 391 [± 132] g day⁻¹, respectively.

In Experiment 2, calves mono-infected with *O. ostertagi* and grazing a pure-chicory sward or ryegrass/clover (controls) had a similar faecal egg excretion until Day 20 p.i. ($P = 0.7$). From Day 22 p.i. a rapid and significant reduction in faecal egg excretion was observed in chicory-fed calves until the end of the trial, and at Day 35 p.i. chicory calves had a 65% lower mean FECDM compared with control animals ($P < 0.01$). Cumulative FECDM during the entire experiment were significantly reduced in the chicory group (mean [\pm S.D.] chicory = 12,870 [$\pm 8,136$] vs. control = 22,260 [$\pm 5,671$] EPG of faecal DM; $P < 0.01$). Post-mortem worm recovery revealed that calves fed pure forage chicory (fresh and silage) had highly significant reductions in the adult worm burdens of *O. ostertagi* (arithmetic/geometric mean reduction of 57/66%; $P < 0.001$) in comparison with control animals. The estimated per capita fecundity of *O. ostertagi* females (FECDM at slaughter/total number of females recovered) was not statistically different between groups ($P = 0.24$). Based on sward measurements, chicory calves had an estimated mean daily intake of 5.7 kg DM calf⁻¹ during the trial, with increasing consumption of chicory silage from Day 16 p.i. onwards; by Day 34 p.i. ~70% of the DM intake in this group was chicory silage. In contrast, control animals had an estimated daily intake of 6.9 kg DM

ryegrass/clover calf¹ during the entire experiment. Calves fed with pure chicory and control animals had growth rates of 374 (± 160) g day⁻¹ and 783 (± 229) g day⁻¹ in CHI₂ and CTL₂ calves, respectively. Thus, close monitoring of the DM available in pure chicory swards grazed by parasitized calves, particularly during periods of rapid reproductive plant growth, seems critical to secure that animal production goals are met.

Our study only considered the direct anthelmintic effects of chicory against experimental infections, while we prevented any reinfection from pasture in Experiment 2, thus excluding the potential reduced infection pressure due to lower number of L3 in chicory pastures as previously reported (Moss and Vlassoff, 1993; Marley et al., 2003). However, with the study designs of Experiment 1 and 2 it is not possible to determine if the reduced *O. ostertagi* counts in chicory-fed animals were a result of reduced worm establishment, due to the expulsion of established adult worms or both. Nevertheless, the similar faecal egg excretion until Day 20 p.i. in chicory and control calves in Experiment 2 suggests that dietary chicory may not prevent the establishment and development of *O. ostertagi* into egg-laying adults; rather, the marked drop in FECDM observed from Day 22 p.i. in chicory-fed calves indicates that dietary chicory may selectively (and rapidly) affect the survival of adult worms. Sesquiterpene lactones were identified in both experiments only in forage chicory (fresh and silage), while CT were not detected in any of the experimental feeds. In Experiment 1, chicory silage had a concentration of 12.3 g total SL kg DM⁻¹, with 11,13-dihydro-8-deoxylactucin (DI-8-DOL) and 11,13-dihydro-lactucopicrin (DI-LCP) as the main SL. In Experiment 2, fresh chicory had a concentration of 22.5 g total SL kg DM⁻¹ and chicory silage of 16.8 g total SL kg DM⁻¹; in fresh chicory, the main SL detected were lactucin (LAC) and 11,13-dihydro-lactucin (DI-LAC), while in chicory silage the predominant SL were DI-LCP and DI-8-DOL. Considering the SL content in fresh/silage chicory, the DM intake per calf in both trials and assuming that dietary SL were not inactivated or metabolised in the forestomach, the concentration of SL reaching the abomasum (assuming an abomasum volume of 7 – 8 L) would roughly correspond to 5.37 mg SL mL⁻¹ (Experiment 1) and 13.25 mg SL mL⁻¹ (Experiment 2) per animal. Undoubtedly, further research is required to directly quantify the concentration of dietary SL reaching different gut compartments, which may influence the anthelmintic activity of SL *in vivo*.

Feeding forage chicory-rich diets resulted in a significant reduction of adult worm burdens of *O. ostertagi* in infected calves in two independent experiments, while no apparent activity was observed against *C. oncophora*. The markedly lower number of *O. ostertagi* L3 detected in chicory calves in the Experiment 1, was likely a result of the reduced worm burden in these animals. This contrasts a previous study reporting that infected steers grazing a mixed chicory/ryegrass pasture (~24% of chicory DM in the field) and animals grazing pure ryegrass had similar numbers of *O. ostertagi* L3 in larval cultures, while worm counts were not reported (Marley et al., 2014). Our results therefore indicate that a higher amount of chicory in the diet ($\geq 70\%$ of chicory DM in the diet) may be necessary to exert

an anthelmintic effect against *O. ostertagi* *in vivo*. Sesquiterpene lactones were identified in fresh chicory and silage, suggesting that these PSM may contribute to the observed anthelmintic effects of chicory. Forage chicory (fresh and ensiled) was readily consumed by calves and we demonstrated that the anthelmintic effects observed were preserved despite ensiling. Essential requirements for any novel approach for parasite control is that farmers are able to access, adopt, adapt and implement these methods to their production systems (Vercruyse and Dorny, 1999; Ketzis et al., 2006). In this regard, the preservation of chicory as silage may be a practical strategy for farmers to use forage chicory as a selective anti-*Ostertagia* feed, independent of seasonal availability of chicory in the field. It is interesting to notice that, in comparison with fresh forage, chicory silage prepared for both experiments (in two consecutive years) contained increased concentrations of DI-LCP and DI-8-DOL. In the *in vitro* studies (Manuscript III), DI-8-DOL was linked with higher anthelmintic potency against *O. ostertagi*. Consequently, further research is needed to clarify how ensiling affects the SL profile and anthelmintic activity of chicory silage in comparison with fresh leaves, as well as the optimal ensiling procedures to preserve the crop's nutritional quality.

3.3 *In vitro* studies of the anthelmintic activity of plant secondary metabolites of forage chicory

3.3.1 *In vitro* studies of the direct anthelmintic effects of purified forage chicory extracts against *Cooperia oncophora*: preliminary investigations

In the preliminary *in vitro* studies using chicory leaves (cv. Spadona) collected in an organic dairy farm, incubation with chicory extracts in the EHA induced a dose-dependent inhibition in the hatching of *C. oncophora* eggs, with more than 95% inhibition at concentrations $\geq 2500 \mu\text{g dry extract mL}^{-1}$ and an EC_{50} of $640 \mu\text{g dry extract mL}^{-1}$ (95% CI = 490 – 830). In the AMIA, after 12 h of incubation in the chicory extract a total inhibition in worm motility was detected at concentrations $> 500 \mu\text{g dry extract mL}^{-1}$ ($\text{EC}_{50} = 190 \mu\text{g dry extract mL}^{-1}$; 95% CI = 170-230), and after 48 h of incubation a complete inhibition of worm motility was observed at concentrations $> 250 \mu\text{g dry extract mL}^{-1}$ ($\text{EC}_{50} = 70 \mu\text{g dry extract mL}^{-1}$; 95% CI = 60-90). Adult worms incubated at 2% DMSO (negative controls) had 100% motility until 48 h of incubation. Chromatographic analyses by HPLC-MS revealed no clear molecular profile and this may be explained, to some extent, by the potential presence of undetectable free SL in this extract, in comparison with unbound (“released”) SL detected in extracts used in Manuscript III and isolated with a different procedure. These direct anthelmintic effects contrast the lack of activity of forage chicory towards *C. oncophora* adults observed in Experiment 1 (Manuscript II), and could reflect that active compounds present in forage chicory do not reach the small intestine in concentrations sufficient to exert their activity against *C. oncophora*.

3.3.2 Sesquiterpene lactone-containing extracts from two chicory cultivars show distinct chemical profiles and direct anthelmintic effects against *O. ostertagi* L1 and adult worms (Manuscript III)

Chemical analyses by HPLC-MS of chicory Spadona and Puna II extracts confirmed the presence of the three well-known guaianolide SL of chicory: lactucin (LAC), 8-deoxylactucin (8-DOL) and lactucopicrin (LCP). In addition, the 11, 13-dihydro (DI) derivatives of the mentioned SL were also detected: DI-LAC, DI-8-DOL and DI-LCP. Between 60 – 65% of the tested extracts was composed of SL, while the remaining (unknown) compounds could not be determined. Comparable levels of total and individual SL and unknown compounds were detected in both extracts, with the exception of DI-LAC and DI-8-DOL which were present in significantly higher concentrations in Puna II and Spadona extracts, respectively.

In the LFIA, Spadona and Puna II extracts induced a dose-dependent inhibition of larval feeding with increasing concentrations; however, Spadona extract exerted a remarkably higher potency than Puna II-extract when tested at equal concentrations, illustrated by significantly lower EC_{50} values ($P < 0.0001$). In the LEIA, no differences were observed in exsheathment rates of *O. ostertagi* L3 exposed to SL-containing extracts (at all tested concentrations) or negative controls ($P > 0.05$). In the AMIA, both SL-containing extracts exerted a rapid and dose-dependent paralysis of *O. ostertagi* adults with increasing concentrations. After only 6 h of incubation with the highest extract concentrations, Spadona and Puna II extracts induced worm paralysis in 100 and 92% of the incubated worms, respectively. However, at lower concentrations Spadona extract showed a higher potency reflected in significantly lower EC_{50} values after 6, 24 and 48 h of incubation, in comparison with Puna II extracts ($P < 0.0001$). Adult worms exposed to 1000 μg dry Spadona or Puna II extract mL^{-1} for 24 h (dead) and examined by SEM showed no obvious structural damage in the buccal opening or cuticle and appeared similar to worms incubated with 1% DMSO (live negative controls) (Fig. 3.2). These findings suggest that the mode of anthelmintic action of SL may be different from that of CT or cysteine proteinases, which are known to induce marked disruptions in the cuticle of exposed worms. Based on results from the *in vitro* assays, our data suggest that the increased concentration of DI-8-DOL in Spadona-extract (3.6-fold increase) coincides, to some extent, with the increased potency and lower EC_{50} values exerted by Spadona-extract in the LFIA (3.3-fold lower EC_{50}) and in the AMIA (2.6-fold lower EC_{50}), in comparison with Puna II-extract. These findings suggest that DI-8-DOL is a likely candidate to explain the higher anthelmintic effects of the Spadona-extract and this molecule has not been previously described to have an anthelmintic effect. Nevertheless, the higher potency of Spadona-extract may also have been related with a synergistic effect of all the compounds present in the extract rather than being caused by a single molecule. As purified DI-8-DOL is not commercially available, subsequent studies were carried out in an attempt to fractionate and isolate DI-8-DOL from

Spadona-extracts and to confirm its anthelmintic activity *in vitro*. Fractionation of purified extracts prepared from chicory cv. Spadona leaves was performed using preparative HPLC, and DI-8-DOL was isolated as pure fractions but at extremely low quantities (<0.5 mg), not sufficient to accurately measure its weight or to perform *in vitro* assays, and therefore additional fractionation work was required. Unfortunately, further work was not possible within the time frame of the present PhD study.

Sesquiterpene lactone-containing extracts from forage chicory inhibited larval feeding of L1 and exerted a lethal effect towards *O. ostertagi* adults, inducing a rapid paralysis, whereas no effects were observed on the exsheathment of L3. We observed substantial differences in the anti-parasitic activity between extracts of two forage chicory cultivars and this may be related with their distinct content of SL. To confirm these findings, further fractionation, isolation and testing of individual SL are needed to identify the most active compound(s).

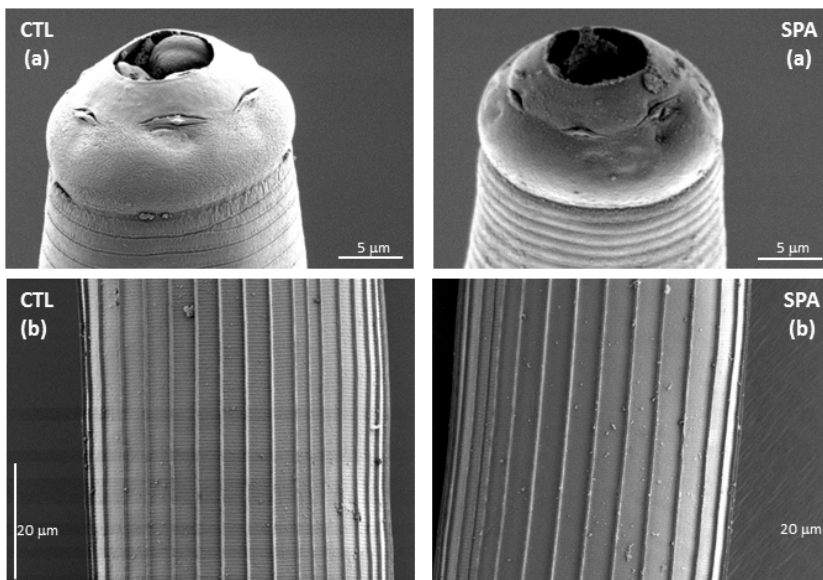


Fig. 3.2. Scanning electron microscopy images of *Ostertagia ostertagi* adults incubated during 24 hours with 1% DMSO in PBS (CTL) or 1000 μg Spadona extract/mL in PBS (SPA). Note the intact mouth opening and circular striations close to the mouth opening (a) and intact prominent longitudinal ridges (b) in all worms.

Chapter 4: Conclusions

The constant risk posed by GI parasites in grazing cattle, the development of AR in bovine nematodes and the continuous pressure to reduce the use of synthetic chemicals in food animals require integrated approaches that combine the field monitoring of AR and the use of novel control methods to safeguard animal health and productivity in a sustainable manner. In the present investigations, a range of field, controlled and laboratory studies were performed to advance our knowledge on the efficacy of the most common anthelmintic drug used in Danish cattle, the potential use of forage chicory as a complementary control of GI nematodes and the role of SL in the anthelmintic activity of chicory.

Field studies (Manuscript I) revealed a reduced efficacy of injectable IVM to control GI nematodes in FSG calves in three of six Danish farms investigated. *Cooperia oncophora* was the main species surviving IVM treatment in all farms, but *O. ostertagi* was also identified post-treatment in one farm with confirmed reduced treatment efficacy. Yet, due to the low number of farms enrolled in the FECRT, this study may not represent the general situation in Denmark. Moreover, the presence of IVM-resistant nematodes suggested by the FECRT in three farms with reduced treatment efficacy should preferably be confirmed by CET. High FEC aggregation markedly affects the performance of the statistical methods used to estimate drug efficacy and results provided by such methods for highly over-dispersed datasets and with low sample size ($n \leq 10$) should be interpreted with caution and standardised guidelines for the design and evaluation of FECRT in cattle are pressingly needed. More than 98% of all anthelmintic treatments performed in the six farms investigated during the 11 years prior to our FECRT were ML, mostly IVM pour-on products. Accordingly, preliminary studies revealed that ML were the predominant anthelmintic drugs in Danish cattle production during 2010 to 2014 (unpublished data). The reduced efficacy of IVM detected in this study and the widespread use of ML in Danish cattle, suggest that farmers and their advisors should be aware of potential lack of efficacy and preferably closely monitor the efficacy of their anthelmintic treatments. Also other initiatives should be implemented in order to slow-down the selection of resistance to IVM and other ML e.g., use of other drug groups or targeted selective treatments with the still effective compounds. In addition, novel parasite control methods are required to reduce the reliance on anthelmintics while decreasing parasite burdens in grazing cattle.

Two independent controlled experiments (Manuscript II) demonstrated that calves fed with $\geq 70\%$ of chicory DM in the diet had significant reductions in the adult worm burden of the pathogenic *O. ostertagi* in the abomasum (60 – 66% reduction), while *C. oncophora* in the small intestine was unaffected. Forage chicory (fresh and silage) was readily consumed by calves and the reduced burden of *O. ostertagi* in these animals was also

associated with a decrease in FEC from Day 22 p.i. onwards, which suggests that dietary chicory selectively affects the adult worms but not the preceding larval stages. Stabled animals fed chicory silage had higher weight gains compared with controls. In contrast calves allocated to a pure chicory pasture in its second year showed lower growth rates due to poor regrowth and low nutritional quality of the chicory crop. Therefore, grazing of calves on pure chicory under Danish conditions seems to require careful planning and monitoring to secure acceptable production gains. Sesquiterpene lactones were identified only in fresh chicory and silage and these PSM may contribute to the observed anthelmintic effects. In addition, the anthelmintic effects of chicory were preserved despite ensiling, and this may be a practical way to conserve and use chicory in farms, independent of season and pasture availability. Pelleting may be another option, which, however, was not tested in the current studies. Furthermore, chicory is well-suited for cultivation in Denmark and other Northern temperate climates and thus show promise as anti-*Ostertagia* feed for use in cattle that may reduce infection levels and the need of drug treatments.

The observed *in vivo* effects of forage chicory against *O. ostertagi* were confirmed by *in vitro* studies. These studies (Manuscript III), demonstrated direct anthelmintic effects of SL-containing extracts from two chicory cultivars against *O. ostertagi* L1 and adults, but not towards L3. Interestingly, chicory extracts from the tested cultivars showed distinctly different potencies and SL profiles; the more potent extract isolated from chicory Spadona had a significantly higher concentration of only one compound, DI-8-DOL. Whether the increased activity of Spadona extract is related only with the higher concentration of DI-8-DOL or to the synergistic effect of all SL in chicory Spadona is not clear. In addition, other molecules with anti-parasitic activity and not detected by chromatographic analyses (i.e. due to poor absorption of UV light) may have been present in the chicory extracts. Moreover, the differences in potency between chicory cultivars *in vitro* should be confirmed *in vivo*. In this context, our results can help to target the responsible compound(s) and to select cultivars with higher anthelmintic activity for cattle and potentially other herbivores.

Chapter 5: Perspectives and future research

Current and future changes in the management of cattle production systems (e.g. due to economic pressure or legislative changes) can have profound impact on the infection dynamics of GI nematodes and the required control interventions. Furthermore, the effects of climate change are expected to affect parasite biology, grassland growth, length of the grazing season and animal husbandry management in very specific and complex interplays that will require a sound understanding of parasite epidemiology and farming practices to maintain optimal parasite control (Morgan and Wall, 2009; van Dijk et al., 2010; Phelan et al., 2016). In Denmark, grazing is usually associated with lower profits and milk production (compared to indoor, loose-housing systems), and despite the lack of official statistics, educated guesses indicate that approximately 35% of all Danish cattle farms still apply grazing in one or more age groups (Reijs et al., 2013). Most of these are organic farms, which by law must provide cattle with access to pasture from mid-April until November (Naturerhvervstyrelsen, 2014). Organic farms are encouraged to apply other means of parasite control than (preventive) use of veterinary drugs. However, due to lack of documentation of other effective parasite control methods, most farms, whether organic or not, still rely on commercial anti-parasitic drugs.

Our results revealed a lower than expected efficacy of injectable IVM confirmed in three of six studied cattle farms, while inconclusive results were observed in the other three farms. Although our results cannot be directly interpreted as cases of IVM-resistance in cattle nematodes and may not reflect the IVM efficacy at the country level, they should stir up further AR surveys in Denmark. This is particularly relevant considering the widespread use of ML in Danish cattle. In this context, it seems relevant to reduce the selection pressure for AR on the still effective anthelmintics, but first those effective drugs should be identified. The selection of suitable cattle farms for the performance of the FECRT proved to be challenging. At the same time, standardise methods for the design and evaluation of FECRT in cattle are required. Once such guidelines are available, the monitoring of the treatment efficacies on a particular farm may be combined with the treatments routinely performed by veterinarians.

One potential alternative to anthelmintics for the control of GI nematodes explored in the present investigations, was the use of forage chicory-rich diets. The selective activity of forage chicory against *O. ostertagi*, the most pathogenic cattle nematode in Denmark and other temperate regions, observed in our controlled experiments is encouraging. However, several issues still need to be investigated before the use of forage chicory can be recommended for on-farm use (see future research below). Grazing cattle of all ages acquire infections with mixed species in the field (although older animals are predominantly infected with *O. ostertagi*), and therefore, it is expected that not all GI nematodes will be

affected by chicory feeding, as observed with *C. oncophora* in our study. Moreover, the economic benefit of including chicory-rich diets in production systems, in terms of animal growth/milk production as a consequence of its anthelmintic effects, needs to be assessed and compared with the extra costs associated with the production and use of chicory. The benefits are expected to vary largely according to different management systems (e.g. availability of [extra] land for cultivation of pure chicory, length of the grazing season and exposure to GI nematodes, etc.) and perhaps age groups (calves, heifers or adult cattle). However, forage chicory, as any other nutraceutical or bioactive forage, is first a feed and therefore both its nutritional and anthelmintic value should be evaluated. In Denmark, chicory is increasingly popular as a component of mixed pastures for cattle grazing (Smidt and Brimer, 2005; Pirhofer-Walzl et al., 2011), although usually sown at a very low rate which may be related to the risk of off-flavour in milk. Our findings indicate that levels of 70% chicory DM in the diet will exert an anthelmintic effect but the lower effective intake is currently not established. Whether sufficiently high concentrations of chicory can be achieved in Danish cattle systems needs further research, but the evaluation of pure swards of chicory for cattle grazing is progressing in other countries (Chapman et al., 2008; Parish et al., 2012; Muir et al., 2014). Moreover, the potential use of chicory silage, which seems to preserve the anthelmintic activity of chicory, should be further explored as a delivery strategy that could be appealing for farmers, even as supplementary feeding for grazing cattle.

Results of the *in vitro* studies testing SL-containing extracts from chicory suggest that different chicory cultivars may have distinct anthelmintic potencies, which may be linked to their particular SL profile, but this remain to be tested *in vivo*. Interestingly, the higher anthelmintic activity of the chicory Spadona extract (rich in DI-8-DOL) compared with the chicory Puna extract against *O. ostertagi* has recently been confirmed by our group against pig nematodes (Williams et al., 2016), suggesting that the molecules in this particular extract have a broad-spectrum anthelmintic activity that warrants further research. It suggests also that the lack of effect against *C. oncophora* may perhaps relate to pharmacokinetics and not pharmacodynamics.

Based on the findings of this PhD thesis, there is a need to further investigate the presence of AR in cattle nematodes in Denmark and to advance our knowledge on the anthelmintic activity of forage chicory and its bioactive compounds. This could be done by:

- Examining the field efficacy of different anthelmintics against GI nematodes infecting Danish cattle and the extent of AR in cattle nematodes through field studies including a larger number of farms across Denmark. However, this requires the development of standardised guidelines for the performance and analysis of FECRT in cattle, which are currently not available.

- Defining the life stage(s) of *O. ostertagi* affected by forage chicory diets and the length of chicory feeding required to exert an anti-*Ostertagia* activity in infected cattle. More specifically, the effects of short-term feeding with pure chicory (e.g. 1, 2 or 5 days) could be studied in animals infected with *O. ostertagi* and slaughtered at different time points to detect the numbers of larval and adult stages. This may allow detecting if chicory primarily exerts its effect by preventing the establishment of *O. ostertagi* L3 or by eliminating established adult worm populations. In addition, the effect of forage chicory on the total egg output of *O. ostertagi*, not only on FEC/FECDM, should be addressed by studying the total faecal output of chicory-fed animals. Moreover, to determine the full potential of chicory as an anthelmintic forage for cattle there is a need to address whether chicory can affect other abomasal nematodes of cattle (e.g. *Haemonchus placei*, *Trichostrongylus axei*).
- Studying whether the ensiling affects the presence of certain SL and/or the anthelmintic potency of forage chicory by comparing the SL profile and *in vivo* anthelmintic activity of fresh and ensiled chicory. Further, there is a need to examine if chicory can be preserved as pellets or by other means of conservation. Before recommending the use of forage chicory as an anthelmintic strategy in cattle farms, the effect of chicory diets against GI nematode should be studied in the field using animals naturally infected with mixed species, and the cost-effectiveness of including chicory in the production system should be addressed. Finally, it would be interesting to explore whether infected cattle voluntarily graze/feed on chicory as a self-medication strategy.
- With the aim to elucidate the role of SL in the anthelmintic activity of chicory and whether SL can reach the abomasum and other compartments of the GI tract to exert their anthelmintic activity, the pharmacokinetics and pharmacodynamics of SL from chicory should be explored. This can be achieved by detecting SL in the feed and in different body compartments and fluids (e.g. plasma, rumen, abomasum, small intestine and faeces) of animals fed with chicory. The molecular mechanisms of the direct activity of SL and their dihydro-derivatives against parasitic nematodes are not known and should be elucidated by exploring the potential inhibitory effects on nematode vital processes, like enzymatic pathways. Furthermore, it is needed to scrutinize if anthelmintic compounds other than SL are present in chicory by studying the nematocidal activity of different fractions from chicory extracts and by describing their components.
- Finally, the potential adaptation of GI nematodes to the activity of SL and other PSM from chicory should be further investigated e.g. by exposing several nematode generations to sub-therapeutic dose levels, leading to a possible “resistance” of nematodes to bioactive plant compounds, or by comparing the activity of chicory PSM against different nematode strains with potentially distinct susceptibilities to bioactive compounds, as earlier described (Calderón-Quintal et al., 2010).

Chapter 6: References

- Agneessens, J., Claerebout, E., Dorny, P., Borgsteede, F.H., Vercruysse, J., 2000. Nematode parasitism in adult dairy cows in Belgium. *Vet. Parasitol.* 90, 83–92.
- Amorim, M.H.R., Gil da Costa, R.M., Lopes, C., Bastos, M.M.S.M., 2013. Sesquiterpene lactones: Adverse health effects and toxicity mechanisms. *Crit. Rev. Toxicol.* 43, 559–579.
- Anderson, R.C., 2000. Chapter 3. Order Strongylida (the Bursate Nematodes). In: *Nematode parasites of vertebrates: their development and transmission*. CABI Publishing, pp. 41– 229.
- Areskog, M., Ljungström, B., Höglund, J., 2013. Limited efficacy of pour-on anthelmintic treatment of cattle under Swedish field conditions. *Int. J. Parasitol. Drugs Drug Resist.* 3, 129–134.
- Areskog, M., Sollenberg, S., Engström, A., von Samson-Himmelstjerna, G., Höglund, J., 2014. A controlled study on gastrointestinal nematodes from two Swedish cattle farms showing field evidence of ivermectin resistance. *Parasit. Vectors* 7, 13.
- Areskog, M., Von Samson-Himmelstjerna, G., Alvinerie, M., Sutra, J.F., Höglund, J., 2012. Dexamethasone treatment interferes with the pharmacokinetics of ivermectin in young cattle. *Vet. Parasitol.* 190, 482–488.
- Armour, J., 1989. The influence of host immunity on the epidemiology of trichostrongyle infections in cattle. *Vet. Parasitol.* 32, 5–19.
- Armour, J., Bairden, K., Holmes, H., Parkins, J.J., Ploeger, H., Salman, S.K., 1987. Pathophysiological and parasitological studies on *Cooperia oncophora* infections in calves. *Res. Vet. Sci.* 42, 373–381.
- Athanasiadou, S., Githiori, J., Kyriazakis, I., 2007a. Medicinal plants for helminth parasite control: facts and fiction. *Animal* 1, 1392–400.
- Athanasiadou, S., Gray, D., Younie, D., Tzamaloukas, O., Jackson, F., Kyriazakis, I., 2007b. The use of chicory for parasite control in organic ewes and their lambs. *Parasitology* 134, 299–307.
- Athanasiadou, S., Kyriazakis, I., 2007. Plant secondary metabolites: antiparasitic effects and their role in ruminant production systems. *Proc. Nutr. Soc.* 63, 631–639.
- Athanasiadou, S., Tzamaloukas, O., Kyriazakis, I., Jackson, F., Coop, R.L., 2005. Testing for direct anthelmintic effects of bioactive forages against *Trichostrongylus colubriformis* in grazing sheep. *Vet. Parasitol.* 127, 233–243.
- Barrera, P., Sülsen, V.P., Lozano, E., Rivera, M., Beer, M.F., Tonn, C., Martino, V.S., Sosa, M. A., 2013. Natural sesquiterpene lactones induce oxidative stress in *Leishmania mexicana*. *Evid. Based. Complement. Alternat. Med.* 2013, 163404.
- Barry, T., 1998. The feeding value of chicory (*Cichorium intybus*) for ruminant livestock. *J. Agric. Sci.* 131, 251–257.

- Barry, T., McNeill, D., McNabb, W., 2001. Plant secondary compounds; their impact on forage nutritive value and upon animal production. Proc. XIX Int. Grassl. Congr. 445–452.
- Bennema, S.C., Vercruyse, J., Morgan, E., Stafford, K., Höglund, J., Demeler, J., von Samson-Himmelstjerna, G., Charlier, J., 2010. Epidemiology and risk factors for exposure to gastrointestinal nematodes in dairy herds in northwestern Europe. Vet. Parasitol. 173, 247–54.
- Bischoff, T., Kelley, C.J., Karchesy, Y., Laurantos, M., Nguyen-Dinh, P., Arefi, A.G., 2004. Antimalarial activity of lactucin and lactucopicrin: sesquiterpene lactones isolated from *Cichorium intybus* L. J. Ethnopharmacol. 95, 455–457.
- Borgsteede, F.H.M., Tibben, J., Cornelissen, J.B.W.J., Agneessens, J., Gaasenbeek, C.P.H., 2000. Nematode parasites of adult dairy cattle in the Netherlands. Vet. Parasitol. 89, 287–296.
- Burow, E., Thomsen, P.T., Sørensen, J.T., Rousing, T., 2011. The effect of grazing on cow mortality in Danish dairy herds. Prev. Vet. Med. 100, 237–241.
- Burow, E., Rousing, T., Thomsen, P.T., Otten, N.D., Sørensen, J.T., 2013. Effect of grazing on the cow welfare of dairy herds evaluated by a multidimensional welfare index. Animal 7, 834–842.
- Burrows, G., Tyril, R.J., 2004. Sesquiterpene lactones. In: Plumlee, K. (Ed.), Clinical Veterinary Toxicology. Mosby, Inc., USA, pp. 416–418.
- Calderón-Quintal, J., Torres- Acosta, J., Sandoval-Castro, C., Alonso, M., Hoste, H., Aguilar-Caballero, A., 2010. Adaptation of *Haemonchus contortus* to condensed tannins: can it be possible? Arch. Med. Vet. 42, 165–171.
- Calvete, C., Uriarte, J., 2013. Improving the detection of anthelmintic resistance: evaluation of faecal egg count reduction test procedures suitable for farm routines. Vet. Parasitol. 196, 438–452.
- Cavin, C., Delannoy, M., Malnoe, a, Debeffe, E., Touché, a, Courtois, D., Schilter, B., 2005. Inhibition of the expression and activity of cyclooxygenase-2 by chicory extract. Biochem. Biophys. Res. Commun. 327, 742–749.
- Chadwick, M., Trewin, H., Gawthrop, F., Wagstaff, C., 2013. Sesquiterpenoids lactones: benefits to plants and people. Int. J. Mol. Sci. 14, 12780–12805.
- Chapman, D., Tharmaraj, J., Nie, Z., 2008. Milk-production potential of different sward types in a temperate southern Australian environment. Grass Forage Sci. 63, 221–233.
- Charlier, J., Höglund, J., von Samson-Himmelstjerna, G., Dorny, P., Vercruyse, J., 2009. Gastrointestinal nematode infections in adult dairy cattle: Impact on production, diagnosis and control. Vet Parasitol 164, 70–79.
- Charlier, J., van der Voort, M., Kenyon, F., Skuce, P., Vercruyse, J., 2014. Chasing helminths and their economic impact on farmed ruminants. Trends Parasitol. 30, 361–367.
- Claerebout, E., Vercruyse, J., 2000. The immune response and the evaluation of acquired immunity against gastrointestinal nematodes in cattle: a review. Parasitology 120, 25–42.

- Clark, D., Anderson, C., Hongwen, G., 1990. Liveweight gain and intake of Friesian bulls grazing "Grasslands Puna" chicory (*Cichorium intybus* L.) or pasture. *New Zeal. J. Agric. Res.* 33, 219–224.
- Coles, G., Bauer, C., Borgsteede, F., Geerts, S., Klei, T., Taylor, M., Waller, P., 1992. World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Vet. Parasitol.* 44, 35–44.
- Coles, G., Jackson, F., Pomroy, W., Prichard, R., von Samson-Himmelstjerna, G., Silvestre, A., Taylor, M., Vercruyse, J., 2006. The detection of anthelmintic resistance in nematodes of veterinary importance. *Vet. Parasitol.* 136, 167–185.
- Coles, G.C., 2002. Cattle nematodes resistant to anthelmintics: why so few cases? *Vet. Res.* 33, 481–489.
- Coop, R.L., Kyriazakis, I., 2001. Influence of host nutrition on the development and consequences of nematode parasitism in ruminants. *Trends Parasitol.* 17, 325–330.
- Coop, R.L., Sykes, A.R., Angus, K.W., 1979. The pathogenicity of daily intakes of *Cooperia oncophora* larvae in growing calves. *Vet. Parasitol.* 5, 261–269.
- Corwin, R.M., 1997. Economics of gastrointestinal parasitism of cattle. *Vet. Parasitol.* 72, 451–460.
- Cully, D.F., Vassilatis, D.K., Liu, K.K., Paress, P.S., Van der Ploeg, L.H., Schaeffer, J.M., Arena, J.P., 1994. Cloning of an avermectin-sensitive glutamate-gated chloride channel from *Caenorhabditis elegans*. *Nature* 371, 707–711.
- Danmap, 2012. The Danish integrated antimicrobial resistance monitoring and research programme. Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark. Copenhagen, Denmark. ISSN 1600-2032.
- De Graef, J., Claerebout, E., Geldhof, P., 2013. Anthelmintic resistance of gastrointestinal cattle nematodes. *Vlaams Diergeneesk. Tijdschr.* 82, 113–123.
- Demeler, J., Kleinschmidt, N., Küttler, U., Koopmann, R., von Samson-Himmelstjerna, G., 2012. Evaluation of the Egg Hatch Assay and the Larval Migration Inhibition Assay to detect anthelmintic resistance in cattle parasitic nematodes on farms. *Parasitol. Int.* 61, 614–618.
- Demeler, J., Küttler, U., von Samson-Himmelstjerna, G., 2010. Adaptation and evaluation of three different in vitro tests for the detection of resistance to anthelmintics in gastrointestinal nematodes of cattle. *Vet. Parasitol.* 170, 61–70.
- Demeler, J., Van Zeveren, A.M.J., Kleinschmidt, N., Vercruyse, J., Höglund, J., Koopmann, R., Cabaret, J., Claerebout, E., Areskog, M., von Samson-Himmelstjerna, G., 2009. Monitoring the efficacy of ivermectin and albendazole against gastrointestinal nematodes of cattle in Northern Europe. *Vet Parasitol* 160, 109–115.
- Denwood, M.J., Reid, S.W.J., Love, S., Nielsen, M.K., Matthews, L., McKendrick, I.J., Innocent, G.T., 2010. Comparison of three alternative methods for analysis of equine faecal egg count reduction test data. *Prev. Vet. Med.* 93, 316–323.

- Desrués, O., Fryganas, C., Ropiak, H.M., Mueller-Harvey, I., Enemark, H.L., Thamsborg, S.M., 2016. Impact of chemical structure of flavanol monomers and condensed tannins on in vitro anthelmintic activity against bovine nematodes. *Parasitology* 143, 444–454.
- Di Grigoli, A., Todaro, M., Di Miceli, G., Genna, V., Tornambè, G., Alicata, M.L., Giambalvo, D., Bonanno, A., 2012. Effects of continuous and rotational grazing of different forage species on ewe milk production. *Small Rumin. Res.* 106, S29–S36.
- Dobson, R., Hosking, B., Jacobson, C., Cotter, J., Besier, R., Stein, P., Reid, S., 2012. Preserving new anthelmintics: a simple method for estimating faecal egg count reduction test (FECRT) confidence limits when efficacy and/or nematode aggregation is high. *Vet. Parasitol.* 186, 79–92.
- Edmonds, M.D., Johnson, E.G., Edmonds, J.D., 2010. Anthelmintic resistance of *Ostertagia ostertagi* and *Cooperia oncophora* to macrocyclic lactones in cattle from the western United States. *Vet. Parasitol.* 170, 224–229.
- Eisler, M.C., Lee, M.R.F., Tarlton, J.F., Martin, G.B., Beddington, J., Dungait, J.A.J., Greathead, H., Liu, J., Mathew, S., Miller, H., Misselbrook, T., Murray, P., Vinod, V.K., Van Saun, R., Winter, M., 2014. Steps to sustainable livestock. *Nature* 507, 32–34.
- Egerton, J.R., Eary, C.H., Suhayda, D., 1981. The anthelmintic efficacy of ivermectin in experimentally infected cattle. *Vet. Parasitol.*
- El-Abdellati, A., Charlier, J., Geldhof, P., Levecke, B., Demeler, J., von Samson-Himmelstjerna, G., Claerebout, E., Vercruyse, J., 2010. The use of a simplified faecal egg count reduction test for assessing anthelmintic efficacy on Belgian and German cattle farms. *Vet. Parasitol.* 169, 352–357.
- EMA, 2009. Zolvix (Monepantel). Eur. Med. Agency EMA/464656/2009; EMA/V/C/154.
- Entrocasso, C., McKellar, Q., Parkins, J.J., Bairden, K., Armour, J., Kloosterman, A., 1986. The sequential development of type I and type II ostertagiasis in young cattle with special reference to biochemical and serological changes. *Vet. Parasitol.* 21, 173–188.
- Ernst, M., Chatterton, N.J., Harrison, P.A., 1995. Carbohydrate changes in chicory (*Cichorium intybus* L. var. *foliosum*) during growth and storage. *Sci. Hortic. (Amsterdam)*. 63, 251–261.
- Eysker, M., Ploeger, H.W., 2000. Value of present diagnostic methods for gastrointestinal nematode infections in ruminants. *Parasitology* 120 Suppl, S109–S119.
- FAO, 2002. Improved animal health for poverty reduction and sustainable livelihoods, FAO 153. ed. Animal Production and Health Division FAO Agriculture Department, Rome, Italy.
- FAO, 2010. Challenges and opportunities for carbon sequestration in grassland systems. A technical report on grassland management and climate change mitigation, Integrated Crop Management. Rome, Italy.
- FAO, 2014. Sustainable production intensification and sustainable food systems. Rome, Italy.
- Feroli, F., D’Antuono, L.F., 2012. An update procedure for an effective and simultaneous extraction of sesquiterpene lactones and phenolics from chicory. *Food Chem.* 135, 243–250.

- Feroli, F., Manco, M., D'Antuono, L., 2015. Variation of sesquiterpene lactone and phenolic content in chicory and endive germplasm. *J. Food Compos. Anal.* 39, 77–86.
- Foley, J.A., Ramankutty, N., Brauman, K.A., Cassidy, E.S., Gerber, J.S., Johnston, M., Mueller, N.D., O'Connell, C., Ray, D.K., West, P.C., Balzer, C., Bennett, E.M., Carpenter, S.R., Hill, J., Monfreda, C., Polasky, S., Rockström, J., Sheehan, J., Siebert, S., Tilman, D., Zaks, D.P.M., O'Connell, C., 2011. Solutions for a cultivated planet. *Nature* 478, 337–342.
- Forbes, A.B., Huckle, C.A., Gibb, M.J., Rook, A.J., Nuthall, R., 2000. Evaluation of the effects of nematode parasitism on grazing behaviour, herbage intake and growth in young grazing cattle. *Vet. Parasitol.* 90, 111–118.
- Foster, J., Cassida, K., Turner, K., 2011. In vitro analysis of the anthelmintic activity of forage chicory (*Cichorium intybus* L.) sesquiterpene lactones against a predominantly *Haemonchus contortus* egg population. *Vet. Parasitol.* 180, 298–306.
- Foster, J.G., Clapham, W.M., Belesky, D.P., Labreux, M., Hall, M.H., Sanderson, M.A., 2006. Influence of cultivation site on sesquiterpene lactone composition of forage chicory (*Cichorium intybus* L.). *J. Agric. Food Chem.* 54, 1772–1778.
- Fox, M.T., 1993. Pathophysiology of infection with *Ostertagia ostertagi* in cattle. *Vet. Parasitol.* 46, 143–158.
- Fox, M.T., 1997. Pathophysiology of infection with gastrointestinal nematodes in domestic ruminants: Recent developments. *Vet. Parasitol.* 72, 285–308.
- Fox, M.T., Gerrelli, D., Pitt, S.R., Jacobs, D.E., Gill, M., Simmonds, A.D., 1989. *Ostertagia ostertagi* infection in the calf: effects of a trickle challenge on appetite, digestibility, rate of passage and liveweight gain. *Res. Vet. Sci.* 47, 299–304.
- Frankena, K., 1987. The interaction between *Cooperia* spp. and *Ostertagia* spp. (Nematoda: Trichostrongylidae) in cattle. PhD thesis. Agricultural University of Wageningen, The Netherlands.
- Fraser, T., Cosgrove, G., Thomas, W., Stevens, D., Hickey, M., 1988. Performance of grasslands puna chicory. In: *Proceedings of the New Zealand Grassland Association* 49. pp. 193–196.
- Gasbarre, L.C., 1997. Effects of gastrointestinal nematode infection on the ruminant immune system. *Vet. Parasitol.* 72, 327–337.
- Gasbarre, L.C., 2014. Anthelmintic resistance in cattle nematodes in the US. *Vet. Parasitol.* 204, 3–11.
- Gasbarre, L.C., Leighton, E., Sonstegard, T., 2001. Role of the bovine immune system and genome in resistance to gastrointestinal nematodes. *Vet. Parasitol.* 98, 51–64.
- Gershenzon, J., Dudareva, N., 2007. The function of terpene natural products in the natural world. *Nat. Chem. Biol.* 3, 408–14.
- Geurden, T., Chartier, C., Fanke, J., di Regalbono, A.F., Traversa, D., von Samson-Himmelstjerna, G., Demeler, J., Vanimisetti, H.B., Bartram, D.J., Denwood, M.J., 2015. Anthelmintic resistance to ivermectin and moxidectin in gastrointestinal nematodes of

- cattle in Europe. *Int. J. Parasitol. Drugs Drug Resist.* 5, 163–171.
- Ghantous, A., Gali-Muhtasib, H., Vuorela, H., Saliba, N. a., Darwiche, N., 2010. What made sesquiterpene lactones reach cancer clinical trials? *Drug Discov. Today* 15, 668–678.
- Gibbs, H.C., 1992. The effects of subclinical disease on bovine gastrointestinal nematodiasis. *Compend. Contin. Educ. Pract. Vet.* 14, 669–677.
- Gill, J.H., Lacey, E., 1998. Avermectin/milbemycin resistance in trichostrongyloid nematodes. *Int. J. Parasitol.* 28, 863–877.
- Githiori, J.B., Athanasiadou, S., Thamsborg, S.M., 2006. Use of plants in novel approaches for control of gastrointestinal helminths in livestock with emphasis on small ruminants. *Vet. Parasitol.* 139, 308–320.
- González Canga, A., Sahagún Prieto, A.M., Díez Liébana, M.J., Fernández Martínez, N., Sierra Vega, M., García Vieitez, J.J., 2008. The pharmacokinetics and interactions of ivermectin in humans--a mini-review. *AAPS J.* 10, 42–6.
- Grabber, J.H., Zeller, W.E., Mueller-Harvey, I., 2013. Acetone enhances the direct analysis of procyanidin- and prodelphinidin-based condensed tannins in lotus species by the butanol-HCl-iron assay. *J. Agric. Food Chem.* 61, 2669–2678.
- Grønvold, J., Høgh-Schmidt, K., 1989. Factors influencing rain splash dispersal of infective larvae of *Ostertagia ostertagi* (*Trichostrongylidae*) from cow pats to the surroundings. *Vet. Parasitol.* 31, 57–70.
- Grønvold, J., Nansen, P., Gasbarre, L.C., Christensen, C.M., Larsen, M., Monrad, J., Midtgaard, N., 1992. Development of immunity to *Ostertagia ostertagi* (*Trichostrongylidae: Nematoda*) in pastured young cattle. *Acta Vet. Scand.* 33, 305–316.
- Hawkins, J., 1993. Economic benefits of parasite control in cattle. *Vet Parasitol* 46, 159–173.
- Heckendorn, F., Häring, D.A., Maurer, V., Senn, M., Hertzberg, H., 2007. Individual administration of three tanniferous forage plants to lambs artificially infected with *Haemonchus contortus* and *Cooperia curticei*. *Vet. Parasitol.* 146, 123–134.
- Heimler, D., Isolani, L., Vignolini, P., Romani, A., 2009. Polyphenol content and antiradical activity of *Cichorium intybus* L. from biodynamic and conventional farming. *Food Chem.* 114, 765–770.
- Henriksen, S., Aagard, K., 1975. A simple flotation and McMaster technique. *Nord. Vet. Med.* 28, 392–397.
- Henriksen, S.A., Jørgensen, R.J., Nansen, P., Sejrsen, K., Brolund Larsen, J., Klausen, S., 1976. Ostertagiasis in calves. I. The effect of control measures on infection levels and body weight gains during the grazing season in Denmark. *Vet. Parasitol.* 2, 259–272.
- Herrero, M., Thornton, P.K., Gerber, P., Reid, R.S., 2009. Livestock, livelihoods and the environment: understanding the trade-offs. *Curr. Opin. Environ. Sustain.* 1, 111–120.
- Herrero, M., Gerber, P., Vellinga, T., Garnett, T., Leip, A., Opio, C., Westhoek, H.J., Thornton, P.K., Olesen, J., Hutchings, N., Montgomery, H., Soussana, J.F., Steinfeld, H., McAllister, T., 2011. Livestock and greenhouse gas emissions: The importance of getting the numbers right. *Anim. Feed Sci. Technol.* 166-167, 779–782.

- Hitova, A., Melzig, M., 2014. *Cichorium intybus* L. Zeitschrift für Phyther. 35, 198–202.
- Hoskin, S.O., Barry, T.N., Wilson, P.R., Charleston, W.A.G., Hodgson, J., 1999a. Effects of reducing anthelmintic input upon growth and faecal egg and larval counts in young farmed deer grazing chicory (*Cichorium intybus*) and perennial ryegrass (*Lolium perenne*)/white clover (*Trifolium repens*) pasture. J. Agric. Sci. 132, 335–345.
- Hoskin, S.O., Barry, T.N., Wilson, P.R., Charleston, W.A.G., Kemp, P.D., 1999b. Growth and carcass production of young farmed deer grazing sulla (*Hedysarum coronarium*), chicory (*Cichorium intybus*), or perennial ryegrass (*Lolium perenne*) white clover (*Trifolium repens*) pasture in New Zealand. New Zeal. J. Agric. Res. 42, 83–92.
- Hoste, H., Jackson, F., Athanasiadou, S., Thamsborg, S.M., Hoskin, S.O., 2006. The effects of tannin-rich plants on parasitic nematodes in ruminants. Trends Parasitol. 22, 253–261.
- Hoste, H., Martinez-Ortiz-De-Montellano, C., Manolaraki, F., Brunet, S., Ojeda-Robertos, N., Fourquaux, I., Torres-Acosta, J.F.J., Sandoval-Castro, C., 2012. Direct and indirect effects of bioactive tannin-rich tropical and temperate legumes against nematode infections. Vet. Parasitol. 186, 18–27.
- Hoste, H., Sotiraki, S., Mejer, H., Heckendorn, F., Maurer, V., Thamsborg, S., 2014. Alternatives to synthetic chemical antiparasitic drugs in organic livestock farming in Europe. In: Bellon, S., Penvern, S. (Eds.), Organic Farming, Prototype for Sustainable Agricultures. Springer Netherlands, Dordrecht, pp. 149–169.
- Hoste, H., Torres-Acosta, J.F.J., 2011. Non chemical control of helminths in ruminants: Adapting solutions for changing worms in a changing world. Vet. Parasitol. 180, 144–154.
- Hoste, H., Torres-Acosta, J.F.J., Sandoval-Castro, C., Mueller-Harvey, I., Sotiraki, S., Louvandini, H., Thamsborg, S.M., Terrill, T.H., 2015. Tannin containing legumes as a model for nutraceuticals against digestive parasites in livestock. Vet. Parasitol. 212, 5–17.
- Høgh-Jensen, H., Nielsen, B., Thamsborg, S.M., 2006. Productivity and quality, competition and facilitation of chicory in ryegrass/legume-based pastures under various nitrogen supply levels. Eur. J. Agron. 24, 247–256.
- Höglund, J., Engström, A., von Samson-Himmelstjerna, G., Demeler, J., Tydén, E., 2013. Real-time PCR detection for quantification of infection levels with *Ostertagia ostertagi* and *Cooperia oncophora* in cattle faeces. Vet. Parasitol. 197, 251–257.
- Ivarsson, E., Liu, H.Y., Dicksved, J., Roos, S., Lindberg, J.E., 2012. Impact of chicory inclusion in a cereal-based diet on digestibility, organ size and faecal microbiota in growing pigs. Animal 6, 1077–1085.
- Jackson, F., Hoste, H., 2010. In vitro screening of plant resources for extra-nutritional attributes in ruminants: nuclear and related methodologies. In: Vercoe, P.E., Makkar, H.P.S., Schlink, A.C. (Eds.), In vitro screening of plant resources for extra-nutritional attributes in ruminants: nuclear and related methodologies. Springer Netherlands, Dordrecht, pp. 25–45.

- Jackson, F.S., McNabb, W.C., Barry, T.N., Foo, Y.L., Peters, J.S., 1996. The condensed tannin content of a range of subtropical and temperate forages and the reactivity of condensed tannin with ribulose-1,5-bis-phosphate carboxylase (Rubisco) protein. *J. Sci. Food Agric.* 72, 483–492.
- Jackson, R., Townsend, K., Pyke, C., Lance, D.M., 1987. Isolation of oxfendazole resistant *Cooperia oncophora* in cattle. *N. Z. Vet. J.* 35, 187–189.
- Jensen, A., Mejer, H., Mølbak, L., Langkjær, M., Jensen, T.K., Angen, Ø., Martinussen, T., Klitgaard, K., Baggesen, D.L., Thamsborg, S.M., Roepstorff, A., 2011. The effect of a diet with fructan-rich chicory roots on intestinal helminths and microbiota with special focus on *Bifidobacteria* and *Campylobacter* in piglets around weaning. *Animal* 5, 851–860.
- Kaminsky, R., Ducray, P., Jung, M., Clover, R., Rufener, L., Bouvier, J., Weber, S.S., Wenger, A., Wieland-Berghausen, S., Goebel, T., Gauvry, N., Pautrat, F., Skripsky, T., Froelich, O., Komoin-Oka, C., Westlund, B., Sluder, A., Mäser, P., 2008. A new class of anthelmintics effective against drug-resistant nematodes. *Nature* 452, 176–180.
- Kanobana, K., Vervelde, L., Van Der Veer, M., Eysker, M., Ploeger, H.W., 2001. Characterization of host responder types after a single *Cooperia oncophora* infection: kinetics of the systemic immune response. *Parasite Immunol.* 23, 641–653.
- Kaplan, R., Vidyashankar, A., 2012. An inconvenient truth: global worming and anthelmintic resistance. *Vet. Parasitol.* 186, 70–78.
- Kaplan, R.M., Vidyashankar, a N., Howell, S.B., Neiss, J.M., Williamson, L.H., Terrill, T.H., 2007. A novel approach for combining the use of in vitro and in vivo data to measure and detect emerging moxidectin resistance in gastrointestinal nematodes of goats. *Int. J. Parasitol.* 37, 795–804.
- Katiki, L.M., Ferreira, J.F.S., Zajac, A.M., Masler, C., Lindsay, D.S., Chagas, A.C.S., Amarante, A.F.T., 2011. *Caenorhabditis elegans* as a model to screen plant extracts and compounds as natural anthelmintics for veterinary use. *Vet. Parasitol.* 182, 264–268.
- Ketzis, J.K., Vercruyse, J., Stromberg, B.E., Larsen, M., Athanasiadou, S., Houdijk, J.G.M., 2006. Evaluation of efficacy expectations for novel and non-chemical helminth control strategies in ruminants. *Vet. Parasitol.* 139, 321–335.
- Kidane, A., Houdijk, J.G.M., Athanasiadou, S., Tolcamp, B.J., Kyriazakis, I., 2010. Effects of maternal protein nutrition and subsequent grazing on chicory (*Cichorium intybus*) on parasitism and performance of lambs. *J. Anim. Sci.* 88, 1513–1521.
- Kidane, A., Sørheim, K., Eik, L.O., Steinshamn, H., 2014. Growth and chemical composition of chicory and performance of lambs grazing chicory relative to grass-clover mixtures. *Acta Agric. Scand. Sect. A — Anim. Sci.* 64, 233–242.
- Kisiel, W., Zielin, K., 2001. Guaianolides from *Cichorium intybus* and structure revision of *Cichorium* sesquiterpene lactones. *Phytochemistry* 57, 523–527.
- Knox, M.R., Besier, R.B., Le Jambre, L.F., Kaplan, R.M., Torres-Acosta, J.F.J., Miller, J., Sutherland, I., 2012. Novel approaches for the control of helminth parasites of livestock VI: Summary of discussions and conclusions. *Vet. Parasitol.* 186, 143–149.

- Kotze, A.C., Hunt, P.W., Skuce, P., von Samson-Himmelstjerna, G., Martin, R.J., Sager, H., Krücken, J., Hodgkinson, J., Lespine, A., Jex, A.R., Gilleard, J.S., Beech, R.N., Wolstenholme, A.J., Demeler, J., Robertson, A.P., Charvet, C.L., Neveu, C., Kaminsky, R., Rufener, L., Alberich, M., Menez, C., Prichard, R.K., 2014. Recent advances in candidate-gene and whole-genome approaches to the discovery of anthelmintic resistance markers and the description of drug/receptor interactions. *Int. J. Parasitol. Drugs Drug Resist.* 4, 164–184.
- Kumarasingha, R., Palombo, E. a, Bhave, M., Yeo, T.C., Lim, D.S.L., Tu, C.L., Shaw, J.M., Boag, P.R., 2014. Enhancing a search for traditional medicinal plants with anthelmintic action by using wild type and stress reporter *Caenorhabditis elegans* strains as screening tools. *Int. J. Parasitol.* 44, 291–298.
- Langworthy, A., Pembleton, K., Rawnsley, R., Harrison, M., Lane, P., Henry, D., Corkrey, R., 2015. Chicory (*Cichorium intybus* L.) can beat the heat during summer drought in southeast Australian dairying regions. In: Proceedings of the 17th ASA Conference. Hobart, Australia.
- Larsson, A., Uggla, A., Waller, P.J., Höglund, J., 2011. Performance of second-season grazing cattle following different levels of parasite control in their first grazing season. *Vet. Parasitol.* 175, 135–140.
- Lee, J.M., Hemmingson, N.R., Minnee, E.M.K., Clark, C.E., 2015. Management strategies for chicory (*Cichorium intybus*) and plantain (*Plantago lanceolata*): impact on dry matter yield, nutritive characteristics and plant density. *Crop Pasture Sci.* 66, 168–133.
- Lee, K., Ibuka, T., Rong-Yang, W., Geissman, T., 1977. Structure-antimicrobial activity relationships among the sesquiterpene lactones and related compounds. *Phytochemistry* 16, 1177–1181.
- Leveck, B., Dobson, R.J., Speybroeck, N., Vercruyse, J., Charlier, J., 2012. Novel insights in the faecal egg count reduction test for monitoring drug efficacy against gastrointestinal nematodes of veterinary importance. *Vet. Parasitol.* 188, 391–396.
- Li, G., Kemp, P., 2005. Forage Chicory (*Cichorium intybus* L.): a review of its agronomy and animal production. *Adv. Agron.* 88, 187–222.
- Lifschitz, A., Virkel, G., Sallovitz, J., Sutra, J.F., Galtier, P., Alvinerie, M., Lanusse, C., 2000. Comparative distribution of ivermectin and doramectin to parasite location tissues in cattle. *Vet. Parasitol.* 87, 327–338.
- Little, P.R., Hodge, A., Maeder, S.J., Wirtherle, N.C., Nicholas, D.R., Cox, G.G., Conder, G.A., 2011. Efficacy of a combined oral formulation of derquantel-abamectin against the adult and larval stages of nematodes in sheep, including anthelmintic-resistant strains. *Vet. Parasitol.* 181, 180–193.
- Liu, H., Ivarsson, E., Dicksved, J., Lundh, T., Lindberg, J.E., 2012. Inclusion of Chicory (*Cichorium intybus* L.) in pigs' diets affects the intestinal microenvironment and the gut microbiota. *Appl. Environ. Microbiol.* 78, 4102–4109.
- Liu, H., Ivarsson, E., Lundh, T., Lindberg, J.E., 2013. Chicory (*Cichorium intybus* L.) and cereals differently affect gut development in broiler chickens and young pigs. *J. Anim.*

Sci. Biotechnol. 4 (1) 50.

- Lombardi, D., Vasseur, E., Berthiaume, R., DeVries, T.J., Bergeron, R., 2015. Feeding preferences and voluntary feed intake of dairy cows: Effect of conservation and harvest time of birdsfoot trefoil and chicory. *J. Dairy Sci.* 98, 7238–7247.
- Lucchin, M., Varotto, S., Barcaccia, G., Parrini, P., 2008. Chicory and Endive. In: Prohens-Tomás, J., Nuez, F. (Eds.), *Vegetables I: Asteraceae, Brassicaceae, Chenopodiaceae and Cucurbitaceae*. Springer New York, pp. 3–48.
- Lyndal-Murphy, M., Rogers, D., Ehrlich, W.K., James, P.J., Pepper, P.M., 2010. Reduced efficacy of macrocyclic lactone treatments in controlling gastrointestinal nematode infections of weaner dairy calves in subtropical eastern Australia. *Vet. Parasitol.* 168, 146–150.
- Lyndal-Murphy, M., Swain, A., Pepper, P., 2014. Methods to determine resistance to anthelmintics when continuing larval development occurs. *Vet. Parasitol.* 199, 191–200.
- Makkar, H.P.S., 2016. Smart livestock feeding strategies for harvesting triple gain – the desired outcomes in planet, people and profit dimensions: a developing country perspective. *Anim. Prod. Sci.* 56, 519–534.
- Mansour, M.M., Dixon, J.B., Rowan, T.G., Carter, S.D., 1992. Modulation of calf immune responses by *Ostertagia ostertagi*: the effect of diet during trickle infection. *Vet. Immunol. Immunopathol.* 33, 261–269.
- Marley, C.L., Cook, R., Barrett, J., Keatinge, R., Lampkin, N.H., 2006. The effects of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) when compared with perennial ryegrass (*Lolium perenne*) on ovine gastrointestinal parasite development, survival and migration. *Vet. Parasitol.* 138, 280–290.
- Marley, C.L., Cook, R., Barrett, J., Keatinge, R., Lampkin, N.H., McBride, S.D., 2003a. The effect of dietary forage on the development and survival of helminth parasites in ovine faeces. *Vet. Parasitol.* 118, 93–107.
- Marley, C.L., Cook, R., Keatinge, R., Barrett, J., Lampkin, N., 2003b. The effect of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) on parasite intensities and performance of lambs naturally infected with helminth parasites. *Vet. Parasitol.* 112, 147–155.
- Marley, C.L., Fychan, R., Davies, J., Scollan, N., Richardson, R., Theobald, V., Genever, E., Forbes, A., Sanderson, R., 2014. Effects of chicory/perennial ryegrass swards compared with perennial ryegrass swards on the performance and carcass quality of grazing beef steers. *PLoS One* 9, e86259.
- McKellar, Q., Duncan, J.L., Armour, J., McWilliam, P., 1986. Response to transplanted adult *Ostertagia ostertagi* in calves. *Res. Vet. Sci.* 40, 29–34.
- McMichael, A.J., Powles, J.W., Butler, C.D., Uauy, R., 2007. Food, livestock production, energy, climate change, and health. *Lancet* 370, 1253–1263.
- Mejía, M., Fernández Igartúa, B., Schmidt, E., Cabaret, J., 2003. Multispecies and multiple anthelmintic resistance on cattle nematodes in a farm in Argentina: the beginning of high resistance? *Vet Res* 34, 461–467.

- Meshnick, S.R., 2002. Artemisinin: mechanisms of action, resistance and toxicity. *Int. J. Parasitol.* 32, 1655–1660.
- Michel, J.F., 1967. Regulation of egg output of populations of *Ostertagia ostertagi*. *Nature* 215, 1001–1002.
- Michel, J.F., 1969. Some observations on the worm burdens of calves infected daily with *Ostertagia ostertagi*. *Parasitology* 59, 575–595.
- Michel, J.F., 1974. Arrested development of nematodes and some related phenomena. *Adv. Parasitol.* 12, 279–366.
- Michel, J.F., Lancaster, M.B., Hong, C., 1978. Arrested development of *Ostertagia ostertagia* and *Cooperia oncophora*. *J. Comp. Path.* 88, 131–136.
- Miller, M.C., Duckett, S.K., Andrae, J.G., 2011. The effect of forage species on performance and gastrointestinal nematode infection in lambs. *Small Rumin. Res.* 95, 188–192.
- Molan, A.L., Duncan, A.J., Barry, T.N., McNabb, W.C., 2003. Effects of condensed tannins and crude sesquiterpene lactones extracted from chicory on the motility of larvae of deer lungworm and gastrointestinal nematodes. *Parasitol. Int.* 52, 209–218.
- Monrad, J., 2010. Roundworm parasites in cattle. In: Thamsborg, S.M., Roepstorff, A., Monrad, J. (Eds.), *Veterinær Parasitologi: Helminnologi for Veterinærstuderende*. Academic Books, Life Sciences, Copenhagen, Chapter 8.
- Morgan, E.R., Wall, R., 2009. Climate change and parasitic disease: farmer mitigation? *Trends Parasitol.* 25, 308–313.
- Moss, R.A., Vlassoff, A., 1993. Effect of herbage species on gastro-intestinal roundworm populations and their distribution. *New Zeal. J. Agric. Res.* 36, 371–375.
- Muir, S., Ward, G., Jacobs, J., 2014. Milk production and composition of mid-lactation cows consuming perennial ryegrass-and chicory-based diets. *J. Dairy Sci.* 97, 1005–1015.
- Muir, S.K., Ward, G.N., Jacobs, J.L., 2015. Herbage intake and milk production of late-lactation dairy cows offered a second-year chicory crop during summer. *J. Dairy Sci.* 98, 8825–8835.
- Nansen, P., 1987. Production losses and control of helminths of temperate regions. *Int. J. Parasitol.* 17, 425–433.
- Nansen, P., 1993. Current and future prospects for control of ostertagiasis in northern Europe — examples from Denmark. *Vet. Parasitol.* 46, 3–21.
- Nansen, P., Foldager, J., Hansen, J.W., Henriksen, S. A, Jørgensen, R.J., 1988. Grazing pressure and acquisition of *Ostertagia ostertagi* in calves. *Vet. Parasitol.* 27, 325–335.
- Nansen, P., Grønvold, J., Jørgensen, R.J., Henriksen, S.A., Foldager, J., Sejrsen, K., 1989. Outbreaks of early-season trichostrongylosis in calves in Denmark. *Vet. Parasitol.* 32, 199–211.
- Nansen, P., Jørgensen, R.J., Hansen, J.W., Sejrsen, K., 1978. Some factors influencing the exposure of grazing cattle to trichostrongyle infection. *Vet. Sci. Commun.* 2, 193–205.
- Nansen, P., Jørgensen, R.J., Henriksen, S.A., Foldager, J., 1987. The effects of late turnout on the epidemiology and control of ostertagiasis in calves. *Vet. Parasitol.* 24, 139–147.
- Nansen, P., Steffan, P.E., Christensen, C.M., Gasbarre, L.C., Monrad, J., Grønvold, J.,

- Henriksen, S.A., 1993. The effect of experimental trichostrongyle infections of housed young calves on the subsequent course of natural infection on pasture. *Int. J. Parasitol.* 23, 627–638.
- Naturerhvervstyrelsen, 2014. Statistik over økologiske jordbrugsbedrifter 2013. Ministeriet for Fødevarer.
- Neves, J.H. das, Carvalho, N., Rinaldi, L., Cringoli, G., Amarante, A.F.T., 2014. Diagnosis of anthelmintic resistance in cattle in Brazil: A comparison of different methodologies. *Vet. Parasitol.* 206, 216–226.
- Nielsen, B.K., Thamsborg, S.M., Hansen, H., Ranvig, H., Høgh-Jensen, H., 2009. Effects of including chicory in perennial ryegrass-white clover leys on production and health in organic lambs. *Livest. Sci.* 125, 66–73.
- Niezen, J., Waghorn, T.S., Raufaut, K., Robertson, H.A., McFarlane, R.G., 1994. Lamb weight gain and faecal egg count when grazing one of seven herbage and dosed with larvae for six weeks. *Proc. New Zeal. Soc. Anim. Prod.* 54, 15–18.
- Niezen, J.H., Charleston, W.A.G., Hodgson, J., Miller, C.M., Waghorn, T.S., Robertson, H.A., 1998. Effect of plant species on the larvae of gastrointestinal nematodes which parasitise sheep. *Int. J. Parasitol.* 28, 791–803.
- Niezen, J.H., Waghorn, G.C., Graham, T., Carter, J.L., Leathwick, D.M., 2002. The effect of diet fed to lambs on subsequent development of *Trichostrongylus colubriformis* larvae in vitro and on pasture. *Vet. Parasitol.* 105, 269–283.
- Novobilský, A., Mueller-Harvey, I., Thamsborg, S.M., 2011. Condensed tannins act against cattle nematodes. *Vet. Parasitol.* 182, 213–220.
- Novobilský, A., Stringano, E., Hayot Carbonero, C., Smith, L., Enemark, H., Mueller-Harvey, I., Thamsborg, S., 2013. In vitro effects of extracts and purified tannins of sainfoin (*Onobrychis viciifolia*) against two cattle nematodes. *Vet. Parasitol.* 196, 532–537.
- O’Grady, J., Kotze, A.C., 2004. *Haemonchus contortus*: in vitro drug screening assays with the adult life stage. *Exp. Parasitol.* 106, 164–172.
- O’Shaughnessy, J., Earley, B., Mee, J.F., Doherty, M.L., Crosson, P., Barrett, D., Prendiville, R., Macrelli, M., de Waal, T., 2014. Detection of anthelmintic resistance on two Irish beef research farms. *Vet. Rec.* 175, doi:10.1136/vr.102556.
- Pandey, V.S., 1972. Effect of temperature on development of the free-living stages of *Ostertagia ostertagi*. *J. Parasitol.* 58, 1037–1041.
- Paolini, V., Fouraste, I., Hoste, H., 2004. In vitro effects of three woody plant and sainfoin extracts on 3rd-stage larvae and adult worms of three gastrointestinal nematodes. *Parasitology* 129, 69–77.
- Parish, J.A., Parish, J.R., Best, T.F., Saunders, J.R., 2012. Comparison of chicory and annual ryegrass for spring stockering of beef steers. *Prof. Anim. Sci.* 28, 579–587.
- Parkins, J.J., Taylor, M., Holmes, P.H., Bairden, K., Salman, S.K., Armour, J., 1990. Pathophysiological and parasitological studies on a concurrent infection of *Ostertagia ostertagi* and *Cooperia oncophora* in calves. *Res. Vet. Sci.* 48, 201–208.

- Peters, A.M., van Amerongen, A., 1997. A study on the effects of sample pre-treatment on the amount of sesquiterpene lactones found in chicory (*Cichorium Intybus* L.) by ELISA and by HPLC. *Zeitschrift für Leb. und Forsch. A* 204, 189–193.
- Petkevicius, S., Knudsen, K.E.B., Murrell, K.D., Wachmann, H., 2003. The effect of inulin and sugar beet fibre on *Oesophagostomum dentatum* infection in pigs. *Parasitology* 127, 61–68.
- Petkevicius, S., Murrell, K.D., Bach Knudsen, K.E., Jørgensen, H., Roepstorff, A., Laue, A., Wachmann, H., 2004. Effects of short-chain fatty acids and lactic acids on survival of *Oesophagostomum dentatum* in pigs. *Vet. Parasitol.* 122, 293–301.
- Phelan, P., Morgan, E.R., Rose, H., Grant, J., O’Kiely, P., 2016. Predictions of future grazing season length for European dairy, beef and sheep farms based on regression with bioclimatic variables. *J. Agric. Sci.* 154, 765–781.
- Pirhofer-Walzl, K., Søgaard, K., Høgh-Jensen, H., Eriksen, J., Sanderson, M., Rasmussen, J., Rasmussen, J., 2011. Forage herbs improve mineral composition of grassland herbage. *Grass Forage Sci.* 66, 415–423.
- Preston, S., Jabbar, A., Nowell, C., Joachim, A., Ruttkowski, B., Baell, J., Cardno, T., Korhonen, P.K., Piedrafitra, D., Ansell, B.R.E., Jex, A.R., Hofmann, A., Gasser, R.B., 2015. Low cost whole-organism screening of compounds for anthelmintic activity. *Int. J. Parasitol.* 45, 333–343.
- Price, K., DuPont, M., Shepherd, R., Chan, H., Fenwick, G., 1990. Relationship between the chemical and sensory properties of exotic salad crops -coloured Lettuce (*Lactuca sativa*) and Chicory (*Cichorium intybus*). *J Sci Food Agric.* 53, 185–192.
- Prichard, R.K., 2002. Resistance against macrocyclic lactones. In: Vercruyse, J., Rew, R.S. (Eds.), *Macrocyclic Lactones in Antiparasitic Therapy*. CABI Publishing, Oxon, UK; New York, USA, pp. 163–182.
- Prichard, R.K., Hall, C.A., Kelly, J.D., Martin, L.A., Donald, A., 1980. The problem of anthelmintic resistance in nematodes. *Aust. Vet. J.* 56, 239–251.
- Ramos, F., Portella, L.P., de Souza Rodrigues, F., Reginato, C.Z., Pötter, L., Cezar, A.S., Sangioni, L.A., Flores Vogel, F.S., 2016. Anthelmintic resistance in gastrointestinal nematodes of beef cattle in the state of Rio Grande do Sul, Brazil. *Int. J. Parasitol. Drugs Drug Resist.* 6, 93–101.
- Rees, S., Harborne, J., 1985. The role of sesquiterpene lactones and phenolics in the chemical defence of the chicory plant. *Phytochemistry* 24, 2225–2231.
- Reijs, J.W., Daatselaar, C.H.G., Helming, J.F.M., Jager, J., Beldman, A.C.G., 2013. *Grazing dairy cows in North-West Europe: Economic farm performance and future developments with emphasis on the Dutch situation*, LEI Wageningen UR, The Hague. The Hague, Netherlands.
- Rinaldi, M., Geldhof, P., 2012. Immunologically based control strategies for ostertagiosis in cattle: Where do we stand? *Parasite Immunol.* 34, 254–264.
- Roberfroid, M.B., 2005. Introducing inulin-type fructans. *Br. J. Nutr.* 93, S13.
- Roca-Fernández, A.I., Peyraud, J.L., Delaby, L., Delagarde, R., 2016. Pasture intake and milk

- production of dairy cows rotationally grazing on multi-species swards. *Animal*, available on CJO2016. doi:10.1017/S1751731116000331.
- Roepstorff, A., Nansen, P., 1998. *Epidemiology, Diagnosis and Control of Helminth Parasites of Swine*. FAO Animal Health Manual, Rome, Italy.
- Rumball, W., 1986. Grasslands Puna' chicory (*Cichorium intybus* L.). *New Zeal. J. Exp. Agric.* 14, 105–107.
- Rumball, W., Keogh, R., Miller, J., Claydon, R., 2003a. "Choice" forage chicory (*Cichorium intybus* L.). *New Zeal J. Agric. Res.* 46, 49–51.
- Rumball, W., Skipp, R., Keogh, R., Claydon, R., 2003b. "Puna II" forage chicory (*Cichorium intybus* L.). *New Zeal. J. Agric. Res.* 46, 53–55.
- Samanta, A.K., Jayapal, N., Senani, S., Kolte, A.P., Sridhar, M., 2013. Prebiotic inulin: Useful dietary adjuncts to manipulate the livestock gut microflora. *Brazilian J. Microbiol.* 44, 1–14.
- Sanderson, M.A., Labreuveux, M., Hall, M.H., Elwinger, G.F., 2003. Nutritive value of chicory and English plantain forage. *Crop Sci.* 43, 1797–1804.
- Sandoval-Castro, C.A., Torres-Acosta, J.F.J., Hoste, H., Salem, A.Z.M., Chan-Pérez, J.I., 2012. Using plant bioactive materials to control gastrointestinal tract helminths in livestock. *Anim. Feed Sci. Technol.* 176, 192–201.
- Sargison, N., Wilson, D., Scott, P., 2009. Relative inefficacy of pour-on macrocyclic lactone anthelmintic treatments against *Cooperia* species in Highland calves. *Vet. Rec.* 164, 603–604.
- Satrija, F., Nansen, P., 1993. Experimental concurrent infections with *Ostertagia ostertagi* and *Cooperia oncophora* in the calf. *Res. Vet. Sci.* 55, 92–97.
- Scales, G.H., Knight, T.L., Saville, D.J., 1995. Effect of herbage species and feeding level on internal parasites and production performance of grazing lambs. *New Zeal. J. Agric. Res.* 38, 237–247.
- Schader, C., Muller, A., Scialabba, N.E.-H., Hecht, J., Isensee, A., Erb, K.-H., Smith, P., Makkar, H.P.S., Klocke, P., Leiber, F., Schwegler, P., Stolze, M., Niggli, U., 2015. Impacts of feeding less food-competing feedstuffs to livestock on global food system sustainability. *J. R. Soc. Interface* 12, 20150891.
- Schmidt, B., Ilic, N., Poulev, A., Raskin, I., 2007. Toxicological evaluation of chicory extract (humans). *Food Chem Toxicol* 45, 1–18.
- Schmidt, T.J., 2006. Structure-activity relationships of sesquiterpene lactones. *Stud. Nat. Prod. Chem.* 33, 309–392.
- Schreurs, N.M., Molan, A.L., Barry, N., McNabb, W.C., 2002. Effects of grazing undrenched weaner deer on chicory or perennial ryegrass/white clover pasture on the viability of gastrointestinal nematodes and lungworms. *Vet Rec* 348–353.
- Seaman, F.C., 1982. Sesquiterpene lactones as taxonomic characters in the Asteraceae. *Bot. Rev.* 48, 121–595.
- Shaw, D.J., Vercruyse, J., Claerebout, E., Dorny, P., 1998. Gastrointestinal nematode infections of first-grazing season calves in Western Europe: Associations between

- parasitological, physiological and physical factors. *Vet. Parasitol.* 75, 133–151.
- Shepley, E., Vasseur, E., Bergeron, R., Villeneuve, A., Lachance, S., 2015. Short communication: Birdsfoot trefoil as a preventative treatment for gastrointestinal nematodes in pastured dairy heifers. *Can. J. Anim. Sci.* 95, 533–537.
- Sievers, G., Nannig, S., 2006. Effect of the supplementary feeding with *Plantago lanceolata* on the egg output of gastrointestinal nematodes in calves. *Arch. Med. Vet.* 38, 233–238.
- Simonsen, H., Weitzel, C., Christensen, S., 2013. Guaianolide sesquiterpenoids: pharmacology and biosynthesis. In: Ramawat, K., Mérillon, J. (Eds.), *Natural Products*. Springer-Verlag Berlin Heidelberg, pp. 3069–3098.
- Simpson, H. V., 2000. Pathophysiology of Abomasal Parasitism: Is the Host or Parasite Responsible? *Vet. J.* 160, 177–191.
- Sinkovič, L., Demšar, L., Žnidarčič, D., Vidrih, R., Hribar, J., Treutter, D., 2015. Phenolic profiles in leaves of chicory cultivars (*Cichorium intybus* L.) as influenced by organic and mineral fertilizers. *Food Chem.* 166, 507–513.
- Slotved, H.C., Barnes, E.H., Bjørn, H., Christensen, C.M., Eriksen, L., Roepstorff, A., Nansen, P., 1996. Recovery of *Oesophagostomum dentatum* from pigs by isolation of parasites migrating from large intestinal contents embedded in agar-gel. *Vet. Parasitol.* 63, 237–245.
- Smidt, N.W., Brimer, L., 2005. The use of herbs in pastures: An interview survey among bio-dynamic and organic farmers with dairy cattle. *Agric. Human Values* 22, 355–363.
- Smith, P., 2014. Do grasslands act as a perpetual sink for carbon? *Glob. Chang. Biol.* 20, 2708–2711.
- Soutello, R.G. V, Seno, M.C.Z., Amarante, A.F.T., 2007. Anthelmintic resistance in cattle nematodes in Sao Paulo State, Brazil. *Vet Parasitol* 148, 360–364.
- Stafford, K., Coles, G.C., 1999. Nematode control practices and anthelmintic resistance in dairy calves in the south west of England. *Vet. Rec.* 144, 659–661.
- Stepek, G., Behnke, J.M., Buttle, D.J., Duce, I.R., 2004. Natural plant cysteine proteinases as anthelmintics? *Trends Parasitol.* 20, 322–327.
- Street, R.A., Sidana, J., Prinsloo, G., 2013. *Cichorium intybus*: Traditional uses, phytochemistry, pharmacology, and toxicology. *Evidence-based Complement. Altern. Med.* 2013, Article ID 579319.
- Sutherland, I.A., Bullen, S.L., 2015. Parasite control in pasture-grazed dairy cattle: Are we at the edge of a precipice? *Anim. Prod. Sci.* 55, 916–921.
- Sutherland, I.A., Leathwick, D.M., 2011. Anthelmintic resistance in nematode parasites of cattle: a global issue? *Trends Parasitol.* 27, 176–181.
- Swanepoel, F., Stroebel, A., Moyo, S., 2010. The role of livestock in developing communities: enhancing multifunctionality. *The Technical Centre for Agricultural and Rural Cooperation (CTA)*.
- Tamaki, H., Robinson, R.W., Anderson, J.L., Stoewsand, G.S., 1995. Sesquiterpene Lactones in Virus-Resistant Lettuce. *J. Agric. Food Chem.* 43, 6–8.

- Taylor, L.M., Parkins, J., Armour, J., Holmes, P., Bairden, K., Ibarra-Silva, A., Salman, S., 1989. Pathophysiological and parasitological studies on *Ostertagia ostertagi* infections in calves. *Res. Vet. Sci.* 46, 218–225.
- Taylor, M., Berry, E., Vickers, M., 2010. Sustainable anthelmintic use in cattle. *Vet. Rec.* 167, 145–146.
- Taylor, M., Hunt, K.R., Goodyear, K.L., 2002. Anthelmintic resistance detection methods. *Vet. Parasitol.* 103, 183–194.
- Taylor, M.A., Coop, R.L., Wall, R.L., 2016. Parasites of cattle. In: Taylor, M.A., Coop, R.L., Wall, R.L. (Eds.), *Veterinary Parasitology*. Wiley & Sons, pp. 352–435.
- Thamsborg, S.M., Jørgensen, R.J., Waller, P.J., Nansen, P., 1996. The influence of stocking rate on gastrointestinal nematode infections of sheep over a 2-year grazing period. *Vet. Parasitol.* 67, 207–224.
- Thamsborg, S.M., Roepstorff, A., Larsen, M., 1999. Integrated and biological control of parasites in organic and conventional production systems. *Vet. Parasitol.* 84, 169–186.
- Thornton, P.K., 2010. Livestock production: recent trends, future prospects. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 365, 2853–2867.
- Torgerson, P.R., Paul, M., Furrer, R., 2014. Evaluating faecal egg count reduction using a specifically designed package “eggCounts” in R and a user friendly web interface. *Int. J. Parasitol.* 44, 299–303.
- Tzamaloukas, O., Athanasiadou, S., Kyriazakis, I., Huntley, J.F., Jackson, F., 2006. The effect of chicory (*Cichorium intybus*) and sulla (*Hedysarum coronarium*) on larval development and mucosal cell responses of growing lambs challenged with *Teladorsagia circumcincta*. *Parasitology* 132, 419–426.
- Tzamaloukas, O., Athanasiadou, S., Kyriazakis, I., Jackson, F., Coop, R.L., 2005. The consequences of short-term grazing of bioactive forages on established adult and incoming larvae populations of *Teladorsagia circumcincta* in lambs. *Int. J. Parasitol.* 35, 329–335.
- van Dijk, J., Sargison, N.D., Kenyon, F., Skuce, P.J., 2010. Climate change and infectious disease: helminthological challenges to farmed ruminants in temperate regions. *Animal* 4, 377–392.
- Van Houtert, M.F.J., Sykes, A.R., 1996. Implications of nutrition for the ability of ruminants to withstand gastrointestinal nematode infections. *Int. J. Parasitol.* 26, 1151–1167.
- Vercruysse, J., Claerebout, E., 2001. Treatment vs non-treatment of helminth infections in cattle: defining the threshold. *Vet. Parasitol.* 98, 195–214.
- Vercruysse, J., Dorny, P., 1999. Integrated control of nematode infections in cattle: A reality? A need? A future? *Int. J. Parasitol.* 29, 165–175.
- Vercruysse, J., Hilderson, H., Claerebout, E., 1994. Effect of chemoprophylaxis on immunity to gastrointestinal nematodes in cattle. *Parasitol. Today* 10, 129–132.
- Vercruysse, J., Rew, R., 2002. Use of macrocyclic lactones to control cattle parasites in Europe. In: Vercruysse, J., Rew, R.S. (Eds.), *Macrocyclic Lactones in Antiparasitic Therapy*. CABI Publishing, Oxon, UK; New York, USA, pp. 223–247.

- Vermuntt, J.J., West, D.M., Pomroy, W.E., 1996. Inefficacy of moxidectin and doramectin against ivermectin-resistant *Cooperia* spp. of cattle in New Zealand. N. Z. Vet. J. 44, 188–193.
- Vlaminck, J., Borloo, J., Vercruyssen, J., Geldhof, P., Claerebout, E., 2015. Vaccination of calves against *Cooperia oncophora* with a double-domain activation-associated secreted protein reduces parasite egg output and pasture contamination. Int. J. Parasitol. 45, 209–213.
- Waghorn, T.S., Leathwick, D.M., Rhodes, A.P., Lawrence, K.E., Jackson, R., Pomroy, W.E., West, D.M., Moffat, J.R., 2006. Prevalence of anthelmintic resistance on 62 beef cattle farms in the North Island of New Zealand. N. Z. Vet. J. 54, 278–282.
- Waller, P., 2006. Sustainable nematode parasite control strategies for ruminant livestock by grazing management and biological control. Anim. Feed Sci. Technol. 126, 277–289.
- Waller, P.J., 1999. International approaches to the concept of integrated control of nematode parasites of livestock. Int. J. Parasitol. 29, 155–164.
- Waller, P.J., Bernes, G., Thamsborg, S.M., Sukura, A., Richter, S.H., Ingebrigtsen, K., Höglund, J., 2001. Plants as de-worming agents of livestock in the Nordic countries: historical perspective, popular beliefs and prospects for the future. Acta Vet. Scand. 42, 31–44.
- Waller, P.J., Thamsborg, S.M., 2004. Nematode control in “green” ruminant production systems. Trends Parasitol. 20, 493–497.
- Wang, Q., Cui, J., 2011. Perspectives and utilization technologies of chicory (*Cichorium intybus* L.): A review. Afrincan J. Biotechnol. 10, 1966–1977.
- Waugh, C., Clark, D., Harris, S., Thom, E., Copeman, P., Napper, A., 1998. Chicory for milk production. In: Proceedings of the New Zealand Grassland Association 60. pp. 33–37.
- Williams, A.R., Peña-Espinoza, M., Boas, U., Simonsen, H.T., Enemark, H.L., Thamsborg, S.M., 2016. Anthelmintic activity of chicory (*Cichorium intybus*): in vitro effects on swine nematodes and relationship to sesquiterpene lactone composition. Parasitology 143, 770–777.
- Williams, J.C., Knox, J.W., Loyacano, A. F., 1993. Epidemiology of *Ostertagia ostertagi* in weaner-yearling cattle. Vet. Parasitol. 46, 313–324.
- Wulfkuehler, S., Gras, C., Carle, R., 2014. Influence of light exposure during storage on the content of sesquiterpene lactones and photosynthetic pigments in witloof chicory (*Cichorium intybus* L. var. *foliosum* Hegi). LWT - Food Sci. Technol. 58, 417–426.
- Xhomfulana, V., Mapiye, C., Chimonyo, M., Marufu, M.C., 2009. Supplements containing *Acacia karroo* foliage reduce nematode burdens in Nguni and crossbred cattle. Anim. Prod. Sci. 49, 646–653.
- Zidorn, C., 2008. Sesquiterpene lactones and their precursors as chemosystematic markers in the tribe Cichorieae of the Asteraceae. Phytochemistry 69, 2270–2296.



Chapter 7: Manuscripts

Manuscript I

Miguel Peña-Espinoza, Stig M. Thamsborg, Matthew J. Denwood, Markus Drag, Tina V. Hansen, Vibeke F. Jensen, Heidi L. Enemark. 2016. Efficacy of ivermectin against gastrointestinal nematodes of cattle in Denmark evaluated by different methods for analysis of faecal egg count reduction. Submitted to International Journal for Parasitology: Drugs and Drug Resistance.

Manuscript II

Miguel Peña-Espinoza, Stig M. Thamsborg, Olivier Desrues, Tina V. A. Hansen, Heidi L. Enemark. 2016. Anthelmintic effects of forage chicory (*Cichorium intybus*) against gastrointestinal nematode parasites in experimentally infected cattle. Parasitology 143, 1279–1293, <http://dx.doi.org/10.1017/S0031182016000706>

Manuscript III

Miguel Peña-Espinoza, Ulrik Boas, Andrew R. Williams, Stig M. Thamsborg, Henrik T. Simonsen, Heidi L. Enemark. 2015. Sesquiterpene lactone containing extracts from two cultivars of forage chicory (*Cichorium intybus*) show distinctive chemical profiles and *in vitro* activity against *Ostertagia ostertagi*. International Journal for Parasitology: Drugs and Drug Resistance 5, 191–200, <http://dx.doi.org/10.1016/j.ijpddr.2015.10.002>

7.1 Manuscript I

Efficacy of ivermectin against gastrointestinal nematodes of cattle in Denmark evaluated by different methods for analysis of faecal egg count reduction

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Efficacy of ivermectin against gastrointestinal nematodes of cattle in Denmark evaluated by different methods for analysis of faecal egg count reduction

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Abstract

The efficacy of ivermectin (IVM) against gastrointestinal nematodes in Danish cattle was assessed by faecal egg count reduction test (FECRT). Six cattle farms with a history of clinical parasitism and macrocyclic lactone use were included. On the day of treatment (Day 0), 20 naturally infected calves per farm (total $n = 120$) were stratified by initial faecal egg counts (FEC) and randomly allocated to a treatment group dosed with $0.2 \text{ mg IVM kg}^{-1}$ body weight s.c. (IVM; $n = 10$) or an untreated control group (CTL; $n = 10$). Individual FEC were obtained at Day 0 and Day 14 post-treatment and pooled faeces by group were cultured to isolate L3 for detection of *Ostertagia ostertagi* and *Cooperia oncophora* by real-time PCR. Treatment efficacies were analysed using the recommended WAAVP method and two Bayesian methods – ‘Bayescount’ and ‘eggCounts’. A simulation study evaluated the performance of the methods to correctly identify the FEC reduction percentages of simulated bovine FEC data representing the observed real data. In the FECRT, reduced IVM efficacy was confirmed in three farms by all methods based on FEC of treated animals only, and in one farm according to the methods including FEC from both treated and untreated cattle. Post-treatment, *C. oncophora* was detected in IVM groups from all herds and *O. ostertagi* was identified in treated animals from two farms. In the simulation study, highly aggregated FEC were poorly analysed by the WAAVP and eggCounts methods, while the Bayescount gave mostly inconclusive results with the simulated treatment group size ($n = 10$). This is the first report of reduced anthelmintic efficacy in Danish cattle and warrants the implementation of drug resistance monitoring and larger surveys in Denmark. Advantages and caveats regarding the use of Bayesian methods and the relevance of including untreated cattle in the FECRT are discussed.

Keywords: gastrointestinal nematodes, cattle, ivermectin, faecal egg count reduction test, real-time PCR, Bayesian methods

1. Introduction

Grazing cattle are continuously exposed to infection with gastrointestinal nematodes (GIN) that can severely impair the health and productivity of pasture-based livestock systems (Corwin, 1997; Shaw et al., 1998; Charlier et al., 2014). In practice, the control of GIN in cattle largely relies on the routine use of anthelmintic drugs, mainly from the macrocyclic lactone (ML) family (Vercruysse and Rew, 2002; Geurden et al., 2015). As a consequence, worm populations resistant to MLs have been selected, and anthelmintic resistance (AR) is now becoming a serious threat to the control of bovine nematodes in several countries (Sutherland and Leathwick, 2011; Gasbarre, 2014; Sutherland and Bullen, 2015). Coinciding with the development of AR, concerns regarding the prophylactic use of veterinary drugs and chemical residues in both food and environment have led to stricter regulations on the use of anthelmintics in some nations (Thamsborg et al., 1999). In 1999, Denmark became the first country to introduce prescription-only use of anthelmintics in livestock, requiring a mandatory veterinary diagnosis before treatment (Anonymous, 1998; 2013). Since 2000, there has been an additional requirement for all prescriptions in production animals to be registered in 'VetStat' – the Danish system for surveillance of the veterinary use of drugs (Stege et al., 2003). Preliminary analyses in the VetStat database indicate that MLs accounted for ~85% of all anthelmintic drugs prescribed for Danish cattle between 2010 and 2012 (Peña-Espinoza et al, unpublished data). Of all prescribed ML treatments, ivermectin (alone or in combination) was the predominant active compound, corresponding to 72% of all anthelmintic prescriptions during the period. Considering the significance of MLs for current parasite control strategies in Danish cattle, it is essential to know their present field efficacy against GIN, but such research has not yet been conducted.

In the absence of quantitative molecular techniques for the detection of ML-resistance, and the high cost of the controlled efficacy test (the current gold standard method for verification of anthelmintic activity; Wood et al., 1995), the only readily available technique for investigating field drug efficacy is the faecal egg count reduction test (FECRT). This technique estimates the efficacy of an anthelmintic to reduce the faecal egg counts (FEC) of infected animals based on measurements pre- and post-treatment, or between treated and untreated individuals. The major advantages of the FECRT are that all drugs can be tested regardless of active compounds and formulation, and it relies on FEC detection methods, which are readily available from most diagnostic laboratories. Current recommendations to conduct FECRT in cattle derive from guidelines by the World Association for Advancement of Veterinary Parasitology (WAAVP; Coles et al., 1992), originally developed for detection of AR in sheep nematodes. Since, potential limitations have been highlighted concerning its use in cattle, mainly due to the lower faecal egg excretion of cattle compared to sheep, the highly aggregated distribution of FEC in cattle herds, and the high degree of density-dependence of bovine nematodes (Coles, 2002; Coles et al., 2006; Demeler et al., 2010; El-Abdellati et al., 2010; Sutherland and Leathwick, 2011). To reduce the uncertainty of the FECRT results, much information on the true anthelmintic efficacy can be gained by studying the 95% confidence interval of a treatment (Levecke et al., 2012). Robust statistical analyses using Bayesian modelling have been advocated to cope with aggregated and low FEC data, and to increase the accuracy in estimating confidence intervals (Denwood et al., 2010; Torgerson et al., 2014). In addition, likelihood-based methods can be used to explicitly describe the Poisson error distribution of nematode eggs present in faecal samples (Torgerson et al., 2014), increasing the robustness of the FECRT if combined with FEC methods of high sensitivity. Bayesian methods are being increasingly used for monitoring AR in livestock helminths (Neves et al., 2014; O'Shaughnessy et al., 2014; Geurden et al., 2015; Fischer et al., 2015; Novobilský and Höglund, 2015; Ramos et al., 2016). However, the performance of these methods and the standard WAAVP method has not yet

been evaluated under varying levels of parasite aggregation commonly found in bovine FEC data. In addition, sensitive and species-specific tests to detect which GIN species survive treatment are critical for the surveillance of AR and are urgently required for cattle (Coles, 2002; Sutherland and Leathwick, 2011).

The objectives of the present study were: 1) to assess the efficacy of the most common MLs used in Danish cattle, ivermectin (IVM), against GIN in naturally infected Danish cattle by FECRT, and 2) to evaluate the performance of different statistical approaches for estimating drug efficacy using simulated bovine FEC data of similar characteristics to those observed in Danish cattle. In addition, we investigated the prescription patterns of anthelmintics in the study farms in order to examine a possible relationship between previous use of ML drugs and drug efficacy in the FECRT.

2. Materials and methods

2.1. Selection of farms

Cattle farms (~50) with a history of clinical parasitism were contacted through local veterinarians across Denmark during spring 2013 and 2014. The farmers were offered free FEC analyses and evaluation of anthelmintic efficacy by FECRT. Farms were selected based on the following criteria: herd size ≥ 20 first-season grazing (FSG) calves with ≥ 4 weeks of grazing and not treated with anthelmintics within 8 weeks prior to sampling. In addition, the availability of a cattle crush or barn was required for the handling of animals. A total of 19 farms (8 in 2013 and 11 in 2014) that fulfilled these criteria accepted the invitation. Individual faecal samples were collected from 20 FSG calves in each farm between mid-June and early September of 2013 and 2014 for analysis of FEC (initial screening). Due to a low number of farms with mean FEC > 150 strongyle eggs per g (epg) of faeces (as recommended by Coles et al., 1992), farms with a mean FEC ≥ 75 epg were selected for the FECRT. Of the six farms finally included in the study, one herd was a conventional

beef farm (farm #1), three were organic dairy farms (#2, #4 and #6), one was an organic beef farm (#5) and one was a conventional dairy farm (#3). The cattle breeds in these farms were Danish Holstein crossbreeds (#1 and #5), Danish Holstein (#2, #3, and #6) and Danish Jersey (#4). All the selected farms were located in the peninsula of Jutland and the FECRT was conducted within one to four weeks after the initial screening.

2.2 Faecal egg count reduction test (FECRT)

The FECRT was performed to test the efficacy of IVM based on WAAVP recommendations (Coles et al., 1992). Pre- and post-treatment faecal samples from treated and untreated animals were included, and a total of 120 FSG calves were enrolled in the FECRT studies. On the day of treatment (Day 0), 20 FSG animals from each farm were stratified by FEC (based on the initial screening) and randomly allocated to a treatment group (IVM; n =10) or an untreated control group (CTL; n =10) of similar (initial) mean FEC. Due to a limited number of animals available in farms #4 and #6 at the start of the FECRT, the CTL groups at these properties consisted of nine calves. Oral formulations of IVM are not registered for use in cattle in Denmark, so injectable IVM was used. At Day 0, individual body weights (BW) were estimated in the IVM group using a girth tape for cattle (Rondo combi®, Kruuse, Denmark), and the calves were injected with the recommended dose of IVM ($0.2 \text{ mg kg}^{-1} \text{ BW s.c.}$, Ivomec® 10 mg/mL, Merial Norden A/S). Faecal samples were collected rectally from all animals on Day 0 and 14 days post-treatment (Day 14). Immediately after collection, the faecal samples were vacuum packed (Freshield Touch, CSE Co, Gyeonggi-do, Korea) to create anaerobic conditions and transported to the laboratory in a cooling box. On all farms, animals in the IVM and CTL groups grazed together on the same pastures until Day 14 when all control calves were treated with the recommended dose of injectable IVM as described above.

2.3 Parasitological analyses

Upon arrival at the laboratory, faecal samples were refrigerated at 5°C until analysis. Individual FEC were determined using an accredited, modified McMaster technique with a sensitivity of 5 egg (Henriksen and Aagard, 1975). At Day 0 and Day 14, pooled larval cultures were prepared from the IVM and CTL groups by mixing 10 g of faeces from each animal of the same group into a pool, which was then cultured according to Roepstorff and Nansen (1998). After 14 days of incubation at 20°C, nematode L3 were recovered by Baermannisation and stored at 12°C. A small number of L3 were harvested from the post-treatment larval cultures from farms #1, #2, #5 and #6 (< 40 larvae per group). All pooled L3 were used for molecular detection of *Ostertagia ostertagi* and *Cooperia oncophora* by real-time quantitative PCR (qPCR).

2.4 Species-specific identification of nematode larvae by qPCR

Molecular detection of *O. ostertagi* and *C. oncophora* in the pooled L3 suspensions was performed using the qPCR method described by Höglund et al. (2013), with modifications. Briefly, all L3 pooled per group were concentrated by centrifugation and transferred into a 2 mL cryotube. Larvae (in 200 µl of tap water) were mixed with 1 mL buffer ATL (QIAGEN, Germany) and 600 µl of 0.5 mm Zirconia beads (BioSpec Products, USA) and homogenised by bead-beating for 1 min at 6.5 m/s (FastPrep®-24, MP Biomedicals, USA). Subsequently, the suspension was digested at 56°C for 60 min using 20 µl of Proteinase K (20 mg/ml, QIAGEN, Germany) following manufacturer's instructions. Genomic DNA was extracted from the digested larval homogenate by QIAamp® DNA Mini Kit (QIAGEN, Germany). For qPCR analyses, primers and probes targeting the second internal transcribed spacer (ITS-2) of the ribosomal DNA of *O. ostertagi* and *C. oncophora* were used (Höglund et al., 2013). ITS-2 copies of both nematode species were quantified by correlating cycle threshold (Ct) values to a standard curve with 2×10^7 , 2×10^6 , 10^5 , 10^4 and 10^3 molecules µl⁻¹ of stock plasmid DNA. The plasmid DNA was made from a pCR® 2.1 vector (Thermo-Fischer

Scientific, USA) that comprised ITS-2 sequences of *C. oncophora* (GenBank[®] accession no. AB245040.2, position 651-729) and *O. ostertagi* (GenBank[®] accession no. AB245021.2, position 1036-1126) (Höglund et al., 2013). The reactions were run in a Rotor-Gene Q RG-6000[®] (QIAGEN, Germany) in total volumes of 25 µL using 2 µl DNA as a template. The PCR mix contained 0.65 U Taq2000[®] polymerase (Agilent Technologies, USA), 0.3 µM forward and reverse primers, 0.2 µM probe, 200 µM dNTP and 5.5 mM MgCl₂. Rotor-Gene Q[®] series software (QIAGEN, Germany) was used to determine Ct values for each run. The cycling conditions were 95°C for 10 min and 50 cycles of amplification (95°C, 15 sec, 62°C, 60 sec). All samples and standards were carried out in technical duplicates, with exception of the standard curves, which were carried out in triplicates.

2.5 Use of anthelmintics in the selected farms between 2002 and 2012

Data of all anthelmintics prescribed between 2002 and 2012 in the six farms selected for the FECRT were extracted from the VetStat database (Stege et al., 2003). Data retrieved included the name and active compound of the prescribed anthelmintic, the formulation and total amount of the drug prescribed, and the targeted cattle age group. In VetStat, cattle are divided into three age groups: 1) calves < 12 months old (heifer and bull calves); 2) young cattle ≥ 12 months old (heifers until first calving and steers until slaughter); 3) adult cattle (cows after first calving). However, the exact number of treatments based on each prescription is not recorded in VetStat. We therefore estimated the number of treatments for each prescription in the ‘targeted’ group (i.e. calves, young cattle or adult cattle) within each farm using the total amount of a prescribed drug, the recommended dose of a given anthelmintic (considering its formulation) and a defined BW for each cattle age group. The defined BW for each age group was estimated based on data from the Danish Cattle Association (Danmap, 2012) and considering common anthelmintic treatment practices in each group as: 200 kg BW for calves, 450 kg BW for young cattle and 620 kg BW for adult cattle.

Due to the defined BWs and the likely variation in the actual amount of active compound used for different animals following each prescription, the calculated number of treatments is only a proxy of the real number of animals treated. In addition, it was assumed that anthelmintics prescribed to farmers were used within a month, which may not always have been the case. However, this potential bias is presumably consistent across the different drugs and age groups. The number of animals per age group in each farm at the prescription date was retrieved from the Central Husbandry Register (Ministry of Environment and Food of Denmark, <http://chr.fvst.dk>, accessed on 15 March 2016). The efficacy of previously used anthelmintics had not been tested in any of the six farms prior to the study.

2.6 Statistical analyses

The efficacy of IVM was analysed by calculating the arithmetic mean FEC reduction percentage (FECR%) with 95% confidence intervals (CI) in the IVM group using different statistical methods, both including and excluding the FEC fluctuations in the CTL group. Initially, the FECR% and 95% CI were determined based on WAAVP guidelines (Coles et al., 1992) as: 1) FECR% (*With* CTL) = $100 \times (1 - [T_2/C_2])$, where T_2 and C_2 are the arithmetic mean FEC of the IVM and CTL group at Day 14, respectively, and 2) FECR% (*No* CTL) = $100 \times (1 - [T_2/T_1])$, where T_1 and T_2 are the arithmetic mean FEC of the IVM group at Day 0 and 14, correspondingly.

Furthermore, we tested two Bayesian modelling approaches to infer the true IVM efficacy. Firstly, we used the Markov chain Monte Carlo (MCMC) method ‘Bayescout’, modified from Denwood et al. (2010) and Geurden et al. (2015). Briefly, this method models the pre-treatment FEC data as a gamma-Poisson (negative binomial) distribution and the post-treatment FEC data as a separate gamma-Poisson distribution; the difference of the mean between these distributions is the FECR%. The 95% CI of the FECR% is also provided. This modelling approach accounts for the FEC aggregation between individuals in a group and for the Poisson errors of the egg counting method.

Moreover, the Bayescount method allows the FEC variability among animals to be different between Day 0 and 14. This increases the uncertainty in the estimations compared with assuming equal FEC variability at Day 0 and 14, but includes the potential differences in drug efficacy between animals and the higher FEC variation commonly observed post-treatment. Further description of the method can be found in Denwood (2015). In the present study, the efficacy of IVM treatments in the Bayescount method was evaluated considering 1) the FEC variation in the IVM and CTL groups between Day 0 and 14 (*With* CTL), and 2) the FEC variation only in the IVM group between Day 0 and 14 (*No* CTL). All FEC data were transformed to the number of eggs counted (FEC divided by 5) for analysis in the Bayescount method.

In addition, we evaluated the efficacy of IVM treatments using the ‘eggCounts’ method described by Torgerson et al. (2014). This Bayesian method is very similar to Bayescount – it also uses a MCMC approach and models a gamma-Poisson distribution for pre- and post-treatment FEC data, thus accounting for FEC aggregation in groups and the Poisson errors of the egg counting process (Torgerson et al., 2014). The differences between these two Bayesian methods are that eggCounts has a web interface, allowing easy access for users not familiar with statistical modelling (available at: <http://www.math.uzh.ch/as/index.php?id=eggcounts&L=1>), as well as differences in the prior distributions used in the modelling. In particular, the eggCounts method defines a single prior to the dispersion parameter (overdispersion), and as a consequence assumes that no variation in drug efficacy exists between animals in the same treated group. Further details of the method can be found in Paul et al. (2014) (available at: <http://arxiv.org/abs/1401.2642>). For eggCounts, a correction factor of 5 epg was applied and FEC data from only the IVM group at Day 0 and 14 were used (*No* CTL), as the method currently does not incorporate the FEC variation of a separated CTL group.

Calculation of FECR% and 95% CI for the WAAVP method was performed according to Coles et al. (1992) in Microsoft Excel® 2010. The modelling with Bayescount and eggCounts was conducted in R version 3.2.2 (R Core Team, 2015) using the freely-available packages ‘Bayescount’ (Denwood, 2015) and ‘eggCounts’ (Torgerson et al., 2014), respectively. The level of FEC aggregation in animal groups at initial sampling and in the FECRT studies was estimated as the (moment estimate) exponent of the negative binomial distribution (k) according to Smith and Guerrero (1993).

2.7 Interpretation of treatment efficacy

The efficacy level of IVM treatments were interpreted based on the FECR% and lower 95% CI as recommended by Coles et al. (1992), as well as the upper 95% CI as suggested by Lyndal-Murphy et al. (2014), from which we categorised three conditions:

- 1) Efficacious, when mean FECR% and upper CI $\geq 95\%$ and lower CI $\geq 90\%$;
- 2) Reduced efficacy, when mean FECR% and upper CI $< 95\%$ and lower CI $< 90\%$;
- 3) Inconclusive, when none of the above conditions were met.

2.8 Simulation study

A simulation study was carried out to compare the performance of the standard WAAVP method and the two Bayesian approaches in estimating drug efficacy under varying FECR% and FEC aggregation scenarios. For this study, we analysed the FECR between Day 0 and 14 only in the simulated treatment groups (i.e. without untreated controls). Datasets were simulated based on pre- and post-treatment FEC aggregation values (k) representing the two most extreme datasets (farms #3 and #5) and one moderate dataset (farm #1) observed in our FECRT studies. Farm #3 was chosen to represent very high FEC aggregation ($k_{\text{Day } 0} = 0.6$ and $k_{\text{Day } 14} = 0.01$), farm #5 was

selected to represent low FEC aggregation ($k_{\text{Day } 0} = 2.6$ and $k_{\text{Day } 14} = 1.4$) and farm #1 represented moderate FEC aggregation ($k_{\text{Day } 0} = 1.2$ and $k_{\text{Day } 14} = 0.8$). For each of these aggregation values (k), FEC datasets were simulated from a negative binomial distribution with a sample size of ten animals and a mean FEC pre-treatment (Day 0) of 34 eggs counted (equal to 170 epg using a FEC method with a detection limit of 5 epg). Post-treatment (Day 14) data were then simulated with mean FEC of 5 eggs counted (equal to 25 epg, representing a simulated FECR = 85%) or 1 egg counted (equal to 5 epg, simulated FECR = 97%) and with the post-treatment aggregation values shown above. All values represented the average of the data observed in the FECRT studies. The datasets were simulated 1,000 times for each of the (two) simulated FECR% and (three) aggregation levels described (i.e. representing the results of 1,000 different treatment groups for each set of parameters). Each of the 6,000 simulated datasets were then analysed with the Bayescount and eggCounts packages in R, using the ‘unpaired’ model (without controls) for the former, and the ‘fecr_mcmc’ function with an ‘unpaired’ model for the latter. All other options and model inference were as described for the FECRT studies (Section 2.6) and the mean FECR% and 95% CI obtained for each simulated dataset were recorded. Simulated datasets were also analysed using the WAAVP method without controls (as described in Section 2.6) and with 95% CI calculated according to Coles et al. (1992). The performance of the methods was investigated in two ways: firstly, by studying the probability of the simulated FECR% being included in the 95% CI provided by each method (i.e. coverage probability of the 95% CI); secondly, through the classification probability that the methods would correctly detect a reduced efficacy (for a simulated FECR = 85%), efficacious treatment (for a simulated FECR = 97%) or inconclusive results using the interpretation described in Section 2.7.

3. Results

3.1. Initial screening of farms

Faecal egg count data observed in groups of 20 FSG calves from the 19 farms initially screened are summarised in Table 1. A total of 380 animals were sampled during mid-June and early September of 2013 and 2014. Strongyle eggs were detected on all properties. Very low egg excretion levels (mean FEC \leq 44 epg) were seen in 12 farms (63%) and highly aggregated FEC ($k < 1$) were observed in 13 farms (68%). All six farms selected for the FECRT had an initial arithmetic mean FEC \geq 75 epg.

3.2 FECRT

Results of the FECRT are presented in Table 2. At Day 0, IVM groups in farms #5 and #6 had a very low mean FEC (\leq 55 epg). Highly aggregated FEC were observed in the IVM group of farm #3 at Day 0 ($k = 0.6$), and in IVM groups from all farms at Day 14 ($k \leq 0.8$, except farm #5). In farm #6, three calves in the IVM group could not be sampled at Day 14 and were excluded from the analyses.

The WAAVP and the Bayescount methods including the FEC of treated and untreated animals (*With CTL*) declared reduced IVM efficacy in farm #2, whereas only the WAAVP method stated reduced drug efficacy in farm #1. Furthermore, the WAAVP method (*With CTL*) indicated efficacious treatment in farm #5, while all other results were inconclusive, as determined by both methods including IVM and CTL groups.

The WAAVP, Bayescount and eggCounts methods unanimously indicated reduced IVM efficacy in farms #1, #2 and #4 when analysing the FEC fluctuations only in the treated groups (*No CTL*). All methods (*No CTL*) yielded inconclusive results in farm #5, and the methods disagreed in two farms: only the two Bayesian methods indicated reduced efficacy in farm #6, while reduced IVM efficacy

was only indicated by the eggCounts method in farm #3; in the latter, a high FEC aggregation was observed in the IVM group ($k < 1$). Furthermore, in farm #1, eight CTL calves with high FEC at Day 14 ($k = 1.9$) were treated with injectable moxidectin (0.2 mg kg⁻¹ BW s.c., Cydectin® inj. 10 mg/mL), and faecal samples analysed 21 days post-treatment revealed a mean FECR = 89% (95% CI = 82 – 93) by the eggCounts method (data not shown).

3.3 Real-time qPCR for detection of *O. ostertagi* and *C. oncophora*

Proportions of ITS-2 copies of *O. ostertagi* and *C. oncophora* detected in pre- and post-treatment pooled larval cultures in the FECRT are presented in Table 3. At Day 0, *O. ostertagi* and *C. oncophora* ITS-2 copies were detected in all groups from all farms, with the exception of the CTL group in farm #2, where only *C. oncophora* was identified. Post-treatment, *O. ostertagi* ITS-2 copies were detected only in IVM groups of farms #3 and #4, while *C. oncophora* ITS-2 copies were identified in treated groups from all six farms. Amplification efficiencies of the standard curves were 96% and 100% for *C. oncophora* and *O. ostertagi*, respectively, with correlation coefficients (R^2) of 0.99 for both species.

3.4 Anthelmintic prescriptions in the farms selected for FECRT

The anthelmintic prescriptions between 2002 and 2012 in the six farms selected for FECRT are shown in Supplementary Table 1. Prescriptions with anthelmintics other than MLs were rarely observed (only in farm #2 with albendazole and farm #3 with levamisole). In calves from all six farms, IVM was the most commonly used ML (87.2% of all prescriptions), followed by moxidectin (12.8%). Similarly, IVM was the predominant ML used in young cattle (84.7% of all prescriptions), followed by moxidectin (14.7%). In adult cattle, IVM constituted 61.5% of all prescriptions in farms #2, #3, #4 and #6, followed by eprinomectin (23.8%) and moxidectin (12.5%). No adult cattle

were recorded in farms #1 and #5 during the period. Topical (pour-on) drugs were the predominant formulations used in all six farms (98.6%, 95.1% and 88.4% of all prescribed treatments in calves, young and adult cattle, respectively). In general, treatments were prescribed at irregular intervals in all farms throughout the decade preceding the FECRT and 61% of all prescriptions were targeted to less than 50% of the animals in a given age group. Nevertheless, evidence suggesting whole-group treatments was observed at all farms, particularly farm #1 (the only conventional beef herd in the study) and farm #6 (organic dairy farm). There were no marked differences in the prescription patterns between beef and dairy or conventional and organic farms.

3.5 Simulation study

The performance of the different methods to estimate drug efficacy is presented in Table 4 as the coverage probability of the 95% CI provided by each method. This describes the ability to include the correct simulated FECR% under varying degrees of FEC aggregation, with the target coverage probability of the 95% CI expected to be close to 95%. The 95% CI provided by the WAAVP and Bayescount methods had an acceptable coverage of the simulated FECR% at low and moderate FEC aggregation levels ($k = 2.6$ and 1.2). In comparison, the 95% CI provided by the eggCounts method had a lower coverage of the simulated drug efficacy and was below the expected target coverage of all simulated parameters. High FEC aggregation ($k = 0.6$) markedly affected the performance of the WAAVP and eggCounts methods. With a simulated FECR = 85% and $k = 0.6$, the 95% CI of the WAAVP and eggCounts methods correctly included the simulated FECR% in only 29% and 4% of the observed iterations, respectively. Poor coverage probability of the WAAVP and eggCounts methods was also observed with simulated FECR = 97% and $k = 0.6$. In comparison, the coverage probability of the Bayescount method was only moderately reduced when the FECs were highly aggregated (89 – 91% coverage).

The classification probabilities of the methods to detect reduced drug efficacy, efficacious treatment and inconclusive results are presented in Table 5 for different levels of simulated FECR% and FEC aggregation. Results are presented as the frequency of iterations classified using the criteria described in Section 2.7. The eggCounts method provided the highest number of correctly identified positive (reduced efficacy when simulated FECR = 85%) and negative (efficacious treatment when simulated FECR = 97%) results in comparison with the WAAVP and Bayescount methods, particularly at low and moderate FEC aggregation levels ($k = 2.6$ and 1.2). However, at high FEC aggregation ($k = 0.6$) and with a simulated FECR = 85%, the WAAVP and eggCounts methods provided a high number of false negative results and erroneously declared efficacious treatments for most of the datasets with simulated treatment inefficacy. The Bayescount method had the lowest number of false positive/negative results, but also the highest frequency of inconclusive results (for all k values and simulated FECR%) compared with the eggCounts and WAAVP methods.

4. Discussion

In the FECRT, reduced IVM efficacy was detected in three farms by all methods based on the FEC of treated cattle only, and in one farm according to the methods including the FEC from treated and untreated animals. Post-treatment, *O. ostertagi* and *C. oncophora* L3 were identified in faeces of treated animals in one and six farms with declared reduced IVM efficacy, respectively. This is the first report of reduced anthelmintic efficacy in Danish cattle. Results provided by the different methods of calculation in the same farm were not always in agreement, and an important number of results were inconclusive, particularly with methods including the FEC of untreated animals. The subsequent simulation study demonstrated that high FEC aggregation can affect the outcome of the FECRT, depending on the method used. Studying the anthelmintic prescription patterns in the

decade previous to the FECRT in the six farms revealed an extensive reliance on MLs in general and IVM in particular.

During the initial screening previous to the FECRT, FSG calves with mean FEC < 100 epg were detected in most farms. Similar observations were reported by El-Abdellati et al. (2010) in a larger number of naturally infected, untreated FSG calves from Belgian and German farms. In addition, a high FEC aggregation ($k < 1$) was observed in the majority of the investigated herds, which agrees with earlier reports of overdispersed FEC in naturally infected cattle (Polley and Bickis, 1987) and sheep (Morgan et al., 2005). At Day 0 of the FECRT, three farms had a mean FEC < 150 epg (farms #4, #5 and #6), which was a consequence of our initial selection of farms with mean FEC ≥ 75 epg. Although the use of a FEC method with high sensitivity (5 epg in the present study) reduces the diagnostic uncertainty in samples with low FEC (El-Abdellati et al., 2010; Levecke et al., 2011), a low mean FEC pre-treatment will likely affect the outcome of a FECRT, particularly when group size ≤ 10 and FEC are highly aggregated ($k = 0.1$) (Levecke et al., 2012). In our case, IVM groups in farms #4, #5 and #6 at Day 0 had a low to moderate FEC aggregation ($k = 1.8, 2.6$ and 1.1 , respectively). However, the reduced drug efficacy detected in herds with initial mean FEC < 150 epg should preferably be confirmed by new FECRTs including animals with higher egg excretion.

The qPCR method applied in the present study was able to detect ITS-2 copies of *O. ostertagi* and *C. oncophora* in pooled larval cultures from naturally infected cattle, as earlier reported by Areskog et al. (2013). In agreement with previous European reports (Demeler et al., 2009; Geurden et al., 2015), *C. oncophora* was the predominant species surviving IVM treatment in all farms. This result was expected as this species is the dose-limiting GIN for IVM (Egerton et al., 1981). The more pathogenic *O. ostertagi* was detected in IVM groups from all six farms at Day 0 and from two

farms post-treatment. However, reduced efficacy by all methods (*No CTL*) was only detected in one of these farms (#4), therefore the possible presence of an IVM-resistant *O. ostertagi* in this herd should be further confirmed. The absence of *O. ostertagi* L3 in post-treatment cultures from most farms reflects the expected high efficacy of injectable IVM against this abomasal species (Egerton et al., 1981; Lifschitz et al., 2000). However, the examination of pooled L3, the low FEC of samples and the poor yield of L3 following culture may have been masking the true ratio of the species surviving treatment. Individual larval cultures could have increased the sensitivity of the test and resulted in a different outcome. However, this was not possible within the timeframe of the project. A major limitation to the routine use of L3 for species detection is the time required to culture L3 and the well-known variability in developmental requirements of different species in larval cultures (Roeber and Kahn, 2014). Other nematode stages could be used for species-specific detection by molecular techniques (Harmon et al., 2006), and recent studies from our group have explored this by effectively quantifying ITS-2 copies in eggs and first-stage larvae of *O. ostertagi* (Drag et al., 2016).

In the FECRT, methods of calculation that included the FEC of untreated animals resulted in a higher number of inconclusive results, in comparison with methods excluding the CTL group. The inclusion of untreated animals in a FECRT has been recommended to detect changes in the FEC of a herd not related with the treatment, and for the correction of such fluctuations in the estimation of drug efficacy (Coles et al., 1992; Lyndal-Murphy et al., 2014). However, the inclusion of a control group is based on the assumption that treated and untreated calves with comparable FEC share similar worm burdens, which may not be always the case (Michel, 1967; 1969). Moreover, the density-dependent control of fecundity in some cattle nematodes, such as in *O. ostertagi*, may reduce the FEC in untreated animals due to newly acquired infections and could increase the egg excretion of female worms in treated groups due to lower worm burden and competition (Dobson et

al., 2012). In practice, when few animals are available for a FECRT ($n \leq 20$), the inclusion of a control group will limit the size of the treatment group and consequently increase uncertainty of the estimated efficacy (Denwood et al., 2010). The low number of FSG calves with positive FEC that could be included in our study suggests that in these circumstances, the inclusion of untreated cattle in the FECRT should not be recommended. Instead, it may be advisable to include more animals in the treatment group in order to increase the certainty of the calculated FECR% (Denwood et al., 2010; Levecke et al., 2012).

In the simulation study, the Bayescount outperformed the other two methods in correctly identifying the simulated FECR% within the 95% CI produced, at varying levels of FEC aggregation. Surprisingly, the WAAVP method showed a better coverage probability than the more sophisticated eggCounts, but the former has the major limitation that it does not provide a 95% CI when post-treatment FEC are zero (FECR = 100%) (Torgerson et al., 2014; Geurden et al., 2015). Highly aggregated FEC data were poorly analysed by both the WAAVP and eggCounts methods, which were unable to estimate an optimal CI for the observed reductions. This inaccuracy was further reflected in the higher frequency of ‘false negatives’ (i.e. declaring efficacious treatment when simulated FECR = 85%) provided by the eggCounts and the WAAVP methods in the scenario of high overdispersion. These results suggest that when analysing highly aggregated data ($k < 1$), the drug efficacy estimated by the WAAVP and eggCounts (with default priors, see below) methods should be interpreted with caution. This was evidenced in the FECRT, where the IVM group in farm #3 had a highly aggregated FEC and only the eggCounts declared reduced drug efficacy in this property, while all other methods indicated inconclusive results. Nevertheless, in cases of simulated low and moderate aggregations the eggCounts provided the higher rate of ‘true positive’ (i.e. declaring reduced efficacy for simulated FECR = 85) and ‘true negative’ (i.e. declaring efficacious treatment for simulate FECR = 97%), followed by the WAAVP and Bayescount methods. In

comparison, the Bayescount method provided no false negative results, despite varying FEC aggregation levels, but also gave the highest rate of inconclusive results. This suggests that a sample size of ten animals is not sufficient to provide conclusive results using the Bayescount and to correctly distinguish between a FECR of 85% and 97% in the presence of high overdispersion. Bayescount and eggCounts methods are extremely similar, but do differ in terms of the default prior distributions that are used by each procedure, with Bayescount using more conservative prior distributions for the parameter controlling FEC aggregation. The relatively informative priors used by the eggCounts method reduce the number of inconclusive results, but at the expense of producing erroneous inference when the prior distribution conflicts with the data. The poor coverage probability of 95% CI by eggCounts therefore derives from the distribution priors set in this method, which exclude the possibility of very high overdispersion as seen in our data. It is possible to adjust the prior distributions for both the eggCounts and Bayescount methods, although this requires some familiarity with Bayesian methods, and is not possible using the web interface for eggCounts. Given the difficulties associated with eggCounts under high FEC overdispersion and Bayescount with a low sample size, we suggest that the use of both Bayesian methods requires some consideration of appropriate priors for the models, as well as an increased sample size of treatment groups. Due to the possible erroneous inference obtained from the use of these complex methods inappropriately, we also strongly recommend that users without the necessary statistical experience seek assistance with implementation and interpretation of both the eggCounts and Bayescount methods.

At the recommended dose of 0.2 mg kg⁻¹ BW, injectable IVM is expected to reduce susceptible *O. ostertagi* adults and L4 stages by $\geq 99\%$ and susceptible *C. oncophora* adults and L4 stages by $\geq 97\%$ (Egerton et al., 1981). However, the current WAAVP guidelines suggests that any treatment with a FECR > 90% in cattle should not be considered a case of drug resistance (Coles et al., 1992).

In practice, most studies evaluating anthelmintic efficacy in cattle declare AR when FECR < 95% and lower 95% CI < 90, as proposed for sheep. Nevertheless, it has been suggested that this criteria is biased towards declaration of AR when there is none, particularly if the mean FECR% is between 90 – 95% and the CI is wide due to small group sizes (Lyndal-Murphy et al., 2014). In the present study, we included the upper 95% CI in the interpretation of the FECRT to increase the certainty of detecting true cases of IVM inefficacy. A similar interpretation for FECRT studies in cattle has been reported in recent publications by Geurden et al. (2015) and Ramos et al. (2016). However, the effect of including the upper 95% CI in the interpretation for estimating drug efficacy using a FECRT, and how this correlates with an actual resistant phenotype confirmed by controlled efficacy tests, warrant further investigation. Notwithstanding, reduced drug efficacy in the FECRT may not necessarily be caused by AR. A lower-than-expected *in vivo* efficacy, or varying drug response between animals, could be the result of under dosing (e.g. due to inaccurate estimation of BW) and/or altered drug pharmacokinetics and pharmacodynamics in different animals (e.g. due to nutrition-related variations in fat reserves that may affect the persistent efficacy of ML, erratic absorption of drugs from the site of injection and/or interactions with other co-administered drugs) (González Canga et al., 2008; El-Abdellati et al., 2010; Areskog et al., 2012; De Graef et al., 2013; Areskog et al., 2014). These factors can impair the correct estimation of drug efficacy and detection of AR, particularly in the dose-limiting species *C. oncophora*. Recently, *C. oncophora* populations that were declared resistant to the recommended dose of injectable IVM by FECRT in two Swedish cattle farms (using girth tape for weight estimation and with FECR% [upper CI] = 78% [97%] and 79% [98%] in each farm; Demeler et al., 2009) were declared IVM-susceptible when tested in calves under controlled conditions (Areskog et al., 2014). Therefore, the presence of IVM-resistant nematodes suggested by our FECRT in three farms, as well as the AR status in the farms with inconclusive results and low initial FEC, should be confirmed by controlled efficacy test.

The use of anthelmintics in the farms included in our FECRT was investigated to potentially detect trends in drug use and the extent of treatments with MLs. Based on VetStat data, it was confirmed that between 2002 and 2012, more than 98% of all anthelmintics prescribed in the six farms were MLs, mostly topical IVM products. A similar reliance on ML drugs has been preliminarily detected in the entire Danish cattle population in the period 2010 – 2014, with ML constituting ~85% of all treatments – of which 61% were IVM, mainly in topical formulations (Peña-Espinoza et al., unpublished data). The irregular prescription of MLs observed in the study farms correlates with the prescription-only regulations in Denmark, illustrated by the treatment of single animals or selected groups of animals in the herds. However, prescription patterns suggesting whole-group treatments in some farms indicate that these may be recommended by veterinarians under certain conditions (e.g. during outbreaks of dictyocaulosis), and the effect of this practice on the selection for AR needs further investigation. In addition, and considering the relevance of MLs for nematode control in Danish cattle, the true extent of AR in bovine nematodes in Denmark needs to be assessed in larger surveys. Until then, producers and veterinarians should be aware of potentially ineffective treatments against GIN of cattle, and closely monitor the efficacy of these treatments, while reducing the reliance on anthelmintics by including other parasite control strategies, such as grazing management and feeding with bioactive forages (Nansen et al., 1987; 1988; Peña-Espinoza et al., 2016).

In conclusion, reduced IVM efficacy was detected in three of six Danish cattle farms investigated. *Cooperia oncophora* was the main species surviving IVM treatment, but *O. ostertagi* was also identified post-treatment in one farm with reduced IVM efficacy. High FEC aggregation markedly affects the performance of the statistical methods used to estimate drug efficacy, and results provided by such methods for highly over-dispersed datasets and low group sizes should be interpreted with caution. The presence of IVM-resistant nematode strains suggested by the FECRT

in three farms should preferably be confirmed by controlled efficacy test. The reduced efficacy of IVM detected in this study and the widespread use of ML drugs in Danish cattle, suggest that farmers and their advisors should be aware of potentially ineffective treatments and should closely monitor the efficacy of anthelmintic prescriptions.

Conflicts of interests

The authors declare that they have no conflicts of interests.

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References

Anonymous, 1998. Lov nr. 1043 af 23/12-1998 om ændring af lov om lægemidler (in Danish).

<https://www.retsinformation.dk/forms/R0710.aspx?id=21108> (visited on 14 January 2016).

Anonymous, 2013. BEK nr. 1671 af 12/12/2013 (in Danish).

<http://www.retsinformation.dk/forms/R0710.aspx?id=160842> (visited on 14 January 2016).

Areskog, M., Ljungström, B., Höglund, J., 2013. Limited efficacy of pour-on anthelmintic treatment of cattle under Swedish field conditions. *Int. J. Parasitol. Drugs Drug Resist.* 3, 129–134.

doi:10.1016/j.ijpddr.2013.06.002

Areskog, M., Sollenberg, S., Engström, A., von Samson-Himmelstjerna, G., Höglund, J., 2014. A controlled study on gastrointestinal nematodes from two Swedish cattle farms showing field evidence of ivermectin resistance. *Parasit. Vectors* 7, 13. doi:10.1186/1756-3305-7-13

Areskog, M., Von Samson-Himmelstjerna, G., Alvinerie, M., Sutra, J.F., Höglund, J., 2012. Dexamethasone treatment interferes with the pharmacokinetics of ivermectin in young cattle. *Vet. Parasitol.* 190, 482–488. doi:10.1016/j.vetpar.2012.07.011

Charlier, J., van der Voort, M., Kenyon, F., Skuce, P., Vercruysse, J., 2014. Chasing helminths and their economic impact on farmed ruminants. *Trends Parasitol.* 30, 361–367. doi:10.1016/j.pt.2014.04.009

Coles, G., Bauer, C., Borgsteede, F., Geerts, S., Klei, T., Taylor, M., Waller, P., 1992. World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Vet. Parasitol.* 44, 35–44.

Coles, G., Jackson, F., Pomroy, W., Prichard, R., von Samson-Himmelstjerna, G., Silvestre, A., Taylor, M., Vercruysse, J., 2006. The detection of anthelmintic resistance in nematodes of veterinary importance. *Vet. Parasitol.* 136, 167–185. doi:10.1016/j.vetpar.2005.11.019

Coles, G.C., 2002. Cattle nematodes resistant to anthelmintics: why so few cases? *Vet. Res.* 33, 481–489.

Corwin, R.M., 1997. Economics of gastrointestinal parasitism of cattle. *Vet. Parasitol.* 72, 451–460. doi:10.1016/S0304-4017(97)00110-6

Danmap, 2012. The Danish integrated antimicrobial resistance monitoring and research programme. Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark. Copenhagen, Denmark. ISSN 1600-2032.

De Graef, J., Claerebout, E., Geldhof, P., 2013. Anthelmintic resistance of gastrointestinal cattle nematodes. *Vlaams Diergeneeskd. Tijdschr.* 82, 113–123.

Demeler, J., Küttler, U., von Samson-Himmelstjerna, G., 2010. Adaptation and evaluation of three different in vitro tests for the detection of resistance to anthelmintics in gastro intestinal nematodes of cattle. *Vet. Parasitol.* 170, 61–70. doi:10.1016/j.vetpar.2010.01.032

Demeler, J., Van Zeveren, A.M.J., Kleinschmidt, N., Vercruysse, J., Höglund, J., Koopmann, R., Cabaret, J., Claerebout, E., Areskog, M., von Samson-Himmelstjerna, G., 2009. Monitoring the efficacy of ivermectin and albendazole against gastro intestinal nematodes of cattle in Northern Europe. *Vet Parasitol* 160, 109–115. doi:10.1016/j.vetpar.2008.10.030

Denwood, M.J., 2015. Bayescount: power calculations and Bayesian analysis of count distributions and FECRT data using MCMC. <https://cran.r-project.org/web/packages/bayescount/index.html>

Denwood, M.J., Reid, S.W.J., Love, S., Nielsen, M.K., Matthews, L., McKendrick, I.J., Innocent, G.T., 2010. Comparison of three alternative methods for analysis of equine faecal egg count reduction test data. *Prev. Vet. Med.* 93, 316–323. doi:10.1016/j.prevetmed.2009.11.009

Dobson, R., Hosking, B., Jacobson, C., Cotter, J., Besier, R., Stein, P., Reid, S., 2012. Preserving new anthelmintics: a simple method for estimating faecal egg count reduction test (FECRT) confidence limits when efficacy and/or nematode aggregation is high. *Vet. Parasitol.* 186, 79–92. doi:10.1016/j.vetpar.2011.11.049

- Drag, M., Höglund, J., Nejsum, P., Thamsborg, S. M., Enemark, H. L., 2016. The level of embryonation influences detection of *Ostertagia ostertagi* eggs by semi-quantitative PCR. *Parasit. Vectors* 9:368. doi: 10.1186/s13071-016-1657-4
- Egerton, J.R., Eary, C.H., Suhayda, D., 1981. The anthelmintic efficacy of ivermectin in experimentally infected cattle. *Vet. Parasitol.* 8, 59-70. doi:10.1016/0304-4017(81)90018-2
- El-Abdellati, A., Charlier, J., Geldhof, P., Levecke, B., Demeler, J., von Samson-Himmelstjerna, G., Claerebout, E., Vercruyse, J., 2010. The use of a simplified faecal egg count reduction test for assessing anthelmintic efficacy on Belgian and German cattle farms. *Vet. Parasitol.* 169, 352–357. doi:10.1016/j.vetpar.2010.01.015
- Fischer, J.K., Hinney, B., Denwood, M.J., Traversa, D., von Samson-Himmelstjerna, G., Clausen, P.-H., 2015. Efficacy of selected anthelmintic drugs against cyathostomins in horses in the federal state of Brandenburg, Germany. *Parasitol. Res.* 114, 4441–4450. doi:10.1007/s00436-015-4685-7
- Gasbarre, L.C., 2014. Anthelmintic resistance in cattle nematodes in the US. *Vet. Parasitol.* 204, 3–11. doi:10.1016/j.vetpar.2014.03.017
- Geurden, T., Chartier, C., Fanke, J., di Regalbono, A.F., Traversa, D., von Samson-Himmelstjerna, G., Demeler, J., Vanimisetti, H.B., Bartram, D.J., Denwood, M.J., 2015. Anthelmintic resistance to ivermectin and moxidectin in gastrointestinal nematodes of cattle in Europe. *Int. J. Parasitol. Drugs Drug Resist.* 5, 163–171. doi:10.1016/j.ijpddr.2015.08.001
- González Canga, A., Sahagún Prieto, A.M., Díez Liébana, M.J., Fernández Martínez, N., Sierra Vega, M., García Vieitez, J.J., 2008. The pharmacokinetics and interactions of ivermectin in humans-a mini-review. *AAPS J.* 10, 42–6. doi:10.1208/s12248-007-9000-9

- Harmon, A.F., Zarlenga, D.S., Hildreth, M.B., 2006. Improved methods for isolating DNA from *Ostertagia ostertagi* eggs in cattle feces. *Vet. Parasitol.* 135, 297–302. doi:10.1016/j.vetpar.2005.10.014
- Henriksen, S., Aagard, K., 1975. A simple flotation and McMaster technique. *Nord. Vet. Med.* 28, 392–397.
- Höglund, J., Engström, A., von Samson-Himmelstjerna, G., Demeler, J., Tydén, E., 2013. Real-time PCR detection for quantification of infection levels with *Ostertagia ostertagi* and *Cooperia oncophora* in cattle faeces. *Vet. Parasitol.* 197, 251–257. doi:10.1016/j.vetpar.2013.04.044
- Levecke, B., Dobson, R.J., Speybroeck, N., Vercruyse, J., Charlier, J., 2012. Novel insights in the faecal egg count reduction test for monitoring drug efficacy against gastrointestinal nematodes of veterinary importance. *Vet. Parasitol.* 188, 391–396. doi:10.1016/j.vetpar.2012.03.020
- Levecke, B., Rinaldi, L., Charlier, J., Maurelli, M.P., Morgoglione, M.E., Vercruyse, J., Cringoli, G., 2011. Monitoring drug efficacy against gastrointestinal nematodes when faecal egg counts are low: do the analytic sensitivity and the formula matter? *Parasitol. Res.* 109, 953–7. doi:10.1007/s00436-011-2338-z
- Lifschitz, A., Virkel, G., Sallovitz, J., Sutra, J.F., Galtier, P., Alvinerie, M., Lanusse, C., 2000. Comparative distribution of ivermectin and doramectin to parasite location tissues in cattle. *Vet. Parasitol.* 87, 327–338. doi:10.1016/S0304-4017(99)00175-2
- Lyndal-Murphy, M., Swain, A., Pepper, P., 2014. Methods to determine resistance to anthelmintics when continuing larval development occurs. *Vet. Parasitol.* 199, 191–200. doi:10.1016/j.vetpar.2013.11.002
- Michel, J.F., 1967. Regulation of egg output of populations of *Ostertagia ostertagi*. *Nature* 215, 1001–1002.

Michel, J.F., 1969. Some observations on the worm burdens of calves infected daily with *Ostertagia ostertagi*. *Parasitology* 59, 575–595.

Morgan, E.R., Cavill, L., Curry, G.E., Wood, R.M., Mitchell, E.S.E., 2005. Effects of aggregation and sample size on composite faecal egg counts in sheep. *Vet. Parasitol.* 131, 79–87. doi:10.1016/j.vetpar.2005.04.021

Nansen, P., Jørgensen, R.J., Henriksen, S.A., Foldager, J., 1987. The effects of late turnout on the epidemiology and control of ostertagiasis in calves. *Vet. Parasitol.* 24, 139–147. doi:10.1016/0304-4017(87)90138-5

Nansen, P., Foldager, J., Hansen, J.W., Henriksen, S. A, Jørgensen, R.J., 1988. Grazing pressure and acquisition of *Ostertagia ostertagi* in calves. *Vet. Parasitol.* 27, 325–335.

Neves, J.H. das, Carvalho, N., Rinaldi, L., Cringoli, G., Amarante, A.F.T., 2014. Diagnosis of anthelmintic resistance in cattle in Brazil: A comparison of different methodologies. *Vet. Parasitol.* 206, 216–226. doi:10.1016/j.vetpar.2014.10.015

Novobilský, A., Höglund, J., 2015. First report of closantel treatment failure against *Fasciola hepatica* in cattle. *Int. J. Parasitol. Drugs Drug Resist.* 5, 172–177. doi:10.1016/j.ijpddr.2015.07.003

O’Shaughnessy, J., Earley, B., Mee, J.F., Doherty, M.L., Crosson, P., Barrett, D., Prendiville, R., Macrelli, M., de Waal, T., 2014. Detection of anthelmintic resistance on two Irish beef research farms. *Vet. Rec.* 175:120. doi:10.1136/vr.102556.

Peña-Espinoza, M., Thamsborg, S.M., Desrues, O., Hansen, T.V.A., Enemark, H.L., 2016. Anthelmintic effects of forage chicory (*Cichorium intybus*) against gastrointestinal nematode parasites in experimentally infected cattle. *Parasitology*, available on CJO2016. doi:10.1017/S0031182016000706

- Polley, L., Bickis, M.G., 1987. Gastrointestinal nematode parasites in Saskatchewan cattle: egg count distributions in beef animals. *Can. J. Vet. Res.* 51, 465–469.
- R Core Team. R: A language and environment for statistical computing, 2015. R Foundation for Statistical Computing, Vienna, Austria. <https://www.r-project.org/>.
- Ramos, F., Portella, L.P., de Souza Rodrigues, F., Reginato, C.Z., Pötter, L., Cezar, A.S., Sangioni, L.A., Flores Vogel, F.S., 2016. Anthelmintic resistance in gastrointestinal nematodes of beef cattle in the state of Rio Grande do Sul, Brazil. *Int. J. Parasitol. Drugs Drug Resist.* 6, 93–101. doi:10.1016/j.ijpddr.2016.02.002
- Roeber, F., Kahn, L., 2014. The specific diagnosis of gastrointestinal nematode infections in livestock: Larval culture technique, its limitations and alternative DNA-based approaches. *Vet. Parasitol.* 205, 619–628. doi:10.1016/j.vetpar.2014.08.005
- Roepstorff, A., Nansen, P., 1998. Epidemiology, diagnosis and control of helminth parasites of swine. *FAO Animal Health Manual*, Rome, Italy.
- Shaw, D.J., Vercruyse, J., Claerebout, E., Dorny, P., 1998. Gastrointestinal nematode infections of first-grazing season calves in Western Europe: General patterns and the effect of chemoprophylaxis. *Vet. Parasitol.* 75, 115–131. doi:10.1016/S0304-4017(97)00214-8
- Smith, G., Guerrero, J., 1993. Mathematical models for the population biology of *Ostertagia ostertagi* and the significance of aggregated parasite distributions. *Vet. Parasitol.* 46, 243–257. doi:10.1016/0304-4017(93)90062-R
- Stege, H., Bager, F., Jacobsen, E., Thougard, a., 2003. VETSTAT - The Danish system for surveillance of the veterinary use of drugs for production animals. *Prev. Vet. Med.* 57, 105–115. doi:10.1016/S0167-5877(02)00233-7

Sutherland, I.A., Bullen, S.L., 2015. Parasite control in pasture-grazed dairy cattle: Are we at the edge of a precipice? *Anim. Prod. Sci.* 55, 916–921. doi:10.1071/AN14881

Sutherland, I.A., Leathwick, D.M., 2011. Anthelmintic resistance in nematode parasites of cattle: a global issue? *Trends Parasitol.* 27, 176–181. doi:10.1016/j.pt.2010.11.008

Thamsborg, S.M., Roepstorff, A., Larsen, M., 1999. Integrated and biological control of parasites in organic and conventional production systems. *Vet. Parasitol.* 84, 169–86.

Torgerson, P.R., Paul, M., Furrer, R., 2014. Evaluating faecal egg count reduction using a specifically designed package “eggCounts” in R and a user friendly web interface. *Int. J. Parasitol.* 44, 299–303. doi:10.1016/j.ijpara.2014.01.005

Vercruysse, J., Rew, R., 2002. Use of macrocyclic lactones to control cattle parasites in Europe, in: Vercruysse, J., Rew, R.S. (Eds.), *Macrocyclic Lactones in Antiparasitic Therapy*. CABI Publishing, Oxon, UK; New York, USA, pp. 223–247.

Wood, I., Amaral, N., Bairden, K., Duncan, J., Kassai, T., Malone, J., Pankavich, J., Reinecke, R., Slocombe, O., Taylor, S., Vercruysse, J., 1995. World Association for the Advancement of Veterinary Parasitology (WAAVP) second edition of guidelines for evaluating the effectiveness of anthelmintics in ruminants (bovine, ovine, caprine). *Vet. Parasitol.* 58, 181–213.

Table 1. Faecal egg counts (FEC) of 20 first-season grazing calves for each of the 19 Danish farms sampled during initial screening in the 2013 and 2014 grazing seasons. A total of 380 first-season grazers were examined. Data are presented as arithmetic mean FEC and range (in egg per gram of faeces, epg), the exponent of the negative binomial distribution (k) and the date of sampling. Six farms (farms #1 – 6) were included in the subsequent faecal egg count reduction tests (FECRT).

Farm	Mean FEC (epg)	Range (epg)	k	Date of sampling ^b
1	438	85 – 975	1.89	16 June 2013
2	101	5 – 205	2.92	24 June 2013
3	184	5 – 730	0.50	20 August 2013
4	273	20 – 960	0.83	27 June 2014
5	75	10 – 225	2.27	09 July 2014
6	75	15 – 215	2.03	14 August 2014
7	8	0 – 40	0.61	02 July 2013
8	37	0 – 135	0.60	04 July 2013
9	44	0 – 135	1.18	23 July 2013
10	3	0 – 30	0.14	05 August 2013
11	17	0 – 80	0.85	14 August 2013
12	17	0 – 60	0.72	24 June 2014
13	221 ^a	5 – 1165	0.40	02 July 2014
14	18	0 – 55	1.72	22 July 2014
15	8	0 – 40	0.69	23 July 2014
16	6	0 – 25	0.59	15 August 2014
17	3	0 – 20	0.19	25 August 2014
18	2	0 – 10	0.67	26 August 2014
19	11	0 – 40	0.47	07 September 2014

^aDespite high FEC this farmer did not want to participate in the FECRT

^bTurn-out of grazing cattle in Denmark is usually around late April/early May

Table 2. Faecal egg count reduction test in calves naturally infected with gastrointestinal nematodes in six Danish cattle farms. A total of 115 first-season

grazers were sampled on the day of treatment (Day 0) and 14 days post-treatment (Day 14). The calves were treated subcutaneously with ivermectin (IVM, 0.2 mg kg⁻¹ body weight) or left untreated (CTL). Treatment efficacies were calculated including the variation of faecal egg counts (FEC) in treated and control groups (*With* CTL) or in treated groups only (*No* CTL).

Group	Farm #1 (Beef, conv.)		Farm #2 (Dairy, org.)		Farm #3 (Dairy, conv.)		Farm #4 (Dairy, org.)		Farm #5 (Beef, org.)		Farm #6 (Dairy, org.)	
	IVM	CTL	IVM	CTL	IVM	CTL	IVM	CTL	IVM	CTL	IVM	CTL
n	10	10	10	10	10	10	10	9	10	10	7	9
FEC Day 0	333	325	76	69	173	209	354	351	55	39	31	77
(k)	(1.2)	(1.1)	(1.8)	(2.3)	(0.6)	(1.1)	(2.1)	(0.8)	(2.6)	(1.4)	(1.1)	(0.4)
FEC Day 14	46	311	14	55	15	286	51	562	4	98	4	126
(k)	(0.8)	(2.4)	(0.3)	(2.2)	(0.01)	(0.6)	(0.7)	(0.6)	(1.4)	(1.2)	(0.01)	(0.1)
<i>With</i> CTL	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]
WAAVP ^a	85 ^I [67 – 95]	75 ^R [27 – 92]	95 ^I [56 – 99]	91 ^I [74 – 97]	96 ^E [92 – 99]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]
Bayescount ^b	85 ^I [53 – 95]	77 ^R [18 – 91]	93 ^I [19 – 99]	91 ^I [57 – 97]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]	97 ^I [89 – 98]
<i>No</i> CTL	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]	FECR% [CI]
WAAVP ^a	86 ^R [66 – 94]	82 ^R [47 – 94]	92 ^I [30 – 99]	86 ^R [67 – 94]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]	94 ^I [87 – 97]
Bayescount ^b	86 ^R [47 – 94]	82 ^R [32 – 93]	91 ^I [6 – 97]	85 ^R [33 – 94]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]	93 ^I [81 – 98]
eggCounts ^c	86 ^R [83 – 89]	82 ^R [73 – 88]	91 ^R [88 – 94]	85 ^R [82 – 88]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]	93 ^I [86 – 97]

FEC = arithmetic mean faecal egg count; k = exponent of the negative binomial distribution; FECR% = FEC reduction percentage; CI = 95% confidence interval

^aColes et al. 1992; ^bDenwood et al. 2010; ^cTorgerson et al. 2014; ^EEfficacious; ^RReduced efficacy; ^IInconclusive; ^{conv.} = conventional; ^{org.} = organic

Table 3. Proportion (percentage) of the second internal transcriber space (ITS-2) copy numbers of *Ostertagia ostertagi* and *Cooperia oncophora* detected by real-time quantitative PCR in pooled L3 cultured from faeces of naturally infected calves. The calves were treated with the recommended dose of injectable ivermectin (IVM) or left untreated (CTL). The samples were collected on the day of treatment (Day 0) and 14 days post-treatment (Day 14).

Farm	Group	Day 0		Day 14	
		O.o.	C.o.	O.o.	C.o.
#1	IVM	42	58	0	100
	CTL	39	61	57	43
#2	IVM	79	21	0	100
	CTL	0	100	2	98
#3	IVM	45	55	85	15
	CTL	78	22	74	26
#4	IVM	8	92	0.5	99.5
	CTL	17	83	6	94
#5	IVM	42	58	0	100
	CTL	84	16	0	100
#6	IVM	3	97	0	100
	CTL	6	94	0	100

O.o. = *Ostertagia ostertagi*; C.o. = *Cooperia oncophora*

Table 4. Coverage probabilities of the 95% confidence intervals (CI) provided by the WAAVP, Bayescount and eggCounts methods with datasets of simulated FECR = 85% or 97% and with varying levels of FEC aggregation at Day 0 ($k_{\text{Day}0}$) and Day 14 ($k_{\text{Day}14}$) post-treatment. Results are presented as the percentage of iterations in which the simulated FECR% was correctly included within the 95% CI provided by the method or above/below the 95% CI. A total of 1,000 iterations were performed for each combination of true FECR% and k (total 6,000 iterations). See text for further details on the simulated parameters.

Method	WAAVP ^a			Bayescount ^b			eggCounts ^c					
	Simulated FECR%	$k_{\text{Day}0}$	$k_{\text{Day}14}$	Within ^d	Above ^e	Below ^e	Within ^d	Above ^e	Below ^e	Within ^d	Above ^e	Below ^e
95% Confidence Interval												
85	2.6	1.4	1.4	93	3	4	96	3	1	58	21	21
85	1.2	0.8	0.8	92	3	5	94	6	1	48	26	25
85	0.6	0.01	0.01	29	4	67	91	9	0	4	16	79
97	2.6	1.4	1.4	96	2	2	99	1	0	85	8	8
97	1.2	0.8	0.8	92	3	5	99	1	0	74	10	15
97	0.6	0.01	0.01	33	4	63	89	11	0	12	16	73

FECR% = faecal egg count reduction percentage simulated; k = exponent of the negative binomial distribution simulated.

^aColes et al. (1992); ^bDenwood et al. (2010); ^cTorgerson et al. (2014); ^dTarget 95%; ^eTarget 2.5%.

Table 5. Classification probabilities for the WAAVP, Bayescount and eggCounts methods to detect a reduced drug efficacy (R), efficacious treatment (E) or inconclusive (I) results of datasets with simulated FECR = 85% or FECR = 97% and varying levels of FEC aggregation on Day 0 ($k_{\text{Day}0}$) and Day 14 ($k_{\text{Day}14}$) post-treatment. Results are presented as the proportion of iterations detected by each method fulfilling the criteria for efficacious treatment (lower CI $\geq 90\%$; FECR% and upper CI $\geq 95\%$), reduced efficacy (FECR% and upper CI $< 95\%$; lower CI $< 90\%$) or inconclusive (none of the conditions met). A total of 1,000 iterations were performed for each combination of simulated FECR% and k (total 6,000 iterations). See text for further details on the simulated parameters.

Method	WAAVP ^a					Bayescount ^b					eggCounts ^c						
	$k_{\text{Day}0}$	$k_{\text{Day}14}$	R	E	I	R	E	I	R	E	I	R	E	I	R	E	I
85	2.6	1.4	0.798	0.002	0.200	0.699	0	0.301	0.955	0.003	0.042	0.955	0.003	0.042	0.955	0.003	0.042
85	1.2	0.8	0.626	0.009	0.365	0.550	0	0.450	0.904	0.020	0.076	0.904	0.020	0.076	0.904	0.020	0.076
85	0.6	0.01	0.108	0.645	0.247	0.188	0	0.812	0.241	0.722	0.037	0.241	0.722	0.037	0.241	0.722	0.037
97	2.6	1.4	0.793	0.001	0.206	0.3	0	0.700	0.871	0.004	0.125	0.871	0.004	0.125	0.871	0.004	0.125
97	1.2	0.8	0.645	0.002	0.353	0.156	0	0.844	0.840	0.018	0.142	0.840	0.018	0.142	0.840	0.018	0.142
97	0.6	0.1	0.733	0.019	0.248	0	0.063	0.937	0.832	0.123	0.045	0.832	0.123	0.045	0.832	0.123	0.045

FECR% = faecal egg count reduction percentage; k = exponent of the negative binomial distribution simulated.

^aColes et al. (1992); ^bDenwood et al. (2010); ^cTorgerson et al. (2014)

Supplementary Table 1. Anthelmintic prescriptions between 2002 and 2012 in the six farms included in the faecal egg count reduction tests. Data are presented as the estimated number of treatments (per prescription^a) / exact total number of cattle (per age group^b) at the date of prescription. A second column within the same year represents the same age group being re-treated during the year in question.

	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	
Farm #1 (Beef, conventional)	Calves - Heifers 1/39 (DOR, inj.)	- - -	- - -	- - -	- - -	- - -	- - -	78/78 (MOX, top) 108/108 (MOX, top)	- 44/98 (IVM, top)	31/56 (IVM, top) 53/53 (IVM, top) 53/53 (IVM, top)	13/73 (IVM, inj.) 69/69 (IVM, inj.)	77/77 (IVM, top) 44/86 (IVM, top)
Farm #2 (Dairy, organic)	Calves - Heifers - Cows 1/100 (ALB, bol.)	- - 16/102 (IVM, inj.)	- 28/51 (IVM, top)	- - -	- 1/58 (DOR, inj.) 16/105 (IVM, top)	30/57 (IVM, top) 56/56 (IVM, top)	- - -	- - -	- - -	- - -	- - -	- 22/54 (IVM, top) 16/102 (IVM, top)
Farm #3 (Dairy, conventional)	Calves - Heifers - Cows -	- - -	- - -	- - -	34/34 (IVM, top) 16/56 (IVM, top) 20/129 (IVM, top)	- - 16/129 (MOX, top)	- - -	- - -	- - -	51/51 (IVM, top)	- - -	- 22/89 (IVM, top)
Farm #4 (Dairy, organic)	Calves - Heifers - Cows -	- - -	- - -	- - -	- - -	- - -	30/30 (IVM, top) 20/57 (IVM, top)	- - -	- - -	- 2/60 (IVM, inj.)	- - -	41/41 (IVM+CLO, top) 56/56 (IVM+CLO, top) 40/151 (EPR, top)
Farm #5 (Beef, organic)	Calves 1/23 (IVM, inj.) Heifers -	- 1/27 (IVM, inj.)	- - -	- 22/77 (IVM, top)	50/89 (MOX, top) 22/69 (MOX, top)	3/86 (IVM, inj.) 24/69 (IVM, top)	1/70 (IVM, top) 1/83 (IVM, inj.)	3/74 (IVM, inj.) 1/107 (IVM, inj.)	57/110 (IVM, top) 26/79 (IVM, top)	33/57 (IVM, top) 47/110 (IVM, top)	- -	50/105 (IVM, top)
Farm #6 (Dairy, organic)	Calves - Heifers - Cows -	13/47 (IVM, top) 44/66 (IVM, top)	- 44/76 (IVM, top)	- 11/88 (IVM, top)	- 44/80 (IVM, top) 3/170 (MOX, top)	- 44/74 (IVM, top)	50/100 (IVM, top)	50/64 (IVM, top)	- 44/126 (IVM, top)	- 44/156 (IVM, top)	- 16/262 (IVM, top)	75/97 (IVM, top) 22/154 (IVM, top)

ALB = albendazole; CLR = clorsulon; DOR = doramectin; EPR = eprinomectin; IVM = ivermectin; LEV = levamisole; MOX = moxidectin; inj. = injectable; top. = topical; bol. = bolus

^aPrescription data were obtained from the Danish system for surveillance of the veterinary use of drugs for production animals (VetStat)

^bNumber of animals per age group in each farm at prescription date was retrieved from the CHR database (Ministry of Environment and Food of Denmark, <http://chr.fvst.dk>, accessed on 15 December 2015).

7.2 Manuscript II

Anthelmintic effects of forage chicory (*Cichorium intybus*) against gastrointestinal nematode parasites in experimentally infected cattle.

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Anthelmintic effects of forage chicory (*Cichorium intybus*) against gastrointestinal nematode parasites in experimentally infected cattle

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SUMMARY

Two experiments studied the effects of dietary chicory against gastrointestinal nematodes in cattle. In Experiment (Exp.) 1, stabled calves were fed chicory silage (CHI₁; $n = 9$) or ryegrass/clover hay (CTL₁; $n = 6$) with balanced protein/energy intakes between groups. After 16 days, all calves received 10 000 *Ostertagia ostertagi* and 66 000 *Cooperia oncophora* third-stage larvae (L3) [day (D) 0 post-infection (p.i.)]. In Exp. 2, calves were assigned to pure chicory (CHI₂; $n=10$) or ryegrass/clover (CTL₂; $n = 10$) pastures. After 7 days, animals received 20 000 *O. ostertagi* L3/calf (D0 p.i.) and were moved regularly preventing pasture-borne infections. Due to poor regrowth of the chicory pasture, CHI₂ was supplemented with chicory silage. At D40 p.i. (Exp. 1) and D35 p.i. (Exp. 2) calves were slaughtered for worm recovery. In Exp.1, fecal egg counts (FEC) were similar between groups. However, *O. ostertagi* counts were significantly reduced in CHI₁ by 60% (geometric mean; $P < 0.01$), whereas *C. oncophora* burdens were unaffected ($P = 0.12$). In Exp. 2, FEC were markedly lowered in CHI₂ from D22 p.i onwards ($P < 0.01$). *Ostertagia ostertagi* adult burdens were significantly reduced in CHI₂ by 66% ($P < 0.001$). Sesquiterpene lactones were identified only in chicory (fresh/silage). Chicory shows promise as an anti-*Ostertagia* feed for cattle and further studies should investigate its on-farm use.

Key words: *Cichorium intybus*, anthelmintic effects, cattle, *Ostertagia ostertagi*, *Cooperia oncophora*, sesquiterpene lactones, silage, ruminant.

INTRODUCTION

Parasitism with gastrointestinal nematodes (GIN) is ubiquitous in grazing cattle, and despite advances in vaccine development (Rinaldi and Geldhof, 2012; Vlaminck *et al.* 2015) on-farm control strategies largely rely on the regular use of anthelmintic drugs. As a result, reduced drug efficacy due to anthelmintic resistance (AR) is increasing among GIN of cattle in several countries (Sutherland and Leathwick, 2011; Gasbarre, 2014; Cotter *et al.* 2015; Geurden *et al.* 2015) threatening efforts to sustain animal health and productivity. The challenge posed by drug resistance urgently requires the development of effective alternatives, aiming at reducing the reliance on anthelmintics and lowering the selection pressure for AR on the still-effective drugs (Waller and Thamsborg, 2004; Sykes, 2010). Among other methods, the use of bioactive plants

with direct *in vivo* anthelmintic activity has shown promise as a complementary parasite control strategy in small ruminants (reviewed by Hoste *et al.* 2006, 2015), but comparable evidence in large ruminants is scarce (Sandoval-Castro *et al.* 2012; Shepley *et al.* 2015).

One bioactive plant investigated as a potential anthelmintic crop in small ruminants, particularly in temperate regions, is chicory (*Cichorium intybus* L., *Asteraceae*). Chicory is a perennial herb that has been traditionally used for human consumption in Europe and other regions of the world, and since the 1980s forage cultivars for livestock have been developed (Rumball, 1986; Barry, 1998; Li and Kemp, 2005). Studies with forage chicory offered at levels $\geq 80\%$ of the diet [based on dry matter (DM) intake] to parasitized lambs have consistently reported anthelmintic effects on abomasal worms (*Teladorsagia circumcincta* and *Haemonchus contortus*), while no apparent activity was observed against small intestinal nematodes (Scales *et al.* 1995; Marley *et al.* 2003; Athanasiadou *et al.* 2005; Tzamaloukas *et al.* 2005; Heckendorn *et al.* 2007). Direct anthelmintic effects of chicory are believed to derive from its content of plant secondary

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metabolites (PSM), namely condensed tannins (CT) and sesquiterpene lactones (SL), but previous research has detected only marginal CT levels in chicory, although *in vivo* anti-parasitic activity was still observed (Tzamaloukas *et al.* 2005; Heckendorn *et al.* 2007). Until now, no studies have investigated the SL content of forage chicory used in anthelmintic trials in ruminants, but *in vitro* effects of SL-containing extracts against GIN have been demonstrated (Foster *et al.* 2011a; Peña-Espinoza *et al.* 2015). Additionally, it is known that high-protein/energy diets can indirectly affect GIN infections in both large and small ruminants by increasing the animals' capacity to cope with, and eventually eliminate, helminth infections (Mansour *et al.* 1992; Van Houtert and Sykes, 1996; Coop and Kyriazakis, 2001; Tzamaloukas *et al.* 2006). Therefore, it is important to discern if the potential anti-parasitic effects of certain feeds are caused by direct anthelmintic activity or by indirect beneficial effects through improved nutrition (Hoste *et al.* 2015).

To date, only a single investigation has examined *in vivo* anthelmintic effects of chicory in cattle; the study reported that steers, naturally infected with GIN and grazing a mixed chicory-ryegrass pasture (24% chicory DM in the field), had no differences in fecal egg counts (FEC) or blood markers for GIN compared with infected controls grazing ryegrass (Marley *et al.* 2014). However, previous *in vivo* studies with sheep suggest that higher levels of chicory (possibly $\geq 50\%$ chicory DM in the diet) may be necessary to affect GIN in the host, but this has yet to be demonstrated in cattle.

The main objective of our study was to investigate the anthelmintic effects of forage chicory-rich diets against experimental infections with GIN in cattle. First, we studied the effect of ensiled chicory in stabled calves infected with *Ostertagia ostertagi* and *Cooperia oncophora*, the most common GIN of cattle in Northern Europe, in comparison with infected animals fed a control and protein/energy-balanced diet [Experiment (Exp.) 1]. Next, we compared the course of mono infections with *O. ostertagi* in calves grazing either pure forage chicory or ryegrass/clover paddocks, while carefully avoiding any re-infections from pasture (Exp. 2). In addition, the chemical profiles of the different feeds were examined, with focus on their content of CT and SL.

MATERIALS AND METHODS

Parasites

For Exp. 1, infective third-stage larvae (L3) were isolated from four nematode-naïve donor calves, orally inoculated with anthelmintic-susceptible strains of *O. ostertagi* and *C. oncophora* (kindly

provided by Prof. Dr. Janina Demeler, Freie Universität Berlin, Germany). Feces from the donor calves were collected rectally and used to prepare larval cultures (Roepstorff and Nansen, 1998). After 14 days, the cultures were baermannized and isolated L3 were pooled and maintained at 12 °C for 3 months until experimental infection. Two days prior to inoculation, motile L3 were selected by a further baermannization and 100 larvae were morphologically identified according to van Wyk and Mayhew (2013); the final inoculum consisted of 13% *O. ostertagi* and 87% *C. oncophora* L3. For Exp. 2, *O. ostertagi* L3 were isolated from two nematode-naïve donor calves mono-infected with an ivermectin susceptible *O. ostertagi* strain (Batch number: OOSG10, Ridgeway Research, UK). *Ostertagia ostertagi* L3 recovered from larval cultures were maintained for 3 months at 12 °C prior to inoculation and infective doses were prepared as described for Exp. 1.

Study designs and animals

Experiment 1. This experiment was performed between October–December 2013 as a controlled trial to study the course of experimental infections with *O. ostertagi* and *C. oncophora* in stabled calves fed ensiled forage chicory or a control roughage diet with balanced protein and energy levels. The study design is summarized in Fig. 1. Fifteen Danish Jersey bull-calves [mean (\pm S.D.) 92 \pm 26 days; body weight (BW): 92 \pm 21 kg], reared indoor on a commercial farm, were included in the trial. Upon arrival at the experimental farm, all calves were orally drenched with fenbendazole (Panacur®Vet, MSD Animal Health, 5 mg kg BW⁻¹) in order to ensure nematode-free conditions (later confirmed with zero FEC in all animals 1 week post treatment). After drenching, calves were stratified by BW and blocked in five subgroups of three calves with similar BW. Calves were housed in straw-bedded pens with concrete floor, daily removal of feces, external feeders and free access to water. Subgroups of three animals were randomly assigned to chicory (CHI₁, $n=9$; three subgroups of three calves) or control (CTL₁, $n=6$; two subgroups of three calves) feeding groups. The calves were fed twice daily *ad libitum* with chicory silage (CHI₁) or ryegrass/clover hay (CTL₁) throughout the 56 days of experiment (see Feeds – Exp. 1). Additionally, both groups were offered a commercial concentrate to balance crude protein (CP) and metabolizable energy (ME) intakes between groups throughout the trial. The DM intake per pen was calculated on a daily basis by subtracting the residual feed unconsumed by each subgroup from the feed that was offered the previous day. The estimated daily DM, CP and ME intakes during the preceding 2/3 days were adjusted for the mean BW of each subgroup and used to define exact

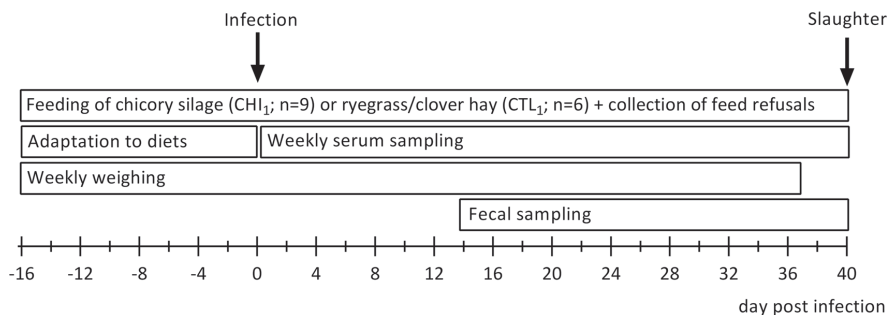


Fig. 1. Study design of Experiment 1 investigating the course of experimental infections with *Ostertagia ostertagi* and *Cooperia oncophora* in stabled calves fed with chicory silage and concentrate (CHI₁) or ryegrass/clover hay and concentrate (CTL₁) during 56 days.

daily amounts of concentrate for the next 2/3 days, in order to balance the expected CP (and ME) intakes between all subgroups. Feeding was introduced with a relatively high level of concentrate at the start of the trial and gradually replaced with increasing amounts of chicory silage or ryegrass/clover hay. At the day of infection, the concentrate represented ~25% of the DM intake in all subgroups. After 16 days of feed adaptation [day (D) 0 post-infection (p. i.)] all calves were orally inoculated using a plastic syringe with a total of 76 000 L3 (10 000 *O. ostertagi* L3 and 66 000 *C. oncophora* L3) of the inoculum described above, administered over three consecutive days. At D40 p.i., all calves were euthanized by captive bolt pistol and exsanguinated for worm recovery.

Experiment 2. This study was designed as a grazing trial to investigate the course of experimental infections with *O. ostertagi* in calves grazing either pure forage chicory or ryegrass/clover (control) pastures, without supplementary concentrate feeding. The trial was conducted during 42 days in August–September 2014 and the study design is outlined in Fig. 2. Twenty Danish Holstein bull-calves (136 ± 16 days; 153 ± 24 kg BW), raised in a commercial farm without previous access to pasture, were used. At arrival, all animals were orally drenched with fenbendazole as described for Exp. 1 to remove any potential nematode infection from the farm of origin (confirmed with zero FEC in all calves 1 week post treatment). After treatment, calves were stratified by BW and randomly assigned to a chicory (CHI₂) or a control (CTL₂) group of ten animals each. The CHI₂ and CTL₂ groups were allocated to a pure forage chicory or ryegrass/clover pasture, respectively (see *Feeds – Exp. 2*). Animals had *ad libitum* access to water throughout the trial. After 7 days of diet adaptation (D0 p.i.), all calves were orally infected using a plastic syringe with a total of 20 000 *O. ostertagi* L3 of the inoculum described above, administered over two consecutive days.

Infective doses were designed according to published guidelines for anthelmintic evaluation (Wood *et al.* 1995). Calves in CHI₂ and CTL₂ groups were strip grazed within their respective swards throughout the experiment. From D14 p.i. until the end of the trial, calves were moved to a new area with 5 days intervals, thus avoiding any potential reinfection on pasture. By the second half of Exp. 2, the pure chicory pasture was at the reproductive (flowering) stage. Calves in the CHI₂ group quickly consumed the chicory leaves and the upper parts of stems and flowers in the new grazing strip areas, which, in addition to poor leaf regrowth, resulted in a low amount of chicory on the pasture by D16 p.i. Therefore, the CHI₂ group was supplemented *ad libitum* with chicory silage (prepared on May 2014, see *Feeds – Exp. 2*) during the last 19 days of the trial (D16–D35 p.i.). The CTL₂ group was not supplemented during the experiment. At D35 p.i., all animals were euthanized by captive bolt pistol and exsanguinated for worm recovery.

Ethics

Animal trials were approved by the Animal Experiments Inspectorate of the Danish Ministry of Environment and Food (License j. No. 2013-15-2934-00763) and were conducted in the experimental farm of the University of Copenhagen (Tåstrup, Denmark). Calves in both experiments were inspected at least twice daily during the entire study periods to detect any unusual behaviour or signs of clinical disease.

Feeds

Experiment 1. Forage chicory (cv. Spadona) was sown on 7 May 2013 as a pure sward [7.8 kg chicory seeds per hectare (ha^{-1})] on a 0.9 ha field at the experimental farm of the University of Copenhagen ($55^{\circ}67'48''\text{N}$, $12^{\circ}29'75''\text{E}$). The soil

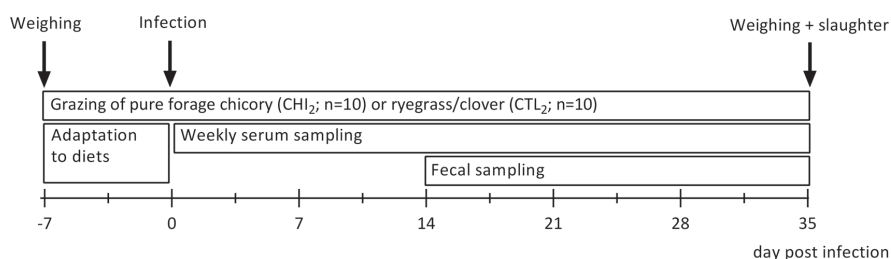


Fig. 2. Study design of Experiment 2 investigating the course of experimental infections with *Ostertagia ostertagi* in calves grazing pure forage chicory (CHI₂) or ryegrass/clover (CTL₂) during 42 days.

(moraine clay loam) was fertilized with 50 kg N ha⁻¹ in early May 2013. No herbicides were used. Chicory leaves were cut on 23 September 2013 and let to wilt for 24 h on the field to prepare chicory silage. No further cuts were performed in the chicory sward during 2013. At harvest, chicory leaves represented 55% of the total DM in the field. Unsovn plant species in the sward were mainly chamomile (*Matricaria recutita*) with some mugwort (*Artemisia vulgaris*) and shepherd's purse (*Capsella bursa-pastoris*). After wilting, a silage inoculant (Silosolve[®]AS, 200 g inoculant 75 L⁻¹ water, CHR Hansen, Denmark) was manually sprayed on the harvested field and the plant material was wrapped in bales of 300 kg. Ensiled chicory underwent anaerobic fermentation for an average of 8.5 weeks until use in the trial. Ryegrass/clover (*Lolium perenne*/*Trifolium repens*) hay was used as control feed. Throughout Exp. 1, all animals were supplemented with a concentrate for calves (Gronmix[®], Danish Agro, Denmark) in order to balance CP and ME intakes between groups.

Experiment 2. In early March 2014, the same forage chicory sward used for Exp. 1 was fertilized with 80 kg N ha⁻¹. On May 2014, when the chicory plants were at the vegetative stage and chicory leaves constituted >90% of the total DM in the field, the sward was cut for ensiling. This chicory silage was produced to provide a potential supplement for calves allocated to grazing on chicory during Exp. 2. By early July 2014, the chicory plants were progressing into their reproductive stage and a cut 40 cm above the soil level was performed in order to remove long stems and flowers and prepare the field for the grazing trial. As a control pasture, a 0.8 ha ryegrass/clover sward located at the same farm and left ungrazed in 2013 was used. The control field was cut in mid-June 2014 and allowed to regrow for 2 months. At the start of Exp. 2, chicory leaves, stems and flowers constituted 90% of the total DM in the sward (chicory leaves = ~60%, chicory stem and flowers = ~40%) while the remaining 10% were a mix of unsovn clover and shepherd's purse. Ryegrass

constituted 80% of the total DM in the control field, with the rest comprised mainly of clover.

Feed sampling and analyses. In Exp. 1, samples from chicory silage, ryegrass/clover hay and concentrate were collected for chemical analyses on D - 9 p.i. Calves in the CTL₁ group consumed hay uniformly, whereas CHI₁ animals selectively consumed the soft chicory leaves in the silage while refusing thicker stems from unsovn plant species (see *Feeds - Exp. 1*). Therefore, samples of the offered chicory silage and of the residual (unconsumed) feed were dried for 24 h at 90 °C for correction of DM chicory intake. In Exp. 2, fresh forage samples from the chicory and ryegrass/clover swards were collected at the start of the trial (D - 7 p.i.) and 1 week before the end of the trial (D28 p.i.) by harvesting 20 samples (50 × 50 cm², cut 5–8 cm above soil level) at similar intervals following a diagonal line across each sward and pooled by field. The supplementary chicory silage prepared in 2014 was sampled at the start of Exp. 2. A rough estimate of the daily DM intake of CHI₂ and CTL₂ groups from pasture was calculated by comparing the available DM in the fields at the first and last sampling. While the intake of chicory silage by CHI₂ calves was inferred from the number of opened silage bales and the estimated silage not consumed by the animals from D16 until D34 p.i. (corrected for DM).

All experimental feeds were submitted to an accredited laboratory (Eurofins Steins Laboratorium Ltd., Holstebro, Denmark) for analyses of DM, CP, net energy lactation (NEL), amino acids absorbed in the intestine (AAT), *in vitro* organic matter digestibility (OMD), ash and neutral detergent fibre (NDF). *In vivo* OMD% of ryegrass/clover (hay and pasture) was estimated by near-infrared spectrometry, while *in vitro* OMD% of chicory (silage and fresh) and concentrate were estimated according to Tilley and Terry (1963) or by the *in vitro* enzyme digestible organic matter (EFOS) method (Volden, 2011), respectively. Feeds were adjusted to *in vivo* OMD% following the regression formulas recommended by the Nordic Feed Evaluation System (NorFor; Volden, 2011) as

follows: (a) chicory silage *in vivo* OMD% = $6.73 + 0.950 \times \text{in vitro OMD\%}$, (b) fresh chicory *in vivo* OMD% = $4.1 + 0.959 \times \text{in vitro OMD\%}$, (c) concentrate *in vivo* OMD% = $5.38 + 0.867 \times \text{in vitro OMD\%}$. ME from all feeds was calculated from NEL values divided by 0.65 (Nielsen *et al.* 2003) and expressed as mega joule (MJ) kg^{-1} DM. The content of CT and SL were investigated in samples of all experimental feeds collected at the start of Exp. 1 and 2; CT were analysed using the acetone–butanol–HCl method (Grabber *et al.* 2013), while total and individual SL were characterized as described by Peña-Espinoza *et al.* (2015).

Animal measurements

In Exp. 1, calves were individually weighed weekly from arrival until D37 p.i., while animals in Exp. 2 were weighed at arrival and on the day of slaughter, using an electronic scale. Blood samples from each calf were collected by jugular venipuncture at weekly intervals from D0 p.i. until slaughter in both trials. Recovered serum was analysed for total protein (TP), albumin (ALB), inorganic phosphate (IP) and serum pepsinogen. Serum pepsinogen was analysed according to Dorny and Vercruyse (1998) and expressed as units of tyrosine (U Tyr/L).

Parasitological analyses

FEC and larval cultures. In Exp. 1 and 2, fecal samples were collected rectally from each animal every 2–3 days from D14 p.i. until slaughter and analysed for strongyle FEC using an accredited, modified McMaster technique with a sensitivity of 5 eggs per g (epg) of feces (Henriksen and Aagard, 1975). The DM percentage of each fecal sample (fecal DM%) was determined by drying 3 g of fresh feces for 24 h at 90 °C. Subsequently, fecal DM% was used to correct individual FEC per g of fecal DM (FEC_{DM}) using the formula:

$$\text{FEC}_{\text{DM}} = \text{FEC} \times (100/\text{fecal DM\%})$$

In Exp. 1, larval cultures were prepared for each subgroup at D19, D26 and D33 p.i. by mixing 10 g of fresh feces from each animal into one pool which was cultured for 14 days as described by Roepstorff and Nansen (1998). After baermannization, 100 L3 from each subgroup were identified as *O. ostertagi* or *C. oncophora* according to van Wyk and Mayhew (2013).

Worm counts. Immediately after slaughter the abomasum and small intestine (Exp. 1) or only abomasum (Exp. 2) from each animal were removed and opened into individual buckets for worm recovery. The organs were thoroughly washed with warm

(38 °C) saline solution (0.9% NaCl) until total volumes of 5 L (abomasum) or 10 L (small intestine). Subsamples of $2 \times 10\%$ and $2 \times 1\%$ of stirred abomasal and small intestinal washed content, respectively (Exp. 1), and subsamples of $2 \times 2\%$ of stirred abomasal washed content (Exp. 2), were collected using a 25- μm -pore sieve. All worms collected in the two abomasal subsamples (Exp. 1 and 2) and on the two small intestinal subsamples (Exp. 1) per animal were counted. Male worms ($n = 20$ per animal) from abomasal and small intestinal samples were further transferred to microscope slides, added a drop of lactic acid (Sigma L1250, 10% v/v in distilled water) and observed by light microscopy to verify the nematode species (Barth and Visser, 1991). In Exp. 1, chemical digestion of abomasal mucosa for detection of inhibited *O. ostertagi* L4 was performed as described by Wood *et al.* (1995) in one animal from each subgroup (three CHI₁ and two CTL₁ calves).

Ostertagia ostertagi female fecundity in mono-infected calves. In Exp. 2, the per capita fecundity of *O. ostertagi* females was calculated by dividing the FEC_{DM} at slaughter by the total number of females recovered from the same animal and was expressed as the number of eggs female⁻¹ per g of fecal DM.

Statistical analyses

All statistical analyses were performed in R version 3.2.2 (R Core Team, 2015). Untransformed FEC_{DM}/worm count data were analysed using generalized linear models, which have been advocated to increase statistical power and reduce risks of type I and II errors when analysing overdispersed (non-normally distributed) data, compared with the modelling of (log-)transformed counts (Wilson and Grenfell, 1997; Paterson and Lello, 2003). In Exp. 1, differences in FEC_{DM} between feeding groups at each sampling point were analysed by generalized linear-mixed effect (*glmer*, R package 'lme4') models assuming a negative binomial (*nb*) distribution and testing the effect of feeding group (CHI₁ or CTL₁) as fixed (main) factor and subgroup of three calves as random factor; the random factor (or random effect) included the variation of data between subgroups within the same feeding group (fixed factors) in the models. In addition, the cumulative FEC_{DM} of each animal at the end of the trial was calculated according to Vercruyse *et al.* (1993) and compared between feeding groups using the Wilcoxon rank sum test. Worm counts were analysed by fitting *glmer.nb* models, testing the effect of feeding group as fixed factor and subgroup as random factor. Fecal DM%, BW gains and serum pepsinogen, TP, ALB and IP levels were analysed as repeated measures by fitting linear mixed-effects

Table 1. Chemical composition of feeds offered to stabled calves infected with *Ostertagia ostertagi* and *Cooperia oncophora* in Experiment 1

	Experimental feeds		
	Chicory silage	Ryegrass/clover hay	Concentrate ^a
DM (%)	35.7	86.5	89.6
CP (g kg ⁻¹ DM)	93	84	204
ME (MJ kg ⁻¹ DM)	6.5	7.2	10.5
AAT (g kg ⁻¹ DM)	54	63	114
OMD (% of DM)	62	59	83
Crude ash (g kg ⁻¹ DM)	207	49	70
Sand (g kg ⁻¹ DM)	72	0.0	0.0
NDF (g kg ⁻¹ DM)	326	577	286
CT ^b (% DM)	n.d.	n.d.	n.d.
SL ^c (g kg ⁻¹ DM)	12.3	0.0	0.0
DI-LAC	2.1	0.0	0.0
LAC	1.8	0.0	0.0
8-DOL	1.2	0.0	0.0
DI-8-DOL	3.8	0.0	0.0
DI-LCP	2.6	0.0	0.0
LCP	0.8	0.0	0.0

DM, dry matter; CP, crude protein; ME, metabolizable energy; AAT, aminoacid absorbed in the intestine; OMD = *in vivo* organic matter digestibility; NDF, neutral detergent fibre; CT, condensed tannins; SL, sesquiterpene lactones; LAC, lactucin; DI-LAC, 11, 13-dihydro-lactucin; 8-DOL, 8-deoxylactucin; DI-8-DOL, 11, 13-dihydro-8-deoxylactucin; LCP, lactucopicrin; DI-LCP, 11, 13-dihydro-lactucopicrin; n.d., not detected.

^a Grønmix[®] (Danish Agro).

^b According to Grabber *et al.* (2013; estimated detection limit = 0.2% of CT in DM).

^c According to Peña-Espinoza *et al.* (2015).

(*lme*, R package 'nlme') models, testing the effect of feeding group, time and their interaction as fixed factors, initial values as covariate and animal within subgroup as random factor (to account for the correlation of repeated measures from the same animal and the variation of data between subgroups of the same feeding group). Data acquired on a subgroup-basis were not subjected to statistical analysis. Total DM, CP and ME intakes recorded daily from each subgroup were adjusted to 100 kg BW and are presented as arithmetic means (\pm s.d.). Percentage of *O. ostertagi* and *C. oncophora* L3 in larval cultures pooled by subgroup are reported as arithmetic means (\pm s.d.). In Exp. 2, differences in FECDM between feeding groups were analysed similarly as described for Exp.1, with the exception that all the models were fitted using generalized linear models (*glm*, package 'MASS'), assuming a *nb* distribution. Worm counts and per capita female fecundity were analysed by *glm.nb* models with feeding group (CHI₂ or CTL₂) as fixed factor. Fecal DM%, BW gains and serum pepsinogen, TP, ALB and IP levels were analysed as repeated measures by fitting *lme* models, including group, time and their interaction as fixed factors, initial values as covariates and animal as random factor (to include correlation of data from the same calf). Repeated measures (not count) data were power-transformed if required to fit normality assumptions using the boxcox command (package 'MASS'). All models were

validated by analysis of residual plots. A level of $P < 0.05$ was considered significant.

RESULTS

Experiment 1

Feed analyses. Chemical compositions of the experimental feeds are presented in Table 1. Chicory silage had the lowest DM% of all feeds. Crude protein, ME, AAT and OMD% were similar in chicory silage and ryegrass/clover hay. Chicory silage had a markedly higher content of crude ash (207 g kg⁻¹ DM) compared with ryegrass/clover hay (49 g kg⁻¹ DM) and concentrate (70 g kg⁻¹ DM). Sand, which was only detected in chicory silage, resulted in corrected ash content (crude ash minus sand) of 135 g kg⁻¹ DM chicory silage. Ryegrass/clover hay had the highest NDF content (577 g kg⁻¹ DM), followed by chicory silage (326 g kg⁻¹ DM) and concentrate (286 g kg⁻¹ DM). Condensed tannins were not detected in any of the experimental feeds. Sesquiterpene lactones were identified only in chicory silage (12.3 g total SL kg⁻¹ DM), with 11,13-dihydro-8-deoxylactucin (DI-8-DOL) and 11,13-dihydro-lactucopicrin (DI-LCP) as the main SL (Table 1).

Feed intake. Mean daily intakes of DM, CP and ME over the experiment are presented in Table 2.

Table 2. Estimated dry matter, crude protein and metabolizable energy intakes in stabled calves fed with chicory silage and concentrate (Chicory) or ryegrass/clover hay and concentrate (Control) in Experiment 1

Group	Daily mean (\pm s.d.) intake 100 kg BW ⁻¹	Day post infection						
		0	7	14	21	28	35	40
Chicory (n = 9)	DM (kg)	1.7 (\pm 0.1)	2.3 (\pm 0.2)	2.4 (\pm 0.1)	3.0 (\pm 0.2)	3.1 (\pm 0.1)	3.1 (\pm 0.2)	3.0 (\pm 0.2)
	CP (g)	192.7 (\pm 26.5)	264.9 (\pm 10.4)	278.8 (\pm 2.9)	356.2 (\pm 17.6)	369.5 (\pm 2.9)	375.3 (\pm 12.7)	362.6 (\pm 12.1)
	ME (MJ)	12.9 (\pm 1.4)	17.8 (\pm 0.9)	18.9 (\pm 0.5)	23.1 (\pm 0.2)	23.0 (\pm 0.2)	23.4 (\pm 1.1)	22.9 (\pm 1.1)
Control (n = 6)	DM (kg)	2.0 (\pm 0.2)	2.5 (\pm 0.2)	2.6 (\pm 0.5)	2.9 (\pm 0.4)	3.1 (\pm 0.5)	3.2 (\pm 0.6)	3.1 (\pm 0.6)
	CP (g)	236.9 (\pm 5.2)	278.6 (\pm 3.4)	298.0 (\pm 31.0)	343.7 (\pm 18.0)	375.6 (\pm 20.9)	372.0 (\pm 17.4)	366.7 (\pm 14.0)
	ME (MJ)	16.8 (\pm 0.9)	20.0 (\pm 0.9)	21.3 (\pm 3.2)	24.3 (\pm 2.3)	26.2 (\pm 2.9)	26.5 (\pm 3.1)	25.9 (\pm 2.9)

Data is based on daily measurements from subgroups of three calves and presented as daily mean intakes (\pm s.d.) of the preceding 5/7 days for a standard calf of 100 kg body weight (BW).

DM, dry matter; CP, crude protein; ME, metabolizable energy.

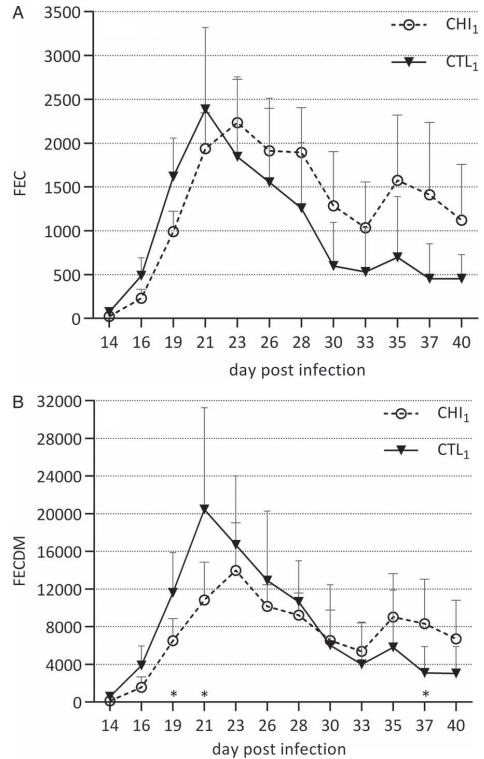


Fig. 3. Arithmetic mean fecal egg counts (FEC; A) and FEC per g of fecal dry matter (FECDM; B) in stabled calves experimentally infected with *Ostertagia ostertagi* and *Cooperia oncophora* and fed with chicory silage and concentrate (CHI₁) or ryegrass/clover hay and concentrate (CTL₁) in Experiment 1. Error bars indicate s.d. *P < 0.05.

A comparable increase in DM intake was recorded in CHI₁ and CTL₁ animals during the trial. At D0 p.i., the DM intake in the CHI₁ group was composed of 76% chicory silage, whereas the CTL₁ consumed 80% ryegrass/clover hay. At D39 p.i. (day before slaughter), the DM intake constituted 74% chicory silage and 67% ryegrass/clover hay in the CHI₁ and CTL₁ groups, respectively. The remaining component of the diets was the concentrate, which was readily eaten by all subgroups. Balancing of protein and energy intakes resulted in marginally higher CP and ME intakes in CTL₁ calves, but with comparable levels between groups throughout the experiment.

Parasitology. Fecal DM% was not significantly affected by feeding group, time or their interaction (P > 0.10), but a significant effect of initial values (covariate) was confirmed (P < 0.001). Arithmetic mean FEC and FECDM are presented in Fig. 3. Fecal strongyle eggs were detected from D14 p.i. onwards in calves from both groups, with significant differences in FECDM between groups only at D19,

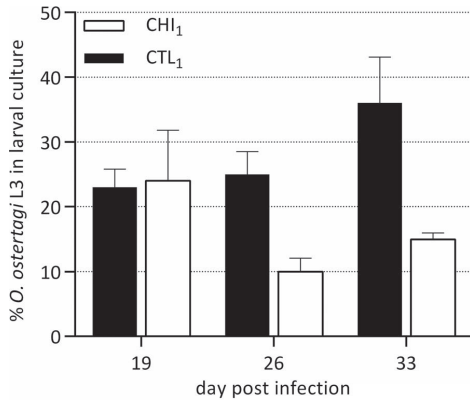


Fig. 4. Arithmetic mean percentage of *Ostertagia ostertagi* third-stage larvae (L3) identified in pooled fecal larval cultures of stabled calves experimentally infected with *O. ostertagi* and *Cooperia oncophora* and fed with chicory silage and concentrate (CHI₁) or ryegrass/clover hay and concentrate (CTL₁) in Experiment 1. Error bars indicate s.d.

D21 and D37 p.i. Higher FECDM was observed initially in CTL₁ (from D14 to D28 p.i.), but following a marked reduction by D33 p.i. this group had the lowest FECDM during the last days of the trial. In comparison, a more stable egg excretion was observed in CHI₁ throughout the study period. However, the cumulative FECDM during the entire experiment was not significantly different between groups [mean (± s.d.) CHI₁ = 204 280 (±65 264) vs CTL₁ = 225 128 (±94 501) epg of fecal DM; *P* = 0.6]. Mean percentages of *O. ostertagi* L3 in pooled larval cultures from each group are presented in Fig. 4. At D19 p.i., similar proportions of *O. ostertagi* L3 were detected in CHI₁ and CTL₁ animals. On D26 and D33 p.i., in contrast, a markedly lower number of *O. ostertagi* L3 was observed in pooled larval cultures from the CHI₁ calves.

Post-mortem worm recovery in CHI₁ and CTL₁ groups is summarized in Table 3. Calves fed with chicory silage had a significant arithmetic mean reduction of 52% (geometric mean reduction 60%) in the number of adult *O. ostertagi*, in comparison with CTL₁ animals (*P* < 0.01). In contrast, adult counts of *C. oncophora* were not statistically different between groups (*P* = 0.12). In CTL₁ calves, the number of adult *O. ostertagi* corresponded to 14% of the total worm counts in this group, which agrees with the proportion of *O. ostertagi* L3 in the original infective dose (13%). No inhibited *O. ostertagi* L4 were observed in the digested abomasal mucosa of the investigated calves.

Biochemical parameters. Very low serum pepsinogen values were detected in CHI₁ and CTL₁ calves throughout the trial (Fig. 5). Mean (±s.d.)

Table 3. Worm counts of adult *Ostertagia ostertagi* and *Cooperia oncophora* detected post-mortem in stabled calves fed with chicory silage and concentrate (Chicory) or ryegrass/clover hay and concentrate (Control) in Experiment 1

Group	<i>O. ostertagi</i> adult worm counts			<i>C. oncophora</i> adult worm counts		
	Males	Females	Total	Males	Females	Total
Chicory (n = 9)	878* (175–1915)	984** (250–1800)	1862* (425–3715)	17 537 (6750–25 450)	19 267 (11 200–25 750)	36 804 (17 950–51 200)
Control (n = 6)	1747 (1205–2235)	2152 (1555–2960)	3899 (2760–5195)	10 392 (800–29 200)	12 750 (800–27 450)	23 142 (1600–56 650)

Results are presented as arithmetic means (range). **P* < 0.05; ***P* < 0.01.

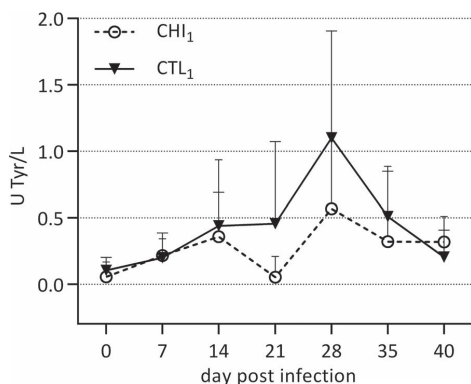


Fig. 5. Arithmetic mean serum pepsinogen levels (in units tyrosine; U Tyr/L) in stabled calves experimentally infected with *Ostertagia ostertagi* and *Cooperia oncophora* and fed with chicory silage and concentrate (CHI₁) or ryegrass/clover hay and concentrate (CTL₁) in Experiment 1. Error bars indicate s.d.

serum pepsinogen levels from D7 until D40 p.i. in CHI₁ and CTL₁ were 0.31 (\pm 0.2) U Tyr/L and 0.48 (\pm 0.2) U Tyr/L, respectively. Time had a significant influence on pepsinogen levels over the study period ($P < 0.001$), with a peak in both groups on D28 p.i., but neither group nor group \times time interaction affected pepsinogen values ($P > 0.18$). Serum TP and ALB levels were not affected by group or group \times time interaction ($P > 0.12$), but were significantly influenced by time ($P < 0.001$), reflected in decreasing TP and ALB values in both groups from D0 p.i. until slaughter (data not shown). Similarly, serum IP levels were significantly influenced by time ($P < 0.001$), but additionally group \times time interaction had a significant effect ($P = 0.006$), with decreasing IP values during the trial in both groups but with consistently lower IP levels in the CTL₁ group (data not shown).

Animal weights. Bodyweight gains were significantly affected by time and group \times time interaction ($P < 0.001$ and $P = 0.004$, respectively), with increasing BW in both groups but higher growth rates in CHI₁ calves. Similar mean (\pm s.d.) daily growth rates were observed in both groups until D19 p.i. [CHI₁ = 383 (\pm 116) *vs* CTL₁ = 325 (\pm 236) g day⁻¹]. In contrast, from D19 until D37 p.i., CHI₁ and CTL₁ groups had mean growth rates of 811 (\pm 59) and 391 (\pm 132) g day⁻¹, respectively. In addition, initial BW (covariate) had a significant effect on growth rates, with a lower weight gain in lighter calves, irrespective of group ($P < 0.001$).

Experiment 2

Feed analyses and feed intake. Chemical profiles of the experimental feeds used in Exp. 2 are presented

in Table 4. The chicory sward had markedly lower DM%, CP and NDF levels compared with the ryegrass/clover pasture. Chicory silage had a slightly higher CP content compared with fresh chicory, which reflects the high protein content of the chicory sward on May 2014 (142.6 g CP kg⁻¹ DM) when it was cut for ensiling. Comparable ME levels were identified in all feeds, while fresh chicory and chicory silage had higher ash content in comparison with fresh ryegrass/clover. No CT were detected in any of the experimental feeds. Sesquiterpene lactones were identified in fresh chicory (22.5 g total SL kg⁻¹ DM) and chicory silage (16.8 g total SL kg⁻¹ DM). In fresh chicory, the main SL detected were lactucin (LAC) and 11,13-dihydro-lactucin (DI-LAC), while in chicory silage the predominant SL were DI-LCP and DI-8-DOL. Based on sward measurements, CHI₂ calves had an estimated mean daily intake of 5.7 kg DM calf⁻¹ during the trial, with increasing consumption of chicory silage from D16 p.i. onwards and ~70% of the DM intake being chicory silage at D34 p.i., whereas CTL₂ animals had an estimated daily intake of 6.9 kg DM ryegrass/clover calf⁻¹ during the entire experiment.

Parasitology. Fecal DM% was significantly affected by group, time and their interaction and covariate ($P < 0.05$). Similar fecal DM content was observed between groups until D22 p.i., followed by markedly lower fecal DM% in CTL₂ animals from D24 p.i. onwards, with a mean (\pm s.d.) fecal DM% at slaughter of 18.72 (\pm 1.9) and 12.40 (\pm 2.2)% in CHI₂ and CTL₂ groups, respectively ($P < 0.001$). Arithmetic mean FEC and FECDM are presented in Fig. 6. Fecal strongyle eggs were detected from D17 p.i. onwards in animals from both groups. At D20 p.i., mean FECDM of CHI₂ and CTL₂ were 1589 and 1474 epg, respectively ($P = 0.7$). From this point, mean FECDM in the CHI₂ group was rapidly and significantly reduced until the end of the trial: at D35 p.i., CHI₂ animals had a 65% lower mean FECDM compared with CTL₂ calves ($P < 0.01$). Cumulative FECDM during the entire experiment was significantly reduced in the CHI₂ group [mean (\pm s.d.) CHI₂ = 12 870 (\pm 8136) *vs* CTL₂ = 22 260 (\pm 5671) epg of fecal DM; $P < 0.01$].

Post-mortem worm recovery of CHI₂ calves revealed significantly lower numbers of *O. ostertagi* males [mean (range) CHI₂ = 2106 (283–5425) *vs* CTL₂ = 4692 (2775–6800) male worms; $P < 0.01$] and females [CHI₂ = 2256 (617–4100) *vs* CTL₂ = 5386 (3700–6700) female worms; $P < 0.001$], compared with CTL₂ animals. Calves fed pure forage chicory had a highly significant arithmetic mean reduction of 57% (geometric mean reduction 66%) in the total worm burden of *O. ostertagi*, compared with control animals [mean (range) CHI₂ = 4362 (900–9525) *vs* CTL₂ = 10 078 (6475–13 325) total

Table 4. Chemical composition of feeds consumed by calves in Experiment 2

	Experimental feeds		
	Fresh forage chicory	Chicory silage	Ryegrass/clover
DM (%)	11.5 (±4.3)	23.5	22.0 (±2.1)
CP (g kg ⁻¹ DM)	98.2 (±32.1)	117.2	159.8 (±4.1)
ME (MJ kg ⁻¹ DM)	7.2 (±0.9)	8.6	8.3 (±0.5)
AAT (g kg ⁻¹ DM)	67.5 (±6.4)	66.0	80.1 (±4.2)
OMD (% of DM)	63.1 (±9.5)	74.5	67.1 (±4.7)
Crude ash (g kg ⁻¹ DM)	150.2 (±30.7)	147.2	108.6 (±5.2)
Sand (g kg ⁻¹ DM)	17.0 (±1.4)	5.0	22.0 (±11.3)
NDF (g kg ⁻¹ DM)	260.5 (±54.5)	326.4	430.2 (±24.3)
CT ^a (% DM)	n.d.	n.d.	n.d.
SL ^b (g kg ⁻¹ DM)	22.5	16.8	0.0
DI-LAC	5.9	3.4	0.0
LAC	6.1	2.7	0.0
8-DOL	3.1	1.5	0.0
DI-8-DOL	3.5	4.1	0.0
DI-LCP	3.5	4.6	0.0
LCP	0.4	0.5	0.0

Results for fresh forage chicory and ryegrass/clover are presented as means (±s.d.), summarizing the samples collected from the swards at D - 7 and D28 p.i.

DM, dry matter; CP, crude protein; ME, metabolizable energy; AAT, aminoacid absorbed in the intestine; OMD, *in vivo* organic matter digestibility; NDF, neutral detergent fibre; CT, condensed tannins; SL, sesquiterpene lactones; LAC, lactucin; DI-LAC, 11, 13-dihydro-lactucin; 8-DOL, 8-deoxylactucin; DI-8-DOL, 11, 13-dihydro-8-deoxylactucin; LCP, lactucopicrin; DI-LCP, 11, 13-dihydro-lactucopicrin; n.d., not detected.

^a According to Grabber *et al.* (2013; estimated detection limit = 0.2% of CT in DM).

^b According to Peña-Espinoza *et al.* (2015).

worms; $P < 0.001$]. Per capita fecundity of *O. ostertagi* females was not statistically different between groups [mean (range) CHI₂ = 0.19 (0.05–0.39) vs CTL₂ = 0.25 (0.12–0.39) eggs female⁻¹ per g fecal DM; $P = 0.24$].

Biochemical parameters. Mean serum pepsinogen levels are illustrated in Fig. 7, which shows almost identical profiles in both groups with a common peak at D21 p.i. and comparable levels at slaughter [mean (±s.d.) CHI₂ = 1.02 ± 0.4 U Tyr/L; CTL₂ = 1.01 ± 0.2 U Tyr/L]. Only time had a significant influence on pepsinogen levels ($P < 0.001$), but not group or group × time interaction ($P > 0.8$). Similarly, serum TP and ALB levels were only significantly affected by time ($P < 0.001$), with equivalent and declining serum concentrations in both groups over the trial, and a significant effect of covariate ($P < 0.05$) (data not shown). Serum IP concentrations were significantly influenced by covariate ($P = 0.005$), group ($P = 0.008$) and time ($P < 0.001$), illustrated by constantly lower IP values in the CTL₂ group, but with decreasing IP levels in both groups until D21 p.i. and comparable values between groups at slaughter (data not shown).

Animal weights. Bodyweight gains were significantly influenced by group, time and group × time interaction ($P < 0.001$), with mean (±s.d.) growth rates of 374 (±160) g day⁻¹ and 783 (±229) g day⁻¹ in CHI₂ and CTL₂ calves, respectively.

DISCUSSION

In the present study, we demonstrated that cattle experimentally infected with GIN and fed forage chicory-rich diets had significant reductions in worm burdens of *O. ostertagi* (Exp. 1 and 2), but not of *C. oncophora* (Exp. 1), compared with infected animals fed control diets. In Exp. 1, calves fed with chicory silage had largely similar fecal egg excretions compared with control animals, but markedly lower numbers of *O. ostertagi* L3 in larval cultures from D26 p.i. These findings were confirmed in Exp. 2, where calves mono-infected with *O. ostertagi* and fed pure forage chicory (fresh and silage) had a significant reduction of nematode eggs in feces from D22 p.i. until slaughter, compared with control animals, without significant differences in the fecundity of female worms. In addition, SL were detected in both experiments only in forage chicory (fresh and silage), while CT were not identified in any of the experimental feeds.

Differences in the FECDM of chicory-fed animals between experiments can be interpreted based on the distinct worm populations in the two trials. In Exp. 1 (animals co-infected with *O. ostertagi* and *C. oncophora*), the similar cumulative FECDM in CHI₁ and CTL₁ calves is explained by the unaffected *C. oncophora* population, which represented 87% of the infective dose and was responsible for approximately 65–90% of the egg excretion (based on larval cultures). In contrast, the lower number of *O. ostertagi* L3 in larval cultures of

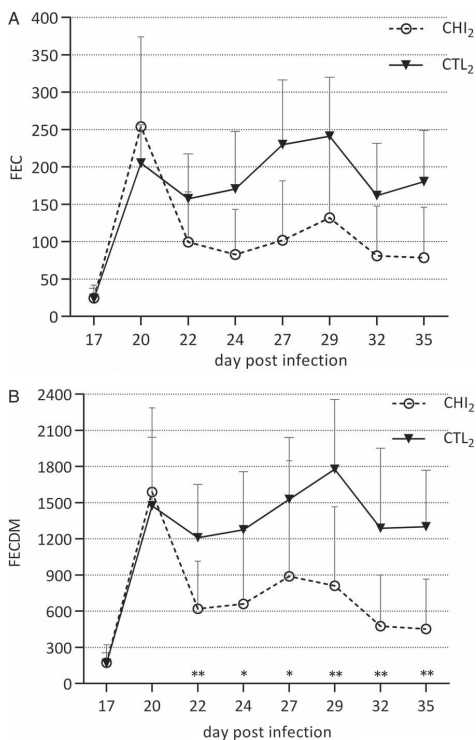


Fig. 6. Arithmetic mean fecal egg counts (FEC; A) and FEC per g of fecal dry matter (FECMD; B) in calves experimentally infected with *Ostertagia ostertagi* and grazing pure forage chicory (CHI₂) or ryegrass/clover (CTL₂) in Experiment 2. Error bars indicate S.D. **P* < 0.05; ***P* < 0.01.

chicory-fed calves from D26 p.i. onwards probably derived from a decreased fecal egg excretion of this species, as a consequence of the reduced number of *O. ostertagi* females in these animals. Towards the end of Exp. 1, CTL₁ calves had a lower mean FECMD compared with CHI₁ animals (only significant at D37 p.i.). Post-mortem analyses demonstrated lower mean *C. oncophora* counts in CTL₁ animals compared with CHI₁ calves, although with large variations in both groups and no statistical difference in *C. oncophora* burdens between groups. While in Exp. 2 (animals mono-infected with *O. ostertagi*), the significantly lower cumulative FECMD of the CHI₂ group was likely a result of the reduced number of female *O. ostertagi* in these calves, later confirmed post-mortem. Nevertheless, FECMD (or FEC) alone as an indicator of egg excretion should be evaluated carefully while it is affected by the fecal output, which in turn is influenced by the DM% and digestibility of the feed and by the DM intake of the animal (Heckendorn *et al.* 2007). In our case, the accurate DM intake and fecal output of the calves in Exp. 2

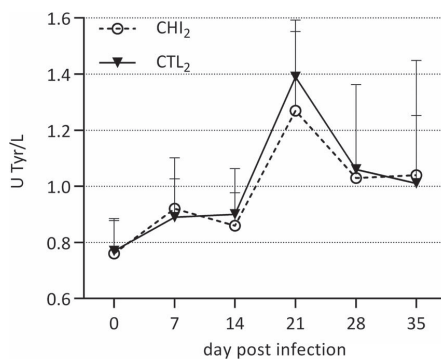


Fig. 7. Arithmetic mean serum pepsinogen levels (in units tyrosine; U Tyr/L) in calves experimentally infected with *Ostertagia ostertagi* and grazing pure forage chicory (CHI₂) or ryegrass/clover (CTL₂) in Experiment 2. Error bars indicate S.D.

was not monitored, and therefore, further studies in which the total daily fecal egg production is examined are required to confirm the effect of chicory feeding on *O. ostertagi* egg excretion and pasture contamination.

Mean serum pepsinogen levels of chicory and controls animals in both trials were <1.5 U Tyr/L, below the threshold of 2.0 U Tyr/L related with subclinical *O. ostertagi* infections (Vercruysse and Claerebout, 2001), reflecting the low infective doses with this species in our study. Consequently, our results are insufficient to determine possible consequences of chicory feeding in the pathophysiology of *O. ostertagi* in the infected animals, although no differences were detected in serum pepsinogen concentrations between feeding groups, despite the significantly lower burden of adult *O. ostertagi* in chicory-fed animals in both experiments.

Considering the comparable protein/energy intakes between groups in Exp. 1 and the low nutritional plane of chicory-fed calves in Exp. 2 (further discussed below), in addition to the slow development of immunity against *O. ostertagi* in parasite-naïve calves (Gasbarre, 1997) and the minimal loss of worms before 50 days p.i. observed in similar infections (Michel, 1969), the present study demonstrates that forage chicory can selectively reduce *O. ostertagi* infections in cattle. Our results are consistent with earlier studies in sheep, which reported that chicory-rich diets induced selective anthelmintic effects only against abomasal nematodes (without affecting per capita female fecundity), but not towards small intestinal species (Scales *et al.* 1995; Marley *et al.* 2003; Athanasiadou *et al.* 2005; Tzamaloukas *et al.* 2005; Heckendorn *et al.* 2007). Similar to our findings in Exp. 1, Marley *et al.* (2014) reported no differences in FECMD and serum pepsinogen levels in cattle naturally infected with mixed species of GIN and grazing either

chicory/ryegrass or pure ryegrass for 126 days. But in contrast to our study, the cited authors did not observe any effect of chicory/ryegrass feeding on the number of *O. ostertagi* L3 in larval cultures. Post-mortem worm counts were not conducted. In the study by Marley *et al.* (2014), the animals grazed a sward with only 24% of chicory DM, much lower than the chicory levels offered in our study ($\geq 70\%$ of the daily DM intake), which may explain the lack of anthelmintic effect in that experiment. Previous studies have also reported a lower development and recovery of infective L3 from forage chicory swards that could reduce the larval challenge of animals grazing chicory (Moss and Vlassoff, 1993; Marley *et al.* 2006). However, this is unlikely to have contributed to the reduction in worm burdens in our Exp. 2 due to the artificial inoculation of the animals and the frequent relocation of the calves to clean areas, thus preventing any reinfection on pasture.

In relation to the PSM analysed in the experimental feeds, SL were only detected in chicory (ensiled and fresh) in both experiments. To our knowledge, this is the first report of the SL profile in forage chicory cv. Spadona. Previous studies have reported SL levels in other forage chicory cultivars, with levels ranging between 1.55 and 15.24 g total SL kg⁻¹ DM (Foster *et al.* 2006, 2011b). However, our findings are not directly comparable with these results because of differences between the studies regarding chicory cultivars, cultivation methods, locations, season, stage of plant growth and screening methods used, all of which can affect the detection and concentration of SL in chicory (Rees and Harborne, 1985; Foster *et al.* 2011b; Ferioli and D'Antuono, 2012). Whereas CT were not identified in any of the experimental feeds; however, we cannot conclude if CT were totally absent in chicory or if CT were present below the estimated detection limit of the method (0.2% CT in DM). Nevertheless, the undetectable levels of CT in all the experimental feeds indicate that CT were not required for the observed anthelmintic effects and suggest that SL and/or other PSM could be responsible for the anti-parasitic activity of forage chicory. Several phytochemicals other than SL have been reported in chicory, e.g. derivatives of hydroxycinnamic acids (e.g. chicoric, chlorogenic and caffeic acid), coumarins (e.g. chicoriin) and flavonoids (Rees and Harborne, 1985; Heimler *et al.* 2009; Sinkovič *et al.* 2015), and their potential significance in the anthelmintic effects of chicory needs further investigation.

Although our study was not designed to identify the specific nematode stages affected by chicory feeding, the similar FECDM in chicory and control groups in Exp. 2 up until D20 p.i. suggests that dietary chicory may not interfere with the exsheathment, establishment and development of the larval stages of *O. ostertagi* into egg-laying

adults; rather, the marked drop in FECDM observed between D20 and D22 p.i. indicates that chicory may selectively (and rapidly) affect the survival of adult worms. Recently, we showed that SL-containing chicory extracts do not interfere with the artificial exsheathment of *O. ostertagi* L3 *in vitro*, but can induce a rapid paralysis of adult *O. ostertagi* in a dose-dependent manner (Peña-Espinoza *et al.* 2015). Previously, the *in vivo* activity of forage chicory against adult stages but not towards incoming L3 has been reported in sheep infected with *T. circumcincta* (Tzamaloukas *et al.* 2005). In Exp. 1, the absence of inhibited *O. ostertagi* L4 in the investigated CHI₁ and CTL₁ calves does not necessarily indicate a lack of arrested larvae in the other (unscreened) animals; however, it suggests that feeding with chicory silage did not interfere with the larval development into adult worms, although this remains to be demonstrated.

The mechanisms behind the selective activity of dietary chicory against abomasal worms but not towards small intestinal nematodes are unknown, but comparable findings in sheep by authors in different countries (Scales *et al.* 1995; Marley *et al.* 2003; Tzamaloukas *et al.* 2005) suggest that the mechanisms are preserved across ruminant species, abomasal nematodes, and perhaps, chicory cultivars and cultivation conditions. In our study, the lack of activity towards *C. oncophora* infections may indicate that there is none in chicory; interestingly, preliminary experiments from our group have demonstrated direct and dose-dependent *in vitro* activity of purified chicory extracts against adult *C. oncophora* (Peña-Espinoza *et al.* unpublished results). Hence, one explanation for the lack of *in vivo* activity against intestinal worms could be that PSM, such as SL, do not reach the small intestine in concentrations sufficient to exert their activity. Differences in the stability of PSM in different gut compartments have been documented for CT (Hoste *et al.* 2006), and studies suggest that SL may be more stable at low pH (Ferreira and Gonzalez, 2008; Saroglou *et al.* 2008). Currently, the understanding of the pharmacokinetics of SL in livestock is rudimentary, but previous studies have indicated that these phytochemicals can be metabolized by ruminants (cited by Barry, 1998; Ferreira and Gonzalez, 2008). Undoubtedly, more research is required to fully elucidate the fate of dietary SL from chicory in the digestive tract of ruminants and to establish how factors such as changing pH and host metabolism can affect the concentrations, and eventually the anthelmintic activity, of SL *in vivo*. Nonetheless, the selective effect of chicory against abomasal nematodes may also be explained by local changes induced only in the abomasum that result in worm expulsion, rather than or in combination with a direct effect on the worms. In traditional medicine, chicory has been described to have

'stomach tonic' effects in humans (Ahmed *et al.* 2003; Hitova and Melzig, 2014), which are believed to stimulate the appetite by increasing gastric secretions in the stomach. However, currently no studies have explored these effects in ruminants.

Experiments 1 and 2 had different study designs, which resulted in distinct voluntary intakes of chicory and animal growth between trials. In Exp. 1, we examined the effects of a chicory-rich diet against GIN while excluding nutritional-related effects by matching protein/energy intakes between groups with a commercial concentrate. As a result, chicory constituted ~70% of the daily DM intake in Exp. 1. In comparison, in Exp. 2 we investigated the anthelmintic effects of a pure chicory diet (>90% chicory DM intake) against *O. ostertagi* infections, without balancing nutritional levels between groups, but aiming for calves to exercise their voluntary feeding intake in the respective swards. In both experiments, chicory was readily eaten by the animals. In Exp. 1, significantly higher growth rates were detected in calves fed chicory silage, particularly from D19 p.i. onwards, and despite the comparable protein/energy intakes in both groups and the short duration of the experiment. Whether the markedly higher weight gain in chicory calves from D19 p.i. was related to the lower *O. ostertagi* worm burden in these animals, and/or to a higher nutritional value (e.g. in macro/micronutrients) of chicory silage compared with ryegrass/clover hay, was not further investigated. In contrast, weight gains in Exp. 2 were significantly lower in chicory-fed animals compared with controls. This was most likely due to the poor nutritional quality and low amounts of chicory on the pasture and thus lower DM intake (i.e. 5.7 vs 6.9 kg DM calf⁻¹ for CHI₂ and CTL₂, respectively). In Exp. 2, our main problems were the rapid progression of the chicory plants into the reproductive stage from June 2014 onwards (2nd year sward), the very limited regrowth following the cut of the chicory sward in early July 2014 and the lower protein content of fresh chicory, as compared with the ryegrass/clover pasture. By the second half of Exp. 2 (late August 2014), the pure chicory sward was in the reproductive stage with few areas still containing chicory leaves, and therefore the calves needed to be supplemented with chicory silage. Despite this supplementary feeding, which also had lower protein content than the control pasture, the poor growth of the CHI₂ group was not reversed during the study period. Thus, close monitoring of the DM available in pure chicory swards grazed by parasitized calves, particularly during periods of rapid reproductive stem growth, seems to be critical to secure that animal production goals are met.

Finally, our findings suggest that the inclusion of a high percentage of fresh or conserved (ensiled) chicory in the diet of infected cattle could represent

a complementary and selective anti-*Ostertagia* strategy that may reduce infection levels and the need of drug treatments. *Ostertagia ostertagi* is considered the most important GIN infecting grazing cattle in temperate regions and cases of AR in this species have been reported (Edmonds *et al.* 2010; Gasbarre, 2014; Geurden *et al.* 2015). Moreover, forage chicory can be cultivated in a wide-range of climates (Li and Kemp, 2005), including areas where the conditions for other bioactive forages like tanniferous plants may be limited, as Northern Europe (Høgh-Jensen *et al.* 2006; Kidane *et al.* 2014). Our results also demonstrated that the conservation of chicory leaves as silage did not obstruct their anthelmintic effects, and therefore, this may be an acceptable method to preserve and use chicory in farms, independent of seasonal availability. Moreover, and in comparison with fresh forage, chicory silage prepared for Exp. 1 and 2 (in two consecutive years) contained increased concentrations of DI-LCP and DI-8-DOL; the latter SL has been recently linked with higher anthelmintic potency against *O. ostertagi in vitro* (Peña-Espinoza *et al.* 2015). Further studies are needed to clarify how ensiling affects the SL profile and the anthelmintic activity of chicory silage in comparison with fresh leaves and to evaluate the most appropriate on-farm use of forage chicory.

In conclusion, feeding forage chicory (≥70% of chicory DM in the diet) led to a significant reduction in worm burdens and FEC of *O. ostertagi* in experimentally infected calves in two independent experiments, while no apparent activity was observed against *C. oncophora*. Sesquiterpene lactones were identified and individually characterized in ensiled and fresh forage chicory, suggesting that these PSM may contribute to the observed anthelmintic effects of dietary chicory *in vivo*. However, further research is needed to fully elucidate the exact compound(s) and mechanism(s) behind the *in vivo* anti-parasitic effects of chicory in ruminants. Forage chicory (fresh and ensiled) was readily consumed by calves and we demonstrated that anti-parasitic effects were preserved despite ensiling, which may facilitate the use of chicory in integrated parasite control strategies on farms to reduce the reliance on anthelmintic drugs.

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REFERENCES

- Ahmed, B., Al-Howiriny, T. and Siddiqui, A.B. (2003). Antihepatotoxic activity of seeds of *Cichorium intybus*. *Journal of Ethnopharmacology* **87**, 237–240.
- Athanasidou, S., Tzamaloukas, O., Kyriazakis, I., Jackson, F. and Coop, R.L. (2005). Testing for direct anthelmintic effects of bioactive forages against *Trichostrongylus colubriformis* in grazing sheep. *Veterinary Parasitology* **127**, 233–243.
- Barry, T. (1998). The feeding value of chicory (*Cichorium intybus*) for ruminant livestock. *Journal of Agricultural Science* **131**, 251–257.
- Barth, D. and Visser, M. (1991). *Magen-Darm-Nematoden des Rindes, Diagnostischer Atlas*. Ferdinand Enke Verlag, Stuttgart, Germany.
- Coop, R.L. and Kyriazakis, I. (2001). Influence of host nutrition on the development and consequences of nematode parasitism in ruminants. *Trends in Parasitology* **17**, 325–330.
- Cotter, J.L., Van Burgel, A. and Besier, R.B. (2015). Anthelmintic resistance in nematodes of beef cattle in south-west Western Australia. *Veterinary Parasitology* **207**, 276–284.
- Dorny, P. and Vercruyse, J. (1998). Evaluation of a micro method for the routine determination of serum pepsinogen in cattle. *Research in Veterinary Science* **65**, 259–262.
- Edmonds, M.D., Johnson, E.G. and Edmonds, J.D. (2010). Anthelmintic resistance of *Ostertagia ostertagi* and *Cooperia oncophora* to macrocyclic lactones in cattle from the western United States. *Veterinary Parasitology* **170**, 224–229.
- Feroli, F. and D'Antuono, L.F. (2012). An update procedure for an effective and simultaneous extraction of sesquiterpene lactones and phenolics from chicory. *Food Chemistry* **135**, 243–250.
- Ferreira, J. and Gonzalez, J. (2008). Chemical and biological stability of artemisinin in bovine rumen fluid and its kinetics in goats (*Capra hircus*). *Brazil Journal of Veterinary Parasitology* **17**, 103–109.
- Foster, J.G., Clapham, W.M., Belesky, D.P., Labreux, M., Hall, M.H. and Sanderson, M.A. (2006). Influence of cultivation site on sesquiterpene lactone composition of forage chicory (*Cichorium intybus* L.). *Journal of Agricultural and Food Chemistry* **54**, 1772–1778.
- Foster, J., Cassida, K. and Turner, K. (2011a). *In vitro* analysis of the anthelmintic activity of forage chicory (*Cichorium intybus* L.) sesquiterpene lactones against a predominantly *Haemonchus contortus* egg population. *Veterinary Parasitology* **180**, 298–306.
- Foster, J., Cassida, K. and Sanderson, M. (2011b). Seasonal variation in sesquiterpene lactone concentration and composition of forage chicory (*Cichorium intybus* L.) cultivars. *Grass and Forage Science* **66**, 424–433.
- Gasbarre, L.C. (1997). Effects of gastrointestinal nematode infection on the ruminant immune system. *Veterinary Parasitology* **72**, 327–337.
- Gasbarre, L.C. (2014). Anthelmintic resistance in cattle nematodes in the US. *Veterinary Parasitology* **204**, 3–11.
- Geurden, T., Chartier, C., Fanke, J., di Regalbono, A.F., Traversa, D., von Samson-Himmelstjerna, G., Demeler, J., Vanimisetti, H.B., Bartram, D.J. and Denwood, M.J. (2015). Anthelmintic resistance to ivermectin and moxidectin in gastrointestinal nematodes of cattle in Europe. *International Journal for Parasitology: Drugs and Drug Resistance* **5**, 163–171.
- Grabber, J.H., Zeller, W.E. and Mueller-Harvey, I. (2013). Acetone enhances the direct analysis of procyanidin- and prodelpinidin-based condensed tannins in lotus species by the butanol-HCl-iron assay. *Journal of Agricultural and Food Chemistry* **61**, 2669–2678.
- Heckendorn, F., Häring, D.A., Maurer, V., Senn, M. and Hertzberg, H. (2007). Individual administration of three tanniferous forage plants to lambs artificially infected with *Haemonchus contortus* and *Cooperia curticei*. *Veterinary Parasitology* **146**, 123–134.
- Heimler, D., Isolani, L., Vignolini, P. and Romani, A. (2009). Polyphenol content and antiradical activity of *Cichorium intybus* L. from biodynamic and conventional farming. *Food Chemistry* **114**, 765–770.
- Henriksen, S. and Aagard, K. (1975). A simple flotation and McMaster technique. *Nordisk Veterinær Medicin* **28**, 392–397.
- Hitova, A. and Melzig, M. (2014). *Cichorium intybus* L. *Zeitschrift für Phytotherapie* **35**, 198–202.
- Hoste, H., Jackson, F., Athanasiadou, S., Thamsborg, S.M. and Hoskin, S.O. (2006). The effects of tannin-rich plants on parasitic nematodes in ruminants. *Trends in Parasitology* **22**, 253–261.
- Hoste, H., Torres-Acosta, J.F.J., Sandoval-Castro, C., Mueller-Harvey, I., Sotiraki, S., Louvandini, H., Thamsborg, S.M. and Terrill, T.H. (2015). Tannin containing legumes as a model for nutraceuticals against digestive parasites in livestock. *Veterinary Parasitology* **212**, 5–17.
- Højh-Jensen, H., Nielsen, B. and Thamsborg, S.M. (2006). Productivity and quality, competition and facilitation of chicory in ryegrass/legume-based pastures under various nitrogen supply levels. *European Journal of Agronomy* **24**, 247–256.
- Kidane, A., Sorheim, K., Eik, L.O. and Steinshamn, H. (2014). Growth and chemical composition of chicory and performance of lambs grazing chicory relative to grass-clover mixtures. *Acta Agriculturae Scandinavica, Section A — Animal Science* **64**, 233–242.
- Li, G. and Kemp, P. (2005). Forage chicory (*Cichorium intybus* L.): a review of its agronomy and animal production. *Advances in Agronomy* **88**, 187–222.
- Mansour, M.M., Dixon, J.B., Rowan, T.G. and Carter, S.D. (1992). Modulation of calf immune responses by *Ostertagia ostertagi*: the effect of diet during trickle infection. *Veterinary Immunology and Immunopathology* **33**, 261–269.
- Marley, C., Cook, R., Keatinge, R., Barrett, J. and Lampkin, N. (2003). The effect of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) on parasite intensities and performance of lambs naturally infected with helminth parasites. *Veterinary Parasitology* **112**, 147–155.
- Marley, C.L., Cook, R., Barrett, J., Keatinge, R. and Lampkin, N.H. (2006). The effects of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) when compared with perennial ryegrass (*Lolium perenne*) on ovine gastrointestinal parasite development, survival and migration. *Veterinary Parasitology* **138**, 280–290.
- Marley, C., Fychan, R., Davies, J., Scollan, N., Richardson, R., Theobald, V., Genever, E., Forbes, A. and Sanderson, R. (2014). Effects of chicory/perennial ryegrass swards compared with perennial ryegrass swards on the performance and carcass quality of grazing beef steers. *PLoS ONE* **9**, e86259.
- Michel, J.F. (1969). The regulation of egg output by *Ostertagia ostertagi* in calves infected once only. *Parasitology* **59**, 767–774.
- Moss, R.A. and Vlassoff, A. (1993). Effect of herbage species on gastrointestinal roundworm populations and their distribution. *New Zealand Journal of Agricultural Research* **36**, 371–375.
- Nielsen, B.K., Thamsborg, S.M. and Kristensen, T. (2003). Feed supplements for young dairy breed calves after turn-out to pasture: effect on weight gain and subclinical coccidiosis in organic production systems. *Acta Agriculturae Scandinavica, Section A—Animal Sciences* **53**, 1–10.
- Paterson, S. and Lello, J. (2003). Mixed models: getting the best use of parasitological data. *Trends in Parasitology* **19**, 370–375.
- Peña-Espinoza, M., Boas, U., Williams, A.R., Thamsborg, S.M., Simonsen, H.T. and Enemark, H.L. (2015). Sesquiterpene lactone containing extracts from two cultivars of forage chicory (*Cichorium intybus*) show distinctive chemical profiles and *in vitro* activity against *Ostertagia ostertagi*. *International Journal for Parasitology: Drugs and Drug Resistance* **5**, 191–200.
- R Core Team (2015). *R: A Language and Environment for Statistical Computing*. 3-2-2. R Foundation for Statistical Computing, Vienna, Austria. <https://www.r-project.org/>
- Rees, S. and Harborne, J. (1985). The role of sesquiterpene lactones and phenolics in the chemical defence of the chicory plant. *Phytochemistry* **24**, 2225–2231.
- Rinaldi, M. and Geldhof, P. (2012). Immunologically based control strategies for ostertagiosis in cattle: where do we stand?. *Parasite Immunology* **34**, 254–264.
- Roepstorff, A. and Nansen, P. (1998). *Epidemiology, Diagnosis and Control of Helminth Parasites of Swine*. FAO Animal Health Manual, Rome, Italy.
- Rumball, W. (1986). Grasslands Puna' chicory (*Cichorium intybus* L.). *New Zealand Journal of Experimental Agriculture* **14**, 105–107.
- Sandoval-Castro, C.A., Torres-Acosta, J.F.J., Hoste, H., Salem, A.Z.M. and Chan-Pérez, J.I. (2012). Using plant bioactive materials to control gastrointestinal tract helminths in livestock. *Animal Feed Science and Technology* **176**, 192–201.
- Saroglou, V., Demetzos, C. and Skaltsa, H. (2008). Effects of pH and temperature on the stability of sesquiterpene lactones. *Planta Medica* **74**, PC102.
- Scales, G.H., Knight, T.L. and Saville, D.J. (1995). Effect of herbage species and feeding level on internal parasites and production performance of grazing lambs. *New Zealand Journal of Agricultural Research* **38**, 237–247.

- Shepley, E., Vasseur, E., Bergeron, R., Villeneuve, A. and Lachance, S. (2015). Short communication: birdsfoot trefoil as a preventative treatment for gastrointestinal nematodes in pastured dairy heifers. *Canadian Journal of Animal Science* **95**, 533–537.
- Sinkovič, L., Demšar, L., Žnidarčič, D., Vidrih, R., Hribar, J. and Treutter, D. (2015). Phenolic profiles in leaves of chicory cultivars (*Cichorium intybus* L.) as influenced by organic and mineral fertilizers. *Food Chemistry* **166**, 507–513.
- Sutherland, I. A. and Leathwick, D. M. (2011). Anthelmintic resistance in nematode parasites of cattle: a global issue? *Trends in Parasitology* **27**, 176–181.
- Sykes, A. R. (2010). Host immune responses to nematodes: benefit or cost? Implications for future development of sustainable methods of control. *Revista Brasileira de Zootecnia* **39**, 376–382.
- Tilley, J. and Terry, R. (1963). A two stage technique for the in vitro digestion of forage crops. *Journal of the British Grasslands Society* **18**, 104–111.
- Tzamaloukas, O., Athanasiadou, S., Kyriazakis, I., Jackson, F. and Coop, R. L. (2005). The consequences of short-term grazing of bioactive forages on established adult and incoming larvae populations of *Teladorsagia circumcincta* in lambs. *International Journal for Parasitology* **35**, 329–335.
- Tzamaloukas, O., Athanasiadou, S., Kyriazakis, I., Huntley, J. F. and Jackson, F. (2006). The effect of chicory (*Cichorium intybus*) and sulla (*Hedysarum coronarium*) on larval development and mucosal cell responses of growing lambs challenged with *Teladorsagia circumcincta*. *Parasitology* **132**, 419–426.
- Van Houtert, M. F. J. and Sykes, A. R. (1996). Implications of nutrition for the ability of ruminants to withstand gastrointestinal nematode infections. *International Journal for Parasitology* **26**, 1151–1167.
- van Wyk, J. and Mayhew, E. (2013). Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: a practical lab guide. *Onderstepoort Journal of Veterinary Research* **80**, Art. #539.
- Vercruysse, J. and Claerebout, E. (2001). Treatment vs non-treatment of helminth infections in cattle: defining the threshold. *Veterinary Parasitology* **98**, 195–214.
- Vercruysse, J., Dorny, P., Hong, C., Harris, T. J., Hammet, N. C., Smith, D. G. and Weatherley, A. J. (1993). Efficacy of doramectin in the prevention of gastrointestinal nematode infections in grazing cattle. *Veterinary Parasitology* **49**, 51–59.
- Vlaminck, J., Borloo, J., Vercruysse, J., Geldhof, P. and Claerebout, E. (2015). Vaccination of calves against *Cooperia oncophora* with a double-domain activation-associated secreted protein reduces parasite egg output and pasture contamination. *International Journal for Parasitology* **45**, 209–213.
- Volden, H. (ed.) (2011). *NorFor - The Nordic Feed Evaluation System*. European Federation of Animal Science publication No. 130. Wageningen Academic Publishers, The Netherlands.
- Waller, P. J. and Thamsborg, S. M. (2004). Nematode control in “green” ruminant production systems. *Trends in Parasitology* **20**, 493–497.
- Wilson, K. and Grenfell, B. T. (1997). Generalized linear modelling for parasitologists. *Parasitology Today* **13**, 33–38.
- Wood, I., Amaral, N., Bairden, K., Duncan, J., Kassai, T., Malone, J., Pankavich, J., Reinecke, R., Slocombe, O., Taylor, S. and Vercruysse, J. (1995). World Association for the Advancement of Veterinary Parasitology (WAAVP) second edition of guidelines for evaluating the effectiveness of anthelmintics in ruminants (bovine, ovine, caprine). *Veterinary Parasitology* **58**, 181–213.

7.3 Manuscript III

Sesquiterpene lactone containing extracts from two cultivars of forage chicory (*Cichorium intybus*) show distinctive chemical profiles and *in vitro* activity against *Ostertagia ostertagi*.

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Sesquiterpene lactone containing extracts from two cultivars of forage chicory (*Cichorium intybus*) show distinctive chemical profiles and *in vitro* activity against *Ostertagia ostertagi*



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ABSTRACT

The study investigated direct anthelmintic effects of sesquiterpene lactones (SL)-containing extracts from forage chicory against free-living and parasitic stages of *Ostertagia ostertagi*. Freeze-dried leaves from chicory cultivars 'Spadona' and 'Puna II' were extracted using methanol/water. Total SL were further fractionated by solid-phase extraction and resulting extracts were characterised by high-performance liquid chromatography (HPLC). *O. ostertagi* eggs from faeces of mono-infected calves were hatched and L1 were used in a larval feeding inhibition assay (LFIA), while cultured L3 were used in a larval exsheathment inhibition assay (LEIA). Adult worms were immediately recovered after slaughter and used for motility inhibition assays (AMIA). Electron microscopy (EM) was performed on adult *O. ostertagi* exposed to 1000 µg extract mL⁻¹ of both chicory cultivars. In all assays, decreasing concentrations of SL-containing extracts in PBS (1% DMSO) were tested in replicates with 1% DMSO in PBS as negative controls. HPLC demonstrated similar concentrations of most SL in both extracts. However, Spadona-extract contained significantly higher concentrations of 11, 13-dihydro-8-deoxylactucin ($P = 0.01$), while Puna II-extract had increased levels of 11, 13-dihydro-8-deoxylactucin ($P < 0.0001$). In the LFIA, both extracts reduced larval feeding at increasing concentrations, but Spadona-extract showed higher potency confirmed by significantly lower EC₅₀ ($P < 0.0001$). In the LEIA, neither of the two extracts interfered with the exsheathment of L3 ($P > 0.05$). In the AMIA, both SL-containing extracts induced a dose-dependent effect but Spadona-extract showed greater activity and exerted faster worm paralysis than Puna II-extract with significantly lower EC₅₀ ($P < 0.0001$). No cuticular damage was observed by EM in worms exposed to any of the extracts. We have demonstrated that SL-containing extracts from forage chicory can inhibit feeding of free-living larvae and exert direct effects against parasitic stages of *O. ostertagi*. Our results may contribute to the identification of natural anti-parasitic compounds and to interpret the *in vivo* anthelmintic effects of forage chicory.

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1. Introduction

In the face of increasing anthelmintic resistance in parasitic nematodes of ruminants, the development of efficacious novel control strategies that can reduce the reliance on anthelmintics are urgently needed (Waller and Thamsborg, 2004). Feeding of animals with certain plants has shown much promise as a complementary

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parasite control option, with *in vivo* results for several forages demonstrating anthelmintic efficacy against a number of gastrointestinal nematode species (Githiori et al., 2006; Hoste et al., 2015). Most of the anti-parasitic activity of these plants is believed to derive from the presence of biologically active compounds, designated as plant secondary metabolites (PSM). Consequently, a comprehensive study of PSM can help to understand the anthelmintic effects of bioactive crops in the field and to potentially detect novel anti-parasitic molecules. One of the plants investigated as a potential anthelmintic forage for ruminants is chicory (*Cichorium intybus*), which has been linked with significant reductions in adult worm burdens and egg excretion of abomasal nematodes in sheep (Scales et al., 1994; Marley et al., 2003; Tzamaloukas et al., 2005; Heckendorn et al., 2007). Chicory is known to produce a group of biologically active terpenoids, the sesquiterpene lactones (SL), in both leaves and roots (Rees and Harborne, 1985; Price et al., 1990; Foster et al., 2011; Graziani et al., 2015). In chicory leaves, SL are present as glycosides (bound to carbohydrates) and as free molecules (Feroli et al., 2015). Sesquiterpene lactones are PSM thought to be involved in the protection of plants against herbivore attacks and for signalling between plants (Gershenzon and Dudareva, 2007). In biological systems, SL have demonstrated antioxidant, antibiotic, anti-cancerigenous and antiprotozoan properties (Bischoff et al., 2004; Cavin et al., 2005; Schmidt, 2006; Barrera et al., 2013).

To date, two studies have provided preliminary evidence of *in vitro* activity of SL from chicory against free-living stages of parasitic nematodes of ruminants. Molan et al. (2003) reported inhibitory effects of crude SL-extracts from chicory roots on free-living larvae of *Dictyocaulus viviparus* and mixed-species of gastrointestinal nematodes isolated from red deer, however, detection and characterisation of SL in the tested extracts was not performed. Foster et al. (2011) analysed extracts containing free SL from two chicory cultivars and found differences in the concentration of individual SL and inhibitory effects towards the eclosion of a predominantly *Haemonchus contortus* egg population. In contrast with free-living stages occurring in the environment, parasitic stages in the host have different biochemical characteristics and detoxification mechanisms, thus the same compound(s)/PSM can exert different effects and affect distinct targets in free-living or adult stages (O'Grady and Kotze, 2004; Hoste et al., 2015). As yet, no studies have explored the direct effects of well-characterised SL-containing extracts from forage chicory on parasitic stages, which are the expected primary targets of dietary SL in the host.

The main objective of our study was to investigate and compare the direct effects of SL-containing extracts isolated from two forage chicory cultivars on key biological processes of free-living and parasitic stages of *Ostertagia ostertagi*. This abomasal nematode is considered to be the most pathogenic and economically important parasite infecting grazing cattle in temperate areas of the world (Nansen, 1993; Fox, 1993; Charlier et al., 2014). Two chicory cultivars were included in the study in order to account for variations in the activity of extracts derived from different plant cultivars, previously described in chicory (Foster et al., 2011). In relation, a detailed characterisation of the SL profile in the tested chicory extracts was performed.

2. Materials and methods

2.1. Plant materials

Chicory cultivar (cv.) 'Spadona' and cv. 'Puna II' were sown on 7 May 2013 in two separate fields at the experimental farm of the University of Copenhagen (Tåstrup, Denmark, 55°67'48"N, 12°29'75"E). Both fields were fertilized with 50 kg N/ha in spring

(early May). The soil type in both fields was moraine clay loam. Chicory cv. Spadona was sown as a pure sward (7.8 kg seeds/ha) whereas cv. Puna II was mixed with timothy (*Phleum pratense*; seed rate: 6 kg chicory + 6 kg timothy seeds/ha). Leaves from both chicory cultivars were hand-picked on 23 July 2013 and stored at -20 °C in the dark until extraction of SL. At the moment of collection, all chicory plants were at the vegetative stage.

2.2. Extraction of sesquiterpene lactones from forage chicory

Sesquiterpene lactones were isolated and purified from chicory leaves according to Feroli and D'Antuono (2012), with some modifications. This extraction method selectively separates total (free and unbound) SL from phenols and other plant compounds. Chicory leaves were freeze-dried overnight, ground into powder, after which pulverized leaf material (2 g) from each cultivar was weighed separately into two 50 mL tubes. Extraction solvent (30 mL) of methanol/Milli-Q-H₂O (4/1; v/v) containing 2% formic acid (v/v) was added to each tube. The tubes were vortexed for 1 min, sonicated in a water bath at room temperature for 10 min and centrifuged (10 min, 1540 g). After centrifugation, supernatants from the same cultivar were pooled in a 200 mL round-bottomed glass flask (1 flask per cultivar). Remaining leaf material in the 50 mL tubes was extracted three more times as described above and supernatants were pooled in the corresponding round-bottomed flask. Collected supernatants were concentrated under reduced pressure at 35 °C to evaporate methanol and formic acid and freeze-dried overnight to remove Milli-Q-H₂O. The resulting crude extracts were resuspended in methanol (5 mL) and dissolved with 70 mL of a cellulase enzyme solution (10 mg cellulase from *Aspergillus niger* [Sigma 22178]/mL Milli-Q-H₂O) in order to release glycoside (bound) SL. The dissolved extracts with cellulase enzyme solution were then transferred to 15 mL tubes and incubated in a water bath for 2 h at 40 °C. After enzymatic treatment, extracts were distributed into 50 mL tubes (3 tubes per cultivar) and ethyl acetate (25 mL) was added to each tube. The tubes were centrifuged (10 min, 1540 g) and the supernatants (ethyl acetate-extracts) were collected in round-bottomed glass flasks (1 flask per cultivar). Remaining sedimented material in the 50 mL tubes was extracted with ethyl acetate (2 × 25 mL) as described above. The collected ethyl acetate-extracts were evaporated to dryness under reduced pressure (35 °C), redissolved in methanol (8 mL) and transferred to 50 mL tubes (1 tube per cultivar). Final purification of SL from phenols and other plant compounds was performed by solid-phase extraction (SPE). Dichloromethane (28 mL) was added to each tube and centrifuged (10 min, 1540 g). A SPE vacuum manifold was equipped with 12 × 6 mL SPE tubes (Supelclean® LC-Si SPE tubes, Supelco 505374). SPE tubes were conditioned with dichloromethane/*i*-propanol (6 mL, 1/1; v/v) and equilibrated with dichloromethane (6 mL). Clean 12 × 15 mL tubes (6 tubes per cultivar) were set in the vacuum manifold for collection of SL. Extract mixed with dichloromethane (6 mL) was loaded in each SPE tube and the obtained liquid fractions were transferred into one tared 50 mL glass flask per chicory cultivar. Collected fractions were dried under reduced pressure (35 °C) and resulting purified extracts were weighed. Obtained extracts were highly viscous with low solubility in Milli-Q-H₂O or PBS. These purified chicory extracts were therefore resuspended in 100% DMSO at a concentration of 100 mg dry weight extract/mL DMSO (stock solution) and stored at -20 °C until use for *in vitro* assays and chemical analyses. High-performance liquid chromatography (HPLC) analyses were performed with stock solution of purified chicory extracts after removal of DMSO by freeze-drying and resuspension in methanol (10 mg dry weight extract/mL methanol). Two additional extractions from the same plant material were performed as described

above and confirmed the repeatability of the extraction method (data not shown).

2.3. Chemical analyses of purified chicory extracts

Compounds present in the obtained purified extracts were characterised by HPLC with mass spectrometry (MS) carried out in a Shimadzu Nexera X2 equipment and a Bruker MicroTOF-Q III mass spectrometer, using an Supelco, Ascentis Express Peptide ES-C18 column (2.7 μm , 160 Å). A 1 mL/min linear gradient from 0 to 100% buffer B over 10 min was employed (buffer A: 0.025% trifluoroacetic acid [TFA] in 10% aqueous acetonitrile-MeCN; buffer B: 0.025% TFA in 90% aqueous MeCN). Molecular mass analysis was carried out in conjunction with HPLC on a quadruple mass spectrometer in positive and negative electron spray ionization modes. Individual SL were identified by comparison of the main fragment ions (m/z) detected in all major peaks and retention times from published reports of SL profiles in chicory (Sessa et al., 2000; Ferioli and D'Antuono, 2012; Graziani et al., 2015). To quantify individual SL, HPLC was carried out at a wavelength where the UV absorptions of the different compounds were approximately the same (i.e. isosbestic point). This approximate isosbestic point was determined from the UV absorption spectra of all compounds detected in the purified extracts and occurred at wavelengths of 271 nm for cv. Spadona and 275 nm for cv. Puna II (Supplementary Figs. 1 and 2, respectively). Consequently, each extract was analysed at the mentioned wavelength for determination of individual peak areas. Of the SL known to be present in chicory, only lactucopicrin was commercially available as a pure standard when the experiments were conducted. Therefore, all detected peak areas were quantified by comparison with an external standard calibration curve using pure lactucopicrin. Here, a two-fold dilution of pure lactucopicrin (Extrasynthese 3813) was prepared in methanol (1000–15.6 μg lactucopicrin/mL) and analysed by HPLC-MS at 271 nm and 275 nm under the same conditions as the purified chicory extracts. Two separate SL quantifications were performed on each purified chicory extract.

2.4. Parasite material

Two Jersey calves, aged 5–6 months, were infected with 10,000 or 20,000 third-stage larvae (L3) of an ivermectin-susceptible *O. ostertagi* strain (Ref Label: OOSG10, Ridgeway Research, UK). The study was approved by the Animal Experiment Inspectorate, Ministry of Food, Agriculture and Fisheries of Denmark (j. No. 2013-15-2934-00763). Nematode eggs were detectable in faeces from both animals at day 16 post-infection. The eggs were recovered and hatched to obtain first-stage larvae (L1) according to Novobilský et al. (2011) and were used immediately after collection for a larval feeding inhibition assay. L3 were obtained from culture of eggs in faeces mixed with vermiculite for 14 days at 25 °C as described by Roepstorff and Nansen (1998). After recovery by baermannisation, L3 were maintained at 12 °C until use in a larval exsheathment inhibition assay. Calves were necropsied at two consecutive days in order to isolate live adult worms for motility inhibition assays. After captive bolt stunning and bleeding, the abomasum was immediately recovered and live *O. ostertagi* adults were isolated using the agar-migration method (Christensen et al., 1995). Adult worms were allowed to actively migrate from digesta into warm saline for 3 h at 37 °C and were then collected using a 20 μm sieve. Subsequently, worms were transferred into a 50 mL tube in warm saline solution (37 °C) and were washed three times in warm, sterile incubation medium (RPMI 1640 with L-glutamine, Gibco 11875-085), supplemented with 200 U/mL penicillin, 200 μg /mL streptomycin and 2 μg /mL amphotericin B. After the last

wash, worms were transferred to a Petri dish from which they were immediately collected for the assays.

2.5. In vitro assays

2.5.1. Larval feeding inhibition assay (LFIA)

The objective of the LFIA was to investigate the effect of SL-containing extracts on the feeding behaviour of *O. ostertagi* L1 and performed according with Jackson and Hoste (2010) and Novobilský et al. (2011), with modifications. Fluorescein isothiocyanate (FITC)-labelled *Escherichia coli* was prepared according to Jackson and Hoste (2010) and preserved at –20 °C until use. After isolation, L1 were diluted in PBS (0.05 M NaCl in Milli-Q-H₂O, pH 6.9) until achieving a suspension of approximately 100 L1/500 μL of PBS. Stock solutions of purified chicory extracts were serially dissolved in PBS and five concentrations (in triplicates) were prepared for each extract in 1.5 mL Eppendorf tubes (500 μL of diluted extract per tube). Approximately 100 L1 (500 μL of larval suspension) were added to each tube, reaching final concentrations of 500, 250, 100, 50, 10 μg dry extract/mL PBS (final concentration 1% DMSO; total volume in tube = 1000 μL). L1 incubated with ivermectin (IVM, Sigma I8898, 1 mg/mL) and 1% DMSO in PBS were run in triplicates as positive and negative controls, respectively. All tubes were gently shaken and pre-incubated horizontally for 2 h at 25 °C. After pre-incubation, 10 μL of FITC-labelled *E. coli* were added to each tube. Then, the tubes were gently shaken and incubated horizontally for 18 h at 25 °C. After incubation, the tubes were centrifuged for 1 min at 6000 g and 800 μL of the supernatant was removed. Sediment with L1 was transferred into microscope slides and observed under a fluorescent microscope with a blue filter (Leica DMR A2, band pass filter 450–490 nm, long pass filter 515 nm) at 200 \times and 400 \times magnifications. All L1 in each tube were counted and classified as: a) fed larvae, with the presence of green fluorescent FITC-labelled *E. coli* in the larval oesophagus and/or intestine or b) unfed larvae, lacking FITC-labelled *E. coli* in the gut or presence of green fluorescence only outside the larval body or around the buccal opening. The number of fed/unfed larvae was used to calculate larval feeding percentages in each replicate, as: % larval feeding = $100 \times [(\text{number of fed L1})/(\text{total number of L1})]$.

2.5.2. Larval exsheathment inhibition assay (LEIA)

This assay measured the potential inhibitory effect of SL-containing extracts on the chemically-induced exsheathment of *O. ostertagi* L3 and was conducted as described by Jackson and Hoste (2010) with minor modifications. Briefly, ensheathed L3 (2–3 months after isolation) were baermannised for 3 h using a 28 μm nylon mesh in order to isolate only live larvae. Obtained L3 were resuspended in PBS (pH 6.9) until a larval suspension of approximately 1650 L3/mL PBS was achieved. Stock solutions of Spadona and Puna II extracts were serially diluted in 100% DMSO and three decreasing concentrations were prepared for each chicory extract in 1.5 mL Eppendorf tubes (10 μL of each decreasing extract concentration per tube). Extra tubes were prepared with 10 μL of 100% DMSO as negative controls. Approximately 1600 L3 larvae (990 μL of the L3 suspension in PBS) were added to each tube, achieving final concentrations of 1000, 500 and 250 μg dry extract/mL PBS (1% DMSO) and 1% DMSO (negative controls). All tubes were vortex agitated for 10 s and pre-incubated horizontally for 3 h at 22 °C. After pre-incubation, the tubes were centrifuged at 6000 g for 2 min and the supernatants were removed. One mL of PBS was added to each tube and the washing procedure was repeated two more times before L3 were finally resuspended in 1 mL of PBS. A set of 24-well plates were prepared for the artificially induced larval exsheathment by adding 1940 μL of an exsheathment solution per well. Exsheathment solution was prepared by dilution of sodium

hypochlorite 10–15% (Sigma 425044) in PBS (1/500; v/v, pH 6.9). Approximately 100 L3 from each pre-incubation tube were then transferred into each well of the 24-well plate containing exsheathment solution (1 well plate per tube). This chemically-induced exsheathment was sequentially stopped by addition of 100 µL of Lugol iodine per well at 15, 30, 45 and 60 min after start of incubation. Exsheathment was replicated four times for each chicory extract concentration and 1% DMSO (negative control) and for each time point. Ensheathed or exsheathed larvae in all wells at each time point were quantified using an inverted microscope at 200 × magnification. Exsheathment rates were calculated for each well using the formula: % exsheathment = $100 \times \frac{(\text{number of exsheathed larvae per well})}{(\text{total number of larvae per well})}$.

2.5.3. Adult motility inhibition assay (AMIA)

The AMIAs were designed to investigate the inhibitory effect of SL-containing extracts on the motility of adult *O. ostertagi*. The assays were carried out following the methods described by Paolini et al. (2004) and Jackson and Hoste (2010), with some modifications. Two independent AMIAs were performed testing decreasing concentrations of each purified chicory extract (four assays in total) using freshly isolated adult worms collected after slaughter (see Section 2.4). For each assay, 48 well-plates were prepared with six decreasing concentrations of Spadona or Puna II extracts dissolved in the incubation medium for adult worms described in Section 2.4. The following concentrations were tested: 1000, 500, 250, 100, 50 and 10 µg dry extract/mL incubation medium (final concentration 1% DMSO; total volume in well = 1000 µL). Every concentration was tested in four replicates per plate. Wells containing IVM (1 mg/mL) and 1% DMSO dissolved in incubation medium were also prepared in four replicates in each plate as positive and negative controls, respectively. Four additional replicate wells were prepared for incubation of worms in 1000 µg dry extract/mL or 1% DMSO for electron microscopy (EM). Immediately after collection from digesta and washing (section 2.4), four to six adult *O. ostertagi* (males and females) were carefully added to each well. The plates were then placed in an incubator at 37 °C and worm motility was checked after 6, 24 and 48 h of incubation using a stereomicroscope. At each time point, the plates were gently hand-agitated to stimulate worm movement and the number of motile and non-motile (no movement detected during 10 s) individuals in each well were recorded. All observations were made by the same person. After 24 h of incubation, the incubation medium was removed from all wells and replaced by fresh incubation medium (1000 µL; 37 °C) containing identical Spadona or Puna II-extract concentrations. Further, at 24 h worms incubated in the extra replicates at 1000 µg dry extract/mL and 1% DMSO were collected, carefully washed in PBS and fixed in 2% glutaraldehyde in PBS for EM (section 2.6). Motility scores obtained in each AMIA were used to calculate worm motility percentage in each well as: % worm motility = $100 \times \frac{(\text{number of motile worms per well})}{(\text{total number of worms per well})}$.

2.6. Electron microscopy

Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) investigations were performed to detect ultra-structural damage in adult *O. ostertagi* exposed to the highest concentration of SL-containing extracts during 24 h. Adult worms incubated with both chicory extracts and 1% DMSO were selected for SEM, while worms exposed to Spadona-extract and 1% DMSO were selected for TEM. Adult worms for SEM were fixed as described by Williams et al. (2014a) and after fixation worms were placed into 99% ethanol and then coated with gold using a Low Vacuum Coater Leica EM ACE200. Scanning electron microscopies

were performed using a Phillips XL 30 FEG SEM and image processing was done with Scandium software. Adult worms for TEM were fixed, processed and analysed as described by Williams et al. (2014a).

2.7. Statistical analysis

Concentrations of individual and total compounds, detected in two separate quantifications on each purified extract, were compared between Spadona and Puna II extracts using a pairwise t-test with Bonferroni's correction. In the AMIA and the LFIA the effective concentration of each SL-containing extract able to inhibit the motility or feeding in 50% of adults or L1 (EC₅₀), respectively, was calculated. Tested extract concentrations were log-transformed and motility/feeding percentages obtained in the assays (eight replicates per concentration for AMIA and three replicates per concentration for LFIA) were analysed by non-linear (least squares) regression using the model *log (inhibitor) vs. response-variable slope* in GraphPad Prism® version 6.05, as described by Demeler et al. (2010). In the AMIA and LFIA the highest extract concentration tested from both extracts achieved a total inhibition (0%) of motility/feeding, with the sole exception of Puna II-extract in the AMIA after 6 h of incubation. Consequently, bottom values were set as 0% and top values were set as the mean motility/feeding percentage observed in negative control replicates. The R squared measure of goodness of fit (R²) was calculated for each dose-response curve. Statistical differences between EC_{50s} obtained with both purified extracts were analysed by extra sum-of-squares F test with a null hypothesis of equal EC₅₀. In the LFIA, the mean exsheathment percentages were calculated for each concentration and time point and were compared with the exsheathment rates obtained in the negative control wells (1% DMSO) by two-way ANOVA. A value of *P* < 0.05 was considered significant.

3. Results

3.1. Chemical analyses of purified chicory extracts

Chromatograms of the purified chicory extracts analysed by HPLC-MS are presented in Fig. 1. Chemical characterisations of detected molecules in two separate quantifications of each chicory extract are presented in Table 1. All peaks in both extracts were detected before 4 min. Identification of *m/z* ion fragments (positive and negative mode) by MS confirmed that a certain degree of hydrolysis occurred in the SL during the extraction process (Supplementary Table 1). Lactucin (LAC), 8-deoxylactucin (8-DOL) and lactucopicrin (LCP) were detected in extracts from both chicory cultivars, as well as their 11, 13-dihydro (DI)-derivatives: DI-LAC, DI-8-DOL and DI-LCP (Table 1, Fig. 2). In addition, four unknown compounds were detected in both chicory extracts. From the original leaf material, mean (±S.D.) total SL recovered were 9.4 (±1.2) g total SL/kg of dried leaves in Spadona and 11.7 (±1.3) g SL/kg of dried leaves in Puna II. Similar levels of total and individual SL and unknown compounds were detected between extracts, with the exception of DI-LAC and DI-8-DOL which were found in significantly higher concentrations in Puna II and Spadona extracts, respectively (Table 1).

3.2. LFIA

In negative controls, 55.2% (±3.3%) of L1 incubated with 1% DMSO had fed at the end of the incubation. All fed larvae of the negative controls were motile while all unfed larvae were dead. In contrast, all L1 incubated with IVM (positive control) were dead and unfed. Larvae exposed to SL-containing extracts, however,

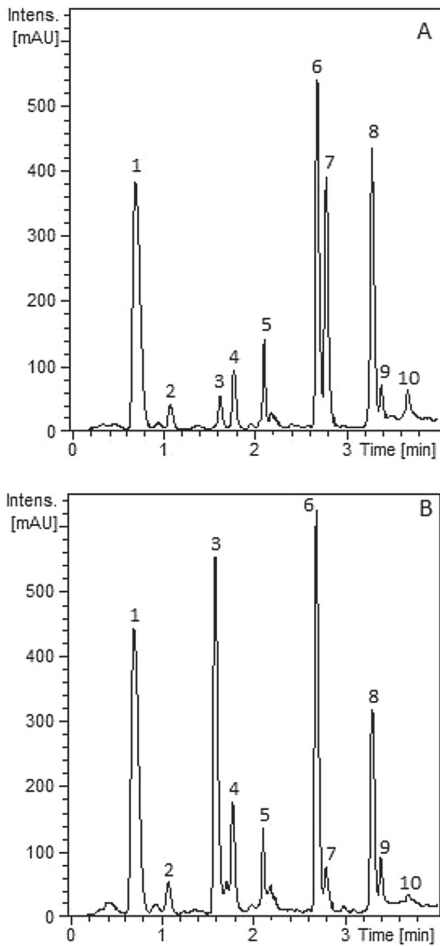


Fig. 1. Chromatograms obtained of purified extracts from chicory cv. Spadona (A) and cv. Puna II (B) used for *in vitro* assays with free-living and parasitic stages of *Ostertagia ostertagi*. Chromatograms are presented at the wavelength where isosbestic points were detected for each extract (271 nm for Spadona and 275 nm for Puna II). 3 = 11, 13-dihydro-lactucin; 4 = lactucin; 6 = 8-deoxy-lactucin; 7 = 11, 13-dihydro-8-deoxylactucin; 8 = 11, 13-dihydro-lactucopicrin; 9 = lactucopicrin. Peaks 1, 2, 5 and 10 = unknown.

showed a distinct pattern as all fed L1 were motile while unfed larvae were either motile or dead. No morphological damage was observed in L1 exposed to the SL-containing extracts at any tested concentration. Dose-response curves obtained in the LFIA with the tested extracts are depicted in Fig. 3. Larvae incubated with the lowest concentration of SL-containing extracts had similar feeding percentage as observed in the negative controls, while the highest concentration tested (500 µg dry extract/mL) of both extracts induced a total suppression of larval feeding. Nevertheless, at intermediate concentrations, Spadona-extract exerted a remarkably higher potency than Puna II-extract, tested at equal concentrations, illustrated by significantly lower EC_{50} values (EC_{50} [95% confidence interval] in µg dry extract/mL: Spadona = 26.9 [22.3–32.4] and Puna II = 108.7 [80.9–146.2], $P < 0.0001$).

3.3. LEIA

No significant differences were observed in the exsheathment rates between SL-containing extracts at all tested concentrations and negative controls ($P > 0.05$). All negative control L3 (pre-incubated with 1% DMSO) were exsheathed after 45 min of incubation with the exsheathment solution. Similarly, >99% of L3 exposed to any of the SL-containing extracts were exsheathed after 45 min of incubation.

3.4. AMIA

In negative controls, a mean worm motility of 100% was observed at 6 and 24 h of incubation with 1% DMSO. After 48 h of incubation, mean (\pm S.D.) worm motility percentage in the negative controls was 94.8% (\pm 2.6%). Dose-response curves obtained in the AMIAs at 6, 24 and 48 h of incubation are presented in Fig. 4. Mean motility of worms incubated with the lowest concentration of both SL-containing extracts was not different from negative controls at any time point. In contrast, and after 6 h of incubation with the highest concentration, Spadona and Puna II extracts induced a paralysis in 100 and 92% of the incubated worms, respectively. However, at intermediate concentrations Spadona-extract induced a greater paralysis in exposed worms, reflected in significantly lower EC_{50} values at all time points, as compared with worms incubated with the same concentrations of Puna II-extract ($P < 0.0001$, Table 2). In addition, a time-dependent effect of the inhibitory activity of the SL-containing extracts was observed, represented as decreasing EC_{50} values for both extracts as the incubation period progressed.

3.5. Electron microscopy

At the time of collection for electron microscopy (after 24 h of incubation) all worms incubated at 1% DMSO were motile and all worms incubated with the SL-containing extracts were dead. Examinations by SEM showed no noticeable differences between negative control nematodes and worms exposed to SL-containing extracts, with no obvious structural damage in the buccal opening or cuticle of worms exposed to Spadona-extract (Supplementary Fig. 3a, b) or Puna II-extract. Ultrastructural examination of thin sections of *O. ostertagi* adults by TEM did not demonstrate substantial differences between nematodes. Negative control nematodes and nematodes incubated with Spadona-extract had integral exterior borders of the cuticle and an intact basal layer and hypodermis. The only apparent difference was a slight decoloration and subtle erosion of the epicuticle in worms incubated with Spadona-extract (Supplementary Fig. 3c).

4. Discussion

Here, we have demonstrated direct and dose-dependent anti-parasitic effects of SL-containing extracts of forage chicory on the motility and feeding of *O. ostertagi* adults and L1, respectively. In contrast, no effects were observed on the exsheathment of L3 at any of the tested concentrations. Marked differences in anthelmintic activity were detected in SL-containing extracts from two forage chicory cultivars, with Spadona-extract exerting higher anti-parasitic effects compared with Puna II-extract. Despite the similar content of total SL and unknown molecules in the two tested extracts, the increased potency of Spadona-extract may be associated with its distinctive composition of individual compounds, which is further discussed below.

In the AMIA, nonetheless the rapid lethal effect of high concentrations of SL-containing extracts on adult worms, EM investigations did not reveal any morphological damage of SL-

Table 1
Chemical characterisation by high-performance liquid chromatography-mass spectrometry of purified extracts from leaves of chicory cv. Spadona and cv. Puna II used for *in vitro* assays with free-living and parasitic stages of *Ostertagia ostertagi*.

Extract		Spadona			Puna II			Pairwise t-test
Peak	Compound	% Mean total peak area ^a	µg Compound/mg dry extract		% Mean total peak area ^a	µg Compound/mg dry extract		
			Q1 ^b	Q2 ^b		Q1 ^b	Q2 ^b	
1	U1	27.4	143.0	117.0	26.4	137.7	106.7	N.S.
2	U2	2.1	9.6	9.4	2.4	17.7	17.3	N.S.
3	DI-LAC	2.3	10.7	11.4	19.9	105.3	87.2	****
4	LAC	4.0	19.7	18.5	6.3	37.3	29.8	N.S.
5	U3	5.1	25.7	23.9	5.0	30.5	23.9	N.S.
6	8-DOL	20.4	105.7	81.2	20.6	108.6	84.2	N.S.
7	DI-8-DOL	14.8	76.5	61.6	3.0	20.4	17.4	***
8	DI-LCP	17.0	87.8	74.8	11.9	65.2	50.9	N.S.
9	LCP	2.4	11.1	12.2	2.8	19.5	23.5	N.S.
10	U4	4.5	22.3	21.6	1.7	13.5	12.7	N.S.
Total SL		60.9	311.5	259.7	64.5	356.3	293.0	N.S.
Unknown		39.1	200.6	171.9	35.5	199.4	160.6	N.S.

Q: quantification; U: Unknown; SL: sesquiterpene lactones; LAC: Lactucin; DI-LAC: 11, 13-dihydro-lactucin; 8-DOL: 8-deoxylactucin; DI-8-DOL: 11, 13-dihydro-8-deoxylactucin; LCP: Lactucopicrin; DI-LCP: 11, 13-dihydro-lactucopicrin.

N.S.: $p > 0.05$; *** $P = 0.01$; **** $P < 0.0001$.

^a Area percentage of the respective peak of the total peak area in the chromatograms presented in Fig. 1.

^b Quantification of individual compounds was performed using an external standard curve with a pure lactucopicrin standard.

exposed worms, with the exception of a decoloured and slightly eroded epicuticle, suggesting a possible early stage of cuticle degeneration. These results are in contrast with studies describing marked disruptions in the cuticle and ultrastructure of nematodes exposed to other bioactive plant compounds like cysteine proteases and condensed tannins (Stepek et al., 2004; Brunet et al., 2011; Martínez-Ortiz-de-Montellano et al., 2013; Williams et al., 2014a, 2014b). Our findings therefore suggest that the mode of action of SL is different from that of the above mentioned PSM and this needs to be further explored.

In the LFIA, mean percentage of fed larvae in the negative controls (1% DMSO in PBS) were similar to published reports of LFIA with *O. ostertagi* L1 using 100% PBS as negative control medium (Novobilský et al., 2011). All unfed larvae in the negative controls were dead and similar low control values were observed by the authors in preliminary studies with L1 from the same *O. ostertagi* strain, but the reasons for this are unknown. In larvae exposed to SL, both extracts demonstrated a dose-dependent inhibitory effect on the feeding of *O. ostertagi* L1. Nonetheless, Spadona-extract inhibited larval feeding at much lower concentrations than Puna II-extract, which correlates with results from the AMIA. EC_{50} values obtained in the LFIA with both chicory extracts were much lower than those observed in the AMIA, confirming that inhibition of larval feeding required a much lower concentration than the inhibition of adult worm motility, as observed previously in studies with IVM (Geary et al., 1993; Gill et al., 1995). This indicates that the inhibition of the pharyngeal muscles appears to occur at a much lower extract concentration than the inhibition of motility, suggesting that pharyngeal muscles could be a more sensitive target for SL.

In the LEIA, SL-containing extracts did not affect the exsheathment kinetics of exposed *O. ostertagi* L3. Exsheathment is a precondition for the later embedding of L3 in the abomasal mucosa and our results agree with *in vivo* findings that showed no effect of forage chicory (cv. Grasslands Puna) on the establishment of *Teladorsagia circumcincta* L3 in infected lambs (Tzamaloukas et al., 2005). In the LEIA, all L3 incubated with the highest concentration of both chicory extracts were alive and motile at the end of the assay. These results are further supported by preliminary data from our group indicating that there is no effect of the same SL-containing extracts on the motility of *O. ostertagi* L3, even after incubation for 24–48 h with very high concentrations (2000 µg dry extract/mL, data not published).

At sampling, chicory plants were at the vegetative stage and leaves from both cultivars were hand-picked on the same occasion in order to standardise the collection procedure. Early findings by Rees and Harborne (1985) described that free SL are present in the latex of chicory roots and leaves and that levels of individual SL vary in different parts of the chicory plant according to the growth stage. Nevertheless, these findings need to be confirmed in different chicory cultivars as well as by screening of other sections of the aerial part of the plant (i.e. stems and flowers), which can also be grazed by cattle (Peña-Espinoza et al., unpublished observations). With respect to the quantification of compounds in the extracts, this was performed using an external calibration curve with pure lactucopicrin, the only SL of chicory commercially available as a pure standard at the time of the experiments. Our approach was a quantification of the compounds in the extracts (unknown concentrations) by comparison with the calibration curve (known concentrations), using peak areas and assuming similar chromatographic properties in increasing concentrations of compounds in the extract and in the standard. A quantification using pure standards of all SL and their dihydro-derivatives would have increased the sensitivity of the method. However, this was an initial attempt to understand the SL profile in these particular extracts using, as a standard, a compound actually present in the extracts (lactucopicrin). For comparison, previous studies have added santonin (another SL not present in chicory) as internal standard for quantification of SL in chicory (Ferioli and D'Antuono, 2012; Graziani et al., 2015).

Our results are the first evidence of anthelmintic activity of SL-containing extracts against larval feeding and adult motility, contributing to expand our knowledge on the role of SL in the anti-parasitic properties of chicory. Previously, Molan et al. (2003) were the first to suggest the potential anthelmintic effect of SL in chicory and reported the inhibitory effects of a crude extract from chicory roots against free-living larvae. However, these authors did not report the SL characterisation of the tested extract or the chicory cultivar used. More recently, Foster et al. (2011) described the dose-dependent inhibition of nematode egg hatching by free SL-containing extracts from forage chicory cv. Grasslands Puna and cv. Forage Feast. These authors reported the presence of only the three major SL of chicory in the tested extracts and detected an increased anthelmintic effect of Grasslands Puna-extract. In contrast to the present investigations, Foster et al. (2011) used much larger test concentrations (0–10 mg dry extract/mL). In our

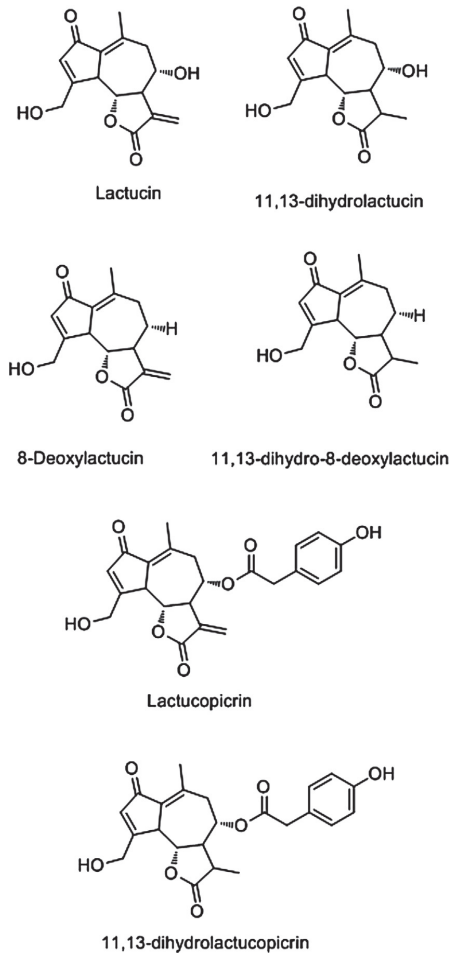


Fig. 2. Structures of sesquiterpene lactones detected in purified extracts from chicory cv. Spadona and cv. Puna II and tested *in vitro* against free-living and parasitic stages of *Ostertagia ostertagi*.

study the range of tested concentrations was based on preliminary *in vitro* studies with *O. ostertagi* by the authors (data not published) and was selected with the aim to detect a dose-dependent response. Scarce literature exists on the metabolism of SL from chicory in the digestive tract of ruminants and therefore, at the moment, it is difficult to accurately estimate the concentration of dietary SL that could reach the abomasum, the predilection site of *O. ostertagi* in cattle. However, in a preliminary study we have observed that 4 months-old calves are able to consume daily up to 3 kg dry matter of chicory silage (with 5.3 g SL/kg dry matter), and assuming an abomasum volume of 5 L (Nickel et al., 1979) and that SL are not inactivated in the forestomach, this would roughly correspond to a maximum of 3.18 mg SL/mL in the abomasum (Peña-Espinoza et al., unpublished results).

In our study, the three well-known guaianolide SL of chicory were also detected in leaf extracts of cv. Spadona and cv. Puna II: lactucin, 8-deoxylactucin and lactucopicrin, as reported for other

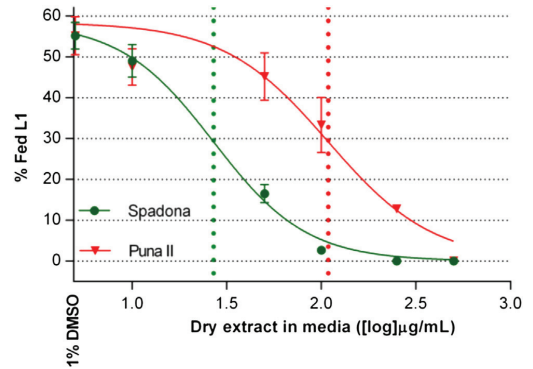


Fig. 3. Dose–response curves obtained in the larval feeding inhibition assay with *Ostertagia ostertagi* L1 incubated at different concentrations of SL-containing extracts (log μg dry extract/ml) from two chicory cultivars. Error bars represent S.D. between replicates (n = 3 X concentration per extract). Data points with no error bars indicate that the variation among values was 0 or close to 0. Dotted vertical lines represent EC₅₀ values for each extract.

chicory cultivars (Rees and Harborne, 1985; Price et al., 1990; Foster et al., 2011; Ferioli and D'Antuono, 2012; Graziani et al., 2015). Furthermore, the 11, 13-dihydro-derivatives of the mentioned SL were also detected: 11, 13-dihydro-lactucin, 11, 13-dihydro-8-deoxylactucin and 11, 13-dihydro-lactucopicrin, which have been also described in salad chicory cultivars (Ferioli and D'Antuono, 2012; Wulfkuehler et al., 2014; Ferioli et al., 2015). Spadona and Puna II extracts induced dose-dependent anthelmintic effects in the LFIA and the AMIA, suggesting that anti-parasitic compounds were present in both extracts. In addition, unknown compounds were also detected in both chicory extracts, which may have contributed to the observed anthelmintic effects. The HPLC-MS analyses revealed similar quantities of unknown compounds and of total and individual SL between the tested extracts, except for 11, 13-dihydro-lactucin and 11, 13-dihydro-8-deoxylactucin, which were found in significantly higher concentrations in Puna II and Spadona extracts, respectively. Based on results from the *in vitro* assays, our data suggest that the increased concentration of 11, 13-dihydro-8-deoxylactucin in Spadona-extract (3.6-fold increase) correlates, to some extent, with the increased potency and lower EC₅₀ values exerted by Spadona-extract in the LFIA (3.3-fold lower EC₅₀) and in the AMIA (2.6-fold lower EC₅₀), in comparison with Puna II-extract. These findings suggest that 11, 13-dihydro-8-deoxylactucin is a likely candidate to explain the higher anthelmintic effects of the Spadona-extract. In contrast, 11, 13-dihydro-lactucin, with increased levels in the less-potent Puna II-extract, seems not to be particularly related with an anthelmintic effect. However, unknown compounds were also present in the extract and no pure SL were tested in our study. Therefore we cannot confirm a direct relation between concentration of a particular SL and anthelmintic effect at the moment. To confirm these findings, further fractionation, isolation and testing of individual molecules in the extracts would identify the most active compound(s). Nevertheless, and to the best of our knowledge, this is the first report of the SL profiles of forage chicory cv. Spadona and cv. Puna II and the first study describing the *in vitro* anti-parasitic activity of forage chicory extracts containing a distinctive profile of dihydro-derivatives of SL. Previously, Foster et al. (2011) linked the higher *in vitro* activity of Grassland Puna-extract to the higher content of 8-deoxylactucin, which in our study was present in similar quantities in Spadona and Puna II extracts. This suggests that, if SL are

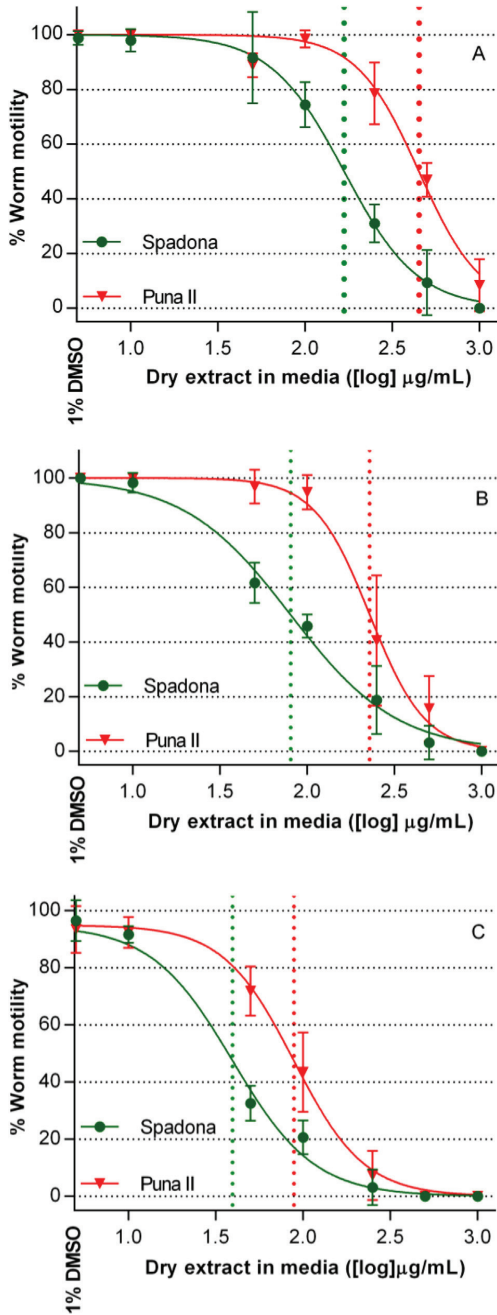


Fig. 4. Dose–response curves obtained in adult motility inhibition assays with *Ostertagia ostertagi* at 6 h (A), 24 h (B) and 48 h (C) after incubation with different concentrations of SL-containing extracts ([log] μg dry extract/ml) from two chicory cultivars. Each data point in the graphs represents the mean motility percentage of 8 replicates (n = 4–6 adult worms per replicate) for each concentration obtained in two independent assays. Error bars represent S.D. between replicates. Data points with no error bars indicate that the variation among values was 0 or close to 0. Dotted vertical lines represent EC₅₀ values for each chicory extract.

responsible for the anti-parasitic effects of chicory *in vivo*, only some SL may be considered as candidates for novel anthelmintic compounds and for selection of forage chicory cultivars with increased anthelmintic activity.

It is known that guaianolide SL can exert potent cytotoxic activities, which have been mainly attributed to the presence of an α -methylene (CH₂) functional group linked to the γ -lactone in the SL molecule (Simonsen et al., 2013). This α -methylene group is known to react with sulfhydryl (thiol) groups in cysteine and cysteine-containing peptides by a Michael-type addition, affecting cell signalling, cell replication, apoptosis and mitochondrial respiration (Schmidt, 2006; Simonsen et al., 2013). As an example, it has been reported that natural SL can reduce the intracellular concentration of free glutathione in *Leishmania mexicana mexicana*, which led to a toxic intra-cellular accumulation of reactive oxygen species and blocked cell proliferation (Barrera et al., 2013). However, dihydro-derivatives of SL lack this α -methylene group and still can exert biological effects (Lee et al., 1977; Schmidt, 2006; Ghantous et al., 2010), which clearly indicates that there is not just one single biological mechanism of effect for all SL. Ren et al. (2005) reported that 11, 13- β -dihydro-lactucopicrin and 11, 13- β -dihydro-lactucin from *Mulgedium tatarica*, missing the α -methylene group but having an ester group at C-8 instead, exerted a higher toxicity towards human nasopharyngeal and liver cancer cells. Currently, the molecular mechanisms of action of SL and their dihydro-derivatives towards nematodes are unknown and need to be further investigated. Moreover, it is uncertain whether the anti-parasitic effects observed in our study correspond to the action of single (known and/or unknown) molecules or a synergistic effect between different plant compounds, as indicated by recent findings with condensed tannins and flavonoids (Klongsiriwet et al., 2015).

Our study indicates that the tested compounds isolated from forage chicory could affect *O. ostertagi* populations in cattle by reducing the feeding and motility/survival of worms. In addition, the higher anti-parasitic effects exerted by Spadona-extract, and the dose-dependent effect of both extracts, suggest that potential *in vivo* effects could be dependent on the chicory cultivar under investigation and the concentration of active molecules that reach the abomasum. However, compounds present in the tested extracts may not be the same molecules available at the site of infection of *O. ostertagi*. Although little is known about the pharmacokinetics of SL in ruminants, Ferreira and Gonzalez (2008) demonstrated that artemisinin, a purified SL widely used to treat malaria in humans, given to goats as an oral capsule (23 mg artemisinin/kg body weight), was partly metabolised in the rumen and was detected as dihydroartemisinin in plasma after 4 h of treatment. Moreover, *in vivo* trials in sheep indicate that the active compounds do not seem to be degraded or inactivated in the rumen and can reach the abomasum to exert their anti-parasitic activity (Scales et al., 1994; Tzamaloukas et al., 2005). Yet, extensive research is needed to elucidate the fate of dietary SL and other bioactive plant compounds in the digestive tract of ruminants.

In conclusion, we have demonstrated that SL-containing extracts from forage chicory can inhibit larval feeding and exert direct anti-parasitic effects against adult stages of *O. ostertagi*, whereas no effects were observed on larval exsheathment. We observed substantial differences in the anti-parasitic activity between extracts of two forage chicory cultivars and this may be related to variations in the concentration of individual SL. The distinctive anti-parasitic activities of extracts from different cultivars with particular molecular profiles can help to target the identification of the responsible compound(s). However, further studies are warranted not only to confirm the PSM/compound(s) responsible for the anti-

Table 2

Effective concentration able to induce 50% inhibition in worm motility (EC₅₀) by purified extracts from leaves of chicory cv. Spadona and cv. Puna II at different incubation times in adult motility inhibition assays with *Ostertagia ostertagi*.

Incubation time	6 h		24 h		48 h	
Extract	Spadona	Puna II	Spadona	Puna II	Spadona	Puna II
EC ₅₀ (µg dry extract/mL)	167.6****	449.4	80.3****	228.2	35.7****	82.4
95% CI	146.7–191.3	406.1–497.4	70.5–91.6	198.6–262.1	31.5–40.3	72.3–93.4
R ²	0.96	0.95	0.97	0.95	0.98	0.96

CI = confidence interval; R² = goodness-of-fit; ****P < 0.0001.

parasitic activity in the tested chicory extracts, but also to assess the *in vivo* anthelmintic effects of a chicory-based diet in cattle infected with gastrointestinal nematodes.

Conflicts of interest

The authors declare that they have no conflicts of interest.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.ijpddr.2015.10.002>.

References

Barrera, P., Sülsen, V.P., Lozano, E., Rivera, M., Beer, M.F., Tonn, C., Martino, V.S., Sosa, M.A., 2013. Natural sesquiterpene lactones induce oxidative stress in *Leishmania mexicana*. *Evid. Based. Complement. Altern. Med.* 2013, 163404. <http://dx.doi.org/10.1155/2013/163404>.

Bischoff, T.A., Kelley, C.J., Karchesy, Y., Laurantos, M., Nguyen-Dinh, P., Arefi, A.G., 2004. Antimalarial activity of lactucin and lactucopicrin: sesquiterpene lactones isolated from *Cichorium intybus* L. *J. Ethnopharmacol.* 95, 455–457.

Brunet, S., Fourquaux, I., Hoste, H., 2011. Ultrastructural changes in the third-stage, infective larvae of ruminant nematodes treated with sainfoin (*Onobrychis viciifolia*) extract. *Parasitol. Int.* 60, 419–424.

Cavin, C., Delannoy, M., Malnoe, A., Debeve, E., Touché, A., Courtois, D., Schilter, B., 2005. Inhibition of the expression and activity of cyclooxygenase-2 by chicory extract. *Biochem. Biophys. Res. Commun.* 327, 742–749.

Charlier, J., van der Voort, M., Kenyon, F., Skuce, P., Vercrusysse, J., 2014. Chasing helminths and their economic impact on farmed ruminants. *Trends Parasitol.* 30, 361–367.

Christensen, C., Barnes, E., Nansen, P., Roepstorff, A., Slotved, H., 1995. Experimental *Oesophagostomum dentatum* infection in the pig: worm populations resulting from single infections with three doses of larvae. *Int. J. Parasitol.* 25, 1491–1498.

Demeler, J., Küttler, U., von Samson-Himmelstjerna, G., 2010. Adaptation and evaluation of three different *in vitro* tests for the detection of resistance to anthelmintics in gastro intestinal nematodes of cattle. *Vet. Parasitol.* 170, 61–70.

Ferioli, F., D'Antuono, L.F., 2012. An update procedure for an effective and simultaneous extraction of sesquiterpene lactones and phenolics from chicory. *Food Chem.* 135, 243–250.

Ferioli, F., Manco, M., D'Antuono, L., 2015. Variation of sesquiterpene lactone and phenolic content in chicory and endive germplasm. *J. Food Compos. Anal.* 39, 77–86.

Ferreira, J., Gonzalez, J., 2008. Chemical and biological stability of artemisinin in bovine rumen fluid and its kinetics in goats (*Capra hircus*). *Braz. J. Vet. Parasitol.* 17, 103–109.

Foster, J., Cassida, K., Turner, K., 2011. *In vitro* analysis of the anthelmintic activity of forage chicory (*Cichorium intybus* L.) sesquiterpene lactones against a predominantly *Haemonchus contortus* egg population. *Vet. Parasitol.* 180, 298–306.

Fox, M.T., 1993. Pathophysiology of infection with *Ostertagia ostertagi* in cattle. *Vet. Parasitol.* 46, 143–158.

Geary, T., Sims, S., Thomas, E., Vanover, L., Davis, J., Winterrowd, C., Klein, R., Ho, N., Thompson, D., 1993. *Haemonchus contortus*: ivermectin-induced paralysis of the pharynx. *Exp. Parasitol.* 77, 88–96.

Gershenzon, J., Dudareva, N., 2007. The function of terpene natural products in the natural world. *Nat. Chem. Biol.* 3, 408–414.

Ghantous, A., Gali-Muhtasib, H., Vuorela, H., Saliba, N.A., Darwiche, N., 2010. What made sesquiterpene lactones reach cancer clinical trials? *Drug Discov. Today* 15, 668–678.

Gill, J., Redwin, J., Van Wyk, J., Lacey, E., 1995. Avermectin inhibition of larval development in *Haemonchus contortus* – effects of ivermectin resistance. *Int. J. Parasitol.* 25, 463–470.

Githiori, J.B., Athanasiadou, S., Thamsborg, S.M., 2006. Use of plants in novel approaches for control of gastrointestinal helminths in livestock with emphasis on small ruminants. *Vet. Parasitol.* 139, 308–320.

Graziani, G., Ferracane, R., Sambo, P., Santagata, S., Nicoletto, C., Fogliano, V., 2015. Profiling chicory sesquiterpene lactones by high resolution mass spectrometry. *Food Res. Int.* 67, 193–198.

Heckendorn, F., Häring, D.A., Maurer, V., Senn, M., Hertzberg, H., 2007. Individual administration of three tanniferous forage plants to lambs artificially infected with *Haemonchus contortus* and *Cooperia curticei*. *Vet. Parasitol.* 146, 123–134.

Hoste, H., Torres-Acosta, J.F.J., Sandoval-Castro, C.A., Mueller-Harvey, I., Sotiraki, S., Louvandini, H., Thamsborg, S.M., Terrill, T.H., 2015. Tannin containing legumes as a model for nutraceuticals against digestive parasites in livestock. *Vet. Parasitol.* 212, 5–17.

Jackson, F., Hoste, H., 2010. *In vitro* methods for the primary screening of plant products for direct activity against ruminant gastrointestinal nematodes. In: Vercoe, P.E., Makkar, H.P.S., Schlink, A.C. (Eds.), *In Vitro Screening of Plant Resources for Extra-nutritional Attributes in Ruminants: Nuclear and Related Methodologies*. FAO/International Atomic Energy Agency, Springer, Dordrecht, pp. 25–45.

Klongsriwet, C., Quijada, J., Williams, A.R., Mueller-Harvey, I., Williamson, E.M., Hoste, H., 2015. Synergistic inhibition of *Haemonchus contortus* exsheathment by flavonoid monomers and condensed tannins. *Int. J. Parasitol. Drugs Drug Resist.* 5, 127–134.

Lee, K., Ibuka, T., Rong-Yang, W., Geissman, T., 1977. Structure-antimicrobial activity relationships among the sesquiterpene lactones and related compounds. *Phytochemistry* 16, 1177–1181.

Marley, C., Cook, R., Keatinge, R., Barrett, J., Lampkin, N., 2003. The effect of birdsfoot trefoil (*Lotus corniculatus*) and chicory (*Cichorium intybus*) on parasite intensities and performance of lambs naturally infected with helminth parasites. *Vet. Parasitol.* 112, 147–155.

Martínez-Ortiz-de-Montellano, C., Arroyo-López, C., Fourquaux, I., Torres-Acosta, J.F.J., Sandoval-Castro, C.A., Hoste, H., 2013. Scanning electron microscopy of *Haemonchus contortus* exposed to tannin-rich plants under *in vivo* and *in vitro* conditions. *Exp. Parasitol.* 133, 281–286.

Molan, A.L., Duncan, A.J., Barry, T.N., McNabb, W.C., 2003. Effects of condensed tannins and crude sesquiterpene lactones extracted from chicory on the motility of larvae of deer lungworm and gastrointestinal nematodes. *Parasitol. Int.* 52, 209–218.

Nansen, P., 1993. Current and future prospects for control of ostertagiasis in northern Europe – examples from Denmark. *Vet. Parasitol.* 46, 3–21.

Nickel, R., Schummer, A., Seiferle, S., 1979. *The Viscera of the Domestic Mammals*, second ed. Springer-Verlag, Berlin Heidelberg.

Novobilský, A., Mueller-Harvey, I., Thamsborg, S.M., 2011. Condensed tannins act against cattle nematodes. *Vet. Parasitol.* 182, 213–220.

O'Grady, J., Kotze, A.C., 2004. *Haemonchus contortus*: *in vitro* drug screening assays with the adult life stage. *Exp. Parasitol.* 106, 164–172.

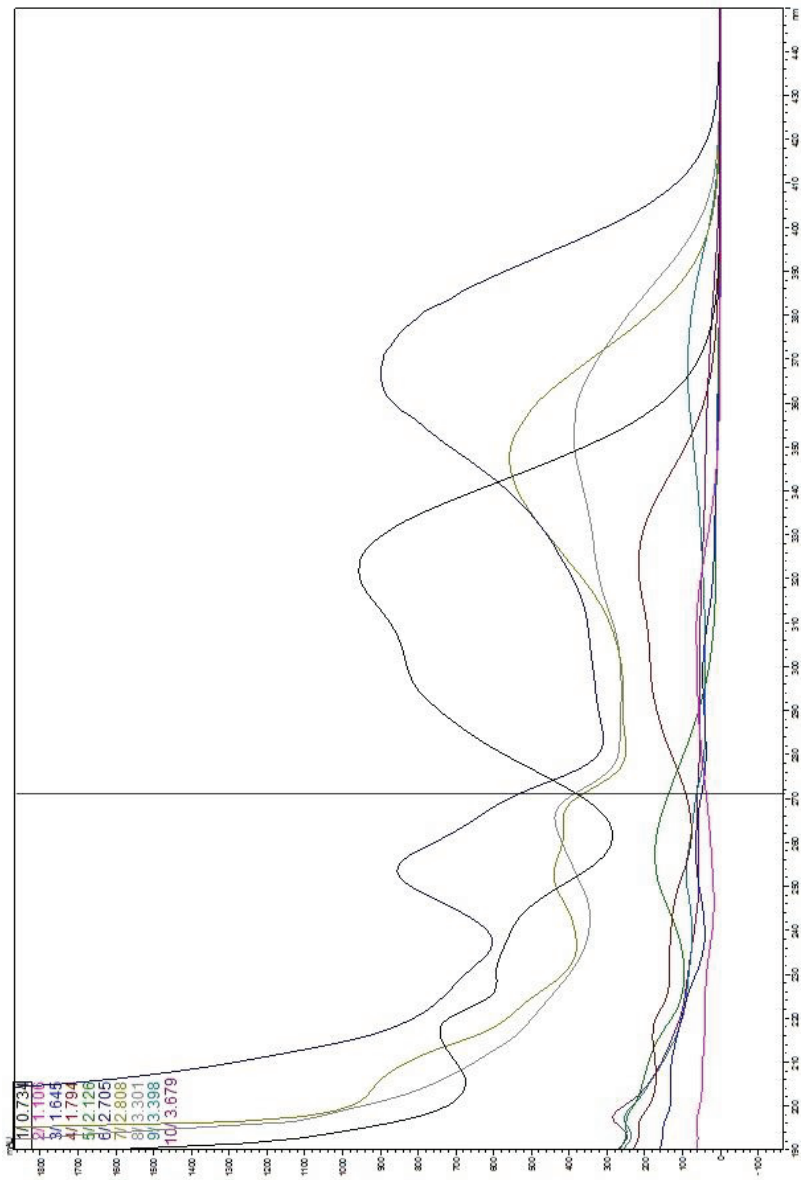
Paolini, V., Fouraste, I., Hoste, H., 2004. *In vitro* effects of three woody plant and sainfoin extracts on 3rd-stage larvae and adult worms of three gastrointestinal nematodes. *Parasitology* 129, 69–77.

Price, K., DuPont, M., Shepherd, R., Chan, H., Fenwick, G., 1990. Relationship between the chemical and sensory properties of exotic salad crops – coloured lettuce (*Lactuca sativa*) and chicory (*Cichorium intybus*). *J. Sci. Food Agric.* 53, 185–192.

Rees, S., Harborne, J., 1985. The role of sesquiterpene lactones and phenolics in the chemical defence of the chicory plant. *Phytochemistry* 24, 2225–2231.

Ren, Y., Zhou, Y., Chen, X., Ye, Y., 2005. Discovery, structural determination and anticancer activities of lactucin-like guaianolides. *Lett. Drug Des. Discov.* 2, 444–450.

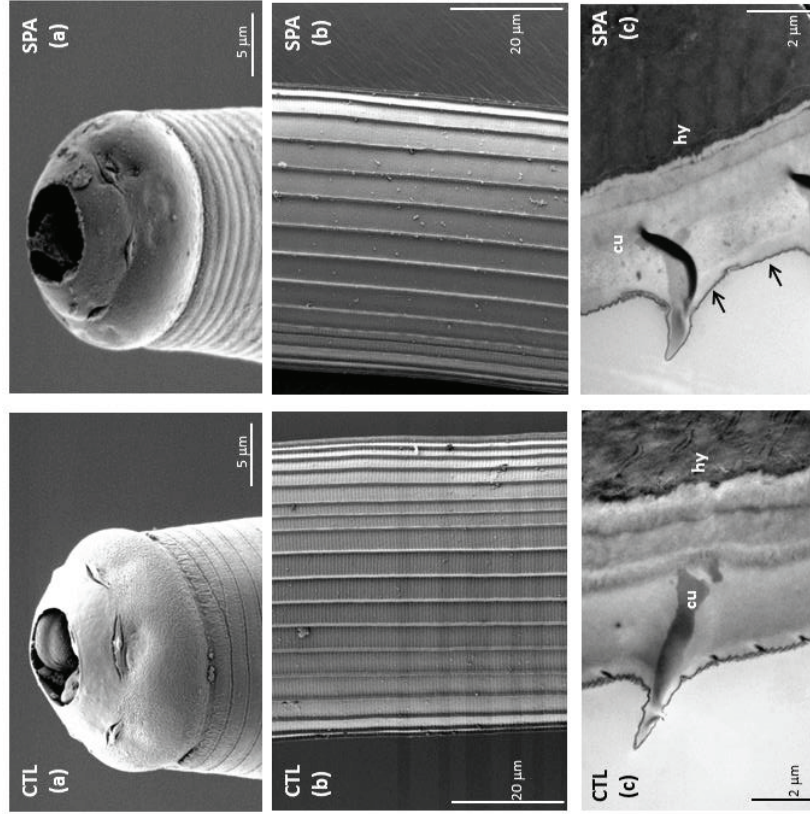
- Roepstorff, A., Nansen, P., 1998. Epidemiology, Diagnosis and Control of Helminth Parasites of Swine. FAO Animal Health Manual, Rome, pp. 56–60.
- Scales, G.H., Knight, T.L., Saville, D.J., 1994. Effect of herbage species and feeding level on internal parasites and production performance of grazing lambs. *N. Z. J. Agric. Res.* 38, 237–247.
- Schmidt, T.J., 2006. Structure-activity relationships of sesquiterpene lactones. *Stud. Nat. Prod. Chem.* 33, 309–392.
- Sessa, R.A., Bennett, M.H., Lewis, M.J., Mansfield, J.W., Beale, M.H., 2000. Metabolite profiling of sesquiterpene lactones from *Lactuca* species. *J. Biol. Chem.* 275, 26877–26884.
- Simonsen, H., Weitzel, C., Christensen, S., 2013. Guaianolide sesquiterpenoids: pharmacology and biosynthesis. In: Ramawat, K., Mérillon, J. (Eds.), *Natural Products*. Springer, Berlin Heidelberg, pp. 3069–3098.
- Stepek, G., Behnke, J.M., Buttle, D.J., Duce, I.R., 2004. Natural plant cysteine proteinases as anthelmintics? *Trends Parasitol.* 20, 322–327.
- Tzamaloukas, O., Athanasiadou, S., Kyriazakis, I., Jackson, F., Coop, R.L., 2005. The consequences of short-term grazing of bioactive forages on established adult and incoming larvae populations of *Teladorsagia circumcincta* in lambs. *Int. J. Parasitol.* 35, 329–335.
- Waller, P.J., Thamsborg, S.M., 2004. Nematode control in “green” ruminant production systems. *Trends Parasitol.* 20, 493–497.
- Williams, A.R., Fryganas, C., Ramsay, A., Mueller-Harvey, I., Thamsborg, S.M., 2014a. Direct anthelmintic effects of condensed tannins from diverse plant sources against *Ascaris suum*. *PLoS One* 9, e97053.
- Williams, A.R., Ropiak, H.M., Fryganas, C., Desrues, O., Mueller-Harvey, I., 2014b. Assessment of the anthelmintic activity of medicinal plant extracts and purified condensed tannins against free-living and parasitic stages of *Oesophagostomum dentatum*. *Parasit. Vectors* 7, 518.
- Wulfkuehler, S., Gras, C., Carle, R., 2014. Influence of light exposure during storage on the content of sesquiterpene lactones and photosynthetic pigments in witloof chicory (*Cichorium intybus* L. var. *foliosum* Hegi). *LWT—Food Sci. Technol.* 58, 417–426.



Supplementary Figure 1. UV absorption patterns of the compounds detected in the Spadona-extract and determination of isosbestic point (271 nm, black line). Numbers represent the molecules presented in Table 1.



Supplementary Figure 2. UV absorption patterns of the compounds detected in the Puna II-extract and determination of isosbestic point (275 nm, black line). Numbers represent the molecules presented in Table 1.



Supplementary Figure 3. Electron microscopy images of *Ostertagia ostertagi* adults incubated during 24 hours with 1% DMSO in PBS (CTL) or 1000 μg Spadona-extract/mL in PBS (SPA). In the scanning electron micrographs (a, b) note the intact mouth opening, circular striations (close to the mouth opening) and intact prominent longitudinal ridges in all worms. In transmission electron micrographs (c) worms exposed to Spadona-extract had a slight decolouration in the cuticle (cu) and a subtle erosion of the epicuticle (black arrows), in comparison with negative controls. Note the intact basal layer separating (cu) and the underlying intact hypodermis (hy) in all worms.

Supplementary Table 1. Molecular fragments (m/z) of sesquiterpene lactones detected by high-performance liquid chromatography – mass spectrometry (positive and negative-ion modes) in selected peaks

Peak	Compound	Molecular weight	Major ion fragments (m/z)
(+)			
3	11, 13-dihydro-lactucin	278	303 [M+Na+2H] ⁺ ; 351 [M+MeCN+MeOH] ⁺
4	Lactucin	276	294 [M+H ₂ O] ⁺ ; 295 [M+H ₂ O+H] ⁺ ; 315 [M+K] ⁺ ; 351 [M+MeCN+MeOH+2H] ⁺ ; 413 [M+TFA+Na] ⁺
6	8-deoxylactucin	261	303 [M+MeCN+H] ⁺
7	11, 13-dihydro-8-deoxylactucin	263	287 [M+Na+H] ⁺ ; 303 [M+K+H] ⁺
8	11, 13-dihydro-lactucopricin	410	317 [M-2HCO ₂ H-H] ⁺ ; 411 [M+H] ⁺
9	Lactucopricin	412	411 [M-H] ⁺
(-)			
			311 [M+MeOH+H] ⁻
			473 [2M-2K-H] ⁻ ; 520 [2M-MeOH] ⁻
			285 [M+Na+H] ⁻ ; 301 [M+MeCN-H] ⁻ ; 415 [M+TFA+Na+H] ⁻
			285 [M+Na-H] ⁻ ; 301 [M+K-H] ⁻ ; 415 [M+TFA+Na-H] ⁻
			399 [M-HCO ₂ +2H+MeOH] ⁻ ; 429 [M+H ₂ O+H] ⁻ ; 523 [M+TFA-H] ⁻ ; 593 [M+pHPA+MeOH] ⁻
			399 [M-HCO ₂ +MeOH] ⁻

HCO₂: formate ion (from formic acid); MeCN: acetonitrile; K: potassium; Na: sodium; MeOH: methanol; TFA: trifluoroacetic acid; pHPA: p-hydroxyphenylacetate

Parasitism with gastrointestinal nematodes is ubiquitous in grazing cattle and its control is critical to sustain satisfactory levels of animal health, welfare and productivity. Conventional control strategies that rely exclusively on anthelmintic drugs are severely threatened by the development of anthelmintic resistance in bovine nematodes. Therefore, alternative control approaches to minimize selection for drug-resistance and that can be combined with the still effective anthelmintics in integrated control strategies are urgently needed. This PhD thesis aims to contribute towards the integrated control of bovine gastrointestinal nematodes by evaluating the field efficacy of ivermectin in naturally-infected Danish cattle and by exploring the direct anthelmintic activity of forage chicory (*Cichorium intybus*) and its bioactive compounds against cattle nematodes.

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