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Hyaluronidase-inhibitory activities of glycosaminoglycans from *Liparis tessellatus* eggs

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Highlights

- ESI-MS/MS showed the presence of uronic acids from *L. tessellatus* eggs
- Hyaluronidase inhibitory effect of sulfated hyaluronic acid from *L. tessellatus* eggs
- Uronic acids from *L. tessellatus* eggs have hyaluronidase inhibitory effects

Abstract

Polysaccharide fractions isolated from *L. tessellatus* eggs were purified and eluted using the DEAE-sepharose fast flow column. These were collected, tested and pooled based on their sugars content: F1, F2, and F3 which contain 26.8, 23.3, and 20.2% sulfated glycans; 34.5, 38.2, and 45.0% uronic acids; and 23.5, 19.0, and 7.5% acetylhexosamines and hexosamines, respectively. Hyaluronidase inhibitory effects of the fractions are in the order F3>F2>F1>Ascorbic acid, with F3 having the highest inhibition among the fractions and that of the standard, ascorbic acid. The ESI-MS/MS confirmed the presence of uronic acids on F3, which could be a ^{0,2}A₂ fragment plus loss of methyl group which is very common among non-methylated, sulfated disaccharides.

Keywords: *L. tessellatus* eggs, hyaluronidase inhibitory effect, hyaluronic acid

1. Introduction

Hyaluronic acid and chondroitin sulfate are constituents of the amorphous substances of connective tissues (Mase, Yamauchi, Kato, Esaki, & Isshiki, 2013). Such compounds are natural glycosaminoglycans consisting of D-glucuronic acid and N-acetyl-D-glucosamine repeating disaccharide units, and a sulfated glycosaminoglycan composed of N-acetylgalactosamine and glucuronic acid, respectively. Hyaluronidase inhibitors act upon hyaluronidase (EC 3.2.1.35) enzyme, which degrades hyaluronic acid and chondroitin sulfate. As a consequence, hyaluronidase inhibitors are effective in suppressing allergies and inflammation (Furusawa, Narita, Iwai, Fukunaga, & Nakagiri, 2011) where the interaction of hyaluronidase with hyaluronic acid disrupts basement membrane integrity and produces an angiogenic response, in which the enzyme mediates inflammation via histamine release from

mast cells (Mase et al., 2013). These compounds are potent regulators that maintain hyaluronidase homeostasis and might serve as anti-inflammatory (Girish, Kemparaju, Nagaraju, & Vishwanath, 2009) and anti-allergen agents (Mase et al., 2013), wherein they counteract the implication of hyaluronidase in tumor invasiveness and homeostasis (Madan et al., 1999).

A number of studies have reported that polysaccharides have hyaluronidase-inhibitory effects. Mase et al. (2013) reported the hyaluronidase-inhibitory activity of a polysaccharide obtained from the unicellular marine alga *Porphyridium purpureum*. A high inhibitory effect of an acidic polysaccharide from coffee silverskin on hyaluronidase was also reported by Furusawa et al. (2011). Hyaluronidase activity from *Naja naja* venom was also inhibited by plant-derived bioactive components and polysaccharides, making possible its use as first aid agents in snakebite therapy (Girish & Kemparaju, 2005).

The fish species, *L. tessellatus* is a very good candidate for polysaccharides extraction, since it has low lipid content (0-0.10%) (Lee et al., 2012) and high carbohydrate content of almost 19.50 ± 1.1 % (Ticar et al., 2015). Its eggs contain mucopolysaccharides which are known for its anticoagulant and anti-thrombotic properties (Arivuselvan et al., 2011; Ticar et al., 2015). Given these data for this fish sample, we subjected the polysaccharide fractions of its eggs for hyaluronidase inhibitory effects. Finally, the fraction with the highest hyaluronidase inhibition was characterized by mass spectrometry to confirm its hyaluronic acid nature.

These polysaccharides from *L. tessellatus* eggs could be used as materials in functional foods with anti-allergenic and anti-inflammatory activities. In addition, among sources of polysaccharides evaluated, specifically on marine organisms, this study is novel, since it made use of fish eggs as another source of bioactive polysaccharides.

2. Materials and methods

2.1. Materials and chemicals

Liparis tessellatus eggs were purchased from a fresh market in Tongyeong, Korea. DEAE-sepharose gel, D-glucose, hyaluronic acid, chondroitin sulfate, and hyaluronidase were obtained from Sigma-Aldrich Chemicals (MO, USA). All other chemicals were of analytical grade.

2.2. Isolation and purification of polysaccharides from *L. tessellatus* eggs (PLE)

Polysaccharides from *Liparis tessellatus* eggs (PLE) were isolated according to the methodology described by Ticar et al. (2015). For purification, a concentration of 10 mg/ml of polysaccharide sample was dissolved in deionized water, filtered in a 0.2- μ m membrane and applied to a DEAE-sepharoseTM fast flow column (2.5 \times 30 cm). The column was equilibrated with 0.5 M sodium phosphate buffer (pH 6.5) and a linear gradient of 0-2.0 mol/L NaCl prepared in the same buffer at a flow rate of 0.5 ml/min. Fractions were collected, checked for total carbohydrate content and pooled together based on their sugars content. Each pooled fraction was dialyzed against deionized water by continuous diafiltration until total dissolved solids was equal to zero. This step was performed using a Pellicon UF membrane (Millipore Corp., Mass., USA) with a molecular weight cut-off of 10 kDa. After dialysis, the fractions were lyophilized and labeled as F1, F2, and F3.

2.3. Analytical methods

2.3.1. Polysaccharides composition

Total carbohydrate content was estimated by the phenol-sulphuric acid method using D-glucose as standard (Dubois et al., 1956). Uronic acids, sulfates, and hexosamines contents in

each dialyzed fraction were measured accordingly by using carbazole (Cesaretti, Luppi, & Volpi, 2003) metachromasia (Keler & Nowotny, 1986) and MBTH (Anthon & Barrett, 2002) assays, respectively.

2.3.2. Hyaluronidase inhibition

The hyaluronidase inhibition assay was performed according to the method described by Yahaya & Don (2012) with some modifications. Three concentrations (2.0, 4.0, and 8.0 mg/ml) of F1, F2, and F3 were prepared. The assay medium consisted of 100 µl of hyaluronidase, 100 µl of sodium phosphate buffer (100 mM, pH 7, 37 °C) with 50 mM NaCl and 0.01% BSA, which were mixed with 50 µl of sample solution and incubated at 37 °C for 10 min. The reaction was then initiated by the addition of 100 µl of the substrate in the form of hyaluronic acid solution (0.03% in 300 mM sodium phosphate, pH 5.35) and incubated at 37 °C for 45 min. The undigested hyaluronic acid was precipitated with 1.0 ml acid albumin solution (0.1% BSA in 24 mM sodium acetate and 79 mM CH₃COOH, pH 3.75). The mixture was left to stand for 10 min at room temperature and the absorbance of the reaction mixtures were measured at 600 nm using Spectramax M2 (Molecular Devices, Sunnyvale, CA, USA). All solutions were freshly prepared before the enzyme assay was performed. The absorbance in the absence of enzyme was used as control for maximum inhibition. Ascorbic acid was used as positive control. The percentage of inhibition was calculated as:

$$\% \text{ Inhibition} = (A_{bc}/A_s) \times 100$$

where; A_{bc} = absorbance of control; A_s = absorbance of sample. All the reactions were performed in triplicate.

2.3.3. Enzyme hydrolysis of polysaccharide fractions and mass spectrometric (MS) analysis

The polysaccharide fractions (F1, F2, and F3) together with the standard hyaluronic acid were dissolved in 100.0 mM sodium phosphate/150.0 mM sodium chloride buffer, pH 5.3. The samples were incubated at 37°C with hyaluronidase from bovine testes and chondroitinase ABC. The digestion sample was heated at 100°C for 5 min, then, freeze-dried.

Freeze-dried samples were then permethylated using NaOH in anhydrous DMSO (dimethyl sulfoxide), followed by treatment with iodomethane. Briefly, in a glass test tube, 100 µl of 50% NaOH and 200 µl of anhydrous methanol were mixed together and vortexed. The NaOH suspension was cleaned with anhydrous DMSO several times. Samples were dissolved in 200 µl of dry DMSO by vortexing, then 300 µl of NaOH suspension in anhydrous DMSO was added, followed by 150 µl iodomethane. After mixing for 5 min by vortex, the mixture was sonicated for 10 min. The reaction was quenched by adding 2 ml of water. The excess iodomethane was removed by bubbling with nitrogen gas. Methylene chloride was added to extract the methylated hyaluronic acid by phase separation. Methylene chloride phase was then evaporated. The methylated polysaccharides were dissolved in 50 µl of 80% methanol in 1.0 mM NaOH for MS analysis on a Thermo LCQ ion trap instrument using direct infusion ESI-MS. Eighty percent methanol in 1.0 mM NaOH was used as spray solvent. The sample was introduced by direct infusion at a flow rate of 1 µl/min using a Harvard syringe pump. The capillary temperature was 200°C and the absolute value of the spray voltage was 3.5 kV in positive ion mode. MSⁿ experiments were carried out in positive ion mode and profile mode with normalized collision energy of 30%.

3. Results and discussion

3.1. Fractionation and chemical composition of polysaccharides from *L. tesellatus* eggs

Polysaccharides isolated from *L. tesellatus* eggs (PLE) were composed of 43.2% total

sugars, 12.0% sulfates, 23.9% uronic acids, and 1.7% hexosamine contents. After elution/fractionation of PLE using a DEAE-sepharose fast flow column chromatography, fractions were collected and pooled according to their sugars compositions obtaining: F1 (fractions 39-48), F2 (fractions 53-69), and F3 (fractions 72-79) (Fig. 1). These fractions were collected, dialyzed, lyophilized, and then tested for total sugar, sulfates, uronic acids, and hexosamines contents (Table 1). The constituents of each pooled fraction were: total sugar (78.0-88.5%), sulfates (20.2-26.8%), uronic acids (34.5-45.0%), and hexosamines (7.5-23.5%) for F1, F2, and F3, respectively. These results indicate variations in the ionic strengths of each fraction, which is in agreement with the study conducted by Tabarsa, Han, Kim, & You (2012) on the water-soluble polysaccharides from *Ulva pertusa*, wherein, fractionation of a crude polysaccharide using ion-exchange chromatography also gave fractions of various ionic strengths with different compositions.

3.2. Hyaluronidase inhibitory effects of polysaccharide fractions

Hyaluronidase is an endo-hexosaminidase that initiates the degradation of high molecular weight hyaluronic acid (Moon et al., 2009). The inhibitory effects of fractions F1, F2, and F3 were evaluated toward hyaluronidase using ascorbic acid as standard (Fig. 2). In this method, the hyaluronidase inhibitory effect is inversely proportional to the transmittance of light as measured at a wavelength of 595 nm. The precipitate formed after treatment of hyaluronidase was the amount of undigested hyaluronic acid which blocked the transmission of light into the solution. Thus, the lower the transmittance means the greater the amount of hyaluronic acid left undigested by hyaluronidase and consequently, higher inhibitory effect. The inhibitory capacity of the three fractions, including the standard is in the order: Ascorbic acid < F1 < F2 < F3. Specifically, all the three fractions have higher inhibitions compared with ascorbic acid, a potent hyaluronidase inhibitor (Botzki et al., 2004), with F3 having the highest activity. From Table 1, it can be observed that F3 has the highest content of uronic

acids (45.0%), when compared to the other two fractions. Thus, the high activity of F3 could be attributed to its uronic acids content. In addition, the presence of sulfated glycans in this fraction (Table 1) might probably have a synergistic action with uronic acids increasing its hyaluronidase inhibition. In a study by Schulz et al. (2009), they noticed an anti-inflammatory potential of a hyaluronic acid containing chondroitin sulfate preparation in the treatment of interstitial cystitis and BPS (bladder pain syndrome), yet an imperfectly understood disease, possibly originating from damage to the glycosaminoglycan layer of the bladder epithelium. This observation can affirm the hyaluronidase inhibitory effect of F3 fraction, since, as a hyaluronidase inhibitor, it can deter HA fragmentation which can promote inflammation (Girish et al., 2009). In line with this observation, sulfated hyaluronic acid (sHA) (Benitez et al., 2011; Furusawa et al., 2011) was found out to inhibit the activity of hyaluronidases by a mixed inhibition mechanism, where the tumor-derived hyaluronidase was found out to be inhibited by sHA, a nontoxic hyaluronidase inhibitor. At the same time, sHA also delineated the molecular mechanism through which hyaluronic acid-hyaluronidase (HA-HAase) system might mediate tumor growth and progression, since HA-HAase is known to promote tumor growth and metastasis (Simpson & Lokeshwar, 2008). Hence, inhibition of HA degradation appears critical and imperative in HAase mediated pathological conditions (Girish et al., 2009).

3.3. Analysis of hyaluronic acids on polysaccharide fractions by mass spectrometry

The polysaccharide fraction, F3, having the highest hyaluronidase-inhibitory activity among the three fractions was characterized by mass spectrometry. Figure 4 shows the full ESI-MS spectra of permethylated hyaluronic acids from hyaluronic acid standard and F3 of *L. tesellatus* eggs. Fig. 4(a) shows the repeat unit of hyaluronic acid and Fig. 4(b) lists the hyaluronic acids according to the number of repeat unit. The mass spectrum of permethylated

hyaluronic acid from standard is depicted in Fig. 4(c) which is presented as the repeat unit from 1 to 4. Figure 4(d) shows the Hyaluronic acids detected in F3 of *L. tessellatus* eggs.

The structures of these hyaluronic acids were confirmed by ESI-MS/MS, which are shown in Fig. 5. The m/z 356.1 in Fig. 5(a) could either be a $^{2,5}A_2$ cross ring fragment or a $^{0,2}A_2$ fragment plus loss of methyl group. In non-methylated sulfated disaccharides the $^{0,2}A_2$ fragment is very common, so the second possibility may be more likely.

The unsaturation into the uronic acid moiety at 4,5 position ($m/z=964.8$, Fig. 4) which was detected on F3, was possibly due to the mechanism of enzyme action which involved splitting of a glycosidic bond by elimination reaction, thereby introducing unsaturation in the liberated molecules (Meyer, Davidson, Linker, & Hoffman, 1956). With this data on the characterization of the F3 fraction, its high hyaluronidase-inhibitory activity can be attributed to its high uronic acids content (Table 1) as confirmed in its mass spectrometric analysis. To relate the significance of the 4,5 position of unsaturation of the HA from the mass spectrometric analysis to the isolated HA from the F3 fraction of *L. tessellatus* eggs, an electrophilic addition reaction will likely occur at the unsaturation site (Carey & Sundberg, 2007) and will yield O-sulfated HA, since sulfate molecules are present (Table 1) in the polysaccharide sample. Relatively, a high hyaluronidase inhibitory effect of chemically-modified O-sulfated HA as compared with other O-sulfated glycosaminoglycans was also observed by Toida et al (1999).

Given the hyaluronidase inhibiting effect of the polysaccharide fraction (F3) from *L. tessellatus* eggs, this could be used as anti-inflammation and anti-allergen agents.

4. Conclusions

This study showed that the polysaccharide fractions isolated from *Liparis tessellatus* eggs exhibit hyaluronidase-inhibitory effects. Hyaluronic acids were confirmed on F3 fraction (having the highest inhibitory effect against hyaluronidase enzyme). It can be concluded too, that sulfates detected on the polysaccharide fraction F3 may have synergistic effect in its inhibition against hyaluronidase enzyme. These results are of importance in the engineering of new anti-inflammation and anti-allergenic health products.

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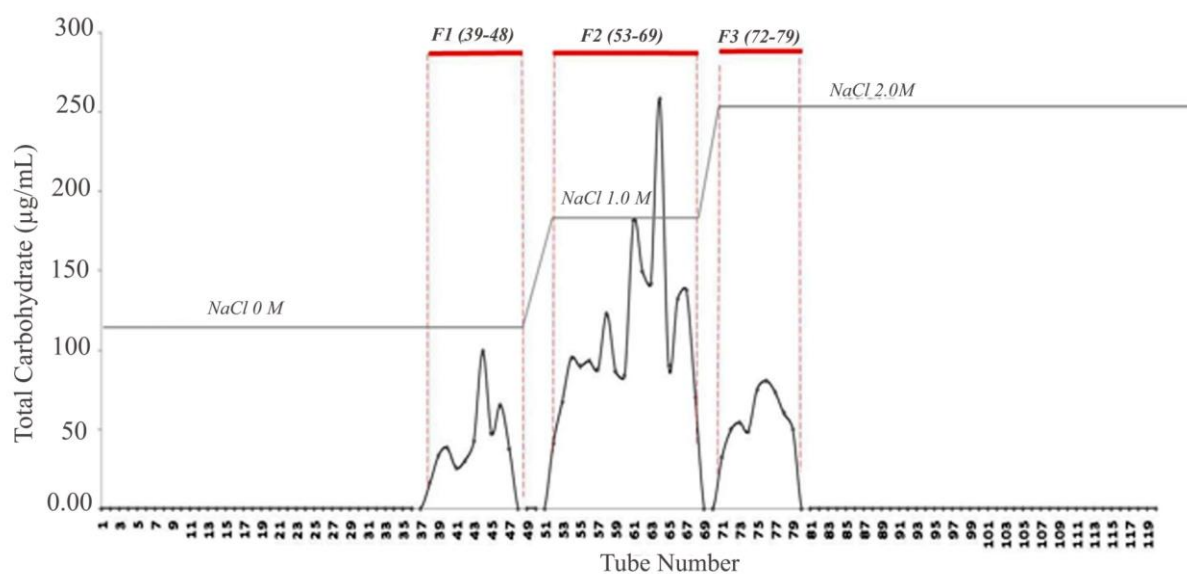


Fig. 1. Elution profiles of polysaccharide fractions isolated from *L. tesellatus* eggs by DEAE-sepharose fast flow column chromatography. Gradient elution was done with (0-2M NaCl) in 0.5 M phosphate buffer. The pooling of fractions was based on the monitoring of total sugar content of individual fractions.

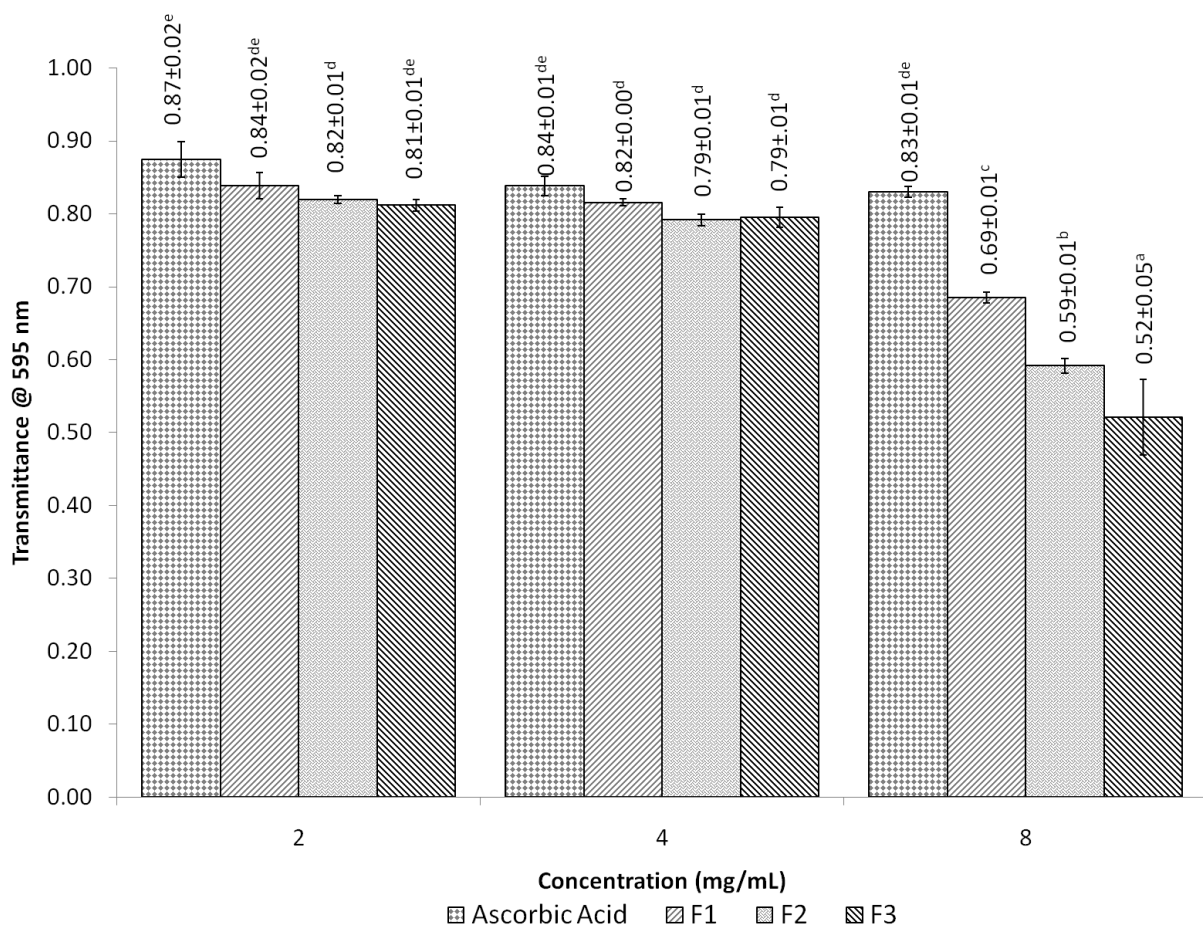


Fig. 2. Hyaluronidase inhibition effects of fraction F1, F2, and F3, obtained by purification of polysaccharides isolated from *L. tesellatus* eggs, compared with ascorbic acid. Values are means ± standard deviation of six replicates. Means that do not share a similar letter are significantly different ($p < 0.05$).

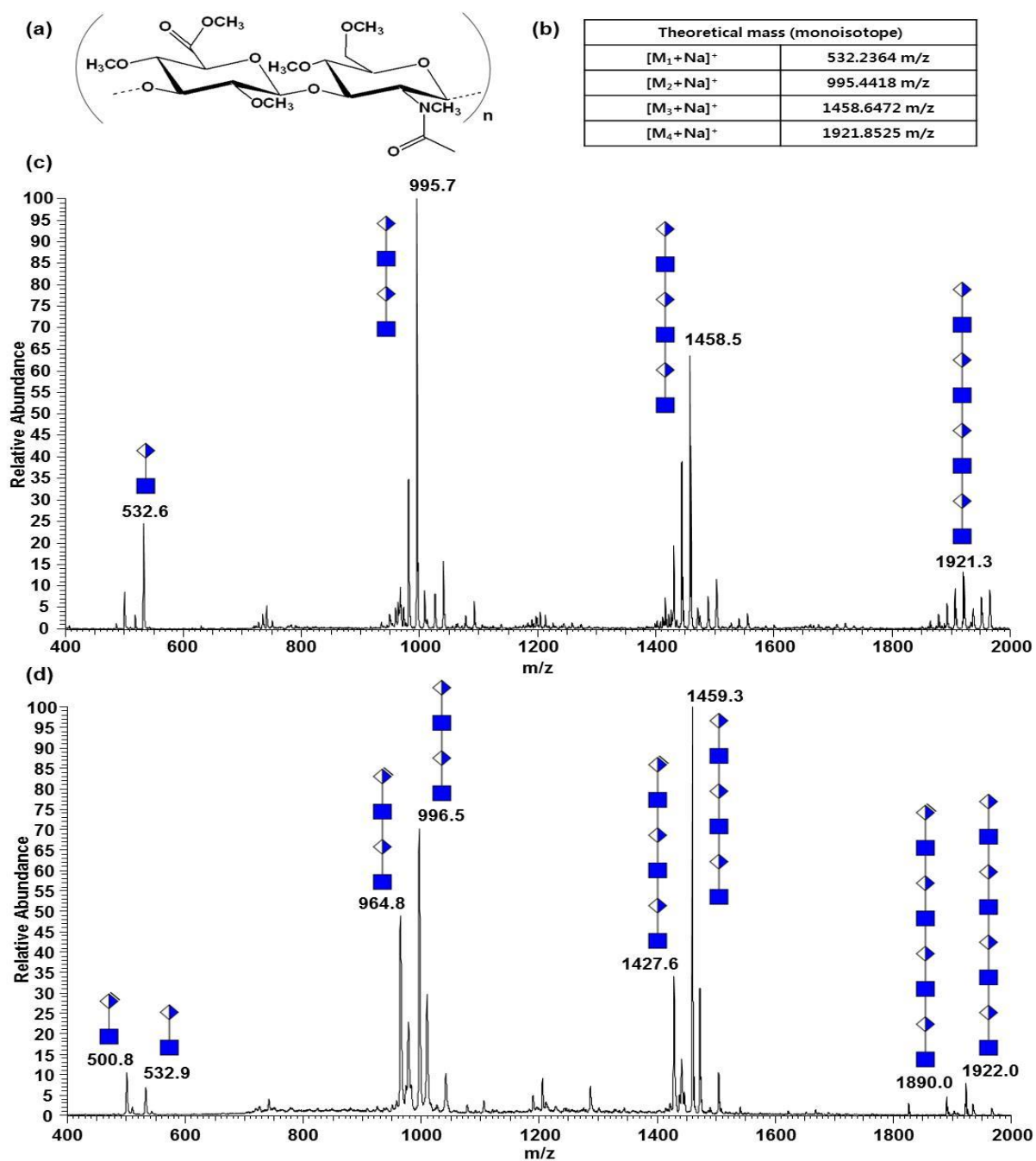


Fig. 4. ESI-MS spectra of permethylated hyaluronic acid (HA). The structure of the oligosaccharide hyaluronic acid (a) and the list of mass (m/z) corresponding to number of repeat unit (b). Mass spectra of permethylated HA from the standard (c) and F3 of *Liparis tesellatus* eggs (d).

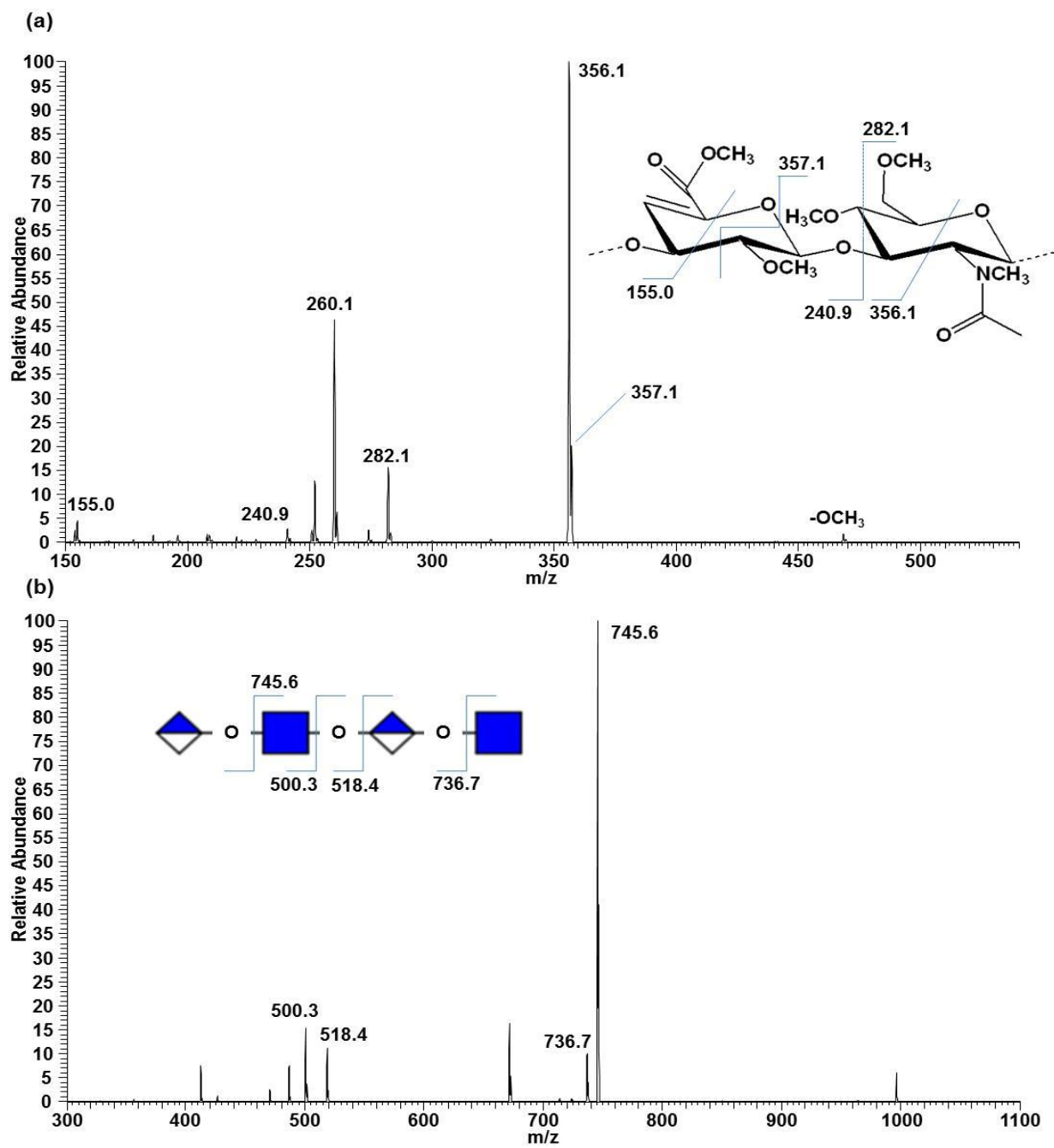


Fig. 5. MS/MS spectra of permethylated hyaluronic acids. MS/MS spectra at m/z 500.8 (a) and at m/z 996.5 (b).

Table 1. Constituents present in the fractions F1, F2, and F3, obtained by purification of polysaccharides isolated from *L. tesellatus* eggs using a DEAE-sepharose fast flow column chromatography.

Fraction	Total sugar (%)	Sulfated glycans (%)	Uronic acids (%)	Acetylhexosamines and hexosamines (%)
F1	88.5±0.3 ^b	26.8±0.5 ^c	34.6±0.1 ^a	23.4±1.2 ^c
F2	81.3±2.6 ^a	22.9±0.1 ^b	35.5±2.6 ^a	19.0±0.1 ^b
F3	78.4±0.8 ^a	20.2±0.1 ^a	45.5±0.8 ^b	7.5±0.1 ^a

Values are means±standard deviation of six replicates. Means in one column that do not share a letter are significantly different ($p < 0.05$).