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Environmental Risk Assessment of Chemicals and Nanomaterials – The Best Foundation for Regulatory Decision-Making?

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Abstract

Environmental risk assessment (ERA) is often considered as the most transparent, objective and reliable decision-making tool for informing the risk management of chemicals and nanomaterials. ERAs are based on the assumption that it is possible to provide accurate estimates of hazard and exposure and, subsequently, to quantify risk. In this paper we argue that since the quantification of risk is dominated by uncertainties, ERAs do not provide a transparent or an objective foundation for decision-making and they should therefore not be considered as a “holy grail” for informing risk management. We build this thesis on the analysis of two case studies (of nonylphenol and nanomaterials) as well as a historical analysis in which we address the scientific foundation for ERAs. The analyses show that ERAs do not properly address all aspects of actual risk, such as the mixture effect and the environmentally realistic risk from nanomaterials. Uncertainties have been recognised for decades, and assessment factors are used to compensate for the lack of realism in ERAs. The assessment factors’ values were pragmatically determined, thus lowering the scientific accuracy of the ERAs. Furthermore, the default choice of standard assay for assessing a hazard might not always be the most biologically relevant, so we therefore argue that an ERA should be viewed as a pragmatic decision-making tool among several, and it should not have a special status for informing risk management. In relation to other relevant decision-making tools we discuss the use of chemical alternative assessments (CAAs) and the precautionary principle.

Keywords

Environmental risk assessment, assessment factors, nonylphenol, nanomaterials, REACH, chemical regulation

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Highlights

- Environmental risk assessments (ERAs) are the preferred choice for informing chemical risk management.
- Two case studies and a historical analysis show that ERAs are not as scientifically well-founded as often perceived.
- ERAs are a pragmatic decision-making tool and should be applied as such, rather than being afforded special status.

1. Introduction

An environmental risk assessment (ERA) is often championed as the preferred decision-making framework for regulators looking to ensure that the regulation and risk management of chemicals and nanomaterials are enforced in the most transparent, objective and reliable way for society (van Leeuwen and Vermeire 1995). An ERA, in many aspects, is regarded as the “holy grail” for addressing risk, one of the major reasons for which is that it is considered the best approach to ensure scientific and evidence-based regulation (Löfsted 2011). In a world where risk perception is believed to be a strong driver of risk management (Slovic 1999), some argue that it is increasingly more important that policymaking is driven by evidence rather than political dogma (Holmes and Clark 2008). ERAs are considered to be a cornerstone in regard to ensuring such evidence-based foundations for regulation, and they now provide the backbone of many pieces of European legislation, such as the water framework directive, biocidal product legislation and chemical legislation known as REACH (EC 2000, 2006 2012).

The fundamental hypothesis on which the ERA paradigm is based is that risk is a function of hazard and exposure. When an ERA is conducted, the hazard and concentration-response assessments, based on the principle that toxicity is concentration-dependent, form the foundation for determining a toxicity threshold. This assessed potency is thereupon used to assess risk by comparing the derived threshold for toxicity with exposure concentrations (EC 2003). This implies that accurate measurements of the hazard and concentration response relationship can be provided, where uncertainties ideally should be negligible or at least well-quantifiable. These experimentally derived assessments thus form the very foundation of ERAs and thereby the “evidence-based” foundations with which they are supposed to provide decision-makers. The four steps of risk assessment (i.e. hazard identification, dose-response assessment, exposure assessment and risk characterisation) were originally proposed by the US National Research Council of the National Academy of Sciences (NRC-NAS) in their landmark 1983 publication “The Red Book” (NRC 1983). During the 1990s, the US EPA adapted the RA framework to ecological risk assessment for assessing risk where human health is not the primary focus. For instance, in 1992, the US EPA published the report *Framework*

1 *for Ecological Risk Assessment*, which proposed principles and terminology for this process (US EPA 1992),
2 which was summarily adopted in the EU via the Technical Guidance Documents (TGDs), although no
3 references are provided within these guidelines (EC 1993a). While its intentions have always been good, the
4 ERA framework has increasingly come under critical scrutiny and has been criticised for not being able to
5 provide the input that risk managers need, and so modifications are currently being discussed in the EU
6 (Scientific Committees 2013).

7 One of the key limitations of the ERA seems to be that risks can only first be truly assessed
8 after an adverse impact has been firmly established scientifically, which is unfortunate when it comes to
9 protecting the environment (EEA 2001, 2013). Article 191 of the Lisbon Treaty states that the protection of
10 the environment ‘shall be based on the precautionary principle and on the principles that preventive action
11 should be taken’ (EU 2007). An important question is therefore whether an ERA can provide sufficient
12 knowledge for decision-makers to, on the one hand, ensure “evidence-based” regulation and on the other
13 hand provide them with enough decision-making support in time to take precautionary preventive actions. In
14 this paper, we argue that the answer to this question is “no.” In order to explain our conclusion, we first
15 analyse how the first two steps of the ERA framework, namely hazard identification and dose-response
16 assessment, are used to inform decision-making in two specific cases. We do this in order to illustrate some
17 of the challenges that ERAs face when it comes to assessing the hazardous nature of chemicals and
18 nanomaterials. The first case considers one of the most comprehensive environmental risk assessments ever
19 performed in the EU, namely in respect to nonylphenol, while the second case examines engineered
20 nanomaterials (ENMs).

21 Based on the nature of the identified challenges, we would argue that they cannot be
22 addressed solely by revising ERAs in the future; rather, they are a reflection of the fundamental limitations
23 of the ERA framework. Via a historical analysis of the development of ERAs, we discuss how these
24 limitations, related to hazard identification and dose-response assessment identified in the two cases, have
25 been well-recognised over time but unfortunately never really addressed. Finally, we discuss how

alternatives such as the precautionary principle and alternative assessment may help to ensure a more timely and transparent foundation for policymaking. First, however, we provide a short introduction to the principles of environmental risk assessment in the EU.

2. Environmental risk assessment in Europe

2.1 Laying down the principles of risk assessment in the EU

Directive 93/67/EEC describes how a risk assessment entails hazard identification, dose (concentration)-response (effect) assessment, exposure assessment for environmental compartments (i.e. aquatic environment, terrestrial environment and air) and risk characterisation (EC 1993b). The objective of the dose (concentration)-response (effect) assessment is to ‘predict the concentration of the substance below which adverse effects in the environmental compartment of concern are not expected to occur’. This concentration is known as the “predicted no-effect concentration” (PNEC) and has to be determined on the basis of information in the notification dossier, e.g. a 21-day study on daphnia magna, testing of higher plant orders and earthworms. A PNEC has to be derived by applying an assessment factor to the values resulting from tests on organisms, e.g. LC50 (median lethal concentration), EC50 (median effective concentration) and NOEL(C) (no-observed-effect level (concentration)) (Table 1). These assessment factors (AFs) are seen as ‘[...] an expression of the degree of uncertainty in extrapolation from test data on a limited number of species to the real environment’, and an AF of the order of 1000 is typically applied to an L(E)C50 value derived from the results of testing for acute toxicity, though it may be reduced in the light of other relevant information. A lower AF is typically applied to a NOEC derived from the results of testing for chronic toxicity, and the AF can be lowered further in cases where more comprehensive data, such as species sensitivity distributions, are available.

Table 1. Assessment factors for deriving a $PNEC_{aquatic}^*$, recommended in Table 16 of the 2003 Technical Guidance document (EC 2003).

Available data	Assessment factor
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At least one short-term L(E)C50** from each of the three trophic levels of the base set (fish, daphnia and algae)	1000
One long-term NOEC*** (either fish or daphnia)	100
Two long-term NOECs from species representing two trophic levels (fish and/or daphnia and/or algae)	50
Long-term NOECs from at least three species (normally fish, daphnia and algae) representing three trophic levels	10
Species sensitivity distribution (SSD) method	5-1 (To be fully justified case by case)
Field data or model ecosystem	Reviewed on a case by case basis

* PNEC_{aquatic} : predicted no effect concentration for the aquatic environment

** L(E)C50 : lethal(effect) concentration for 50% of the test specimens

*** NOEC: no observed effect concentration

The final step in the risk assessment methodology entails comparing the predicted exposure concentration (PEC) with the PNEC for any given compartment, so that a PEC/PNEC ratio may be derived. If the PEC/PNEC ratio is ≤ 1 , it implies that there is no immediate concern according to the available information. If the ratio is ≥ 1 , the competent authority shall judge whether: 1) the substance is of concern and further information is required for the revision of the assessment, but it shall defer a request for that information until the next tonnage threshold is reached, 2) the substance is of concern and further information shall be requested immediately or 3) the substance is of concern and the competent authority shall immediately make recommendations for risk reduction. The type and amount of data required for the ERA is based on the production volume of the chemical, because with a greater production volume comes a greater demand for experimental data. The tonnage threshold thus refers to a production volume where the required set of data is expanded to include additional data. If it has not been possible to derive a PEC/PNEC ratio, the risk characterisation shall entail a qualitative evaluation of the likelihood that an effect will occur under the expected conditions of exposure.

2.2 The emergence of the Technical Guidance Documents

Although Directive 93/67, Regulation 1488/94 and Directive 98/8 lay down general principles for the risk assessment of new substances, existing substances and biocidally active substances or substances of concern present in a biocidal product, they do not include extensive technical details for conducting such risk assessments. In 1993 and 1994, the European Chemical Bureau of the European Commission published the

1 first set of “Technical Guidance Documents on Risk Assessment” (EC 1993a, 1994), which were prepared in
2 order to support Commission Directive 93/67/EEC on Risk Assessment for newly notified substances,
3 Commission Regulation (EC) No 1488/94 on Risk Assessment for existing substances and Directive 98/8/EC
4 of the European Parliament and of the Council concerning the placing of biocidal products on the market.
5 Key aspects of the risk assessment methodology, such as PEC/PNEC-derivation, the use of assessment
6 factors (Table 1) and the four steps of risk assessment, were described in these first TGD versions.
7 Regrettably these TGDs provide no discussion on the uncertainties or the scientific foundations of these
8 significant methodological choices, and it is unclear on which kinds of evidence and insight these are based.
9 In 2003, the TGDs were updated with respect to ERAs, and the effect assessment took on major
10 improvements (e.g. a new chapter on marine risk assessment was added). In 2007, a new regulation for
11 industrial chemicals, called REACH (short for: Registration, Evaluation, Assessment and Restriction of
12 Chemicals), was adopted (EC 2006). Under REACH, manufacturers and importers of chemical substances
13 are required to carry out chemical safety assessments (CSAs) when producing or importing chemicals in
14 quantities of 10 tonnes or more per year. The European Chemical Agency has provided substantial guidance
15 on how it would like industry to prepare these CSAs, and although the terminology is different, the key
16 aspects of the CSA are similar to the chemical risk assessment methodology described in the TDG prepared
17 by the ECB (ECHA 2008).

18 The (largely) experimentally derived PNECs thus form the foundation of the “evidence-
19 based” input into the decision-making process in Europe, in both a historical and a future context. It is
20 therefore interesting to analyse the nature of the science that is being applied within ERAs aiming at deriving
21 PNECs.

3. Hazard and dose-response assessment of nonylphenol

Nonylphenol is an industrial chemical mainly used in nonylphenol ethoxylates production (NPEO) (85% of total production) (Nielsen et al. 2000), which in turn is used as a detergent for industrial cleaning, as a stabiliser in plastics such as PVC and NPEO and in paint formulations (ECHA 2014). In 1997, nonylphenol had a yearly production volume of 73,500 tonnes (EC 2002), making it a high-volume production chemical at the time. It was chosen as one of the substances to be risk assessed under the old European Regulation 793/93 (EC 2002), due to its high production volume and toxic properties. The final risk assessment was published in 2002, building on a review of the scientific data completed in 1999. The RA provides a ‘comprehensive risk assessment of 4-nonylphenol (branched) and nonylphenol’ (EC 2002), and so it is therefore well-suited to illustrating some of the challenges in environmental risk assessments, even for ERAs that were considered and portrayed as scientifically “state-of-the-art” at the time.

3.1 Hazard identification of nonylphenol

ERA hazard identification was based on a review of studies conducted by the EU rapporteur (UK authorities). The report concludes that endpoints such as growth and survival were the most sensitive, and these endpoints are therefore used for assessing PNECs (Table 2) (EC 2002).

3.1.1 Endocrine disrupting effects and “new” versus traditional endpoints

Even though the reported endocrine disrupting effects of nonylphenol are discussed in the ERA report, it is concluded that their threshold values are higher than those for more “traditional” endpoints (EC 2002). The ERA arrives at this conclusion because oestrogenic effects started around 10-20 µg/l whereas the PNEC was estimated based on a long-term NOEC for algae of 3.3 µg/l (see below). The risk assessment of endocrine disruptive chemicals (EDCs) has evolved markedly since the ERA was conducted, and further research into the endocrine disruptive properties of nonylphenol indicates that not all isomers are capable of inducing oestrogenic activity (Soares et al. 2008). If this is not accounted for in the experimental set up it can influence the NOEC, since a mix of isomers will result in lower exposure to the isomer that

induces oestrogenic activity (i.e. the *para*-position isomer). It is furthermore debatable whether EDCs have thresholds or whether they should be treated as being similar to non-threshold genotoxins (EC 2013). If the latter is actually the case, a single *para*-position isomer would, in theory, be sufficient to cause risk. Lee et al. (2003) also found that nonylphenol has anti-androgenic properties, which illustrates that a threshold based on oestrogen-like properties alone might underestimate the real-life risk to populations that is a function of all endocrine disruptive properties. Nonylphenol has recently been re-evaluated under REACH, and based on this evaluation it was concluded that ‘4-Nonylphenol, branched and linear[...] are identified as substances of very high concern (SVHC) in accordance with Article 57 (f) of Regulation (EC) 1907/2006 (REACH) because they are substances with endocrine disrupting properties’ (ECHA 2012). This case thus serves as an example of the limitations that the boundaries of our scientific understanding set for our ability to assess hazard. Today, it would be much more controversial to argue that endocrine disruptive properties are properly accounted for with the survival and growth type of endpoints.

3.1.2 Non-standard tests vs. standard tests

Another related discussion concerns whether data from non-standard tests that are more sensitive than standard tests should be used in ERAs for chemicals. In this respect, the focus has recently been on Bisphenol-A, whereby the US Food and Drug Administration and the European Food Safety Authorities were criticised for neglecting relevant scientific findings because they were not conducted under good laboratory practice (GLP) (Myers et al. 2009). A similar discussion is relevant for nonylphenol. In a 2013 hearing on the decision to classify nonylphenol as an SVHC, several stakeholders argued that the classification of nonylphenol as an EDC was not justified, since it was based on “poor” studies, as defined by the Klimisch score (ECHA 2013), an approach published by Klimisch et al. (1997) for evaluating experimental data for RAs. The principle is that studies are evaluated based on a number of parameters and then categorised within one of four categories, where categories 1 and 2 are considered ‘reliable without and with restrictions’, respectfully. Category 3 is considered ‘not reliable’, whereas category 4 studies are

deemed ‘not assignable’ (Klimisch et al. 1997). High emphasis is placed on studies conducted with standard protocols – and preferably under GLP. Among different responses the ECHA states that a non-monotonic dose-response relationship should not disqualify a study, since such relationships are documented for EDCs and thus should not be characterised as experimental artefacts (ECHA 2013). This illustrates that a very rigid interpretation of what constitutes reliable science may have a tendency to disqualify novel findings thus inhibiting well-timed decision-making.

3.2 Dose response assessment of nonylphenol

The second step in the ERA towards quantifying the hazardous potential of nonylphenol was the dose response assessment aimed at deriving PNECs. Two aspects of the 2002 ERA, which are addressed below, are of specific interest in the context of this paper.

3.2.1 Dismissal of indicative studies and studies with unknown biological significance

The lowest toxicity values for fish, invertebrates and algae (Table 1) formed the basis of the derived PNEC values, following the TGD procedure (EC 1993a). It is interesting to analyse the scientific foundation behind PNECs, in order to evaluate to what extent they are based on the ‘high level of accuracy’ that ERAs should provide (EC 2013).

A chronic NOEC for invertebrates of 24 µg/l originates from a daphnia reproduction study (Table 1), even though this study does not provide the lowest NOEC among the assessed studies. A study by Kahl et al. (1997) found NOEC in the range of 14-45 µg/l for irregularly shaped egg masses, even though the effect was not statistically verified. However, since the biological significance of this observation was unknown, these data were not used further (EC 2002). In another study of the rainbow trout, *Oncorhynchus mykiss*, significant reductions in body weight were observed at 1 µg/l (Ashfield et al. 1998). Since this experiment was conducted with nominal concentrations, and experimental verifications of the concentrations were made, it was only used as an indicative study (EC 2002). The data used to determine the PNEC were

EC₁₀ of 3.3 µg/l from a chronic algae test with *S. subspicatus* (EC 2002), thus ensuring that the PNEC were lower than that derived from the egg mass study, albeit not the *O. mykiss* study.

Table 2. Top part: effect data from the studies that were used to derive PNECs in the European RA of nonylphenol. Bottom part: other scientific studies that could have altered the conclusion of the RA, if they had been available for consideration.

Type of test species	Endpoints for short-term tests	Endpoints for long-term tests
Fish	96 h LC ₅₀ : 0.128 mg/l (<i>P. promelas</i>)	33 d NOEC _{survival} : 0.0074 mg/l (<i>P. promelas</i>)
Invertebrate	96h. EC ₅₀ : 0.0207 mg/l (<i>H. Azteca</i>)	21 d. NOEC _{survival offspring} : 0.024 mg/l (<i>D. magna</i>)
Algae	72 h EC _{50(biomass)} : 0.0563 mg/l (<i>S. subspicatus</i>)	72 h EC _{10(biomass)} : 0.0033 mg/l (<i>S. subspicatus</i>)
Other relevant studies (post 1999)		
SSD	HC ₅ : 1.43 µg/l (freshwater) and 0.84 µg/l (seawater) Gao et al. (2014)	
Genotoxicity	Photoactivated nonylphenol induce oxidative DNA damage (Okamoto et al. 2006)	
Mixture effects	nonylphenol interact in mixtures causing deviation from additivity (Rajapakse et al. 2004)	
EDC	Isomer specific oestrogen-like properties (Soares et al. 2008) Anti-androgen properties (Lee et al. 2003)	

3.2.3 Disregarding real exposure to mixtures

Yet another aspect that is not accounted for in the ERA is to what extent real-life exposure to mixtures containing nonylphenol affects the threshold for toxicity. Such mixture effects will vary depending on mixture composition, as shown by Rajapakse et al. (2004), who illustrate that PNECs based on single chemical ERAs are artefacts that do not reflect real-life toxicity. Oestrogenic effects, measured with the E-SCREEN assay of a mixture containing nonylphenol, deviated from additivity, indicating that it could be problematic to assess mixture effects on the basis of single chemical toxicities (Rajapakse et al. 2004). In another study, Kwak et al. (2001) showed that the mixture effects of nonylphenol and Bisphenol-A were more potent than single chemicals when effects were measured with an array of EDC-sensitive endpoints in the viviparous fish *Xiphophorus helleri*. These studies were published after the RA, but the problem caused by oestrogenic mixtures active substances had been highlighted earlier (Arnold et al. 1996), and participants in a US-EPA sponsored workshop concluded that mixture effects represented one of primary research focus areas for EDCs (Kavlock et al. 1996). Since the PNECs derived in the ERA are based on single toxicity studies, none of them can be said to account for the combined toxicity.

4. Hazard and dose-response assessments of Engineered Nanomaterials

The environmental risks of nanomaterials have been subjected to increased levels of attention in recent years. Risk assessments have repeatedly been proposed by expert committees, policymakers, industry organisations and non-governmental organisations as the central means for informing decision makers about the risks of engineered nanomaterials (ENMs) (Nordan et al. 2006, SCENIHR 2007, US EPA 2007, ED & DuPont 2007, CCA 2008, EFSA 2008, Hankin et al. 2011). An important question is therefore whether the existing ERA framework provides experimental tools that adequately ensure a proper scientific foundation for regulating ENMs within a reasonable period of time.

4.1 Hazard identification of Engineered Nanomaterials

4.1.1 Recognising ignorance about the hazard characteristics of emerging materials and substances, but still ignoring it

When it comes to nanomaterials, there seems to be a general agreement that hazards depend on surface area, surface charge, surface chemistry, state of agglomeration as well as chemical composition (Hansen et al. 2007, Hankin et al. 2011), and especially surface area/reactivity has been mentioned as the newest “nano-relevant” properties for inclusion in hazard identification. This is in contrast to regular chemical risk assessments, which are based on the fact that chemical identity governs the fate and effects of a chemical.

However, all of the mentioned particle characteristics may affect the overall hazard, and since causal relationships still need to be discovered, further research is needed in this area before relevant data demands for hazard identification purposes can be defined. It is further discussed whether the unique properties of each type of nanoparticle make their hazard unique (Hansen et al. 2007; Hartmann et al. 2014). Toxicodynamics for hazards is yet another important area largely governed by uncertainty. Several studies have shown that the uptake and depuration of ENMs is different from results observed for other particles of similar metal composition (e.g. Dai et al. 2013, Khan et al. 2013). To what extent these differences influence the long-term toxicity of ENMs is still unknown, and this lack of understanding stresses the importance of

1 defining the most relevant endpoints for determining ENM hazard. The equivocal identification of hazards
2 for most nanoparticles is furthermore hampered by substantial limitations in our ability to determine
3 individual and multiple particle characteristics simultaneously and in a consistent manner, both prior to and
4 during tests, when using different characterisation techniques and/or across laboratories (RCEP 2008, JRC
5 2014).

6 All the unknowns about hazard characteristics have been well-recognised when it comes to
7 nanomaterials, and this area could therefore be classified as ‘recognised ignorance’ (Hansen et al. 2013).
8 Although they are often recognised in the few risk assessments that have been published (WHO 2013), the
9 problem is that they are continuously ignored in these risk assessments when it comes to actual hazard
10 identification, and so most often classical hazard characteristics are relied upon.

11 **4.1.2 Most sensitive endpoint?**

12 It is furthermore unclear as to whether or not we can rely on knowledge from industrial
13 chemicals when selecting the most sensitive endpoints, since particle hazards have not been considered
14 historically for risk assessment. As noted in the second REACH Implementation Project on Nanomaterials
15 (RIP-oN 2) (Hankin et al. 2011), some novel endpoints and observations in fish and invertebrate tests may
16 provide useful information when it comes to nanomaterials. For fish tests these novel endpoints included fish
17 ventilation rates, gill pathologies, mucus secretion, brain pathology and animal behaviour based on the
18 findings proffered by Schmidt (2007) and Federici et al. (2007). For invertebrates, Lovorn et al. (2007) found
19 that the heart rate, hopping frequency and appendage movement cycle frequency of daphnia were affected by
20 exposure to nanomaterials, which again is not something normally considered in chemical risk assessment.

21 Therefore, it remains unknown as to whether or not the endpoints currently tested and
22 reported on are actually the most sensitive or the most relevant examples, or whether new biological
23 endpoints might actually be more relevant (RCEP 2008, Hartmann et al. 2014).

4.2 Dose-response assessment of Engineered Nanomaterials

4.2.1 When the mass-based dose does not make the poison

Several ecotoxicological studies have reported dose-response relationships for tests with nanomaterials (Hund-Rinke and Simon 2006 and Heinlaan et al. 2008, Kühnel et al. 2009), whereas others have found none (Adams et al. 2006). It is therefore still debatable whether it is possible to establish reliable dose-response scenarios for ENM, due to a number of factors. First of all, nanomaterials have consistently been observed to be present in various environmental compartments in the form of individual primary particles as well as agglomerates and aggregates, whereas specific surface area is constant for individual particles and agglomerates, since the particles bind loosely together and aggregation causes a decrease in specific surface area due to the much stronger chemical bounds between the particles (Oberdörster et al. 2007, Aitken et al. 2011). It is well-known that aggregation and agglomeration are dependent on the coating and functionalisation of the nanomaterials, and that the level of agglomeration and aggregation varies between different test media due to a number of biotic and abiotic factors such as pH, salinity, exposure to sunlight, stirring activities and the presence of natural organic matter. Although the latter is not normally considered an element of standardised ecotoxicological testing of chemicals, it might be very important for understanding agglomeration and aggregation behaviour in the environment itself (Fortner et al. 2005, Hartmann et al. 2014). For most nanomaterials there is a general hypothesis that ecotoxicity is linked to surface area (Dai et al. 2013), which might be due to increased surface reactivity and/or an increase in ion release from the surface as a function of the larger surface area. Normally, one would assume that the higher the concentration of nanoparticles, the higher the available specific surface area (SSA) and, subsequently, ecotoxicity, but this assumption is not necessarily true, as aggregate formation can be concentration-dependent, meaning that in the presence of a higher concentration of nanoparticles the likelihood of aggregation increases, thereby resulting in a decrease in available SSA. This means that we are likely to see increasing levels of ecotoxicity as concentrations of individual particles and agglomerates increase, but only until a certain level, when the concentration becomes so high that concentration-dependent aggregation is

initiated and overall ecotoxicity decreases (Baun et al. 2009). This would mean that a traditional s-shaped dose-response relationship would not be observed, since more than one concentration would be linked to specific effect concentrations (e.g. EC50), thereby complicating the derivation of the PNEC. The hearing on the NP RA, described in the earlier case, illustrates that a move away from the monocausal relationship between concentration and effect lowers the credibility of the data – at least for some stakeholders.

4.2.2 Limited exploration of other suitable dose descriptors

Yet another aspect that complicates the dose-response assessment of ENM relates to particle hazards. Since there is no common agreement on the most important drivers for ENM hazards, it is debatable which metric is most suitable as a dose descriptor. Several alternatives to traditional mass have been proposed, such as surface area or number of particles, but only a few authors have actually investigated alternative metrics for expressing the dose. A number of studies have reported observing a size-dependent effect. Templeton et al. (2006), for instance, compared “as prepared” and purified SWCNT and observed a difference in adverse effects in an estuarine copepod, which they attributed to a partly size-dependent and partly aggregation-dependent difference. For non-aggregating SiO₂ nanoparticles, van Hoecke et al. (2008) showed that increasing effects on the growth rate of algae with decreasing particle sizes can be explained by an increase in specific surface area (van Hoecke et al. 2008).

5. The Origin of the Environmental Risk Assessment

The cases above illustrate some of the challenges that ERAs face when it comes to hazard identification, dose-response assessment and addressing uncertainties and ignorance. Although one could argue that these challenges are related only to nonylphenol and nanomaterials, we would contend that this is not the case. In 2001 and 2013, the European Environment Agency published two reports, “Late Lessons from Early Warning,” exploring numerous cases where regulators had failed to regulate hazardous chemicals in time to protect human health and the environment (EEA 2001, 2013). Many of the cases analysed in the EEA reports were very similar to the ones highlighted herein when it comes to realising the limitations of risk assessment. In order to understand the origin of these systemic challenges, one has to understand the origin of PNEC derivation and how it has helped the ERA to develop into one of the most used decision-support frameworks.

Key aspects of the risk assessment methodology, outlined in the Directives laying down the principles for risk assessing chemical substances and the TGD, such as PEC/PNEC-derivation, the use of extrapolation factors and the four steps of risk assessment, were introduced into the Directives and the first version of the TGDs from 1993 and 1994. Prior to 1993, discussions on how to assess the environmental risk of toxic chemicals had been going on in the United States of America and in many European countries such as Denmark, Germany, Spain and the Netherlands (van Straalen and van Leeuwen 2002). A thorough analysis of the scientific discussion in these historical documents can therefore provide a better understanding of the scientific foundation for the TGD – and thus the foundation of European environmental risk assessment.

5.1 The Dutch Health Council, minimal data requirements and reliable procedures for the derivation of PNEC

In the Netherlands, the Health Council of the Netherlands established a scientific advisory committee to give advice on ecological risk assessments of chemicals (HCN 1989). In 1989, the council published an English version of its advice, termed “Assessing the risk of toxic chemicals for ecosystems,” which provides insights into the now well-established minimal data requirements, the use of PNEC and the development of procedures for deriving PNEC.

The request for advice on ecological risk assessment, dating back to December 1987, came from the Minister of Welfare, Health and Cultural Affairs in the Netherlands, and the task of the committee was to assess “a procedure” to derive recommended values for ecosystems. The “procedure” had been proposed in August 1987 by an interdepartmental working party, “Risk Management for Ecosystems,” and it outlined the minimum data required and the methods which would be applied to derive recommended values for (part of) ecosystems, despite limited knowledge. The proposal made by the working party involved recommendations in regard to water, soil and air, and although it is now almost four decades since their inception, these minimum data requirements and methods provide the backbone of existing chemical ERAs in the EU.

For water, the Working Party Risk Management for Ecosystems (1987) proposed that acute toxicity data for algae, daphnia and fish, according to the OECD test guidelines, are to be a minimum requirement. The lowest LC(E)50 should then be compared with (expected or measured) exposure concentration. If the ratio is > 100 , a recommended value is to be derived by using the Kooijman (1985 and 1987), the Van Straalen (1987) and the Slooff et al. (1986) methods (Working Party Risk Management for Ecosystems 1987) (see Table 3). If the ratio is < 100 , long-term experiments should be carried out, e.g. reproduction tests on daphnia and an egg larva test with fish according to OECD test guidelines, and the recommended value should be derived using the above methods (Working Party Risk Management for

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Ecosystems 1987). The approach was thus comparable to those used in current regulations (e.g. REACH), where risk quotients form a basis for assessing risk.

For soil, acute toxicity data are negligibly required for plants, earthworms and soil arthropods, whereas an inhalation test with a mammal, according to OECD guidelines, and fumigation experiments with plants and insects should be carried out for air quality. If a ratio > 100 , a recommended value is to be derived by using the Kooijman (1985 and 1987) and the Van Straalen (1987) methods. If the ratio is < 100 , long-term experiments should be carried out with the abovementioned organisms, and the recommended value should be derived using the above methods as for water (Working Party Risk Management for Ecosystems 1987).

For mixture toxicity, it is noted that it is an important aspect of risk assessment but could not be taken into account at the time (i.e. August 1987) in the derivation of recommended values, because there are no instruments that deal properly with this problem, which again is noted as an undesirable situation (Working Party Risk Management for Ecosystems 1987).

The methods put forth by Kooijman (1985 and 1987), Van Straalen (1987) and Slooff et al. (1986) (see Table 3) play a very important role in the proposed procedure, and they were used to derive reliable procedures for extrapolating experimental data to environmental effects that the Health Council committee was asked to evaluate.

Table 3. Overview of the three extrapolation concepts that were the foundation of the Dutch committee's recommendations on how PNEC should be derived. The general approach and basic scientific foundations are listed.

Name	Approach	Scientific foundation	Reference
Kooijman method	Extrapolate from LC_{50} values from most sensitive species	- HCS* derived from geometrical mean LC_{50} value from several species. - HCS is divided with a safety factor that is dependent on the number of species observed in the community.	Kooijman (1987)
Van Straalen method	Estimate where NOAEL is not exceeded for 95% of species	- HCS similar to Kooijman method is calculated, but NOAEL** values are used instead of LC_{50} values. - Safety factor is constant, since the HCS is derived in regard to 95% of all species in the ecosystem.	Van Straalen (1987)
Slooff et al. method	Applies regression analysis to large numbers of toxicity data	- Regression analysis allows for predictions of: - LC_{50} effects on one species if the LC_{50} is known for another species. - Chronic NOEC*** can be derived from acute LC_{50} - NOECeco**** based on lowest $L(E)C_{50}$ or lowest chronic NOEC.	Slooff et al. (1986)

* HCS: Hazardous concentration for sensitive species

1 ** NOAEL: No Observed Adverse Effect Level

2 *** NOEC: No Observed Effect concentration

3 *** NOECeco: No Observed Effect concentration for the ecosystem, based on a calculated relationship between field data and acute
4 tests.

5
6
7 Besides evaluating these three methods, the committee also included US EPA (1984), Stephan et al. (1985)
8 and Blanck (1984) in their review, concluding that:

- 9 1) All methods offer some possibilities for including ecological aspects, for example in selecting test
10 species and confidence limits.
- 11 2) None of the methods takes interaction between species into account, even though it is a key feature
12 of ecosystems, except for the method posited by Sloof et al. (1986), which uses field data.
- 13 3) The Kooijman, Van Straalen and US EPA methods can be used for all environmental compartments,
14 whereas the methods employed by Sloof et al. (1986) and Blanck (1984) are only applicable for the
15 aquatic compartment.
- 16 4) The Kooijman (1987) method yielded the lowest results in all cases when used on the chemicals γ -
17 hexachloro-cyclohexane and cadmium, due to a stricter protection principle and the fact that results
18 decreased more substantially as input data variance increased.

19 The committee expressed serious objections to the applicability of the methods developed by the US EPA
20 (1984) and Blanck (1984). In regard to the US EPA method, the committee objected to the use of the
21 triangular (probabilistic) distribution of toxicity data, which implied the existence in ecosystems of
22 concentration thresholds below which the probability of adverse effects are zero. This assumption the
23 Committee found to be too rigorous (HCN 1989). Blanck (1984) used seven species whose EC50 values he
24 had derived, and the committee found it to be an extremely drastic assumption that no species was capable of
25 reacting more sensitively than these seven (HCN 1989).

26 Besides the conclusion related to methods for deriving reliable procedures for the
27 extrapolation of experimental data to environmental effects, the committee also discussed and noted a
28 number of aspects related to the proposed procedure and ecological risk assessment.

1 First, they noted that the purpose of the procedure was to predict the effects of toxic chemicals
2 on ecosystems and to establish boundary conditions for the protection of ecosystems despite the limited
3 knowledge available on ecotoxicology at the time (HCN 1989). Nonetheless, according to the committee,
4 there were insufficient indications relating to the possibilities and limitations of such a procedure.

5 Second, the key to making such an assessment, according to the committee, was the
6 extrapolation from experimental data to effects in ecosystems. The proposed procedure and risk assessment
7 thus aimed at protecting the characteristics of the species composition of the ecosystem but, according to the
8 committee, it was not possible to conclude from the available data whether an ecosystem would react more
9 sensitively to a toxic chemical than individual species. The committee stressed that no safe values could be
10 derived with the procedure but only limits above which certain effects would occur (HCN 1989).

11 Finally, the committee discussed to what extent a risk assessment can make scientifically
12 certain recommendations to policymakers. The committee concluded that none of the reviewed ERA
13 procedures could claim to safeguard ecosystems against the adverse effects of toxic chemicals. However,
14 they provided a first impetus to putting present knowledge to use in a practical way for the protection of
15 ecosystems (HCN 1989). The committee stated that a complete scientific foundation for risk regulation was
16 not possible, and policy could therefore not be based on such a requirement (HCN 1989). The committee
17 also highlighted some areas that were not covered properly in any of the approaches, concluding that the risk
18 assessment procedure could not be applied in the case of mixed toxic chemicals or for carcinogenic or
19 mutagenic chemicals (HCN 1989). The committee recommended that the effects of exposure to chemical
20 mixtures should be included as far as possible (HCN 1989).

21 Advice from the Health Council was later implemented in the Netherlands in the form of a policy
22 paper on risk management and fed into processes that were going on at the time at the OECD and in the EU
23 (HCN 1989, Premises for Risk Management 1989; OECD 1992; Straalen and van Leeuwen (2002)).

5.3 The OECD, the US EPA and three tiers of extrapolation factors

The three tiers of extrapolation factors represent another key aspect of the environmental risk assessment framework and helps explain the systemic challenges that we have observed in the case of nonylphenol and nanomaterials. In the early 1990s, transatlantic dialogue was initiated also under the umbrella of the Hazard Assessment Advisory Board of the Organisation for Economic Co-operation and Development (OECD). When it comes to risk assessment, the proceedings of an OECD workshop in Arlington, USA, are noteworthy, as they were published at the same time as the TGDs were being prepared (OECD 1992). The aim of the workshop was to present state-of-the-art extrapolation methods, to identify areas of uncertainty and to reach consensus on practical, available and validated extrapolation methods (OECD 1992). According to van Straalen and van Leeuwen (2002), the methodology was implemented on a wider scale after the OECD workshop.

Participants in the OECD workshop recommended three tiers of extrapolation factors, each with a factor of ten. The derivation of the three assessment factors was based on empirical experience rather than a theoretical model. The approach was inspired by the approach that the US EPA (1984) had developed in 1984, published in a report named “Estimating concern levels for concentrations of chemical substances in the environment” (US EPA 1984), in which they evaluated how uncertainty can be addressed by applying assessment factors (AF). The publication discussed the scientific rationale behind applying four tiers of AF (Table 4), which have subsequently become more or less standardised AFs for environmental risk assessment. The authors stressed that using AFs did not identify a safe exposure level in ecological risk assessment (comparable to considerations made by the Dutch committee on risk assessment analysed above), and thus it was not directly comparable to the margin of safety (MOS) derived for human health RAs (US EPA 1984). An MOS is used when there is sufficient data to determine a safe level of use, whereas AFs are used when data are absent. The AF was created to account for many experimental and environmental factors that alter test results away from field-level effects, but due to convenience all these differences were grouped

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in three categories (species-to-species sensitivity, chronicity and laboratory-to-field differences). For the purposes of ease and simplicity, all AFs were also rounded off to the nearest power of ten (US EPA 1984).

Table 4: The three assessment factors recommended by the US EPA as a means of handling extrapolation from different data scenarios to real life.

Categories	Level of scientific data at hand	Recommended AF	Scientific rationale
	Field data	1	Field studies measure effects that are expected to occur in natural environments.
Laboratory-to-field	LOEC based on chronic study	10	a) Other species may be more sensitive. b) Test conditions may not represent natural environment. c) Other effects may occur at lower levels. d) The presence of other chemicals in the environment may amplify the effect.
Acute-to-chronic	LOEC based on many acute concentrations	100	e) At least one acute test for each of the three taxonomic groups (fish, invertebrates, algae) or f) At least five acute tests divided among two of the three taxonomic levels. g) Use of juvenile organisms can underestimate effects on eggs and embryos.
Species-to-species	One acute concentration or QSAR	1000	h) No one or two species are consistently more sensitive to a broad array of chemicals. i) Actual vs. estimated (QSARs) LC ₅₀ value may vary up to 10 fold.

The scientific rationale behind these three categories of AFs is blurry. In order to have a species-to-species ratio of 10, the US EPA (1984) argued that data from three fish species (e.g. fathead minnow) and two crustacean species (e.g. *Daphnia magna*) were largely representative of all relevant species' sensitivity. It was further argued that test requirements could therefore be limited to fish and crustaceans (the so-called "cluster concept"), since it was not reasonable to require tests for all relevant species. Algae were subsequently added as a third group, even though no specific data support the inclusion (US EPA 1984).

For the acute-to-chronic ratio (ACR), it was concluded that the data supported an additional AF of 10 despite substantial variations (US EPA 1984). The largest ratio reported was 17.551 for the herbicide Propanil, which was tested on fathead minnow (Call et al. 1983). On a more general level a statistical study of 95 chemicals showed that the median ACR was 8.46, leading to the conclusion that a factor of 10 represented "typical" ACRs.

When the laboratory-to-field ratio (LFR) was derived by comparing experimental acute LC₅₀ data with field toxicity data, the derived LFR spanned from 12-5300. The available field data were relatively

1 limited, and the US EPA described the outcome of the process comparing field to acute LC₅₀ data as a
2 working model, for which further data were needed to refine the projections. With the knowledge they had at
3 hand they found a factor of 1000 from acute-to-field to be reasonable. QSAR data were evaluated as being
4 comparable to acute LC₅₀ data within one order of magnitude, and it was therefore recommended as being
5 treated as being equal to a single LC₅₀. A final interesting detail in regard to the initial development of AFs is
6 that the authors explicitly pointed out that further research should improve and refine them (US EPA 1984).
7 It may seem that this recommendation has been forgotten, though, since it was made more than three decades
8 ago.

9 If a large dataset from long-term tests for different taxonomic groups is available, the use of
10 the species sensitivity distribution (SSD) method is currently perceived as less uncertain compared to the
11 extrapolation from data from the base set of test organisms, and an AF of between 5-1 can be used, if
12 justification is provided (ECHA 2008). Participants in the OECD workshop in 1992 also discussed a
13 relatively large dataset focused on the use of species sensitivity distribution (SSD) methods and regression
14 models. SSD or NOECs were proposed as alternatives to the subjective assessment factors that were
15 otherwise used to extrapolate from small datasets. Despite the recognition that SSD provides a better
16 scientific foundation for hazard evaluation, the derived safe concentrations may not protect all functions of
17 the ecosystem, due to a number of scientific assumptions: a) species in ecosystems can be divided into a
18 small group consisting of every sensitive species and a large group including the less sensitive species; b) the
19 distribution of sensitivity in lab approximates distribution in the ecosystem; c) the sensitivity of a species
20 isolated in a lab is equal to its sensitivity in a complex ecosystem; d) sensitivity can be characterised as
21 (normal) distribution, and such a characterisation made under laboratory conditions can be directly
22 extrapolated to field conditions and e) interactions between species are not accounted for when applying
23 SSDs, and because an SSD is based on existing standard tests the protection level depends on the
24 relationship between these tests and their biological and ecological significance, which can be limited
25 (OECD 1992).

Under REACH, and according to the guidance documents provided by ECHA (2008), the assessment factor to be used on mesocosm studies, or (semi-) field data, needs to be reviewed on a case-by-case basis. This is very much in line with the OECD's workshop report which states that comprehensive ecosystem studies should only be conducted when a hazard is perceived to be high, as such studies can further indicate how fast populations may recover from exposure to a stressor and provide more realistic exposure evaluations. This is the first time that the OECD report has addressed a risk scenario as is it generally understood today (i.e. as the ratio obtained by dividing hazard and exposure assessments), noting that standard laboratory tests cannot predict effects on the ecosystem level due to, for instance, the influence of interspecies interactions, the indirect effects of chemicals, effects on ecosystem processes and recovery rates and finally the cumulative effects of multiple stressors. Even though these considerations are more than 20 years old, some of them are still relevant – as seen in the case studies on nonylphenol and nanoparticles.

6. Discussion

Our analysis of the two case studies illustrates some of the challenges that ERAs face when it comes to assessing chemicals and nanomaterial risk. Based on the nature of these identified challenges, we would argue that they cannot be addressed by revising the ERA; instead, they are a reflection of the fundamental limitations of the ERA framework, and these limitations have been well recognised over time but never really addressed in regulatory application.

6.1 Hazard identification

In the case of nonylphenol and nanoparticles, we identified a number of challenges, including the disregard for endocrine-disrupting effects and “new” endpoints in favour of traditional endpoints, as well as the disregard for findings stemming from non-standard tests in favour of standard tests.

From the case of nonylphenol it is clear that if there are indications that a chemical has properties of very high (or unknown) concern, such as EDCs and carcinogens, it should not be overlooked, even if it seems that other endpoints are more sensitive – especially if the scientific understanding is still in the development phase. In the 2002 ERA for nonylphenol it was concluded that oestrogenic properties are covered by the most sensitive endpoint (a 72h algae growth test) (EC 2002). Since the collection of data for the ERA was finalised in 1999, new studies have been published, as a function of scientific progress, which have led the European Chemical Agency to propose that nonylphenol be classified as SVHC, based on its ED properties (ECHA 2012). We do not highlight this development in order to insinuate that the 2002 ERA was not conducted properly; the work with the 2002 nonylphenol ERA was indeed comprehensive and lived up to the standards that can be expected from such an assessment. However, the case serves as an example of the limitations that the boundaries of our scientific understanding set on our ability to assess risk, if we insist that risk must be quantified with standardised assays. Today it would be much more controversial to argue

1 that disruptive endocrine properties are properly accounted for through the survival and growth type of
2 endpoints. A valuable lesson in this regard is that we should not disregard studies that show the effects of
3 unknown biological importance at very low concentrations but rather highlight any uncertainty in
4 determining the importance of such findings.

5 In regard to the case of ENM it remains to be discovered whether there are nano-specific
6 effects that are not properly accounted for within the current ERA framework, though it is still too early to
7 conclude that the existing framework is sufficient (WHO 2013, Scientific committees 2013). Limits to our
8 understanding of such nano-specific effects, as well as limitations in regard to our technological capabilities,
9 result in high amounts of uncertainty in any ENM hazard identification. This makes it questionable whether
10 we are able to provide evidence-based assessments of ENM hazards that can be used to quantify risk.
11 Looking at the hindsight lessons taken from the ERA of nonylphenol, it might not be feasible to rely on ENM
12 hazard data until we are more certain about the best suitable approach for such hazard identifications. It
13 would therefore seem feasible if the search for novel findings were better and more systematically integrated
14 in the risk assessment process, not least in regard to new and emerging risks such as those from ENMs.

15 The preference for traditional endpoints and standard tests is not something that is specific to
16 nonylphenol and nanoparticles – it can be traced back to the origins of ERAs in the EU. The endpoints that
17 the Working Party Risk Management for Ecosystems in the Netherlands proposed and asked for back in
18 1989 regarded acute toxicity and long-term experiments such as reproduction tests on daphnia and an egg-
19 larva test for fish. Preferences for such quite crude endpoints have led over time to the unfortunate dismissal
20 of indicative studies of new endpoints as well as studies with unknown biological significance. This could
21 explain the reluctance to accept endocrine-disrupting effects and “new” versus traditional endpoints in the
22 2002 ERA of nonylphenol. Similarly, from the outset, the Working Party Risk Management for Ecosystems
23 (1987) referred to OECD standard test guidelines, thereby giving priority to standardised tests over non-
24 standard tests.

The minimum data requirements posited by the Working Party Risk Management for Ecosystems (1987) for air, water and soil are the same as those that are used in REACH, which again could help explain resistance to accepting endocrine-disrupting effects and “new” endpoints versus the more traditional examples that have now become the norm. Furthermore, it also explains why there is a tendency to ignore aspects that have been recognised as “known unknowns” (i.e. uncertainties that we are aware of but are unable to address properly) when it comes to the hazard characteristics of emerging materials and substances. In order to provide the best scientific foundation for risk management, such uncertainties should therefore be highlighted rather than ignored.

6.2 Dose-response assessment

Prior to 1993, discussions on how to assess the environmental risk of toxic chemicals were taking place in the United States of America and in many European countries, such as Denmark, Germany, Spain and the Netherlands (van Straalen and van Leeuwen 2002). Especially the work conducted by the US EPA (1984) and in the Netherlands laid the foundations for the chemical RA framework. The Health Council of the Netherlands established a scientific advisory committee to give advice on the ecological risk assessment of chemicals, and in 1989, this committee published the report “Assessing the risk of toxic chemicals for ecosystems” (HCN 1989) in which minimal data requirements, the use of the term “PNEC” and the development of procedures for the derivation of PNEC, which are well established now, were described.

Although the HCN (1989) defined the minimum data requirements, over time these have become standard data requirements, and very seldom are other data considered in the final dose-response assessment. This has led to the dismissal of indicative studies and studies with unknown biological significance. Moreover, in hindsight, it seems that the PNEC has always been set too low, as ERAs have failed to identify the most sensitive species as well as limitations in the use of extrapolation methods.

1 These limitations of the ERA method were well-recognised by the experts that suggested the
2 approach back in the late 1980s. The committee noted that the purpose of the procedure was to predict the
3 effects of toxic chemicals on ecosystems and to establish boundary conditions for the protection of these
4 ecosystems, despite limited scientific knowledge available at the time on ecotoxicology (HCN 1989). They
5 also noted that the key to making such assessments was extrapolation from experimental data to effects in
6 ecosystems. To obtain sufficient protection this requires that data are derived from tests on the most sensitive
7 species in the ecosystem. Ecological risk assessments aim at protecting the characteristics of the species
8 composition of the ecosystem rather than all individual species. According to the committee, it was not
9 possible to conclude from the available data whether an ecosystem would react more sensitively to a toxic
10 chemical than individual species (HCN 1989). Hence, there was no certainty that protecting individual
11 species would lead to the protection of the entire ecosystem, so the risk assessment paradigm fundamentally
12 aimed at protecting species and not ecosystems (HCN 1989). The committee concluded that none of the RA
13 procedures reviewed could lay claim to safeguarding ecosystems against the adverse effects of toxic
14 chemicals. Furthermore, the committee stated that complete scientific foundation for risk regulation was not
15 possible, and policy should therefore not be based on such a requirement (HCN 1989). The advice from the
16 Health Council was later implemented in the Netherlands in the form of a policy paper on risk management
17 (HCN 1989, Premises for Risk Management 1989) and subsequently implemented in the Technical Guidance
18 Documents (EC 2003) and the Guidance documents used under REACH (ECHA 2008).

19 Several aspects in regard to dose-response assessment and subsequent PNEC derivation are
20 relevant to discuss when it comes to mixtures. It is well documented that mixture toxicity often exceeds the
21 toxicity of individual components in the mixture (e.g. Levis 1992, Syberg et al. 2009, Kortenkamp et al.
22 2009). The Dutch Working Party Risk Management for Ecosystems noted that mixture toxicity is an
23 important aspect of risk assessment, and it was further noted that mixtures could not be taken into account at
24 the time (i.e. August 1987), since there were no instruments that could deal properly with this problem,
25 which again was noted to be an undesirable situation (Working Party Risk Management for Ecosystems

1 1987). Despite this recognition, back in the late 1980s, and empirical evidence on the importance of mixture
2 toxicity that has built up over decades, RAs are primarily carried out solely for single chemicals (EC 2003,
3 EC 2006). The nonylphenol risk assessment serves as an illustration of this problem, since mixture effects
4 are not quantified at all in this RA. Even though mixture toxicity is a complex challenge, solutions have been
5 discussed over the last decades, and have recently been reviewed in (Kortenkamp et al. 2009).

6 7 **6.3 Improving scientific foundation and ensuring timely protection**

8 Our analysis demonstrates that the ERA framework should rather be considered as a pragmatic set of tools
9 that provides a systematic approach to determining risk rather than an evidence-based foundation for
10 decision making. There are several areas where it fails to provide evidence of the real-life hazards of
11 chemicals and nanomaterials, as discussed above. Apart from these gaps there is also a tendency for ERAs to
12 focus too rigidly on achieving transparency and reliability through demands for GLP and standard tests
13 (Myers et al. 2009). Whereas such demands make good sense in relation to data provided by industry, they
14 nevertheless pose a problem in regard to data produced by academia and published in peer-reviewed
15 journals. GLP was introduced to ensure high quality data produced by industry, but the scientific literature
16 already has such a system in place (i.e. the peer review system). There are several problems with this
17 approach that our analysis helps to illuminate. First of all, the most important role of science is to address
18 new questions and to generate a novel understanding of complex issues. If such novel findings are not
19 accounted for, ERAs may fail to provide the foundation for precautionary preventive action as required by
20 the European Treaty (EU 2007). Both the nonylphenol and the ENM cases are illustrations of this problem.
21 Even though the nonylphenol ERA was finalised in 2002 (EC 2002), a recommendation for classifying the
22 compound as SVHC was not made until 2012 (ECHA 2012). And even at this point some the European
23 Council for Alkylphenols and Derivatives argued that it was premature to classify nonylphenol as an SVHC
24 based on its endocrine-disruptive properties, since no general EDC definition was agreed upon (ECHA
25 2013). A possible alternative to this prolongation of the decision-making process would have been to

1 conduct a chemical alternative assessment (CAA) (Lavoie et al. 2010) at a much earlier stage. The outcome
2 of a CAA is meant to provide stakeholders with an assessment of alternatives to the chemicals in question,
3 thus facilitating the substitution process (Lavoie et al. 2010). Illuminating such substitution pathways seems
4 to be a good and viable step in ensuring more timely decision making and therefore protection. This
5 approach would be more in line with commitments under the European Treaty to ensure precautionary
6 preventive action.

7 For ENMs we still rely on ERAs to provide the foundation for decision making, even though
8 it is generally accepted that our ability to assess ENM hazards and subsequently PNECs is poor. It is
9 therefore relevant to call attention to the fact that ERAs should not be considered as a “holy grail” for
10 informing decision making but rather be used as one particular tool in a larger decision-making toolbox.
11 Interestingly, the US EPA (1992) discusses the relationship between the risk assessor and the risk manager –
12 something not normally considered in the EU when it comes to discussions about risk assessment, and
13 definitely something from which European decision-makers could learn. In this discussion it is stated that
14 “The results of the risk assessment serve as input to the risk management process, where they are used along
15 with other inputs defined in EPA statutes, such as social and economic concerns, to evaluate risk
16 management options” (US EPA 1992). It is, however, paramount that the foundation for management is
17 presented in a very transparent manner, as this will ensure that it is clear if socio economic interests are
18 perceived as more important than a specified risk, or if uncertainties regarding the actual risk prevent a
19 reliable assessment of risk. Such transparency is needed if stakeholders and the public are to have the ability
20 to judge if risk assessors live up to their political mandate.

21 Our analysis shows the derivations of PNECs with ERAs are governed substantially by
22 pragmatic decisions, even for substances that can be properly assessed within the ERA framework. When
23 uncertainties that are linked specifically to ENMs are considered along with the limited scientific foundation
24 for the assessment factors applied, it seems questionable whether ERAs can provide the ‘evidence-based
25 foundation’ for decision making that the framework is expected to deliver (Löfsted 2011). There is no doubt

that science is progressing within this field and that the work in understanding ENM hazards and subsequent risk should continue, but until the uncertainties within this field have been further limited it would be more feasible to let decision making rely on alternative frameworks that also include other societal considerations, as mentioned by the US EPA (1992), as well as scientific evaluations that acknowledge uncertainty rather than ignore it and ethical discussions conducted in the light of the precautionary principle.

A final note in this paper concerns transparency. None of the methodologies mentioned here can be said to provide a rigorous scientific solution to inform decision-making. This fact stresses the point that regulatory action is a political responsibility, and so in order for science to help inform such political decisions, transparency is paramount. While the European Commission's European Chemical Bureau focused on updating the TGD in 1996 and in 2003, the US EPA (1998) focused on 'new approaches to complex risk problems by delineating the need for "planning and problem formulation" to address technically challenging assessments of ecosystems, chemical mixtures, and cumulative risk' (US EPA 1998). An important aspect of this approach is that stakeholders are involved in the initial stages of the assessment. Although this approach might not lead to a more accurate estimation of the PNEC, it does provide for better decision support, as it delivers a more in-depth and transparent evaluation of a given risk as well as the pros and cons of various management options.

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References

- Adams, L.K., Lyon, D.Y., Alvarez, P.J.J., 2006. Comparative eco-toxicity of nanoscale TiO₂, SiO₂, and ZnO water suspensions. *Water Res.* 40, 3527-3532
- Aitken, R.A, Bassan, A., Friedrichs, S., Hankin, S.M., Hansen, S.F., Holmqvist, J., Peters, S.A.K., Poland, C.A., Tran, C.L. 2011. Specific Advice on Exposure Assessment and Hazard/Risk Characterisation for Nanomaterials under REACH (RIP-oN 3) Final Project Report . Document reference RNC/RIP-oN3/FPR/1/FINALBrussels: European Commission. Available: http://ec.europa.eu/environment/chemicals/nanotech/pdf/report_ripon3.pdf
- Arnold, S.F., Klotz, D.M., Collins, B.M., Vonier, P.M., Guillette Jr., L.J., McLachlan, J.A. 1996. Synergistic activation of estrogen receptor with combinations of environmental chemicals. *Science*, 272 (5267), pp. 1489-1492.
- Ashfield, L.A., Pottinger, T.G., Sumpter, J.P. 1998. Exposure of female juvenile rainbow trout to alkylphenolic compounds results in modifications to growth and ovosomatic index. *Environ. Toxicol. Chem.* 17(4), 1552-8618
- Baun, A., Hartmann, N.B, Greiger, K.D., Hansen, S.F. 2009. Setting the Limits for Engineered Nanoparticles in European Surface Waters. *Journal of Environmental Monitoring* 11, 1774 – 1781.
- Blanck, H. Species dependent variation among organisms in their sensitivity to chemicals. 1984. *Ecological Bulletins* 36, 107-119
- Call, D.J., Brooke, L.T., Raymond, J.K., Knuth, M.L., Anderson, C., Moriarity, C. 1983. Toxicity, bioconcentration, and metabolism of the herbicide Propanil (3',4'-dichloropropionanilide) in freshwater fish. *Arch Environ Contam Toxicol.* 12, 175-182
- CCA. 2008. Small Is Different: A Science Perspective On The Regulatory Challenges of the Nanoscale. Ottawa: The Council of Canadian Academies, Ottawa, Canada

Field Code Changed

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 Dai, L., Syberg K., Banta, G.T., Selck, H., Forbes, V.E. 2013. Effects, Uptake, and Depuration Kinetics of
- 2 Silver Oxide and Copper Oxide Nanoparticles in a Marine Deposit Feeder, *Macoma balthica*. *ACS Sustain*
- 3 *Chem Eng*, 1(7), 760-767
- 4 EC 1993a. Technical guidance documents in support of the risk assessment Directive (93/67/EEC) for new
- 5 substances notified in accordance with the requirements of Council Directive 67/548/EEC. European
- 6 Commission, Bruxelles, Belgium
- 7 EC 1993b. Commission directive 93/67/EEC of 20 July 1993 laying down the principles for assessment of
- 8 risks to man and the environment of substances notified in accordance with Council Directive 67/548/EEC.
- 9 European Commission, Bruxelles, Belgium
- 10 EC 1994. Technical guidance documents in support of the risk assessment Regulation (1488/94) for existing
- 11 substances in the context of Council Regulation 793/93/EEC. European Commission, Bruxelles, Belgium
- 12 EC 2000. Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000
- 13 establishing a framework for Community action in the field of water policy. European Commission,
- 14 Bruxelles, Belgium
- 15 EC 2002. European Union Risk Assessment Report. 4-Nonylphenol (Branched) and Nonylphenol. CAS Nos:
- 16 84852-15-3 and 25154-52-3 EINECS Nos: 284-325-5 and 246-672-0. Environment Agency Chemicals
- 17 Assessment Section. Oxfordshire, United Kingdom. Can be accessed at:
- 18 <http://echa.europa.eu/documents/10162/43080e23-3646-4ddf-836b-a248bd4225c6>
- 19 EC 2003. Technical Guidance Document on Risk Assessment in support of Commission Directive
- 20 93/67/EEC on Risk Assessment for new notified substances, Commission Regulation (EC) No 1488/94 on
- 21 Risk Assessment for existing substances, Directive 98/8/EC of the European Parliament and of the Council
- 22 concerning the placing of biocidal products on the market. European Commission, Joint Research Center,
- 23 Ispra, Italy.

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 EC 2006. Regulation (EC) No 1907/2006 of the European Parliament and of the Council of 18 December
- 2 2006 concerning the Registration, Evaluation, Authorisation and Restriction of Chemicals (REACH),
- 3 establishing a European Chemicals Agency, amending Directive 1999/45/EC and repealing Council
- 4 Regulation (EEC) No 793/93 and Commission Regulation (EC) No 1488/94 as well as Council Directive
- 5 76/769/EEC and Commission Directives 91/155/EEC, 93/67/EEC, 93/105/EC and 2000/21/EC. European
- 6 Commission, Bruxelles, Belgium

- 7 EC 2012. Regulation (EU) No 528/2012 of the European Parliament and of the Council of 22 May 2012
- 8 concerning the making available on the market and use of biocidal products. European Commission,
- 9 Bruxelles, Belgium

- 10 EC 2013. Minutes of the expert meeting on endocrine disruptors. Meeting on endocrine disruptors . Office of
- 11 the Chief Scientific Adviser, European Commission, Brussels, Belgium.

- 12 ECHA 2008. Guidance on information requirements and chemical safety assessment Chapter R.10:
- 13 Characterisation of dose [concentration]-response for environment. Europeans Chemicals Agency, Helsinki,
- 14 Finland

- 15 ECHA 2012. Support document for identification of 4-Nonylphenol, Branched and linear, Ethoxylated as
- 16 substances of very high concern because, due to their degradation to substances of very high concern (4-
- 17 Nonylphenol, Branched and linear) with endocrine disrupting properties, they cause probable serious effects
- 18 to the environment which gives risk to an equivalent level of to those of CMR and PBT/vPvB substances.
- 19 ECHA, Helsinki, Finland.

- 20 ECHA 2013. Comments on an Annex XV dossier for identification of a substance as SVHC and response to
- 21 these comments - 4-Nonylphenol, branched and linear, ethoxylated [substances with a linear and/or branched
- 22 alkyl chain with a carbon number of 9 covalently bound in position 4 to phenol, ethoxylated covering
- 23 UVCB- and well-defined substances, polymers and homologues, which include any of the individual isomers
- 24 and/or combinations thereof]. ECHA, Helsinki, Finland.

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 ECHA 2014. Draft background document for 4-Nonylphenol, branched and linear, ethoxylated (4-NPnEO).
- 2 European Chemicals Agency, Helsinki, Finland.
- 3 ED & Dupont 2007. Nano Risk Framework. Environmental Defense – DuPont Nano Partnership.
- 4 Washington DC, USA
- 5 EEA 2001. Late lessons from early warnings: the precautionary principle 1896-2000. Environmental issue
- 6 report No 22/2001. European Environmental Agency, Copenhagen, Denmark
- 7 EEA 2013. Late lessons from early warnings: science, precaution, innovation. EEA Report No 1/2013.
- 8 European Environmental Agency, Copenhagen, Denmark
- 9 EFSA 2008. Draft Opinion of the Scientific Committee on the Potential Risks Arising from Nanoscience and
- 10 Nanotechnologies on Food and Feed Safety (Question No EFSAQ-2007-124). European Food Safety
- 11 Authority, Bruxelles, Belgium
- 12 EU 2007. The Lisbon Treaty. The Treaty on European Union and the Treaty on the Functioning of the
- 13 European Union. European Union, Bruxelles, Belgium
- 14 Federici, G., Shaw, B., Handy, R. 2007. Toxicity of titanium dioxide nanoparticles to rainbow trout
- 15 (*Oncorhynchus mykiss*): Gill injury, oxidative stress, and other physiological effects. *Aquatic Toxicology*
- 16 84(4), 415-430
- 17 Fortner, J. D., Lyon, D. Y., Sayes, C. M., Boyd, A. M., Falkner, J. C., Hotze, E. M., Alemany, L. B., Tao, Y.
- 18 J., Guo, W., Ausman, K. D., Colvin, V. L., Hughes, J. B. 2005. C-60 in water: Nanocrystal formation and
- 19 microbial response. *Environ Sci Technol.* 39(11), 4307-4316
- 20 Gao, P., Li, Z., Gibson, M., Gao, H.-2014. Ecological risk assessment of nonylphenol in coastal waters of
- 21 China based on species sensitivity distribution model. *Chemosphere.* 104, 113-119

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 Hankin S.M., Peters S.A.K., Poland C.A., Foss Hansen S., Holmqvist J., Ross B.L., Varet J. and Aitken R.J.
- 2 2011. Specific Advice on Fulfilling Information Requirements for Nanomaterials under REACH (RIP-oN 2)
- 3 Final Project Report
- 4 Hansen, S. F., Larsen, B. H., Olsen, S. I., Baun, A. 2007. Categorization framework to aid Hazard
- 5 Identification of Nanomaterials. *Nanotoxicology* 1, 243-250.
- 6 Hansen, S.F., Maynard, A., Baun, A., Tickner, J.A., Bowman, D.M. 2013. Nanotechnology — early lessons
- 7 from early warnings. In: Gee et al. (Eds.). Late lessons from early warnings: science, precaution, innovation.
- 8 European Environment Agency, Copenhagen, Denmark. Available at:
- 9 <http://www.eea.europa.eu/publications/late-lessons-2>
- 10 Hartmann, N.B., Skjolding, L.M., Hansen, S.F., Kjølholt, J., Gottschalck, F., Baun, A. 2014. Environmental
- 11 fate and behaviour of nanomaterials New knowledge on important transformation processes. Environmental
- 12 Project No. 1594, 2014. Copenhagen: The Danish Environmental Protection Agency
- 13 HCN 1989. Assessing the risk of toxic chemicals for ecosystems. Report 1988/28E. Health Council of the
- 14 Netherlands. The Hague, The Netherlands
- 15 Heinlaan, M., Ivask, A., Blinova, I., Dubourguier, H., Kahru, A. 2008. Toxicity of nanosized and bulk ZnO,
- 16 CuO and TiO₂ to bacteria *Vibrio fischeri* and crustaceans *Daphnia magna* and *Thamnocephalus platyurus*.
- 17 *Chemosphere* 71, 1308–1316
- 18 Holmes, J., Clark, R. 2008. Enhancing the use of science in environmental policymaking and regulation.
- 19 *Environ Sci Policy*. 11, 702-711
- 20 Hund-Rinke, K., Simon, M. 2006. Ecotoxic effect of photocatalytic active nanoparticles TiO₂ on algae and
- 21 daphnids. *Environ Sci Pollut R* 13(4), 225-232.
- 22 JRC 2014. Towards a review of the EC Recommendation for a definition of the term "nanomaterial" Joint
- 23 Research Centre, Ispra, Italy

Field Code Changed

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 Kahl, M.D., Makynen, E.A., Kosian, P.A., Ankley, G.T. 1997. Toxicity of 4-Nonylphenol in a Life-Cycle
- 2 Test with the Midge *Chironomus tentans*. *Ecotoxicol. Environ. Saf.* 38(2), 155-160
- 3 Kavlock, R.J., Daston, G.P., DeRosa, C., Fenner-Crisp, P., Gray, L.E., Kaattari, S., Lucier, G., Luster, M.,
- 4 Mac, M.J., Maczka, C., Miller, R., Moore, J., Rolland, R., Scott, G., Sheehan, D.M., Sinks, T., Tilson, H.A.
- 5 1996. Research needs for the risk assessment of health and environmental effects of endocrine disruptors: A
- 6 report of the U.S. EPA-sponsored workshop. *Environmental Health Perspectives*, 104 (SUPPL. 4), pp. 715-
- 7 740.
- 8 Khan, F.R., Schmuecking, K., Krishnadasan, S.H., Berhanu, D., Smith, B.D., deMello, J.C., Rainbow, P.S.,
- 9 Luoma, S.N., Valsami-Jones, E. 2013. Dietary bioavailability of cadmium presented to the gastropod
- 10 *Peringia ulvae* as quantum dots and in ionic form. *Environ. Toxicol. Chem.* 32(11), 2621–2629
- 11 Klimisch, H.J., Andreae, M., Tillmann, U. 1997. A Systematic Approach for Evaluating the Quality of
- 12 Experimental Toxicological and Ecotoxicological Data. *Regul. Toxicol. Pharm.* 25(1), 1-5
- 13 Kooijman, S.A.L.M. 1985. Toxicity at population level. In J. Cairns, editor, *Multispecies toxicity testing*.
- 14 Pergamon Press, New York
- 15 Kooijman, S.A.L.M. 1987. A safety factor for LC50 values allowing for differences in sensitivity among
- 16 species. *Water Res.* 21, 269-276
- 17 Kortenkamp, A., Backhaus, T., Faust, M. 2009. State of the Art Report on Mixture Toxicity. Final Report.
- 18 European Commission, Bruxelles, Belgium
- 19 Kühnel, D., Busch, W., Meißner, T., Springer, A., Potthoff, A., Richter, V., Gelinsky, M., Scholz, S.,
- 20 Schirmer, K. 2009. Agglomeration of tungsten carbide nanoparticles in exposure medium does not prevent
- 21 uptake and toxicity toward a rainbow trout gill cell line. *Aquat Toxicol.* 93(2–3), 91-99

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 Kwak, H., Bae, M., Lee, M., Lee, Y., Lee, B., Kang, K., Chae, C., Sung, H., Shin, J., Kim, J., Mar, W.,
- 2 Sheen, Y., Cho, M. 2001. Effects of nonylphenol, bisphenol a, and their mixture on the viviparous swordtail
- 3 fish (*Xiphophorus helleri*). *Environ. Toxicol. Chem.* 20(4), 1552-8618
- 4 Lavoie, E.T., Heine, L.G., Holder, H., Rossi, M.S., Lee, R.E., Connor, E.A., Vrabel, M.A., DiFiore, D.M.,
- 5 Davies, C.L. 2010. Chemical Alternatives Assessment: Enabling Substitution to Safer Chemicals. *Environ.*
- 6 *Sci. Technol.* 44(24), 9244-9249
- 7 Lee H.J., Chattopadhyay S., Gong E.Y., Ahn R.S., Lee K. 2003. Antiandrogenic effects of bisphenol A and
- 8 nonylphenol on the function of androgen receptor. *Toxicol Sci.* 75(1), 40-46
- 9 Levis, M.A. 1992. The effects of mixtures and other environmental modifying factors on the toxicities of
- 10 surfactants to fresh-water and marine life. *Water Res.*, 26(8), 1013-1023
- 11 Löfsted, R.E. 2011. Risk versus Hazard – How to Regulate in the 21st Century. *Eur J Risk Regul.* 2, 149-168
- 12 Lovern, S. B., Strickler, J. R., Klaper, R. 2007. Behavioral and Physiological Changes in *Daphnia magna*
- 13 when Exposed to Nanoparticle Suspensions (Titanium Dioxide, Nano-C60, and C60HxC70Hx). *Environ Sci*
- 14 *Technol* 41(12), 4465-4470.
- 15 Myers J.P., vom Saal F.S., Akingbemi B.T., Arizono K., Belcher S., Colborn T., Chahoud I., Crain D.A. ,
- 16 Farabollini F., Guilleffe L.J., Hassold T., Ho S.M., Hunt P.A., Iguchi T., Jobling S., Kanno J., Laufer H.,
- 17 Marcus M., McLachlan J.A., Nadal A., Oehlmann J., Olea N., Palanza P., Parmigiani S., Rubin B.S.,
- 18 Schoenfelder G., Sonnenschein C., Soto A.M., Taisness C.E., Taylor J.A., Vandenberg L.N., Vandenberg
- 19 J.G., Vogel S., Watson C.S., Welshons W.V., Zoeller R.T. 2009. Why public health agencies cannot depend
- 20 upon ‘Good Laboratory Practices’ as a criterion for selecting data: the case of bisphenol-A. *Environ. Health*
- 21 *Perspect.* 117(3), 309-315.
- 22 Nielsen E., Østergaard G., Thorup I., Ladefoged O., Jørgensen J.E. 2000. Toxicological evaluation and
- 23 limit values for nonylphenol ethoxylates, tricresyl, phosphates and benzoic acid. Environmental Project
- 24 (Miljøprojekt), No. 512. The Danish Environmental Protection Agency, Denmark.

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 Nordan, M.M., Holman, M.W., Bünger, M. 2006. Nanotech Commercialization Has Advanced, but
- 2 Government Action to Address Risk Has Not. House Committee on Science. USA
- 3 NRC 1983. Risk assessment in the federal government. Managing the process. National Research Council.
- 4 National Academy Press, Washington, DC.
- 5 Oberdörster G, Oberdörster E, Oberdörster J. 2007. Concepts of Nanoparticle Dose Metric and Response
- 6 Metric. *Environ Health Perspect* 115(6), A 290.
- 7 OECD 1992. Report of the OECD Workshop on the Extrapolation of Laboratory Aquatic Toxicity Data to
- 8 the Real Environment. OECD Environment Monographs No. 59, Organisation for Economic Co-operation
- 9 and Development, Paris.
- 10 Okamoto, Y., Hayashi T., Matsunami S., Ueda K., Toda C., Kawanishi S., Kojima N. 2006. Formation of
- 11 DNA Damaging Nonylphenol Product from Light-Irradiated Nonylphenol. *Journal of health science* 52(1),
- 12 91-95
- 13 Premises for Risk Management (1989) Risk limits in the context of environmental policy. Directorate-
- 14 General for Environmental Protection at the Ministry of Housing, Physical Planning and Environment.
- 15 Second Chamber of the State General, 1988-89 session, 21137, no.s 1–2. The Hague. Available at:
- 16 <http://www.coenrady.com/downloads/handig/pdf017.pdf>
- 17 Rajapakse N, Silva E, Scholze M, Kortenkamp A 2004. Deviation from additivity with estrogenic mixtures
- 18 containing 4-nonylphenol and 4-tert-octylphenol detected in the E-SCREEN assay. *Environ Sci Technol* 38:
- 19 6343–6352.
- 20 RCEP 2008. Twenty-seventh Report Novel Materials in the Environment: The case of nanotechnology Royal
- 21 Commission on Environmental Pollution. The Stationery Office, Norwich, UK
- 22 SCENIHR. 2007. Scientific Committee for Emerging and Newly-Identified Health Risks (SCENIHR) The
- 23 appropriateness of the risk assessment methodology in accordance with the Technical Guidance Documents

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 for new and existing substances for assessing the risks of nanomaterials. European Commission, Bruxelles,
2 Belgium
- 3 Schmidt, K. 2007. Nanofrontiers Visions for the future of Nanotechnology. PEN 6. Washington, D.C.:
4 Project for Emerging Nanotechnologies, Woodrow Wilson International Center for Scholars.
- 5 Scientific Committees 2013. Addressing the New Challenges for Risk Assessment. Scientific Committee on
6 Health and Environmental Risks SCHER, Scientific Committee on Emerging and Newly Identified Health
7 Risks SCENIHR, Scientific Committee on Consumer Safety SCCS, European Commission, Bruxelles,
8 Belgium
- 9 Slooff, W., Van Oers, J.A.M., De Zwart, D. 1986. Margins of uncertainty in ecotoxicological hazard
10 assessment. *Environ. Toxicol. Chem.* 5(9), 1552-8618
- 11 Slovic, P. 1999. Trust, Emotion, Sex, Politics, and Science: Surveying the Risk-Assessment Battlefield. *Risk*
12 *analysis*, 19(4), 689-701.
- 13 Soares, A., Guieysse, B., Jefferson, B., Cartmell, E., Lester, J.N., 2008. Nonylphenol in the environment: a
14 critical review on occurrence, fate, toxicity and treatment in wastewaters. *Environ. Int.* 34, 1033-1049
- 15 Stephan, C. E., Mount, D. I., Hansen, D. J., Gentile, J. H., Chapman, G. A., Brungs, W. A. 1985. Guidelines
16 for deriving numerical national water quality criteria for the protection of aquatic organisms and their uses.
17 US Environmental Protection Agency. PB85-227049, Springfield, VA
- 18 Syberg, K. Jensen, T.S., Cedergreen, N., Rank, J. 2009. On the use of mixture toxicity assessment in
19 REACH and the water framework directive: a review. *Hum Ecol Risk Assess.* 15(6), 1257-1272
- 20 Templeton, R. C., Ferguson, P. L., Washburn, K. M., Scrivens, W. A., Chandler, G. T. 2006. Life-cycle
21 effects of single-walled carbon nanotubes (swnts) on an estuarine meiobenthic copepod. *Environ. Sci.*
22 *Technol.* 40 (23), 7387-7393

This is a post-print version of Syberg, K., & Hansen, S. F. (2016). Environmental risk assessment of chemicals and nanomaterials — The best foundation for regulatory decision-making? *Science of the Total Environment*, 541, 784-794. The printed version of the paper is available at: DOI: 10.1016/j.scitotenv.2015.09.112

- 1 US EPA 1984. Risk Assessment and Management: Framework for Decision Making. EPA 600/9-85-002. U.
- 2 S. Environmental Protection Agency, Chicago, USA
- 3 US EPA 1992. Framework for ecological risk assessment. EPA/630/R-92/001. U.S. Environmental
- 4 Protection Agency, Washington, DC
- 5 US EPA 1998. Guidelines for Ecological Risk Assessment. Federal Register 63(93):26846-26924. U.S.
- 6 Environmental Protection Agency, Washington, DC
- 7 U.S. EPA 2007. Nanotechnology White Paper. EPA 100/B-07/001. U.S. Environmental Protection Agency,
- 8 Washington, DC
- 9 Van Hoecke K., De Schamphelaere K. A., Van der Meeren P., Lucas S., Janssen C. R., 2008. Ecotoxicity of
- 10 silica nanoparticles to the green alga *Pseudokirchneriella subcapitata*: importance of surface area. *Environ.*
- 11 *Toxicol. Chem.* 27(9), 1948–1957.
- 12 van Leeuwen, C.J., Vermeire, T.G. (Eds.) 1995. Risk Assessment of Chemicals: An Introduction. Springer
- 13 Netherlands.
- 14 Van Straalen, N.M. (1987). Stofgehalten in de bodem- (geen) effecten op bodemdieren. In Symposium
- 15 Bodemkwaliteit, Ede. Rapport VTCB M86/44, Leidschendam, pp. 75-84.
- 16 Van Straalen, N. and Van Leeuwen, C. 2002. European history of species sensitivity distributions. In:
- 17 Species Sensitivity Distributions in Ecotoxicology (Eds. L. Posthuma, G. W. Suter II & T. P. Traas). Lewis
- 18 Publishers, Boca Raton. pp.19-34
- 19 WHO 2013. Nanotechnology and human health: Scientific evidence and risk governance. Report of the
- 20 WHO expert meeting 10–11 December 2012, Bonn, Germany.
- 21 Working Party Risk Management for Ecosystems 1987. Integrated Criteria Documents: Toxicological Data
- 22 Required for the Risk Assessment for Ecosystems. In: HCN 1989, 103-107