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1 Optical technologies applied alongside on-site and remote approaches for

2 climate gas emission quantification at a wastewater treatment plant

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16 Highlights

- Emissions of CH_4 , N_2O , and NH_3 were quantified from a wastewater treatment plant
- 18 Whole-plant and on-site emissions were measured using optical analytical techniques
- 19 Biosolid stockpiles accounted for 70% of total CH₄ emission
- N₂O was principally (about 82%) emitted from nitrifying trickling filters
- Important NH₃ emission sources were biosolid stockpiles and mechanical dewatering

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23 Keywords: Methane, nitrous oxide, ammonia, biosolid stockpiles, sewage sludge, emission factors

25 Abstract

26	Plant-integrated and on-site gas emissions were quantified from a Swedish wastewater treatment
27	plant by applying several optical analytical techniques and measurement methods. Plant-integrated
28	CH ₄ emission rates, measured using mobile ground-based remote sensing methods, varied between
29	28.5 and 33.5 kg CH ₄ h ⁻¹ , corresponding to an average emission factor of 5.9% as kg CH ₄ (kg CH ₄
30	$_{\text{production}}$) ⁻¹ , whereas N ₂ O emissions varied between 4.0 and 6.4 kg h ⁻¹ , corresponding to an average
31	emission factor of 1.5% as kg N ₂ O-N (kg TN $_{influent}$) ⁻¹ . Plant-integrated NH ₃ emissions were around
32	0.4 kg h ⁻¹ , corresponding to an average emission factor of 0.11% as kg NH ₃ -N (kg TN $_{removed}$) ⁻¹ .
33	On-site emission measurements showed that the largest proportions of CH_4 (70%) and NH_3 (66%)
34	were emitted from the sludge treatment line (mainly biosolid stockpiles and the thickening and
35	dewatering units), while most of the N_2O (82%) was emitted from nitrifying trickling filters. In
36	addition to being the most important CH ₄ source, stockpiles of biosolids exhibited different
37	emissions when the sludge digesters were operated in series compared to in parallel, thus slightly
38	increasing substrate retention time in the digesters. Lower CH_4 emissions and generally higher N_2O
39	and NH ₃ emissions were observed when the digesters were operated in series. Loading biosolids
40	onto trucks for off-site treatment generally resulted in higher CH ₄ , N ₂ O, and NH ₃ emissions from
41	the biosolid stockpiles. On-site CH_4 and N_2O emission quantifications were approximately two-
42	thirds of the plant-integrated emission quantifications, which may be explained by the different
43	timeframes of the approaches and that not all emission sources were identified during on-site
44	investigation. Off-site gas emission quantifications, using ground-based remote sensing methods,
45	thus seem to provide more comprehensive total plant emissions rates, whereas on-site
46	measurements provide insights into emissions from individual sources.

48 1 Introduction

Wastewater treatment is an anthropogenic source of atmospheric emissions of both methane (CH₄) 49 50 and nitrous oxide (N₂O). Furthermore, ammonium (NH_4^+) is a compound generated and transformed in biological processes occurring at wastewater treatment plants (WWTPs) 51 (Kampschreur et al., 2009) and could potentially be volatilized as ammonia (NH₃). Methane and 52 N₂O are potent greenhouse gases contributing to climate change (Stocker et al., 2013), which is why 53 their quantifications are important in supporting emission reporting and mitigation. Ammonia is a 54 55 plant nutrient, and thus emissions in this regards contribute to environmental eutrophication (Jenkinson, 2001). However, quantifying air emissions from WWTPs is a challenging undertaking, 56 as these discharges are diffuse by nature because they emanate from several diverse process units 57 58 and technologies replete with different physical shapes and emission heights, which, when 59 combined, form a large heterogeneous area source.

In the last 20 years, emission measurements from WWTPs have been performed mainly using 60 61 on-site point measurement methods. Floating chambers combined with liquid sample analysis have been the most common way of measuring emissions of N₂O and CH₄ from wastewater treatment 62 units with open air basins (Ahn et al., 2010; Czepiel et al., 1995; 1993; Ren et al., 2013; Ye et al., 63 2014). Lately, long-term investigations have been carried out along different stages of wastewater 64 reactors, in order to shed light on the temporal and spatial variability of emissions (Rodriguez-65 Caballero et al., 2014; Yan et al., 2014). Although these approaches have increased knowledge 66 about the mechanisms involved in greenhouse gas emissions, they measure only a portion of the 67 emissions from the reactor's surface. A few studies on covered reactors with an air collection 68 69 system (Daelman et al., 2013; Toyoda et al., 2011) have tried to fill this gap, thereby obtaining a 70 larger dataset with diurnal and annual emission changes. However, this approach can be applied

71

72

only to enclosed plants or wastewater treatment units with a ventilation system, and it does not consider physical leakages from pipes and fittings, or any other incidental releases.

73 In the last few years, ground-based optical remote sensing approaches have also been used for greenhouse gas quantification. The vertical radial plume mapping method, using an open-path gas 74 analyzer, can identify elevated concentrations of CH₄ and N₂O from a wastewater reactor, but this is 75 often limited to quantifying CH₄ emissions only, due to background concentrations of N₂O relative 76 to instrument sensitivity (Modrak et al., 2006). Finally, a highly sensitive mobile analytical 77 78 platform, applied in conjunction with the tracer dispersion method, has provided plant-integrated greenhouse gas emission rates for several Scandinavian open-air WWTPs, by performance of 79 downwind plume measurements of both CH₄ and N₂O (Yoshida et al., 2014; Delre et al., 2017). 80 81 Ground-based optical remote sensing methods are considered to provide a more comprehensive 82 overview of whole-site CH₄ emissions from large area sources (Reinelt et al., 2017; Yver Kwok et al., 2015). For ammonia, the scientific literature, to the best knowledge of the authors, does not 83 84 currently report atmospheric emissions from full-scale urban wastewater treatment plants.

Earlier studies have indicated that greenhouse gas emissions from process units such as biosolid stockpiles and nitrifying trickling filters could be significant (Mønster et al., 2014a; Delre et al. 2017). However, to date, only Majumder et al. (2014) have quantified these emissions from biosolid stockpiles. Air emission process data are important in environmental assessments comparing the environmental performances of different treatment technologies.

This paper aims at presenting for the first time a novel multiple measurement approach for air emission quantifications from a full-scale Swedish WWTP, using several optical analytical technologies and measurement methods, thereby allowing quantifications of air emissions from individual process units as well as the whole plant. Emissions of CH₄, N₂O, and NH₃ were measured, and process parameters, which potentially could influence air emissions, were explored.

95 Furthermore, we quantified air emissions from biosolid stockpiles and from nitrifying trickling

96 filters used for urban wastewater nitrification. To the best knowledge of the authors NH₃ emissions

97 from various on-site sources as well as plant-integrated emissions have not been measured at a

- 98 WWTP before.
- 99

100 2 Material and methods

101 **2.1** Site description

The investigated Swedish WWTP treats about 147 Mm³ wastewater per year, corresponding to a 102 pollution load of about 806,000 population equivalent (PE), 6% of which comes from industry and 103 the rest from households. The facility is divided into three main stages: A mechanical pre-treatment 104 105 line, a wastewater treatment line, and a sewage sludge treatment line. Incoming pollution is 106 mechanically removed with coarse and fine bar screens, a sand trap, a fat oil and grease trap, and primary settling tanks. The wastewater treatment line is a combination of several process units, 107 108 starting with a high-loaded activated sludge unit involving pre-denitrification, simultaneous phosphorus precipitation, and biochemical oxygen demand (BOD) removal, followed by secondary 109 settlers. Nitrification is carried out in nitrifying trickling filters, while post-denitrification is 110 performed in moving bed biofilm reactors (MBBRs). Disc filters remove any remaining suspended 111 solids before the treated wastewater is finally discharged into a river. Sewage sludge is thickened 112 before undergoing stabilization through mesophilic anaerobic digestion. Subsequently, the digestate 113 is mechanically dewatered on a centrifuge. Thickening and dewatering operations occur in a 114 building where a ventilation system assures a safe working environment. There is no treatment of 115 116 the vented air. Centrifuges for dewatering are located in a specific room, separated from the other machines but connected to the same ventilation system. Biosolids obtained after dewatering 117 118 digested sludge are stored for about three weeks in open-air stockpiles before being transported off-

119 site. Reject water from the sludge treatment line is recirculated back to the wastewater treatment line. Mechanical and biological treatments are supported, at different stages, through the addition of 120 coagulants and flocculants for phosphorus and solids removal, and methanol for microorganism 121 carbon supply. The plant receives sewage sludge from smaller WWTPs and co-digests industrial 122 food waste. Produced biogas is stored on-site in a gasholder, and routed in pipes to an off-site 123 124 upgrading facility and used as vehicle fuel. During treatment, sewage sludge can be named differently. Hereafter, the following nomenclature is used to refer to the different treatment stages. 125 126 After being removed from wastewater, sewage sludge is called "substrate" when entered into the digester. The liquid output of the digestion is called "digestate," which, after increasing its solid 127 content via centrifugation, is called "biosolids". Table 1 provides an overview of key plant-specific 128 129 information related to the measurement year 2015.

Over 2015, the digesters were run in two different modes: In parallel and in series. Usually, digesters work in parallel with a volatile solids (VS) load of 2.4 kg VS m⁻³ day⁻¹. From June until August, the serial mode was employed, resulting in a sludge retention time of 22 days instead of 20 days. When running in series, the VS load into the first digester increased to 4.4 kg VS m⁻³ day⁻¹, while the second digester had an average load of 2.2 kg VS m⁻³ day⁻¹.

135 **2.2 Optical analytical techniques applied**

Several optical analytical technologies were used to detect different compound concentrations in real time: A Fourier transform infrared spectroscopy analyzer (FTIR), for measuring concentrations of C_2H_2 , N_2O , CH_4 , NH_3 , and C_2H_4 , and two cavity ring down spectroscopy (CRDS) analyzers, for measuring C_2H_2 , N_2O , and CH_4 . Table 2 summarizes details regarding the analytical technologies applied for gas concentration measurements.

141 The FTIR instrument consists of an infrared spectrometer (Bruker Optics GmbH, Matrix-M
142 IRCube) connected to a closed long-path sample cell (Infrared Analysis Inc., model 107-V).

143	Compound-specific concentrations were determined by infrared absorption spectroscopy, whereby
144	CH ₄ , N ₂ O, NH ₃ , C ₂ H ₄ , and C ₂ H ₂ were measured simultaneously and analyzed at 3.3, 4.5, 10.3, 10.5,
145	and 13.7 μ m wavelengths, respectively. In mobile mode, the FTIR analyzer logged every ninth
146	second at a precision of 1.7, 0.3, 2.0, 1.8, and 4.7 ppb for CH_4 , N_2O , NH_3 , C_2H_2 , and C_2H_4 ,
147	respectively (C_2H_2 and C_2H_4 were used as dispersion tracer gases). More information about the
148	instrument can be found in Galle et al. (2001) and Scheutz et al., (2011), and about the FTIR
149	technology in general in the USA Environmental Protection Agency Handbook (Mikel and Merrill,
150	2011).

151 CRDS uses an optical technology in which gas concentration is obtained by measuring directly the "ring-down," or decay, of laser light in a sample cell. One instrument was equipped 152 with lasers detecting CH₄ and C₂H₂ (G2203, Picarro, Inc., Santa Clara, CA), and another was set up 153 to identify N₂O and C₂H₂ (S/N JADS2001, Picarro, Inc., Santa Clara, CA). The CH₄/C₂H₂ analyzer 154 logged twice per second with a precision of 0.77 ppb and 0.06 ppb for CH_4 and C_2H_2 , respectively. 155 The N_2O/C_2H_2 analyzer recorded every 3 seconds with a precision of 7.7 ppb and 0.6 ppb for N_2O 156 and C₂H₂, respectively. More detailed instrument descriptions can be found in Mønster et al. 157 (2014b) and Yoshida et al. (2014). 158

159 2.3 Measurement methods applied

Gas concentration detection, using different optical analytical techniques, was combined withdifferent measurement methods to obtain the emission rate of the target compounds.

Emission rates were obtained mainly by applying the tracer gas dispersion method (TDM), which is based on the principle that gases (with long atmospheric lifetime) disperse in the same way as far as mixing and transport are concerned. Therefore, when good mixing between the two gases is assured, the real-time emission rate of the target gas can be obtained from the relationship between the downwind concentrations of the target gas and the tracer gas. The tracer gas is

167 constantly released from the emitting source and the emission rate is calculated as shown by Eq. 1. 168 In this study, acetylene (C_2H_2) was used as main tracer gas, and ethylene (C_2H_4) was occasionally 169 used as a second tracer to pinpoint a specific source (nitrifying trickling filter).

170
$$E_g = Q_t \frac{(C_g - C_{g \ background})MW_g}{(C_t - C_{t \ background})MW_t}$$
(Eq. 1)

 E_g is the target gas emission in mass per time; Q_t is the tracer release in mass per time; C_g and C_t 171 are the measured off-gas concentrations in parts per billion (ppb); $C_{g \ background}$ and $C_{t \ background}$ 172 are the background concentrations of the target gas and tracer (ppb); and MW_g and MW_t are the 173 molar weights of target gas and tracer gas, respectively (Scheutz et al., 2011). The method can be 174 applied in static or mobile mode. In this study, the static tracer dispersion method was applied at a 175 176 ventilated duct in the thickening and dewatering building. In this case, C₂H₂ was released upstream in the enclosed ventilation duct, prior to a fan passage, so that proper mixing of tracer and target 177 gases was assured from the gas release point to the concentration sampling point. During 178 measurement, the analytical platform was positioned at a fixed location, and so the mode of the 179 tracer dispersion method is referred to as "static" (STDM). A detailed description of the static mode 180 can be found in Fredenslund et al. (2010). Conversely, the mobile mode of the method relies on 181 dynamic downwind concentration measurements of the mixed target and tracer gases, performed 182 across the plume by using the analytical platform in mobile mode. Concentrations in the mobile 183 184 mode in this study were integrated over the plume's cross-section to minimize the effects of any improper source simulation and gas mixing (Mønster et al., 2014b). Moreover, in this study, the 185 mobile approach was applied for specific on-site sources and to the whole WWTP for plant-186 integrated measurements (Table 3). Figure 1 shows an example of downwind plumes used for plant-187 integrated quantifications. A detailed description of the mobile mode (MTDM) can be found in 188 Yoshida et al. (2014) and Delre et al. (2017). 189

190 In the tracer dispersion method's mobile mode, concentration sampling is done at a distance away from the source, and the source emission is inherently dispersed to lower concentration levels 191 192 substantially at the sampling point compared to nearby the source. Despite the high sensitivity of the analytical instrument used, in cases where the emission rate of one of the target gases is very 193 low, the corresponding downwind plume cannot be properly distinguished from the background 194 195 concentration when measurements are performed at a long distance to the source. In this case, the 196 emission rate of the less abundant target gas can instead be inferred from the concentration ratio of 197 the less abundant gas to the more abundant target gas as being sampled closer to the source where concentrations are higher, combined with the emission rate of the more abundant target gas as 198 established with the MTDM (see Eq. 2). When measuring the target gas close to the source, care 199 200 must be taken not to prejudice the ratio, in case the target gases are not homogeneously mixed, and 201 the ratio should be based on an average sampling extended in both space and time.

202
$$\hat{E}^{i} = \overline{E}^{j} \cdot \frac{1}{k} \sum_{k} \int_{t1}^{t2} \frac{(c_{g}^{i} - c_{g background}^{i}) \cdot MW_{g}^{i}}{(c_{g}^{j} - c_{g background}^{j}) \cdot MW_{g}^{j}} dt \qquad \text{Eq. 2.}$$

203 where:

204 \hat{E}^{i} = the inferred emission rate of the secondary (less abundant) target gas species *i*

205 \overline{E}^{j} = the average emission rate of the main (more abundant) target gas species *j* obtained from

206 multiple plume transects as measured by MTDM,

207 $c_a^{i,j}$ = measured off-gas concentration (ppbv) of the main target gas *j* or the secondary target gas *i*,

208 $c_{g \ background}^{i,j}$ = measured background concentration (ppbv) of the main target gas *j* or the secondary 209 target gas *i*,

210 k = the number of gas ratio measurements, each integrated over a time window t_1 to t_2

211 $MW_{a}^{i,j}$ = molar weights of the measured main target gas *j* or the secondary target gas *i*,

212 For gaseous ammonia, the TDM method was complemented by measurements made with the optical remote sensing solar occultation flux (SOF) method (Johansson et al. 2014; Mellqvist et al. 213 2010, 1999; Mikel and Merrill, 2011). In this study, SOF was used for plant-integrated NH₃ 214 quantification. In Europe, the SOF technique is considered best available technology (BAT) 215 (Brinkmann et al., 2016) for measurements of diffuse emissions of volatile organic compounds 216 (VOCs) in the chemical sector. Recently, the SOF method has been used for NH₃ fugitive emissions 217 characterization from agricultural sources (Kille et al., 2017). 218 219 The SOF system applied herein is based on an identical FTIR spectrometer previously described for the TDM method, but instead of an internal infrared source (glowbar), it uses infrared 220

radiation from the sun. Solar light is continuously directed into the FTIR spectrometer by means of
a solar tracker as the measurement vehicle moves through the cross-section of the source emission
plume, and infrared spectra are then recorded sequentially. The spectra are analyzed for infrared
absorption by molecular species present in the source emission plume. The principle for retrieving
the target gas emission rate from a source via SOF measurement is given in Eq. 3.

226
$$E_{g} = \int_{Plume \ start}^{Plume \ stop} \left(\int_{0}^{Plume \ top} c_{g}(z) \cdot u'(z) \cdot dz\right) \cdot dx = \overline{u'_{mass}} \cdot \int_{Plume \ start}^{Plume \ stop} column(x) \cdot dx$$

227 Eq. 3

- where
- 229 E_a = emission rate of the target gas
- 230 $c_a(z)$ = concentration of the target gas at height (z) above the ground

231 u'(z) = wind speed at height (z) above the ground, specifically the wind speed component

232 orthogonal to the horizontal travel direction (*x*) through the plume

233 u_{mass} = plume mass weighted average wind speed, the component orthogonal to the travel direction

234 (x)

column (x) = concentration integrated in the vertical through the plume (e.g. along the slant beam of the sun, compensated with a cosine factor of the solar zenith angle).

To obtain gas emissions from a target source, the SOF instrument vehicle is driven crosswind through the plume of the source, so that solar light cuts through the plume and vertically integrated gas concentration columns are recorded. The measurement starts with an atmospheric background spectrum sampled outside the source plume, and consecutive spectra are then analyzed by reference to the atmospheric background spectrum. By adding consecutive gas column measurements, the integrated mass of the target species across the source plume is hence obtained. The source flux is retrieved by multiplying the integrated plume mass with the average wind speed of the plume.

The FTIR instrument was operated at 0.5 cm⁻¹ wavenumber resolution and a 5-second time 244 245 resolution. Precision in the measured NH₃ column was 0.08 mg/m². A prop and vane wind monitor (R.M. Young Wind Monitor, model 05103), mounted on a 4-meter mast at the measurement site, 246 measured wind speed and direction. Wind direction and speed were averaged over 1 minute and 247 recorded by a data logger (Campbell Scientific model CR200). The wind monitor had a stated 248 accuracy of $\pm 3^{\circ}$ and $\pm 0.3 \text{ ms}^{-1}$. Uncertainty in the flux estimate obtained by the SOF method was 249 dominated by uncertainty in the wind field (e.g. vertical plume distribution and plume transport 250 speed) and was typically $\pm 30\%$. 251

252 **2.4 Measurement campaigns**

In total, 13 measurement campaigns spread over three seasons in 2015 were performed under normal plant operations. Table 3 provides an overview of these campaigns, including measurement date and time intervals, applied measurement methods, adopted optical techniques, and target gases.

257 **3** Results and discussion

258 **3.1** Plant-integrated emission quantification

During the night between August 28th and 29th, plant-integrated measurements were performed by 259 applying MTDM, with two different analytical platforms (FTIR and CRDS) detecting CH₄, N₂O, 260 and NH₃ (Table 3). Three C₂H₂ gas cylinders, each with a different flow rate, were used to simulate 261 CH₄ emissions from the whole facility. Although part of the CH₄ was emitted from the activated 262 sludge reactors, the dominant CH₄ source was the sludge treatment line. CH₄ emission rates from 263 the two analytical platforms provided comparable results: 28.5 ± 3.1 kg CH₄ h⁻¹ and 33.5 ± 3.0 kg 264 CH_4 h⁻¹ for FTIR and CRDS, respectively (Table 4). Values are expressed as the average and 265 standard deviations of transects performed (AV \pm SD). Figure 2 shows the time series of the plant-266 integrated CH₄ emission quantification. CH₄ emission rates varied between 38.8 kg CH₄ h⁻¹ and 267 268 23.4 kg CH₄ h⁻¹, and they did not show any specific release trend throughout the 4 hours of measurements. No unusual operation was reported during the measurement campaign, such as 269 270 activation of the plant flare system for surplus biogas combustion or emergency venting of the digesters, which may have influenced emission patterns (Reinelt et al., 2015; Yoshida et al., 2014). 271 By normalizing emission rates to plant operation parameters so-called emission factors are 272 obtained, which are used for plant and technology intercomparison accounting for differences in 273 material throughput and process efficiencies. Furthermore, emission factors are often applied for 274 estimation of plant emissions (by multiplying with plant specific parameters e.g. material input or 275 removal efficiencies). Emission reporting based on emission factors is common (e.g. Doorn et al., 276 2006; Kampschreur et al., 2009) and there is a great need for providing reliable emission factors 277 representing current technologies to improve emission reporting. Table 5 reports CH₄ emissions 278 normalized by CH₄ production from anaerobic sludge digestion (e.g. kg CH₄ emitted per kg CH₄ 279 produced) and by organic load (chemical oxygen demand COD) to the plant (e.g. kg CH₄ emitted 280

per kg COD_{influent} into the plant), thereby providing CH₄ losses for analysis from an energetic or a 281 treatment capacity perspective, respectively. Since CH₄ was mainly emitted from the sludge 282 283 treatment, the measured CH₄ emissions were normalized using average CH₄ production and COD_{influent} into the plant recorded during the two months prior to the measurement campaign, in 284 order to account for the plant retention time of carbon entering the plant as COD in the influent and 285 being emitted as CH₄ from the biosolid stockpiles. CH₄ production is recorded every minute 286 whereas the COD in the plant inlet is measured five times per week. EFs were about 5.9% as kg 287 CH_4 (kg CH_4 prod.)⁻¹ and about 0.7% as kg CH_4 (kg COD influent)⁻¹, using an average whole site 288 CH₄ emission of 31.0 ± 3.1 kg CH₄ h⁻¹ (Table 5). Comparing these results with the previous two 289 studies performing plant-integrated CH₄ emission quantification from a Scandinavian WWTP 290 291 (Yoshida et al., 2014; Delre et al., 2017), CH₄ EF falls into the lower-to-medium part of the reported range (1.1 - 32.7% as kg CH₄ (kg CH₄ prod.)⁻¹ and 0.2 - 9.1% as kg CH₄ (kg COD 292 influent)⁻¹). 293

294 To measure N₂O emissions from the whole facility, one C₂H₂ tracer gas cylinder and one C₂H₄ gas cylinder were placed by the nitrifying trickling filters, as these were by far the most 295 significant N₂O source at the plant. Also for this target gas, the two analytical platforms provided a 296 297 comparable result: 4.0 ± 0.8 kg N₂O h⁻¹ and 6.4 ± 2.1 kg N₂O h⁻¹ for FTIR and CRDS, respectively (Table 4). Figure 3 shows the time series of the plant-integrated N₂O emission quantifications, 298 which varied between 2.4 kg N₂O h⁻¹ and 11.8 kg N₂O h⁻¹, and no clear trend in the emission was 299 observed. Figure 3 also shows variation in inlet wastewater flowing into the nitrifying trickling 300 filters, as well as the influent of nitrate (NO_3) and NH_4^+ , albeit no correlation with the measured 301 plant-integrated N₂O emissions could be seen. Similarly, no correlation of plant-integrated N₂O 302 emissions was found either with the drop in NO₃⁻ formation in the first hour of measurements or 303 with the peaks of NH₄⁺ removal occurring during nitrification at the nitrifying trickling filters 304

(Figure 4). N₂O emissions were normalized according to total nitrogen (TN) loaded into the plant 305 (kg N₂O-N (kg TN influent)⁻¹) and to TN removed from the plant (kg N₂O-N (kg TN removed)⁻¹). 306 307 The measured N₂O emission was normalized using the average TN influent and the average TN removed recorded during the week prior to the measurement campaign, to account for the plant 308 retention time of nitrogen entering the plant with the influent and being emitted as N₂O from the 309 nitrifying trickling filters. This was done as the nitrifying trickling filters were found to be main 310 N₂O emission source. No daily variation in TN influent into the plant was considered, as the 311 312 WWTP only records this data on a weekly basis. The reject water flow made up 2% of the total wastewater flow treated in the nitrifying trickling filters. It was not possible to relate the measured 313 N₂O emissions to the treatment of nitrogen in the reject water as the nitrogen content in the reject 314 315 water was not measured during the measurement campaign. Only nitrogen in the combined inlet 316 flow to the nitrifying trickling filters (reject water mixed into the main water flow) was measured (as shown in Figure 3 - NO_3^- and NH_4^+ influent flow). On average, the two analytical platforms 317 provided an EF of 0.9% as kg N₂O-N (kg TN influent)⁻¹ and an EF equal to 1.2% as kg N₂O-N (kg 318 TN removed)⁻¹ (Table 5) when using an average whole-site N₂O emission of 5.2 ± 1.5 kg N₂O h⁻¹. 319 These EFs fall somehow in the middle-lower part of the range (0.1 - 5.2%) as kg N₂O-N (kg TN 320 influent)⁻¹ and 0.1 - 5.9% as kg N₂O-N (kg TN removed)⁻¹)—as reported by previous plant-321 integrated N₂O emission quantifications of Scandinavian WWTPs (Yoshida et al., 2014; Delre et 322 323 al., 2017).

Plant-integrated NH₃ measurements were performed in two different campaigns using two different measurement methods but the same optical analytical technology, namely MTDM and SOF, with both using FTIR (Table 3). The main NH₃ source was the sludge treatment line. On July 2^{nd} , SOF measurements provided emissions of 0.4 ± 0.1 kg NH₃ h⁻¹, which were similar to measurements performed with MTDM during the night between August 28th and 29th and resulting in an emission of 0.4 ± 0.2 kg NH₃ h⁻¹ (Table 4).

330 Figure 5 shows plant-integrated NH₃ emission rates over time, recorded in the August campaign, which varied between 0.24 kg NH₃ h⁻¹ and 0.67 kg NH₃ h⁻¹, with two peak emissions 331 measured around 00:30 and 02:00 (August 28th and 29th). The main NH₃-emitting units were 332 biosolid stockpiles and thickening and dewatering buildings. Figure 5 shows no correlation between 333 the recorded peaks of plant-integrated NH₃ emissions and variation in the flow of digestate to the 334 centrifuges, or the flow of biosolids to the stockpiles. After almost one hour of measurements on 335 July 2nd, plant-integrated NH₃ emission quantifications did not show any trend (results not shown). 336 Considering the different nitrogen species in the wastewater and the strict relationship between NH₃ 337 and NH₄⁺, NH₃ emissions were normalized not only with TN influent and TN removed to and from 338 339 the plant, but also with NH₄⁺ influent and NH₄⁺ removed (Table 5). The measured NH₃ emission was normalized using the average TN influent and TN removed recorded during the two months 340 341 prior to the measurement campaign, in order to account for the plant retention time of nitrogen entering the plant and being emitted as NH₃ from the biosolid stockpiles. This was done because 342 sludge treatment processes (sludge dewatering and biosolid storage) were found to be the most 343 important NH₃ emission sources. NH₃ EFs were 0.09% as kg NH₃-N (kg TN influent)⁻¹, 0.11% as kg 344 NH₃-N (kg TN removed)⁻¹, 0.12% as kg NH₃-N (kg NH₄⁺ influent)⁻¹, and 0.15% as kg NH₃-N (kg NH₄⁺ 345 removed)⁻¹ (Table 5). Since this is the first time that NH₃ air emissions have been quantified from a 346 full-scale WWTP, no literature comparison is possible. 347

349 3.2 On-Site emission quantifications

350 3.2.1 On-site CH₄ emission sources and rates

351 Table 4 reports emission rates for all on-site CH₄ emitting process units, while Figure 6 provides the 352 average contribution of each process unit to the total CH₄ emission quantified from on-site sources. For each process unit, the average emission rate (Figure 6) was calculated based on emission rates 353 obtained in the different measurement campaigns (Table 4). The average emission value for the 354 biosolid stockpiles was calculated as a weighted value according to the operation of digesters, 355 356 namely in parallel and serial modes. The most important on-site CH₄ sources included the biosolid stockpiles (70%), ventilation from the thickening and dewatering building (11%), the sand trap inlet 357 (9%), and the activated sludge reactors (5%) (Figure 6). Overall, about 81% of the CH₄ emission 358 359 quantified on-site was released from the sludge treatment line.

360 Figure 7 shows CH₄ emissions from the biosolid stockpiles normalized by the amount of stored material. CH₄ emissions from still stockpiles were quantified over five measurement 361 362 campaigns, the first two of which were performed in winter when biosolids were produced with digesters operated in the parallel mode, while the last three measurement campaigns were 363 performed in summer when the biosolids were produced with digesters operated in the serial mode. 364 In the first two summer campaigns, measurements were performed when stockpiles were still and 365 during truck loading of biosolids for off-site treatment. Higher emissions were obtained from 366 biosolids produced when the digesters were operated in parallel (on average $8.77 \cdot 10^{-6}$ kg CH₄ (kg 367 TS biosolids)⁻¹ h^{-1}) in comparison to when they were operated in the series mode (on average 368 $3.77 \cdot 10^{-6}$ kg CH₄ (kg TS biosolids)⁻¹ h⁻¹). The lower CH₄ emission obtained when operating the 369 digesters in series mode was most likely a result of the higher share of input sludge CH₄ potential 370 being realized as the retention time for a larger share of the substrate in the digesters increased. In 371 fact, the CH₄ production was 7% higher when operating the reactors in series mode, in comparison 372

373	to when operated in parallel. A comparison of emissions escaping from still stockpiles and during
374	truck loading showed a CH ₄ release about 1.5-2.2 times higher as soon as the piles were moved
375	during loading. We suspect the higher release during loading to be caused by stripping of CH4
376	locked up in the sludge pore space or dissolved in the liquid phase of the sludge.
377	The second most important CH ₄ source was the ventilation exhaust of the thickening and
378	dewatering building, which resulted in an average release of 2.3 kg CH ₄ h ⁻¹ and represented 11% of
379	the total CH ₄ emissions measured from on-site sources.
380	$\rm CH_4$ stripping from the sand trap inlet resulted in an average release of 1.8 kg $\rm CH_4$ h ⁻¹ (or
381	about 9% of total CH ₄ emissions). Activated sludge reactors covered about 5% of the total CH ₄
382	emission quantified on-site with an average release of 1 kg CH ₄ h ⁻¹ . Primary and secondary settling
383	tanks covered about 4% and 2% of the total CH_4 emissions quantified on-site with an average
384	release of 0.8 and 0.4 kg CH_4 h ⁻¹ , respectively. Less than 1% of the total CH_4 emissions quantified
385	on-site was provided by nitrifying trickling filters and post-denitrifying MBBR tanks.
386	The sum of the emission for the on-site sources was 20.8 kg CH ₄ h ⁻¹ . Although measurements
387	were not performed at the same time, a qualitative comparison between on-site and off-site
388	measurements showed that the on-site CH4 emission rate was about two-thirds of the plant-
389	integrated CH ₄ emission quantification (on average about 31.0 kg CH ₄ h ⁻¹). The lower emission rate
390	obtained by measuring individual on-site sources in comparison to the plant-integrated emission,
391	apart from different timeframes, was most likely a result of not identifying all on-site sources. In
392	this study, potential missing sources could be CH ₄ slippages from the sludge treatment line and
393	other unknown sources. Previous studies have reported significant CH4 losses from the sludge
394	treatment line (Yoshida et al., 2014; Delre et al. 2017), underlining the reasonably higher value of
395	plant-integrated emission rates than on-site measurements, because the former decrease the risk of

missing important emission sources at the facility (Jensen et al., 2017; Reinelt et al., 2017; Yver
Kwok et al., 2015).

398 Furthermore, other studies that have included an investigation of both wastewater and sewage sludge lines have reported that the largest part of the CH₄ emitted from WWTPs is lost from the 399 sludge treatment line (Daelman et al., 2012; Delre et al., 2017). Both Mønster et al. (2014a) and 400 Delre et al. (2017) cited biosolid stockpiles as an important CH₄ source in WWTPs, but they 401 reported only plant-integrated measurements. Conversely, Majumder et al. (2014) investigated 402 403 seasonal CH₄ emissions from large biosolid stockpiles produced at a WWTP in Australia. Using a closed static chamber method, emissions were measured from stockpiles of different ages: More 404 than 3 years old, 1-3 years old, and less than 1 year old. CH₄ emissions measured from the less than 405 1-year old biosolid stockpiles were about 8.15.10⁻¹⁰ kg CH₄ (kg TS biosolids)⁻¹ h⁻¹, which is lower 406 than the biosolids CH_4 emissions measured in our study (which varied between 1.68 $\cdot 10^{-6}$ and 407 9.29.10⁻⁶ kg CH₄ (kg TS biosolids)⁻¹ h⁻¹ (Figure 7)). The higher emissions measured in our study are 408 most likely a result of the shorter storage time (about 3 weeks) in comparison to those in Majumder 409 et al. (2014) (less than one year). Larsen et al. (2017) measured CH₄ emissions from biosolid 410 stockpiles of mechanical dewatered sludge and reported emissions of 1.98.10⁻⁵ kg CH₄ (kg TS 411 412 biosolids)⁻¹ h⁻¹, which were measured from mechanical dewatered surplus activated sludge (less than one week old) that had not been stabilized by anaerobic digestion prior to dewatering, thus 413 potentially explaining the relatively high emission rates. Our emission rates are important for plant 414 emission reporting while for environmental assessment the emissions from off-site long-term sludge 415 416 storage should also be considered. In Scandinavia, the sludge storage time could be up to about 6 417 months as sludge is most often applied in spring and autumn. It is possible that the emission rates reported by Majumder et al. (2014) are more representative for longer-term (up to one year) sludge 418 419 storage. Larsen et al. (2017) also reports CH₄ emission rates for longer-term (4 months) sludge

storage $(3.55 \cdot 10^{-6} \text{ kg CH}_4 \text{ (kg TS biosolids)}^{-1} \text{ h}^{-1})$, which are lower than for short-term (less than a 420

one week old) storage of mechanical dewatered sludge. CH₄ emissions from mechanical pre-421

treatment have been reported (Wang et al., 2011; Yan et al., 2014) and explained as the 422

423 consequence of CH₄ formation in the sewer network due to anaerobic conditions (Liu et al., 2015).

424

3.2.2 On-site N₂O emission sources and rates

Table 4 reports emission rates for all N₂O-emitting process units, while Figure 6 shows the average 425 426 contribution of all process units to the total N₂O emissions quantified from on-site sources. For each process unit, the average emission rate (Figure 6) was calculated based on emission rates obtained 427 in the different measurement campaigns (Table 4). The total on-site N₂O emission quantification 428 (3.3 kg N₂O h⁻¹) was about two-thirds of the plant-integrated emission quantification (5.2 kg N₂O h⁻¹) 429

¹), which was similar to the CH₄ results comparing on-site emissions with total emissions. 430

The most relevant N₂O emission sources were the nitrifying trickling filters, which emitted 431 about 2.7 kg N₂O h⁻¹, covering about 82% of the total N₂O emission quantified on-site (Figure 6). 432 Secondary settling tanks and biosolid stockpiles each covered about 5% of the total N₂O emissions 433 quantified on-site - each with an average annual release of 0.2 kg N_2O h⁻¹. Figure 7 describes N_2O 434 emissions from biosolids normalized by the amount of material stored. In general, lower N₂O 435 emissions were obtained from the produced biosolids when the digesters were operated in parallel 436 (on average 1.69·10⁻⁸ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹), in comparison to when the digesters were 437 operated in the series mode (on average 7.16.10⁻⁸ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹). This variation 438 in emissions cannot be explained by variations in concentrations of TS, TN, and NH₄-N in the 439 440 produced biosolids, as these did not change significantly between the two different digestion modes: Average TN concentrations were 43.7 and 43.5 g (kg TS)⁻¹ for the serial and the parallel mode, 441 respectively, while average NH₄-N concentrations were 12.4 and 12.7 g (kg TS)⁻¹ for the serial and 442 the parallel mode, respectively (Figure 8). The small change in TS content was caused mainly by 443

the centrifuge's dewatering capabilities. It is possible that higher temperatures during summer in 444 comparison to winter resulted in higher microbial activity and thus caused higher emissions during 445 446 summer. However, this was not the case for CH₄, as the highest emissions were recorded during winter when the reactors were operated in parallel. During truck loading, N₂O emissions were 447 almost four times higher than from the still piles; however, in July, N₂O emissions during truck 448 loading were comparable to the still piles (Figure 7). Considering the emission of both CH4 and 449 N₂O, the total greenhouse gas emission from biosolids reported in CO₂-equivalents was lower 450 451 $(1.35 \cdot 10^{-4} \text{ kg CO}_2 \text{ eq.} (\text{kg TS biosolids})^{-1} \text{ h}^{-1})$ when the reactors were operated in a serial mode in comparison to a parallel mode $(2.53 \cdot 10^{-4} \text{ kg CO}_2 \text{ eq.} (\text{kg TS biosolids})^{-1} \text{ h}^{-1})$. 452

453 Post-denitrifying MBBR, primary settling tanks, activated sludge reactors, the sludge 454 treatment building, and the sand trap inlet had average emission rates lower than 0.1 kg N_2O h⁻¹ and 455 collectively made up only a small share (8%) of the total N_2O emission quantified on-site (Figure 456 6).

Different N_2O emission rates recorded by different campaigns at nitrifying trickling filters and biosolid stockpiles (Table 4) could be justified by the complex relationship between N_2O generation and different factors such as oxygen availability, nitrite content, COD, and TN ratio (Kampschreur et al., 2009).

Majumder et al. (2014) investigated N₂O emissions from large still biosolid stockpiles less than 1 year old and reported their emissions as $1.64 \cdot 10^{-8}$ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹, which is within the range measured in our study (from $3.3 \cdot 10^{-9}$ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹ to $1.25 \cdot 10^{-7}$ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹ (Figure 7)). Similarly, Larsen et al. (2017) measured N₂O emissions escaping from biosolid stockpiles of mechanically dewatered sludge and reported emissions of $1.44 \cdot 10^{-7}$ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹ for material less than one week old while the emission rate of older material (stored for 4 months) was $3.97 \cdot 10^{-8}$ kg N₂O-N (kg TS biosolids)⁻¹ h⁻¹.

469 **3.2.3** On site NH₃ emission sources and rates

470 Table 4 reports emission rates for all NH₃ emitting process units, while Figure 6 shows the average 471 contribution of all process units to the total NH₃ emission quantified on-site. For each process unit, the average emission rate (Figure 6) was calculated based on emission rates obtained in the different 472 473 measurement campaigns (Table 4). The most important NH₃-releasing process units were the biosolid stockpiles and the exhaust used to ventilate the thickening and dewatering building, 474 contributing 44% and 22% of the total NH₃ emissions quantified on-site and emitting 0.17 and 0.09 475 476 kg NH₃ h⁻¹, respectively. Lower NH₃ emissions were in general obtained from biosolids produced when the digesters were operated in parallel (on average 4.87.10⁻⁸ kg NH₃-N (kg TS biosolids)⁻¹ h⁻¹) 477 in comparison to when they were operated in series mode (on average 6.94.10⁻⁸ kg NH₃-N (kg TS 478 biosolids)⁻¹ h⁻¹). This difference cannot be explained by changes in concentrations of TS, TN, and 479 NH₄-N in the produced biosolids (Figure 8). In the campaign performed in June, NH₃ emissions 480 were almost the same during truck loading and still biosolids (Figure 7), whereas in the July 481 campaign, they emissions were almost three times higher during truck loading in comparison to the 482 still stockpiles (Figure 7). 483

Settling tanks (primary and secondary), the sand trap inlet, activated sludge reactors, and nitrifying trickling filters all contributed to the remaining percentage (34%) of the total NH₃ emissions quantified on-site (Figure 6). The sum of the average NH₃ emissions quantified on-site from different process units provided approximately the same emission rate obtained when quantifying plant-integrated NH₃ emissions (0.4 kg NH₃ h⁻¹) (Table 4), although the measurements represent different timeframes.

491 **4** Conclusions

492	٠	Plant-integrated CH ₄ emission rates measured using ground-based remote sensing methods
493		varied between 28.5 and 33.5 kg CH_4 h ⁻¹ , corresponding to an average emission factor of 5.9%
494		as kg CH ₄ (kg CH _{4 production}) ⁻¹ , whereas N ₂ O emissions varied between 4.0 and 6.4 kg h ⁻¹ ,
495		corresponding to an average emission factor of 0.9% as kg N ₂ O-N (kg TN $_{influent}$) ⁻¹ .
496	•	Plant-integrated NH ₃ emissions were around 0.4 kg h ⁻¹ , corresponding to an average emission
497		factor of 0.11% as kg NH ₃ -N (kg TN $_{removed}$) ⁻¹ .
498	•	On-site measurements showed that the largest proportion of the CH ₄ was emitted from the
499		sludge treatment line, with about 70% coming from biosolid stockpiles. While about 82% of the
500		N ₂ O was emitted from nitrifying trickling filters, the most relevant NH ₃ sources were the
501		biosolid stockpiles (44%) and the thickening and dewatering building (22%).
502	•	Biosolids showed different emissions when the sludge digesters were operated in series rather
503		than in parallel mode, as usually done at the WWTP. When the digesters were operated in series
504		mode, lower CH_4 emissions and generally higher N_2O and NH_3 emissions were observed.
505		Emissions of CH ₄ , N ₂ O, and NH ₃ tended to be higher while loading biosolids onto trucks.
506	•	On-site CH_4 and N_2O emission quantifications accounted for approximately two-thirds of the
507		plant-integrated emission quantifications. The difference could be a combined effect of different
508		survey timeframes and that no sources were identified during the on-site investigation. Off-site
509		air emission quantifications, using ground-based remote sensing methods, thus seem to provide
510		more comprehensive total plant emission rates, whereas on-site measurements provide insights
511		into emissions from individual sources.
512		

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630 TABLES

631	Table 1. Plant key	parameters in the invest	igated year.
	Year		2015
	Population Equivaler	nt (PE)	805,000
	Treated wastewater ($(m^3 yr^{-1})$	147,300,000
		BOD	20555
	Influent wastewater	COD	43431
	(Mg yr ⁻¹)	TN	3366
		NH_4^+ -N	2367
		BOD	1150
	Effluent	COD	5916
	(Mg yr ⁻¹)	TN	1169
		NH_4^+ -N	823
		BOD	94.4
	Pollutant removal	COD	86.4
	(%)	TN	65.3
		NH_4^+ -N	65.2
	External material	Food waste	13,783
	(Mg yr ⁻¹)	Sewage sludge	79,220
	Plant	Biosolids (Mg TS yr ⁻¹)	14,846
	n raduction	Biogas (m ³ yr ⁻¹)	11,496,818
	production	CH ₄ content in biogas (%)	63.1

Optical technique	Measuring principle	Model	Measured gas	Used absorption wave length	Precision	Detection frequency
			CH ₄	3.3 µm	1.7 ppb	9 s
		IDCube Metric M. Druhen	N_2O	4.5 μm	0.3 ppb	9 s
FTIR	Infrared absorption spectroscopy	Optics GmbH, Ettlingen,	NH ₃	10.3 µm	2.0 ppb (MTDM) 0.08 mg/m ² (SOF)	9 s (MTDM) 5 s (SOF)
		Germany	C_2H_2	13.7 μm	1.8 ppb	9 s
			C_2H_4	10.5 μm	4.7 ppb	9 s
	Gas concentration is obtained by	G2203, Picarro, Inc., Santa	CH ₄	NA	0.77 ppb	0.5 s
CDDC	measuring directly the "ring-down," or	Clara, CA	C_2H_2	NA	0.06 ppb	0.5 8
CKDS	decay of laser light in a sample cell.	S/N JADS2001, Picarro,	N ₂ O	NA	7.7 ppb	2 с
		Inc., Santa Clara, CA	C_2H_2	NA	0.6 ppb	38

633 Table 2. Overview of applied optical analytical techniques.

634 FTIR: Fourier Transform Infrared. CRDS: Cavity Ring-Down Spectroscopy. NA: Information not available. Precision: the capability to reproduce the same

635 concentration measurement and is expressed as the standard deviation of concentrations recorded during a specific time frame (FTIR: 10 minutes, CRDS: 60 minutes)
 636 when a gas with a constant concentration goes through the instrument.

Date (dd/mm)	Campaign starting time (hh:mm)	Campaign ending time (hh:mm)	Waste water treatment process unit investigated	Optical analytical technology applied	Investigated emissions and measuring method (as superscript, STDM ¹ , MTDM ² , SOF ³ , inferred flux ⁴ using STDM, MTDM, or SOF of main target gas combined with mass ratio measurement of the less abundant gas in plume versus the main target gas)
12/01	11:28	14:40	Ventilation exhaust of thickening and dewatering building	FTIR	N ₂ O ⁻¹ , CH ₄ ⁻¹ , NH ₃ ⁻¹
12/01	12:26	13:48	Biosolid stockpiles	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
13/01	14:24	14:48	Primary settlers	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
14/01	11:45	12:03	Biosolid stockpiles	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
14/01	13:47	14:45	Sand trap inlet	FTIR	N_2O^2 , CH_4^2 , NH_3^2
	13:19	14:19	Nitrifying trickling filters	FTIR	N ₂ O ²
19/02	13:18	14:19	Post-denitrifying MBBR	FTIR	N ₂ O ² , CH ₄ ⁴ , NH ₃ ⁴
	13:17	14:14	Secondary settlers	FTIR	N_2O^2 , CH_4^2 , NH_3^2
09/03	11:39	12:13	Nitrifying trickling filters	FTIR	N ₂ O ² , CH ₄ ⁴ , NH ₃ ⁴
11/02	10:53	11:58	Sand trap inlet	FTIR	N ₂ O ² , CH ₄ ² , NH ₃ ²
11/05	11:05	13:15	Primary settlers	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
17/06	10:24	11:16	Ventilation exhaust of thickening and dewatering building	FTIR	N ₂ O ¹ , CH ₄ ¹ , NH ₃ ¹
	11:54	12:15	Centrifuge room	FTIR	N ₂ O ¹ , CH ₄ ¹ , NH ₃ ¹
22/06	10:37	12:25	Nitrifying trickling filters	FTIR	N ₂ O ²
25/00	13:30	14:22	Activated sludge reactors	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
24/06	10:21	13:01	Biosolid stockpiles	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
02/07	10:45	11:37	Whole plant (plant-integrated measurements)	FTIR	NH ₃ ³
06/07	11:42	14:52	Biosolid stockpiles	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
28/08	23:18	23:32	Ventilation exhaust of thickening and dewatering building	FTIR	CH ₄ ²
	23:51	00:44	Nitrifying trickling filters	FTIR	N ₂ O ²
28 8-20/09	23:22	00:58	Biosolid stockpiles	FTIR	CH ₄ ² , N ₂ O ⁴ , NH ₃ ⁴
200229/08	22.20	02.20	Whole plant (plant-integrated	FTIR	N ₂ O ² , CH ₄ ² , NH ₃ ²
	22.30	02.50	measurements)	CRDS	N_2O^2 , CH_4^2

Table 3. Overview of performed measurement campaigns in 2015.

638 MTDM: Mobile Tracer Dispersion Method. STDM: Static Tracer Dispersion Method. SOF: Solar Occultation Flux. FTIR: Fourier Transform Infrared. CRDS: Cavity

639 Ring-Down Spectroscopy. MBBR: Moving Bed Bioreactors. Please note that measurements were done interchangeably between different units during specified time

640 frames.

Investigated	D (CH4	N ₂ O	NH ₃
wastewater treatment	Date	$(kg h^{-1})$	$(kg h^{-1})$	$(kg h^{-1})$
process unit	(dd/mm)	$AV \pm SD$	$AV \pm SD$	$AV \pm SD$
Sand tran inlat	14/01	3.3±1.5	0.01±0.02	0.05±0.10
Sand trap inter	11/03	0.25±0.11	0.01±0.02	0.01±0.02
Drimory sottlars	13/01	0.64±0.22	0.08±0.05	0.08 ± 0.07
Fillinary settlers	11/03	0.98±0.46	0.09±0.07	0.04±0.03
Activated sludge reactors				
Terrvated studge reactors	23/06	0.99±0.79	0.060±0.059	0.013±0.011
	19/02	n.m.	1.9±1.3	n.m.
Nitrifying trickling filters	09/03	0.10 ± 0.05	1.8±0.6	0.001 ± 0.001
withing theking inters	23/06	n.m.	4.0±0.8	n.m.
	28&29/08	n.m.	3.2±0.2	n.m.
Post-denitrifying MBBR	19/02	0.008 ± 0.004	0.08±0.03	< 0.002
Secondary settlers	19/02	0.4±0.17	0.18±0.11	0.03±0.02
Vantilation and and afthislaning	12/01	1.65±0.11	0.041 ± 0.004	0.041±0.010
ventilation exhaust of thickening	17/06	1.74±0.16	0.005±0.001	0.13±0.01
and dewatering building	28/08	3.4±1.4	n.m.	n.m.
Centrifuge room ^b	17/06	0.43±0.01	0.00080±0.00002	0.0043±0.0001
	13/01	22.5±6.4	0.05±0.06	0.12±0.10
	14/01	20.4±4.5	0.08 ± 0.06	0.17±0.09
Biosolid stockpiles ^c	24/06	6.5±2.8 [½14.3±3.5] ^a	0.28±0.15 [1.07±0.38] ^a	0.18±0.11 [0.19±0.09]
-	06/07	13.4±2.5 [19.4±5.2] ^a	0.36±0.16 [0.25±0.15] ª	0.28±0.09 [0.81±0.56]
	28&29/08	9.7±2.5	0.03±0.01	0.12±0.07
Whole plant	02/07	n.m.	n.m.	0.4±0.1 [FTIR-SOF]
(plant-integrated measurements)	28&29/08	28.5 ± 3.1 [FTIR] 33.5 ± 3.0 [CRDS]	4.0 ± 0.8 [FTIR] 6.4 ± 2.1 [CRDS]	0.4±0.2 [FTIR-MTDM

^a Values in the brackets refer to quantifications during truck loading. ^b Notice that the centrifuge room is placed inside the thickening and dewatering building. 642

643 FTIR: Fourier Transform InfraRed. CRDS: Cavity Ring-Down Spectroscopy. For results based on inferred flux using STDM or MTDM combined with mass ratios,

the given uncertainty corresponds to the combined uncertainty of the STDM/MTDM and the uncertainty in the mass ratio. ° Note that the sludge digesters were 644

645 operated in parallel mode January-May, and serial mode June-August. N.m. denotes not measured.

S C

Car	Naunalization	EF	Method and	Average	
Gas	Normalization	$AV \pm SD$	analytical technology	EF	
	$k_{\alpha} CH (k_{\alpha} COD) \rightarrow 1 (9/)$	0.6 ± 0.1	MTDM and FTIR	0.7	
വ	Kg CH_4 (Kg COD influent) ⁻ (70)	0.7 ± 0.1	MTDM and CRDS	0.7	
_п4	$k \alpha C H (k \alpha C H)^{-1} (0/)$	5.4 ± 0.6	MTDM and FTIR	5.0	
	Kg CI14 (Kg CI14 production) (70)	6.4 ± 0.6	MTDM and CRDS	5.9	
kg N2O-N (kg TN influent) ⁻¹ (%)	0.7 ± 0.1	MTDM and FTIR	0.0		
IO	kg N ₂ O-N (kg IN influent) $(\%)$ 1.1 ± 0.4 MTDM and CRDS	MTDM and CRDS	0.9		
N20	$kg N_{0} O_{1} N (kg T N_{1})^{-1} (\%)$	1.0 ± 0.2	MTDM and FTIR	12	
	kg 1120-11 (kg 111 removed) (70)	1.5 ± 0.5	MTDM and CRDS	1.2	
	$kg NH_{2}N (kg TN + g -)^{-1} (%)$	0.09 ± 0.04	MTDM and FTIR	0.00	
	Kg [11]-10 (Kg 110 influent) (70)	0.09 ± 0.02	SOF and FTIR	0.07	
	$kg NH_{2}-N (kg TN _{2})^{-1} (\%)$	0.11 ± 0.05	MTDM and FTIR	0.11	
JH-	Kg 1113 1 (Kg 111 removed) (70)	0.11 ± 0.03	SOF and FTIR	0.11	
11 3	kg NH ₂ -N (kg NH ₄ ⁺ induce) $^{-1}$ (%)	0.12 ± 0.06	MTDM and FTIR	0.12	
	ing total a transfer (10)	0.12 ± 0.03	SOF and FTIR	0.12	
	kg NH ₂ -N (kg NH ₄ ⁺ removed) $^{-1}$ (%)	0.15 ± 0.07	MTDM and FTIR	0.15	
	(/v)	0.15 ± 0.04	SOF and FTIR	0.10	

648 Table 5. Plant-integrated Emission Factors (EFs).

649 AV: average. SD: Standard Deviation. FTIR: Fourier Transform Infrared. CRDS: Cavity Ring-Down Spectroscopy.

650 SOF: Solar Occultation Flux. EFs for CH₄ and NH₃ were normalized to the COD and TN influent content of the

wastewater and thus did not consider the COD and TN content in the food waste occasionally fed to the anaerobic

digester. Due to the low amounts of food waste fed to the anaerobic digested this would have only minor effect on the

653 EFs (<0.05%).

655 **FIGURES**

656

657



Figure 1. Downwind methane and acetylene plumes along a road 350 m from the main methane

- sources measured at 01:15 on 29/08. Yellow triangles mark tracer gas positions. The methane
- 660 plume is reported in red, while the acetylene plume is depicted with a yellow line. WWTP borders 661 are marked in blue.



- 663 Figure 2. CH₄ plant-integrated emissions over quantification time (28&29/08) measured with FTIR-
- MTDM and CRDS-MTDM. FTIR: Fourier Transform InfraRed. CRDS: Cavity Ring-Down
 Spectroscopy. MTDM: Mobile Tracer Dispersion Method.





Figure 3. N_2O plant-integrated emissions over quantification time (28&29/08) measured with FTIR-MTDM and CRDS-MTDM. The graph shows the variation in the inlet wastewater flow to the

nitrifying trickling filters, as well as the influent of NO₃⁻ and NH₄⁺. FTIR: Fourier Transform

670 InfraRed. CRDS: Cavity Ring-Down Spectroscopy. MTDM: Mobile Tracer Dispersion Method.





Figure 4. N₂O plant-integrated emissions over quantification time (28&29/08) measured with FTIR-MTDM and CRDS-MTDM. The graph shows NH₄⁺ removed and NO₃⁻ formation based on on-line measured inlet and outlet concentrations at the nitrifying trickling filters. FTIR: Fourier Transform
 InfraRed. CRDS: Cavity Ring-Down Spectroscopy. MTDM: Mobile Tracer Dispersion Method.





Figure 5. NH₃ plant-integrated emissions over quantification time (28&29/08) measured with FTIR-

679 MTDM. The graph shows the quantity of digestate processed in the centrifuges and the quantity of

680 produced biosolids sent to the stockpiles. FTIR: Fourier Transform InfraRed. CRDS: Cavity Ring-

681 Down Spectroscopy. MTDM: Mobile Tracer Dispersion Method.



Figure 6. Average contribution of all process units to the total CH₄, N₂O and NH₃ emission

685 quantified from on-site sources.



Figure 7. Air emissions from biosolid stockpiles normalized by the amount of stored material. The first two campaigns were performed when digesters were run in parallel, while the remaining were

690 first two campaigns were performed when digesters were run in parallel, while the remaining were 691 performed when the digesters were run in series. For two measurement campaigns, comparison

between still and loading of biosolids is showed. Emission factors (EF) are reported with

693 uncertainty representing the standard deviation of several successful transects. Campaign date is

reported as dd/mm since all measurements referred to 2015.

Figure 8. Concentrations of TS, TN and NH₄-N in the produced biosolids in 2015. 701