Toxic phytoplankton as a keystone species in aquatic ecosystems: stable coexistence to biodiversity

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1 **Toxic Phytoplankton as a Keystone Species in**
2 **Aquatic Ecosystems: Stable Coexistence to**
3 **Biodiversity***

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Abstract

The effect of allelochemicals released by toxic species in plankton community is often taken into account to reveal plankton biodiversity. Using a minimal chemostat model we show that the interaction between toxic and non-toxic phytoplankton species with changing competitive effects among species due to allelopathy helps to promote the stable coexistence of many species on a single resource and hence can solve the paradox of plankton. We emphasize toxic phytoplankton as a keystone species that strongly uncovers its allelochemicals on other non-toxic phytoplankton and enhances the species persistence and diversity in aquatic ecosystems. In addition, we analyze the consistency of ecosystem functioning and species diversity using a number of approaches, such as sampling hypothesis with selection and complementarity effects, cascading extinction-reinvasion, and examining system dynamics at different enrichment levels and toxicity. Our results suggest that chemostats with one toxic and one or more nontoxic phytoplankton species can be used for the experimental verification of the stable coexistence of many species on a single resource in aquatic ecology.

39 1 Introduction

40 Plankton biodiversity has fascinated ecologists for the past few decades. It has become a major
41 topic of research in plankton ecology. Although the principle of competitive exclusion (Hardin
42 1960) tells us that the number of surviving species cannot exceed the number of limiting resources
43 at equilibrium, still many plankton species coexist depending on a limited number of resources. In
44 the famous paper “The paradox of plankton”, Hutchinson (1961) first posed the classical problem
45 “... how it is possible for a number of species to coexist in a relatively isotropic or unstructured
46 environment all competing for the same sorts of materials”. Numerous attempts have been made to
47 explain this paradoxical diversity. In isotropic environments, non-equilibrium dynamics (i.e. limit
48 cycle or chaotic oscillations) has been suggested to play a leading role in maintaining the desired
49 coexistence of many species on limited variety of resources (Huisman and Weissing 1999, Czárán et
50 al. 2002, Revilla and Weissing 2008, Dutta et al. 2014). Though the approach of non-equilibrium
51 dynamics as a possible solution of the paradox of plankton is fascinating, these findings are still
52 debated in literature (Schippers et al. 2001, Shores et al. 2008, Barton et al. 2010). The
53 explanation of the paradoxical diversity of phytoplankton in isotropic environments governed by
54 equilibrium dynamics is still an unachieved goal. A detailed review of various factors responsible
55 for the plankton biodiversity has been given in the papers by Scheffer et al. (2003) and Wilson
56 (2011).

57 Allelopathy has a great impact on plankton ecosystems. By definition, allelopathy describes
58 any inhibitory or stimulatory effect of one species on another species mediated by the production of
59 some chemical compounds (Rice 1984). Nowadays, however, the term allelopathy is often used to
60 represent an inhibitory effect of biochemical compounds (Cembella 2003). Allelochemical
61 interactions between different populations can have different ecological consequences (Rice 1984).
62 Regarding species distribution in aquatic ecosystems, allelochemicals produced by some species of
63 phytoplankton (generally known as toxin producing phytoplankton) play a crucial role in shaping

66 ecological communities (Macias et al. 2008). In fact, these toxic species act as keystone species
67 where keystone species can be defined as the one whose impact is large on the ecosystem and its
68 communities, disproportionate with its abundance and plays a structural role in the ecosystem (Mills
69 et al. 1993). As a keystone species, toxic phytoplankton negatively affect species interactions by
70 producing toxic chemicals and hence influence other competing species and grazers. However,
71 laboratory experiments and modeling results suggest that allelopathy becomes important and
72 effective only when toxic species abundance is very high (Schmidt and Hansen 2001, Solé et al.
73 2005) and thus allelopathy can have a profound role in toxic bloom prolongation. Recently, Van de
74 Waal et al. (2014) studied the stoichiometric regulation of phytoplankton toxins which may help us
75 to predict toxin-producing phytoplankton blooms and also can manage to control the overall
76 toxicity in natural waters.

77 However, the role of allelopathy (i.e. the presence of a toxic species in the community) in
78 maintaining the large scale biodiversity of phytoplankton is not well established. Although, most of
79 the competition results demonstrate the exclusion of one of the species (Adams et al. 1979, Chao
80 and Levin 1981), only a few empirical data reported the coexistence of toxin producing and
81 sensitive strains in natural and laboratory systems (Abranches et al. 1997, Ruiz-Barba et al. 1994).
82 Such production of toxins is known to reduce the competitive effectiveness among different species
83 either by reducing the abundance of other species (Folt and Goldman 1981, Solé et al. 2005) or by
84 creating spatial separation between species (Frank 1994), which ultimately results coexistence.
85 Previous theoretical studies show the coexistence in non-homogeneous environments or governed
86 by non-equilibrium dynamics (Tilman 1982, Huisman and Weissing 1999, Czárán et al. 2002,
87 Revilla and Weissing 2008, Dutta et al. 2014). In the present work, using a minimal mechanistic
88 modelling approach, theoretically we explore the role of allelopathy on the biodiversity of
89 phytoplankton in a homogeneous environment governed by equilibrium dynamics.

90 We extend the well known Droop model (Droop 1973) in the context of multiple species

91 and single resource in the presence of allelopathy among phytoplankton. The Droop model is the
 92 standard quota model of resource competition (Grover 1991, Grover and Wang 2014). Previously,
 93 Revilla and Weissing (2008) studied the effects of resource storage by considering a similar model
 94 for multiple resources. In a homogeneous environment, they found the coexistence of many species
 95 on few resources governed by non-equilibrium dynamics and this is possible only when the number
 96 of limiting resources is either three or more. There has been a long-standing debate to identify the
 97 relationship between stability and diversity. Trait based approach plays a key role in predicting the
 98 diversity effects on the stability of ecosystems (Loreau and Mazancourt 2013). In order to get a
 99 stable ecological community, species traits must differ in specific ways (Clark et al. 2007). In the
 100 present paper, considering allelopathy as an important trait, we investigate how the presence of a
 101 single toxic species promotes the biodiversity of phytoplankton community under single resource.
 102 Using chemostat approach, first we find the species diversity by simulation technique and then
 103 check the stability of those existing species. Secondly, to identify the impact of allelochemical
 104 interactions on phytoplankton biodiversity, we find the supersaturation region for different toxicities
 105 and biodiversity measures like net effect, selection effect, complementarity effect and resource use
 106 efficiency. Further, cascading extinction patterns following the removal and the reinvasion of a
 107 species are also investigated. In addition, we discuss α -species diversity in fluctuating environmental
 108 conditions by varying the nutrient concentration at different levels.

109 **2 Model description**

110 The model we analyze here is derived from the standard Droop model of resource competition
 111 (Droop 1973). The Droop model describes a well-mixed bioreactor (i.e. a chemostat) that contains
 112 phytoplankton cells with density N_i (10^9 cells l^{-1}) in a growth medium of a limiting nutrient with
 113 concentration R ($\mu\text{mol } l^{-1}$). Each phytoplankton cell is assumed to possess an internal pool of stored

114 nutrients (the so-called cell quota) Q_i (10^{-9} $\mu\text{mol cell}^{-1}$). The chemostat is supplied with nutrients
 115 at an input concentration S ($\mu\text{mol l}^{-1}$) from an external medium. The outflow contains
 116 both medium and phytoplankton cells. Inflow and outflow are characterized by the dilution rate D
 117 (day^{-1}). In the present model, one of the species (N_1) produces toxin that induces additional
 118 mortality in all the other non-toxic species ($N_2, N_3, \dots, N_n$) via allelopathy parameter θ_i
 119 ($\text{l}^3 (10^9 \text{cells})^{-3} \text{day}^{-1}$). Specifically, here θ_i measures the strength of allelopathic effect of N_1 on
 120 the i -th species. Under these assumptions, the modified Droop model for n -species and one-resource
 121 takes the following form:

$$122 \quad \frac{dR}{dt} = D(S-R) - \sum_{i=1}^n f_i(R) N_i, \quad i=1, \dots, n, \quad (1a)$$

$$123 \quad \frac{dQ_i}{dt} = f_i(R) - \mu(Q_i) Q_i, \quad i=1, \dots, n, \quad (1b)$$

$$124 \quad \frac{dN_1}{dt} = [\mu(Q_1) N_1 - m_1] N_1, \quad (1c)$$

$$125 \quad \frac{dN_i}{dt} = [\mu(Q_i) - m_i] N_i - \theta_i N_1^2 N_i^2, \quad i=2, \dots, n, \quad (1d)$$

126 with

$$127 \quad f_i(R) = \frac{v_i R}{K_i + R} \quad \text{and} \quad \mu(Q_i) = r_i \left(1 - \frac{q_i}{Q_i}\right).$$

128 Uptake of the resource R from the external medium is assumed to be an increasing and a
 129 saturating function $f_i(R)$ of the external resource concentration, where v_i (10^{-9} $\mu\text{mol cell}^{-1}$
 130 day^{-1}) is the maximum uptake rate and K_i ($\mu\text{mol l}^{-1}$) is the half-saturation constant for the i -th
 131 species. It is assumed that the phytoplankton growth rate $\mu(Q_i)$ depends entirely on the cell quota
 132 Q_i , where r_i (day^{-1}) is the maximum growth rate under quota saturation and q_i (10^{-9} μmol

cell⁻¹) is the minimum amount of subsistence quota per phytoplankton cell. For $Q_i > q_i$ the growth rate is positive. However, it is set to zero if $Q_i < q_i$. The species specific mortality rate is given by m_i (day⁻¹), which is considered to be same with the dilution D for all the species.

It is evident from Eqs. (1) that we introduce additional mortality to the non-toxic phytoplankton due to allelopathy. From a theoretical point of view, allelopathic interaction was first introduced in a two species Lotka-Volterra competition model by Maynard-Smith (1974). He considered a constant release of toxic chemicals by one species appearing harmful to the other one. After necessary modifications, this model was applied in several directions including allelopathy in bacterial (Nakamaru and Iwasa 2000) and plant community (Dubey and Hussain 2000, An et al. 2003). Concerning allelopathy among phytoplankton, Chattopadhyay (1996) applied this concept in a general two species competition system where each species produces a substance toxic to the other, but only when the other is present. Mathematically, if N_1 and N_2 are the population densities of the toxic and the non-toxic phytoplankton respectively, then the allelopathic interaction term was considered as $\theta_2 N_1 N_2^2$, where $\theta_2(>0)$ is the allelopathy parameter representing the strength of the allelopathic effect of species N_1 on N_2 .

There is evidence that allelopathy among phytoplankton becomes important only when the toxic species reaches high density. Maestrini and Graneli (1991) suggested that at a relatively low population density (10^4 cells/l), the toxin produced by *C. Polylopes* helps only in repelling grazers, but at high cell density ($> 10^6$ cells/l), the concentration of accumulated toxin becomes high and starts affecting competitors. Based on 21 published experimental works, Jonsson et al. (2009) performed a meta-analysis and detected significant allelopathic effect only in studies using high but not low *Chlorophyll a* content. Allelopathic term used by Chattopadhyay (1996) does not exactly satisfy the above mentioned criteria and shows proportional allelopathic effect with all concentrations of cell density of the toxic species. Solé et al. (2005) verified the expression

157 representing allelopathic interaction used by Chattopadhyay (1996) with the help of experimental
 158 data taken from a laboratory study where total 15 species of marine phytoplankton were cultured
 159 together with the toxic *Chrysocromulina polylepis* (Schmidt and Hansen 2001). They found that the
 160 model with the allelopathic term similar to Chattopadhyay (1996) showed discrepancies at low
 161 initial concentrations of the toxic species and suggested that the allelopathic effect should be a
 162 nonlinear function of the amount of toxic cells present in the medium rather than the linear one.
 163 While modifying the expression by $\theta_2 N_1^2 N_2^2$ they found a better agreement between the
 164 theoretical outcome and the experimental data at both high and low concentrations of toxic cells. In
 165 accordance with the previous findings, we also consider a similar type of expression to represent the
 166 allelopathic effect. In our model, the first species, N_1 , is assumed to be toxic and it produces
 167 toxin which increases the mortality of all the remaining species by the quantity
 168 $\theta_i N_1^2 N_i^2$, $i=2, \dots, n$. It is natural that the amount of toxin around the toxic phytoplankton
 169 would be greater than that of the non-toxic phytoplankton, suggesting that the toxic phytoplankton
 170 must have evolved resistance to the toxin and that's why it is having no extra mortality due to the
 171 toxin. This argument also leads to the fact that there is a strong selection pressure on the non-toxic
 172 phytoplankton to evolve resistance to the toxin after coexisting with the toxic phytoplankton for
 173 some period (Hairston et al. 2001). However, the effect of evolution on biodiversity falls outside the
 174 scope of this paper.

175 **3 Results**

176 **3.1 Allelopathy and the coexistence of many species on single resource**

177 We use the modified Droop model with allelopathy to explore the possibility of stable equilibrium
 178 coexistence of many species on limited number of resources. For this reason, a series of simulations
 179 was carried out considering randomly chosen half-saturation constants K_i for different species

180 N_i ($i = 1, \dots, 8$) and random allelopathic effects θ_i ($i = 2, \dots, 8$) of N_1 on the other
 181 species N_i ($i = 2, \dots, 8$). Apart from the values of θ_i , all the other parameter values and their
 182 ranges of variations are taken from the paper by Klausmeier et al. (2004). The set of all the
 183 parameter values corresponding to each of the figures presented in this paper is given in Online
 184 Supplementary Material I.

185 For the model given in Eqs. (1) altogether, 100 simulations are performed, each starting with
 186 a single resource and 8 species with initial species density 0.01. We carry our numerical simulations
 187 for 20 consecutive years to exclude the possibility of transient steady states (Göthlich and Oschiles
 188 2012). In Fig. 1(a), we plot the number of surviving species (along the y -axis) for each of the 100
 189 different simulations (along the x -axis). Clearly, each simulation corresponds to different parametric
 190 setup thus representing different species combinations. The positions of the circles represent the
 191 number of surviving species for different simulation runs. In the case of the survival of a single non-
 192 toxic species (depicted by the plus signs in Fig. 1(a)), the toxic species extinct from the system and
 193 thus leaving the possibility of the coexistence of more species on a single resource in equilibrium.
 194 The red circles represent the situation when the toxic species survives in the community and hence
 195 can promote the possibility of the stable equilibrium coexistence of many species only on one
 196 resource (see Online Supplementary Material II for the mathematical derivation which shows the
 197 coexistence of two species on a single resource and can be extended for more species).
 198 Interestingly, in most of the cases (almost 70 – 80% of our simulations with stochastically chosen
 199 parameters), the number of surviving species exceeds the number of resources (remember only one
 200 resource in our case) in the presence of the allelopathic effect by toxic phytoplankton (see for
 201 example Fig. 1(b)). One should note that the existence of the toxic species in an initial community
 202 does not always guarantee the stable equilibrium coexistence of many species on less number of
 203 resources. The principle of competitive exclusion holds for some of the cases even in the presence
 204 of toxic species. Approximately, in 20 – 25% of our simulations, toxic species is the winner (by

205 outcompeting all other non-toxic competitors) when many species are competing for a common
206 resource (see Fig. 1(b)).

207 Considering a particular species composition (i.e. only one specific set of parameter values)
208 where eight species coexist in one resource, we check the stability of the existing species. Whether
209 the existing species reach an equilibrium state or not, that can be identified by the trajectories using
210 simulation or by the stability analysis. Here we adopt the first one. In this model, the stable
211 coexistence occurs for both toxic and non-toxic species which is shown in Fig. 2. Since eight
212 species are competing for a single resource, the allelopathic interactions between the toxic and the
213 non-toxic species help in the coexistence of many non-toxic species with equilibrium density
214 (Chesson 2000). The allelopathic interactions shape the community structure and also decide the
215 species richness and abundances of the populations. Here the toxic one always dominates in the
216 chemostat. There are evidences that the toxic species exists with high biomass in ecosystems due to
217 the defense mechanism against predation and the availability of rich nutrition (Irigoiien et al. 2005).
218 Toxic bloom occurs due to the growing environmental conditions and the relationship between the
219 cell size and nutrient helps the toxic species to dominate in the ecosystem. Subsequently, these
220 blooming species strongly influence the coexistence of many non-toxic species and further shape
221 the ecosystem processes. In Monchevaa et al. (2001), the blooming species is considered as
222 *keystone species* or *ecological engineers* that helps to maintain local biodiversity within a
223 community. In our model, the toxic species density settles in higher equilibrium density (approx 5.4
224 $\times 10^6$ cells/ml in Fig. 2) in comparison to the non-toxic species equilibrium densities. In addition,
225 the non-toxic species density initially increases, but once the toxic species reaches high density,
226 allelopathic effect comes into play which suppresses the non-toxic species abundances and finally
227 all the non-toxic species settle down in equilibrium with low population densities. Therefore, we
228 also find a similar kind of dominance and high abundance of the toxic species in the presence of
229 allelopathic effect on other species. Moreover, the biodiversity is maintained for a longer time scale

230 by most of the species, even in an equilibrium state.

231 To study how allelopathy influences the coexisting steady state with changing environments,
232 first we consider a two species-one resource system where one of the species is toxic and we
233 perform an analysis in a two-dimensional parameter space spanned by the two experimentally
234 accessible parameters, the dilution rate D and the concentration of the resource S in the inflow
235 medium. In literature, the coexistence of more species in less number of resources is termed as
236 supersaturated coexistence (Revilla and Weissing 2008). The regions of supersaturation in the S – D
237 parameter space for three different values of the allelopathy parameter θ ((a) $\theta = 8 \times 10^{-5}$, (b) $4 \times$
238 10^{-4} , and (c) 5×10^{-3}) are shown in Fig. 3 (marked by gray shaded regions). The other parameter
239 values are same as in Fig. 1, whereas the values of q , v and K 's are given in Table 3 of Online
240 Supplementary Material I-. We consider that a species exists if its concentration is greater than 10^4
241 cells/ml (Prince et al. 2008). Here, the break-even resource concentrations for each species above
242 which the toxic and the non-toxic species survive in isolation, are not indicated in Fig. 3 as they are
243 very small. ~~Therefore, in the white region, either of the species survives depending on the initial~~
244 ~~conditions.~~ In Fig. 3, in the white regions, either of the species survives depending upon the initial
245 conditions whereas in the blue shaded regions both the species are extinct from the system due to
246 high dilution rate. Interestingly, the presence of high allelopathy helps in the coexistence at low
247 resource supply concentrations. However, an increase in the resource input concentration increases
248 the toxic species abundance, which suppresses the non-toxic phytoplankton abundance. As a result,
249 at a very high resource input concentration, the non-toxic species extinct from the system (not
250 shown in Fig. 3). Therefore, very high allelopathy again disturbs the coexistence of toxic and non-
251 toxic phytoplankton. Such coexistence in the presence of allelopathy is pointing towards the fact
252 that the interactions between phytoplankton species may be linked to the evolution of a mechanism
253 for maintaining stable supersaturated biodiversity. By considering more species and one resource in
254 the presence of allelopathy, it is also possible to find a robust region of supersaturation.

255 3.2 Impact of phytoplankton diversity on biomass production

256 The relationship between biodiversity and productivity (i.e. biomass production) of species can be
 257 explained by two hypotheses, called “sampling effect” and “niche complementarity” (Loreau 1998,
 258 Loreau 2000, Loreau and Hector 2001, Fox, 2005, Fargione and Tilman 2005). The sampling effect
 259 reveals the possibility of biodiversity due to one or few dominant high biomass species or species
 260 with particular traits being present in the polyculture. On the other hand, the niche complementarity
 261 hypothesis suggests that resource partitioning interactions between species lead to an increase in
 262 total resource use. These effects can be observed by identifying the species biomass in monoculture
 263 and polyculture. Here, the coexistence of both the toxic and non-toxic species is considered as
 264 polyculture, whereas monoculture represents the presence of either only the toxic or only the non-
 265 toxic species. Using sampling hypothesis, we calculate the diversity effects such as net effect,
 266 selection effect and complementarity effect as a function of nutrient supply (S) using the technique
 267 of Loreau and Hector (2001).

268 The net effect of biodiversity is defined as $\sum Y - \bar{M}$, where Y is the observed biomass of
 269 each species in polyculture ($\sum Y = \sum_{i=1}^n Y_i$ is the biomass of all the species in polyculture) and
 270 $\bar{M} = \left(\frac{1}{n} \sum_{i=1}^n M_i\right)$ is the average monoculture biomass of all the species, where M_i is the
 271 monoculture biomass of the i -th species. The net effect of biodiversity can be partitioned into two
 272 additive components: the selection effect and the complementarity effect. It is given by:

$$\begin{aligned} \sum Y - \bar{M} &= \underbrace{\tilde{D} \times cov(M, \Delta RY)}_{\text{Selection effect}} + \underbrace{\tilde{D} \times \bar{M} \times \overline{\Delta RY}}_{\text{Complementary effect}}, \\ &= \text{Selection effect} + \text{Complementary effect}, \end{aligned}$$

273

274 where $\tilde{D}$ is the number of species in polyculture, 'cov' is the covariance operator, M represents
 275 the biomass of a species in monoculture, ΔRY is the difference between the observed relative
 276 yield Y/M and the expected relative yield $1/\tilde{D}$, and $\overline{\Delta RY}$ is the average ΔRY of all the
 277 species in the considered community. The relative yield and the performance of the species both in
 278 monoculture and polyculture depend on species growth. This species growth can be described in
 279 many ways. It can be a function of the nutrient supply, proportion of resources used by species, and
 280 the efficiency with which a species uses resources (Binkley et al. 2004). Resource use efficiency is
 281 defined as the amount of biomass produced per unit resource (Pastor 2008). Here, we find the
 282 efficiency with which a resource is used by the species in polyculture -in the presence and absence
 283 of allelopathic effect. The resource use efficiency (RUE) is calculated by the

284 sum of biomass of all the species over nutrient supply (i.e. $RUE = \frac{\sum_{i=1}^n N_i}{S}$).

285 In Fig. 4, we plot the total biomass, total resource, resource use efficiency, net effect,
 286 selection effect and complementary effect as a function of nutrient supply concentration for two
 287 species-one resource system where one of the species is toxic. Here we choose $\theta = 1 \times 10^{-3}$ and
 288 the other parameter values are same as in Fig. 3. The solid (dashed) curve represents the measures
 289 in the presence (absence) of allelopathic effect. Clearly, total biomass of the species and RUE
 290 increase (see Fig. 4(a) and (c)) in the presence of allelopathic effect. Also, the net effect and the
 291 selection effect increase during the increase of nutrient supply (see Fig. 4(d) and (e)). A positive
 292 selection indicates that a species achieves greater abundance in polyculture than in monoculture. A
 293 change in average relative yield is measured by the complementarity effect. Negative
 294 complementarity effect in Fig. 4(f) indicates that there might be a lower yield on average than
 295 expected in monoculture. Since, the net effect is the addition of selection and complementarity
 296 effects, overall it is positive even though complementarity has small negative values. Without
 297 allelopathy, an increase in the nutrient supply results in decrease in the selection effect.

3.3 Chance of cascading extinction and reinvasion

Due to complex physical and chemical interactions between species in communities, the event of sudden extinction of one species can lead to a cascade of additional species loss (Lundberg et al. 2000, Fowler 2010). Here, we investigate how the diversity promoted by allelochemicals affects the pattern of cascading extinction following the removal of a non-toxic species in the supersaturated community. Similar to the situation in Fig. 1, here also we maintain the same parametric setup and construct 50 communities for each size of surviving species j ranging from 2 to 8. In total, we collect 350 communities. From each such community, a surviving species (assuming the surviving species density is more than 10^2 cells/ml) is randomly removed (except the removal of the toxic species in order to take into account the effect of allelopathy and hence to retain the supersaturation). Moreover, we set the corresponding maximum uptake rate (v) and the maximum growth rate (r) of the removed species equal to zero and the remaining system is allowed to run for another 20 years. Before removing the species randomly, all the existing species densities are maintained in an equilibrium state. We check the new community sizes and plot them (along y -axis) with respect to their initial community sizes before random removals take place (along x -axis) (see Fig. 5(a)). The open circles represent the expected number of species ($j - 1$) just after the removal of one of the surviving species and filled circles indicate the number of species after cascading extinctions, i.e. when the resulting community consists of less than $j - 1$ species. Here, the size of the circles corresponds to the number of communities for each community size. It is clear that, in general, for small j , the final community size is same as the expected community size, $j - 1$, just after the removal of one surviving species. Moreover, for higher community sizes, generally cascading extinction occurs, but just only one more species than the target species extinct for each case, although the total number of cases of cascading extinction is also very less (0.9 % approximately out of 350 communities).

322 We also investigate the cascading extinction patterns (Lundberg et al. 2000) for reinvasion
323 of the previously removed species into a new community formed after its disappearance (see Fig.
324 5(b)). For this reason, first we reach at the equilibrium after the removal of the species and then
325 reintroduce it to the system with a population density of 10^4 cells/ml. Note that, here we assume if
326 one species goes extinct (density less than 10^2 cells/ml), it cannot return to the system, and we
327 assign a zero value for that extinct species. The result shows that only 0.3 % of the former member
328 of the community were unable to reinvade and interestingly no cascading extinction occurs. Also,
329 the removal and the reintroduction of species do not affect the stability of the remaining coexisting
330 species at the end of the simulation (Rohr et al. 2014). Hence, here the species loss and the
331 appearance of new invaders do not lead to community closure.

332 **3.4 Role of increased nutrient supply on species richness**

333 Now, we investigate the role of nutrient supply on the number of surviving species. In an
334 ecosystem, environmental conditions change due to seasonal succession and many other factors.
335 These environmental fluctuations or changes in the season result in the variation of nutrient level in
336 ecosystems. To take such variations of environment into account, we change the nutrient supply into
337 the system with fixed half saturation constants and fixed allelochemical effects of species. Initially
338 we set all the parameters in such a way that 8 species coexist for constant nutrient supply (i.e. $S=19$
339 $\mu\text{mol/l}$). Now, nutrient supply is increased (i.e. at a certain time, change the constant nutrient
340 supply) and the system is allowed to run for another 20 years. We find that the number of coexisting
341 species in the community (i.e. species richness) decreases with increasing nutrient supply (see Fig.
342 6). Note that, again if we reintroduce the extinct species in the chemostat by changing the nutrient
343 supply to its previous rate ($S = 19 \mu\text{mol/l}$), the species survives. Specifically, the extinct species
344 reinvades in the system during the same amount of nutrient supply. Therefore, species coexistence
345 depends on the supply of nutrient in the chemostat model. In Fig. 6, the blue and the red circles

346 represent two communities with 8 and 6 species in the initial stage, respectively, for the nutrient
 347 supply $S = 19 \mu\text{mol/l}$ for the fixed K_i 's and fixed θ_j 's. The parameter values used are given in
 348 Table 4 of Online Supplementary Material I-. We further increase the nutrient supply in the
 349 chemostat –and find the nutrient supply where the number of coexisting species changes. We
 350 marked those nutrient supply rates where exactly one species becomes extinct from the previous
 351 community size. Clearly, the number of coexisting species in the community (i.e. species richness)
 352 decreases with increasing nutrient supply (see Fig. 6). Therefore, we see biodiversity loss due to the
 353 increase in the supply of nutrient in the chemostat.

354 As an explanation we can say that, due to the allelopathic effect, the abundance of all non-
 355 toxic phytoplankton decreases which results in decrease in competition between species and helps
 356 in species coexistence. However, an increase in nutrient supply in the system favors the growth of
 357 those species which are superior competitor for resource and results in the extinction of inferior
 358 resource competitors. Specifically, the benefit provided by allelochemicals in reducing differences
 359 in competitive abilities between different species nullifies under high nutrient input in the system.
 360 As a result, an increase in nutrient supply exhibits the loss of species diversity and hence shows the
 361 paradox of enrichment (Rosenzweig 1971). Although several other environmental factors affect the
 362 diversity, this eutrophication (changing the system from oligotrophic to eutrophic through resource
 363 enrichment) actually weakens the species diversity with an equilibrium density of the existing
 364 species (Sperfeld et al. 2010).

365 **3.5 Effect of variations in allelopathy on plankton biodiversity**

366 The competition for resources and trait variations among species are the major selective forces to
 367 maintain species diversity. The variations in the allelopathic effect influence the competitive ability
 368 of species and also the species composition. By varying the strength of allelopathic effect in the
 369 community, it largely reflects in species richness. The relationship between the interspecific

370 competition and the evolutionary dynamics of allelopathy can be explained by phenotypic plasticity
371 (Mougi 2013). Due to changing environmental conditions, species phenotype makes an impact in
372 the community (i.e. changes in the growth, morphology, behaviour, etc.) as well as in the
373 ecosystem.

374 Here, we change the allelochemical effects (θ_i) on each species proportionally, i.e. each
375 θ_i is multiplied by a unique constant, say θ_c . We identify the changes in the species diversity
376 by varying the constant θ_c . Initially we start with a community of eight species where one is
377 toxic and rest of the species are non-toxic. At first, θ_c is varied between 0 and 1 which captures
378 the scenario of the reduction in allelopathic effect on each species. As we decrease θ_c from 1 to
379 0, the number of species suddenly drops down from 8 to 1 as θ_c crosses 0.46 (cf. Fig. 7). On the
380 other hand, to take into account the effects of an increase in the strength of allelopathy, θ_c is
381 varied from 1 to 200. Clearly, the increase in θ_c gradually decreases the number of coexisting
382 species.

383 4 Discussion

384 Allelopathy is one of the important biological factors that strongly influences the structure of
385 plankton community. In this paper, we investigated the role of phytoplankton allelopathy on
386 biodiversity of plankton community. According to the principle of competitive exclusion (Hardin
387 1960, Armstrong and McGehee 1980), in homogeneous, well-mixed environments, species
388 competing for the same resource cannot coexist, and the final equilibrium consists of a single
389 species. Indeed, simple competition models and laboratory experiments also suggest that the
390 number of coexisting species in equilibrium cannot be greater than the number of limiting factors
391 (Tilman 1977, 1981, Sommer 1985, 1986, Rothhaupt 1988, 1996). In this paper, it is found that the

equilibrium coexistence of more species is possible on just a single resource in the presence of allelopathy. We checked the coexistence up to 8 species on single resource, although one can further increase the number of coexisting species. In real world, hundreds of species coexist on a small number of resources. The present study suggests that allelopathy can be one of the factors responsible for the coexistence of many species on a limited number of resources. The robustness of this result was shown in Fig. 1 by plotting the number of coexisting species at different species combinations. To structure plankton community, trait based approach has been used (Litchman et al. 2010) and the trade off among those traits helps to promote the functional diversity of plankton community (Edwards et al. 2013). Considering allelopathic effect as a trait, we found the diversity of species. As long as the toxic species remains present in the community, many species can coexist at equilibrium even in a single resource. So it's a kind of trade-off between the resource limitation and the toxicity with the number of coexisting species.

Another well known trade-off associated with allelopathy is the trade-off between benefits in competition or nutrient availability and the metabolic costs of toxin production (Lewis 1986). Allelopathy reduces competitor abundance either by resulting direct mortality to the competing cells or by reducing competitor population growth rate, which ultimately provides a relief mechanism to the toxin producing species from resource competition. Moreover, the toxins produced by some species, make holes in the cell membrane of other species, resulting in a transient nutrient leakage (known as dasmotrophy), which is then taken up by the toxin producing species via osmotrophic uptake. Furthermore, toxins produced by some mixotrophic species, helps in prey capture by paralyzing the prey species. However, according to the evolutionary theory, no benefit comes without a cost. Since, the production of toxin needs allocation of acquired nutrient as well as the maintenance of the synthetic machinery for toxin production, it results in a lower competitive ability or the lower growth rate of the toxin producing species. Moreover, auto-toxicity bears the risk of damaging the toxin producing cells from its own released toxic compounds and thus may

involve some further costs to acquire immunity. In addition to this, the release of toxins in the surrounding water increases the risk of predation as most planktonic grazers use chemical cues to locate their prey. Some recent modeling studies with toxin production by phytoplankton incorporated such trade-offs (Grover and Wang 2014, Chakraborty and Feudel 2014). In the present paper, although the cost-benefit analysis is not taken into account explicitly, but a trade-off is assumed between the growth advantage due to allelopathy and the competitive ability of the toxin producing species. However, the toxin production as well as the trade-offs associated with allelopathy varies with species and several other factors (Brönmark and Hansson 2012). Thus, the robustness of our result were examined by plotting the number of coexisting species at different species combinations (see Fig. 1). Because of this trade-off, the R^* value of the non-toxic species becomes lower compared to the R^* value of the toxic species. Here, R^* value of a species is the break-even resource concentration where the mortality rate equals its reproduction rate and resource concentration below this value forces the species to get extinct. During the competition for a single resource, at equilibrium, theory predicts that the single species with lowest R^* value (lowest requirement for resources) would displace all other species and win the competition (Tilman 1977, 1982). According to our formulation, the non-toxic species would win the competition in the absence of allelopathy because of its lower R^* value. However, the presence of allelopathy suppresses the growth of the non-toxic species, which increases equilibrium resource concentration and sets it to the R^* value of the toxic species. This opens the possibility of the stable coexistence of both the non-toxic and the toxic species on single resource. In the presence of sufficient allelopathic effect, more species can coexist on single resource. In other words, the presence of allelopathy manages to keep the resource equilibrium level high, thus enables the invasion of more species on less number of resources.

The role of competition on phytoplankton biodiversity was previously investigated by many researchers. With the help of a resource competition model (based on a standard model for

442 phytoplankton competition), Huisman and Weissing (1999) showed that the number of coexisting
443 species can exceed the number of resources in a constant and homogeneous environment via non-
444 equilibrium conditions, only if the number of resources is either three or more. A modified version
445 of the model of Huisman and Weissing (1999) by incorporating cell quota also gives similar kind of
446 results (Revilla and Weissing 2008). In the current paper, we examined the diversity of plankton
447 ecosystems considering a similar model like Revilla and Weissing (2008), but with a single resource
448 in the presence of allelopathy. On one hand, the presence of allelopathic interaction relaxed the
449 restriction of the number of minimum resources needed for the coexistence of more species than the
450 number of resources. On the other hand, allelopathy allows more species to coexist in stable
451 equilibrium. However, such stable coexistence always needs the presence of toxic phytoplankton at
452 high density. A high abundance of the toxic species reduces the competitive interaction between
453 different species of phytoplankton by reducing other species abundances and helps to maintain
454 phytoplankton biodiversity. All the other species remain in the system in a stable steady state with
455 low abundances. In natural ecosystems, there are evidences of the dominance of toxic species in a
456 region for a long time period. For example, toxic *Aureoumbra lagunensis* (Buskey and Hyatt 1995)
457 formed a massive long-lasting bloom event in the Laguna Madre, Texas (USA) from the time period
458 1990 to 1997 (Buskey and Stockwell 1993, Buskey et al. 2001). According to our study, we expect
459 that all the other species of phytoplankton coexist with the toxic species at low abundances during
460 that time period and maintain relatively stable abundances.

461 Although, frequent in natural systems, the amount of empirical data showing the coexistence
462 of toxic and non-toxic species is limited. Only a few experiments from various communities are
463 documented showing the coexistence in the presence of toxic effect. The coexistence in the
464 experiments with killer toxin producing yeasts and toxin sensitive yeasts can occur because of the
465 reduction in competition due to the spatial separation in microhabitats or temporal separation in
466 different stages of successions (Abranches et al. 1997, Starmer et al. 1987). Although, our result

also shows a similar kind of reduction in competition between toxic and non-toxic phytoplankton, such reduction occurs due to the suppression of the abundance of the other non-toxic competitors in response to allelopathic interaction. As a result, the toxic phytoplankton dominates and coexists with other non-toxic species (with low abundance) in the system. Similar kind of dominance of the toxin producing species over the non-toxic competitors and their stable coexistence has been observed in an experiment with different bacterial strains. Ruiz-Barba et al. (1994) evaluated Bacteriocin-producing *Lactobacillus plantarum* LPCO10 and its non-bacteriocin-producing, bacteriocin-immune derivative, *L. plantarum* 55-1, to investigate their growth and persistence in natural Spanish-style green olive fermentation. During the experiment, they found the stable coexistence of both the strains and the dominance of bacteriocin producing strain with high population levels. A spatially explicit model of a multispecies bacterial community in the presence of antibiotic interactions within the community also found the stable coexistence of huge numbers of different species even in a temporally constant and spatially homogeneous environment (Czárán et al. 2002). Such stable coexistence occurred due to the local interference competition resulting from the excretion of antibiotic compounds and the resource competitive effects caused by the associated metabolic costs. However, the basic difference with our system is that the previous study was based on a spatially explicit game theoretical model with multiple cyclic dominance structures of different strains within a species where one of the strains are resistant to toxin and the self-organized spatial pattern of the system played a crucial role in determining the dynamics of the system. In comparison, the present system considers competition between two different species of phytoplankton in the presence of allelopathy and no immunity factor is involved.

Not only stabilizing effects, allelopathy can also result in the destabilization of the coexistence steady state. Recently, Grover and Wang (2014) studied a competition model where two toxin producing phytoplankton species are competing for two essential resources with cell quota for each species and found the destabilization of the coexisting equilibria in the presence of

492 allelopathy. They explicitly considered the dynamics of toxins by considering extra equations for
493 them. In comparison, our study is mainly based on the assumption that allelopathic interactions
494 become significant only when the abundance of the toxic species becomes very high and shows
495 stabilizing effects of allelopathy. In natural ecosystems, several factors influence toxin production
496 and different species produce toxins at different rates at different environmental conditions. Thus,
497 depending on the conditions, allelopathy can have different effects, or no effects on the coexisting
498 steady state (Czárán et al. 2002, Grover and Wang 2014, Chakraborty et al. 2008).

499 Furthermore, in this study, we examined how the resource supply affects the ecosystem
500 functioning in the presence and the absence of allelopathy using biodiversity measures (Loreau and
501 Hector 2001). These biodiversity measures show a positive effect of allelopathy on plankton
502 ecosystems. We find positive measures in selection and net effects. Moreover, in the presence of
503 different species combinations in the polyculture, these biodiversity effects may suggest which
504 species dominance affects the relative yield and which one helps to maintain the species
505 composition and diversity (Loreau 1998, Loreau et al. 2009, Turnbull et al. 2012). In general,
506 separating the selection and the complementarity effects in biodiversity experiments, it is possible
507 to assemble the communities with relative performing species. Although negative effects do not
508 contribute to ecosystem functioning, but dominant species significantly identify the species
509 performance with lack of efficiency in resource usage. These effects provide a clear evidence for the
510 importance of toxic species in aquatic ecosystems. The strong relationship between diversity and
511 stability of interacting species makes a positive effect to the ecosystems (Ives and Carpenter 2007).

512 The patterns of cascading extinction are examined following the removal and invasion of
513 species. We find that the rate of cascading extinction is very less in the presence of allelopathy, even
514 for an extremely diverse system. Moreover, community restoration can also be possible if we
515 reintroduce the species in the ecosystem with appropriate environmental conditions. On the other
516 hand, the removal of a species from a community happens due to the inability of the species to

517 make use of the available resources at some particular environmental condition. Removal of species
518 can also occur in nature through selective predation by grazers or seasonal changes in the
519 environment. However, the presence of allelopathy or negative competitive effect of species
520 weakens the interspecific interaction among phytoplankton and helps to maintain the biodiversity
521 for longer time scale. Therefore, we can suggest that environmental factors those are having
522 negative effects on the competition between different phytoplankton species can also have a deep
523 impact on plankton biodiversity.

524 In this study, the nutrient enrichment explains the relationship between ecosystem
525 functioning and species diversity through the competition among phytoplankton species. The
526 strength of the ecological community is explained by considering the toxic phytoplankton as a
527 keystone species. In addition, how nutrient enrichment can influence the structure and pattern of
528 phytoplankton species diversity are illustrated using this model. Although, an increase in the
529 nutrient input enhances species abundances, the competition among species leads to species
530 extinction and results in the loss of biodiversity. The existence of multiple species in different
531 environmental conditions emphasizes the possibility of supersaturation coexistence although the
532 number of existing species varies with the degree of nutrient enrichment (Sperfeld et al. 2010).
533 Further, the invasion of species shows the diversity-stability relationship among communities (Ives
534 and Carpenter 2007).

535 Similar to the nutrient enrichment scenario, the loss of biodiversity can also occur due to the
536 increase in allelopathic interactions among species in the ecosystem. More toxicity in the
537 community leads to loss of species abundance. Therefore, from the biodiversity point of view,
538 nutrient enrichment and allelopathic interactions among different species of phytoplankton can have
539 similar kind of impact on plankton community although they affect plankton populations
540 differently. Nutrient enrichment increases competition among phytoplankton by increasing their
541 abundances (Tubay et al. 2013) whereas allelopathy decreases competition by reducing population

542 abundances of non-toxic phytoplankton (Prince et al. 2008). However, in natural plankton systems,
543 these two factors are interrelated. In general, allelopathy is not effective when the abundance of the
544 toxic phytoplankton remains low. However, under nutrient enrichment, when the abundance of toxic
545 phytoplankton become very high, the aggregated amount of toxin in the water column becomes
546 significant and as a result, allelopathy appears important. Thus, nutrient enrichment increases the
547 competition between phytoplankton species which can be neutralized by an increase in allelopathy,
548 and this can open the possibility for the coexistence of many species. As a consequence of this
549 study, we consider an analogy related to the toxicity. If we consider zooplankton as a keystone
550 predator, we can easily compare the grazing on phytoplankton by the zooplankton with the
551 allelopathic effect. Due to their grazing activity, the zooplankton suppress phytoplankton
552 abundances which helps in the biodiversity of plankton (Leibold 1996). Similarly, we argue that
553 allelopathy also acts in the same way and plays an important role in maintaining biodiversity in
554 plankton systems. However, it is not only grazing rate or allelochemical effects, overall, any kind of
555 negative effect on phytoplankton helps to maintain a stable as well as diverse ecological
556 communities. Further, these diversity-stability relationships can be maintained for a longer time
557 scale in the presence of negative effect. Moreover, variation in the environmental factors can also be
558 responsible for the plankton biodiversity (Stomp et al. 2011). We suggest that, in the low-
559 seasonality subtropical oceans, where non-equilibrium dynamics are less important, allelopathy can
560 play a major role in maintaining the plankton biodiversity.

561 To summarize, number of approaches have been used to find the effects of toxic
562 phytoplankton on a community of different phytoplankton. Each approach suggests that the toxic
563 phytoplankton as a keystone species contributes to have diverse ecological communities. However,
564 still we need to identify the mechanisms to control the toxic phytoplankton bloom with diverse
565 plankton communities. Further extension has to be identified across multiple trophic levels
566 controlling the toxic phytoplankton blooms by predation techniques and food-web patterns.

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573 **References**

- 574 Abranches, J. et al. 1997. The incidence of killer activity and extracellular proteases in tropical
575 yeast communities. – Canadian Journal of Microbiology 43(4): 328–336.
- 576 Adams, J. et al. 1979. Frequency-dependent selection for plasmid-containing cells of *Escherichia*
577 *coli*. – Genetics 91: 627–637.
- 578 An, M. et al. 2003. Mathematical modelling of allelopathy II The dynamics of allelochemicals from
579 living plants in the environment. – Ecological Modelling 161: 53–56.
- 580 Armstrong, R. A. and McGehee, R. 1980. Competitive exclusion. – The American Naturalist 115:
581 151–170.
- 582 Barton, A. D. et al. 2010. Patterns of Diversity in Marine Phytoplankton. – Science 327(5972):
583 1509– 629 1511.
- 584 Binkley, D. et al. 2004. Thinking about efficiency of resource use in forests. – Forest Ecology and
585 Management 193: 5–16.
- 586 Brömark, C. and Hansson, L. 2012. Chemical Ecology in Aquatic Systems. –Oxford University
587 Press.

- 588 Buskey, E. J. and Hyatt, C. J. 1995. Effects of the Texas (USA) brown tide algal on planktonic
589 grazers. – *Marine Ecology Progress series* 126: 285–292.
- 590 Buskey, E. J. et al. 2001. The decline and recovery of a persistent Texas brown tide algal bloom in
591 the Laguna Madre (Texas, USA). – *Estuaries* 24: 337–346.
- 592 Buskey, E. J. and Stockwell, D. A. 1993. Effects of a persistent 'brown tide' on zooplankton
593 populations in the Laguna Madre of south Texas. – In: J., S. T. and Y., S. (eds.), *Toxic*
594 *phytoplankton blooms in the sea*. Elsevier, pp. 659–666.
- 595 Cembella, A. D. 2003. Chemical ecology of eukaryotic microalgae in marine ecosystems. –
596 *Phycologia* 42(4): 420–447.
- 597 Chakraborty, S. and Feudel, U. 2014. Harmful algal blooms: combining excitability and
598 competition. – *Theoretical Ecology* 7: 221–237.
- 599 Chakraborty, S. et al. 2008. Nutrient-limited toxin production and the dynamics of two
600 phytoplankton in culture media: A mathematical model. – *Ecological Modelling* 213: 191–
601 201.
- 602 Chao, L. and Levin, B. R. 1981. Structured habitats and the evolution of anticompetitor toxins in
603 bacteria. – *Proceedings of the National Academy of Sciences of the USA* 78(10): 6324–
604 6328.
- 605 Chattopadhyay, J. 1996. Effect of toxic substances on a two-species competitive system. –
606 *Ecological Modelling* 84: 287–291.
- 607 Chesson, P. 2000. Mechanisms of maintenance of species diversity. – *Annual Review of Ecology,*
608 *Evolution, and Systematics* 31: 343–366.
- 609 Clark, J. S. et al. 2007. Resolving the biodiversity paradox. – *Ecology Letters* 10: 647–662.
- 610 Czárán, T. L. et al. 2002. Chemical warfare between microbes promotes biodiversity. – *Proceedings*
611 *of the National Academy of Sciences of the USA* 99: 786–790.
- 612 Droop, M. R. 1973. Some thoughts on nutrient limitation in algae. – *Journal of Phycology* 9: 264–

613 272.

614 Dubey, B. and Hussain, J. 2000. A model for the allelopathic effect on two competing species. –
615 Ecological Modelling 129: 195–207.

616 Dutta, P. S. et al. 2014. Multiple resource limitation: nonequilibrium coexistence of species in a
617 competition model using a synthesizing unit. – Theoretical Ecology 7: 407–421.

618 Edwards, K. F. et al. 2013. A three-way trade-off maintains functional diversity under variable
619 resource supply. – The American Naturalist 182: 786–800.

620 Fargione, J. E. and Tilman, D. 2005. Diversity decreases invasion via both sampling and
621 complementarity effects. – Ecology Letters 8: 846–856.

622 Folt, C. and Goldman, C. R. 1981. Allelopathy between zooplankton: A mechanism for interference
623 competition. – Science 213: 1133–1135.

624 Fowler, M. S. 2010. Extinction cascades and the distribution of species interactions. – Oikos 119:
625 864–873.

626 Fox, J. W. 2005. Interpreting the 'selection effect' of biodiversity on ecosystem function. – Ecology
627 Letters 8: 846–856 .

628 Frank, S. A. 1994. Spatial polymorphism of bacteriocins and other allelopathic traits. –
629 Evolutionary Ecology 8: 369–386 .

630 Göthlich, L. and Oschiles, A. 2012. Phytoplankton niche generation by interspecific stoichiometric
631 variation. – Global Biogeochem. Cy. 26: 1–8.

632 Grover, J. P. 1991. Dynamics of competition among microalgae in variable environments:
633 experimental tests of alternative models. – Oikos 62: 231– 243.

634 Grover, J. P. and Wang, F. 2014. Competition and allelopathy with resource storage: Two resources.
635 – Journal of Theoretical Biology 351: 9–24.

636 Hairston, N. G. et al. 2001. Natural Selection for Grazer Resistance to Toxic Cyanobacteria:
637 Evolution of Phenotypic Plasticity?. – Evolution 55(11): 2203–2214.

- 638 Hardin, G. 1960. The Competitive Exclusion Principle. – *Science* 131(3409): 1292–1297.
- 639 Huisman, J. and Weissing, F. J. 1999. Biodiversity of plankton by species oscillation and chaos. –
640 *Nature* 402: 407–410.
- 641 Hutchinson, G. E. 1961. The paradox of the plankton. – *The American Naturalist* 95: 137–145.
- 642 Irigoien, X. et al. 2005. Phytoplankton blooms: a loophole in microzooplankton grazing impact?. –
643 *Journal of Plankton Research* 27: 313–321.
- 644 Ives, A. R. and Carpenter, S. R. 2007. Stability and diversity of ecosystem. – *Science* 317: 58–62.
- 645 Jonsson, P. R. et al. 2009. Formation of harmful algal blooms cannot be explained by allelopathic
646 interactions. – *Proceedings of the National Academy of Sciences of the USA* 106: 11177–
647 11182.
- 648 Klausmeier, C. A. et al. 2004. Phytoplankton growth and stoichiometry under multiple nutrient
649 limitation. – *Limnology and Oceanography* 49: 1463–1470.
- 650 Leibold, M. A. 1996. A Graphical model of keystone predators in food webs: Trophic regulation of
651 abundance, incidence, and diversity patterns in communities. – *The American Naturalist*
652 147: 784–812.
- 653 Lewis, W. M. 1986. Evolutionary interpretations of allelochemical interactions in phytoplankton
654 algae. – *The American Naturalist* 127: 184–194.
- 655 Litchman, E. et al. 2010. Linking traits to species diversity and community structure in
656 phytoplankton. – *Hydrobiologia* 653: 15–28.
- 657 Loreau, M. 1998. Separating sampling and other effects in biodiversity experiments. – *Oikos* 82:
658 600–602.
- 659 Loreau, M. 2000. Biodiversity and ecosystem functioning: recent theoretical advances. – *Oikos* 91:
660 3–17.
- 661 Loreau, M. and Hector, A. 2001. Partitioning selection and complementarity in biodiversity
662 experiments. – *Nature* 412: 71–76.

663 Loreau, M. and Mazancourt, C. 2013. Biodiversity and ecosystem stability: a synthesis of
 664 underlying mechanisms. – *Ecology Letters* 16: 106–115.

665 Loreau, M. et al. 2009. Biodiversity and ecosystem functioning. – Oxford University Press.

666 Lundberg, P. et al. 2000. Species loss leads to community closure. – *Ecology Letters* 3: 465–468.

667 Macias, F. A., et al. 2008. Allelopathic agents from aquatic ecosystems: potential biopesticides
 668 models. – *Phytochemistry Rev* 7: 155–178.

669 Maestrini, S. and Graneli, E. 1991. Environmental conditions and ecophysiological mechanisms
 670 which led to the 1988 *Chrysocromulina polylepis* bloom: an hypothesis. – *Oecologia*
 671 *Aquatica* 14: 397–413.

672 Maynard-Smith, J. 1974. Models in ecology. – Cambridge University Press.

673 Mills, L. S. et al. 1993. The keystone-species concept in ecology and conservation. – *Bioscience* 43:
 674 219–224.

675 Monchevaa, S. et al. 2001. Phytoplankton Blooms in Black Sea and Mediterranean Coastal
 676 Ecosystems Subjected to Anthropogenic Eutrophication: Similarities and Differences. –
 677 *Estuarine Coastal and Shelf Science* 53: 281– 295.

678 Mougi, A. 2013. Allelopathic adaptation can cause competitive coexistence. – *Theoretical Ecology*
 679 6: 165–171.

680 Nakamaru, M. and Iwasa, Y. 2000. Competition by allelopathy proceeds in travelling waves:
 681 colicin-immune strain aids colicin-sensitive strain. – *Theoretical Population Biology* 57:
 682 131–144.

683 Pastor, P. 2008. Mathematical Ecology of Populations and Ecosystems. – Wiley–Blackwells
 684 Publishing.

685 Prince, E. K. et al. 2008. Competing phytoplankton undermines allelopathy of a bloom-forming
 686 dinoflagellate. – *Proceedings of the Royal Society B: Biological Sciences* 275(1652):
 687 2733–2741.

688 Revilla, T. and Weissing, F. J. 2008. Nonequilibrium coexistence in a competition model with
689 nutrient storage. – Ecology 89: 865–877.

690 Rice, E. L. 1984. Allelopathy. – Academic Press.

691 Rohr, R. P. et al. 2014. On the structural stability of mutualistic systems. – Science 345: 1253497(1–
692 9).

693 Rosenzweig, M. L. 1971. Paradox of enrichment: destabilization of exploit ative ecosystem in
694 ecological time. – Science 171: 385–387.

695 Rothhaupt, K. O. 1988. Mechanistic resource competition theory applied to laboratory experiments
696 with zooplankton. – Nature 333: 660–662.

697 Rothhaupt, K. O. 1996. Laboratory experiments with a mixotrophic and phototrophic competitors. –
698 Ecology 77: 716–724.

699 Ruiz-Barba, J. L. et al. 1994. Use of *Lactobacillus plantarum* LPCO10, a Bacteriocin Producer, as a
700 Starter Culture in Spanish-Style Green Olive Fermentations. – Applied and Environmental
701 Microbiology 60(6): 2059–2064.

702 Scheffer, M. et al. 2003. Why phytoplankton communities have no equilibrium: solutions to the
703 paradox. – Hydrobiologia 491: 9–18.

704 Schippers, P. 2001. Does "supersaturated coexistence" resolve the "paradox of the plankton"? –
705 Ecology Letters 4(5): 404–407.

706 Schmidt, L. and Hansen, P. 2001. Allelopathy in the prymnesiophyte *Chrysocromulina polylepis*:
707 effect of cell concentration, growth phase and pH. – Marine Ecology Progress Series 216:
708 67–81.

709 Shores, N. et al. 2008. Evolution exacerbates the paradox of the plankton. – Proceedings of the
710 National Academy of Sciences of the USA 105(34): 12365–12369.

711 Solé, J. et al. 2005. Modelling allelopathy among marine algae. – Ecological Modelling 183: 373–
712 384.

713 Sommer, U. 1985. Comparison between steady state and nonsteady state competition: experiments
 714 with natural phytoplankton. – *Limnology and Oceanography* 30: 335–346.

715 Sommer, U. 1986. Nitrate- and silicate-competition among Antarctic phytoplankton. – *Marine*
 716 *Biology* 91: 345–351.

717 Sperfeld, E. et al. 2010. Productivity, herbivory, and species traits rather than diversity influence
 718 invasibility of experimental phytoplankton communities. – *Oecologia* 163: 997–1010.

719 Starmer, W. T. et al. 1987. The ecological role of killer yeasts in natural communities of yeasts. –
 720 *Canadian Journal of Microbiology* 33: 783–796.

721 Stomp, M. et al. 2011. Large-scale biodiversity patterns in freshwater phytoplankton. – *Ecology*
 722 92(11): 2096–2107.

723 Tilman, D. 1977. Resource competition between planktonic algae: an experimental and theoretical
 724 approach. – *Ecology* 58: 338–348.

725 Tilman, D. 1981. Test of resource competition theory using four species of Lake Michigan algae. –
 726 *Ecology* 62: 802–815.

727 Tilman, D. 1982. *Resource Competition and Community Structure*. – Princeton University Press.

728 Tubay, J. M. et al. 2013. The paradox of enrichment in phytoplankton by induced competitive
 729 interactions. – *Scientific Reports* 3:2835: 1–8.

730 Turnbull, L. A. et al. 2012. Coexistence, niches and biodiversity effects on ecosystem functioning. –
 731 *Ecology Letters* 16-s1: 116–127.

732 Van de Waal, D. B. et al. 2014. Stoichiometric regulation of phytoplankton toxins. – *Ecology*
 733 *Letters* 17: 736–742.

734 Wilson, J. B. 2011. The twelve theories of co-existence in plant communities: the doubtful, the
 735 important and the unexplored. – *Journal of Vegetation Science* 22: 184–195.

736 **Figure captions**

737 **Figure 1:** (a) Number of surviving species in 100 different communities after 20 years of
738 simulations. For each simulation, half-saturation constants, K_i , for the different species N_i ($i =$
739 $1, \dots, 8$) and allelopathic effects, θ_i , of N_1 on other species N_i ($i = 2, \dots, 8$) are chosen as
740 random and the other parameter values are given in Online Supplementary Material I in Table 1.
741 This figure depicts that the number of coexisting species varies from 1 to 8 only depending on
742 single resource. (b) Histogram showing the frequency of occurrence of different communities with
743 equal number of coexisting species out of the 100 different communities.

744 **Figure 2:** Trajectories of the toxic and the non-toxic species. All the 8 species are coexisting
745 together in a stable equilibrium state only depending on one resource. Parameter values are given in
746 Online Supplementary Material I in Table 2 .

747 **Figure 3:** The region of supersaturations (shaded in gray) at (a) $\theta = 0.00008$, (b) $\theta = 0.0004$, and (c)
748 $\theta = 0.005$ when one toxic and one non-toxic species are competing for a single resource. ~~In the~~
749 ~~white region, either of the species exists depending on the initial conditions.~~ In the white regions,
750 either of the species exists and in the blue shaded regions both the species are extinct. Clearly, the
751 coexistence of both the species starts to occur at comparatively lower values of the resource supply
752 under high toxic effects. Parameter values are given in Table 3 in Online Supplementary Material I.

753 **Figure 4:** Plot of (a) total biomass of the species, (b) resource, (c) resource use efficiency, (d) net
754 effect, (e) selection effect and (f) complementary effect as a function of the nutrient supply
755 concentration (S) for two species-one resource system where one of the species is toxic. The solid
756 (dashed) curve represents the measurements in the presence (absence) of allelopathic effect. In this
757 figure, we considered $D = 0.5$ and $\theta = 10^{-3}$ and the other parameter values are same as in Fig. 3.

758 **Figure 5:** (a) Pattern of cascading extinction following the removal of a randomly chosen species
759 (50 communities each of all sizes of surviving species). The open circles represent the expected
760 number of species ($j - 1$) just after the removal of one of the surviving species and filled circles
761 indicate cascading extinctions. The size of the circles corresponds to the number of communities for
762 each community size. For higher community sizes cascading extinction occurs but only one more
763 species than just the target species extinct. (b) Reinvasion of the removed species with density
764 10^4 cells/ml into the new community formed by its disappearance. The open circles represent the
765 expected number of species just after the reinvasion and filled circles indicate cascading extinctions.
766 The size of the circles corresponds to the number of communities for each community size.

767 **Figure 6:** Blue and red circles represent two communities with 8 and 6 species in the initial stage
768 for the nutrient supply $S = 19$, respectively. Increase in the nutrient supply (along x -axis) to find
769 where one coexisting species (along y -axis) extincts. Each point represents the nutrient supply rate
770 where the number of species decreases by one from the larger number. Parameter values are given
771 in Online Supplementary Material I in Table 4.

772 **Figure 7:** Diagram shows the proportion of allelochemical effect in the chemostat model (along x -
773 axis) with the number of coexisting species (along y -axis). Parameter values are given in Online
774 Supplementary Material I in Table 5.

Figure 1

Figure 2

Figure 3

Figure 4

780 (a)

(b)

781

Figure 5

782

Figure 6

Figure 7