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Jain-Schlaepfer, Sofia; Midwood, Jonathan D.; Larsen, Martin Hage; Aarestrup, Kim; King, Greg; Suski, Cory; Cooke, Steven J.

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Relationship of baseline and maximum glucocorticoid concentrations to migration propensity –
a field test with wild sub-adult brown trout (*Salmo trutta*)

S.M.R. Jain-Schlaepfer^{*a}, J.D. Midwood^b, M.H. Larsen^{cd}, K. Aarestrup^c, G.D. King^e, C.D. Suski^e and
S.J. Cooke^a

^aEcology and Conservation Physiology Laboratory, Department of Biology and Institute of
Environmental Science, Carleton University, 1125 Colonel By Dr. Ottawa, ON K1S 5B6, Canada

^b Great Lakes Laboratory for Fisheries and Aquatic Science. Fisheries and Oceans Canada,
Burlington, ON, L7S1A1, Canada

^c National Institute of Aquatic Resources, Freshwater Fisheries, Technical University of
Denmark, Vejlsøvej 39, 8600 Silkeborg, Denmark

^d Danish Centre for Wild Salmon, Brusgårdsvej 15, DK-8960 Randers SØ, Denmark.

^eDepartment of Natural Resources and Environmental Sciences, University of Illinois, 1102 S.
Goodwin Ave, Urbana, IL 61801, USA.

*sofiaj@rogers.com, 0467694922, 32 McKinley Street, North Ward 4810, Australia

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21 S.M.R. Jain-Schlaepfer, J.D. Midwood, M.H. Larsen, K. Aarestrup, G.D. King, C.D. Suski and S.J.
22 Cooke

23 **Abstract**

24 There is considerable variation in individuals' glucocorticoid (GC) baseline status and stress
25 responses, yet the cause and consequence of this variation remains ambiguous. Attempts to
26 relate GC levels to fitness and life-history tradeoffs have yielded variable results. In this study,
27 we evaluated whether baseline and post-stressor GC hormone concentrations predicted
28 migration strategy (i.e., resident or migrant) and successful seaward migration in a partially
29 migrating population of juvenile brown trout (*Salmo trutta* (Linnaeus, 1758)). Baseline ($N=99$)
30 or post-stressor ($N=102$) plasma cortisol concentrations were obtained from trout and they
31 were tagged with passive integrated transponder (PIT) and released in a natural Danish stream.
32 Subsequently, fish were tracked with PIT reader systems and the stream was resampled for
33 resident individuals. GC levels were not found to be associated with recapture of resident
34 individuals or migration propensity to our first tracking station (S1), but increased baseline (and
35 not post-stressor) GC levels were associated with increased passage from S1 to our second
36 tracking station, which anecdotally was an area of high predation or challenge. Our study found
37 no evidence to suggest that cortisol regulates the migration life-history in juvenile brown trout,
38 but intermediate increases in baseline GC (and not post-stressor GC) levels may favor migration
39 performance.

40

41 **Key Words**

42 Cortisol, Fish, Glucocorticoid, Migration, Stress response, Brown trout, *Salmo trutta*

43

44 **Abbreviations**

45 GC, glucocorticoid; PIT, passive integrated transponder; S1, tracking station 1; S2, tracking

46 station 2; HPA, hypothalamic–pituitary–adrenal axis; HPI, hypothalamic–pituitary–interrenal

47 axis; K, Fulton’s condition factor.

48 **Introduction**

49 The study of the physiological basis of stress is useful for understanding physical, chemical,
50 social and perceived challenges faced by organisms, and the functional responses employed by
51 organisms to deal with those stressors (Denver 2009). In vertebrates, real or perceived
52 stressors stimulate the hypothalamic–pituitary–adrenal axis (HPA; or the hypothalamic–
53 pituitary–interrenal (HPI) axis in fish) resulting in the release of glucocorticoid (GC) hormones
54 into the blood stream (Sapolsky et al. 2000; Barton 2002). Thus, circulating concentrations of
55 GC hormones are often measured to understand an organism’s stress response (Dantzer et al.
56 2014). Within a species, individuals often show repeatable baseline (prior to a stressor) GC
57 levels and a repeatable GC response following a stressor (Pottinger and Carrick 2001;
58 Ouedraogo et al. 2004; Wada et al. 2008; Rensel and Schoech 2011; Cook et al. 2012; Narayan
59 et al. 2013; Love et al. 2015) resulting in different reactive scope profiles (difference between
60 GC response and baseline) between individuals (Romero et al. 2009). The biological
61 functionality, as well as the individual and evolutionary consequences of such variability among
62 individua, are still debated (Breuner et al. 2008; Bonier et al. 2009).

63
64 To define the biological consequences of individual variation in GC levels during the stress
65 response, researchers have attempted to relate inter-individual differences in baseline and
66 post-stressor GC concentrations to performance (e.g., migration, parental care, territorial
67 behavior) and fitness (e.g. Comendant et al. 2003; Cabezas et al. 2007; Breuner et al. 2008).
68 From a theoretical point of view, it is often hypothesized that individuals with greater post-
69 stressor GC levels should have increased long term fitness (Breuner et al. 2008), because an

individual's stress response increases chances of surviving the stressor (e.g. predation attempt) and mediates trade-offs between reproduction and survival. Indeed, experimental studies show variability in the relationship between the magnitude of the stress response and performance or fitness measures (reviewed by Breuner et al. 2008). This relationship is increasingly understood as context dependent, because an elevated stress response in an emergency situation shifts resources from reproduction to survival (Wingfield et al. 1998); however, an elevated stress response when the perception of stressors is exaggerated can lead to the misallocation of resources, thereby limiting investment in growth and reproduction (Hawlena and Schmitz 2010). There is a paucity of studies directly examining inter-individual differences in stress response while concurrently measuring performance or fitness (Romero and Wikelski 2001; Blas et al. 2007; Cook et al. 2011).

In contrast to the theorized positive association between elevated post-stressor GC levels and performance or fitness, higher baseline GC levels are often inferred to indicate reduced condition, because it suggests an organism is experiencing greater environmental stressors (Cort-Fitness hypothesis; reviewed by Bonier et al. 2009). Higher GC levels are in circulation when exposed to a stressor and are associated with the reallocation of resources from fitness-related factors, such as growth and reproduction, towards the restoration of physiological homeostasis. Despite the potential negative consequences associated with higher baseline GC levels, some studies have found that moderate long term increases in GCs can actually increase individual performance (Comendant et al. 2003; Pravosudov 2003; Cabezas et al. 2007). Further

91 investigation is therefore warranted to understand the relationship between baseline GC
92 hormones and performance.
93

94 Both baseline and post-stressor GC concentrations have a heritable component in some species
95 (Evans et al. 2006; Jenkins et al. 2014) and are associated with a suite of other traits that can
96 lead to different life history and coping strategies (Koolhaas et al. 1999; Pottinger and Carrick
97 1999; Blas et al. 2007; Øverli et al. 2007; Wada et al. 2008). Selection on heritable variation in
98 GC levels and response is thought to drive the evolution of life-history traits (Bolnick et al. 2003;
99 Williams 2008), such as migration behavior. Migration in particular, requires large amounts of
100 energy and often exposes individuals to many stressors including predation and changes in
101 environmental condition (Dingle 2014). Thus, GC levels have the potential to be an important
102 factor determining successful migration (Holberton 1999; Piersma et al. 2000; Angelier et al.
103 2009), which could affect fitness and potentially drive the evolution of migration strategies.
104 Migration success, meaning successfully reaching the migration destination, has been linked to
105 GC levels in a single species of fish, adult sockeye salmon (*Oncorhynchus nerka* (Walbaum,
106 1792)), where upriver migration success was found to increase with stress responsiveness but
107 not baseline stress levels (Cook et al. 2014). GC levels have also been related to the
108 smoltification process in anadromous salmonids. Smolting fish generally have higher baseline
109 and peak GC, which appears to induce the physiological changes required to cope with the
110 transition from freshwater to saltwater (e.g., increased osmoregulatory capacity; Barton et al.
111 1985; Redding et al. 1991; Björnsson et al. 2011). Stressors that cause a GC response may cause
112 a reallocation of energy towards immediate survival resulting in a change in life history and

113 coping strategy (Landys et al. 2006; Shepard et al. 2008). Thus, GC variation resulting from past
114 or current conditions may influence the “decision” to migrate. Alternative migration strategies
115 have the potential to be heritable through heritable GC regulation (Zera and Harshman 2001;
116 Williams 2008).

117
118 For our study species, brown trout (*Salmo trutta* (Linnaeus, 1758)), migration success provides a
119 measure of performance, and also acts as an indicator of alternate life history strategies. More
120 specifically, European populations of brown trout show a continuum of migration strategies,
121 where subpopulations are capable of anadromous activity, smaller scale migrations within
122 freshwater systems (potamodromous), and residency in natal streams (Cucherousset et al.
123 2005). This phenomenon is termed partial migration and reflects the fact that only a
124 component of the population engages in migration, a common trait among some salmonid
125 species (Jonsson and Jonsson 1993). Previous studies on the propensity of migration in brown
126 trout have found that individuals with high metabolisms and high growth rates tend to migrate,
127 likely due to higher resource requirements (Jonsson and Jonsson 1993; Forseth et al. 1999).

128
129 The goal of this study is to define relationships between GC levels (baseline and post-stressor),
130 and migration propensity in juvenile brown trout. Given the role of GCs in smoltification, we
131 would expect to see higher post-stress and baseline GC levels in migrating individuals. The
132 effects of individual body condition and size (length, mass) in relation to migration propensity
133 will also be quantified.

134

Methods

Experimental Design

Brown trout were captured using backpack electroshocking (Scubla ELT 60 II GI; run at 300 volts) in four regions of the Gudsø Stream, Jutland, Denmark (Figure 1) between 11 March and 18 March 2013. This time period was selected as it fell prior to the spring migration, which typically occurs in mid-April (Midwood et al. 2014). Individuals between 120 and 182 mm in total length (TL) were used to minimize the effects of allometry on results. To evaluate an individual's resting plasma cortisol levels (baseline), blood was collected (~0.2-0.3 ml) from a subset of fish within 3 min of capture via caudal puncture of the haemal arch using a 1 ml syringe and 25-gauge needle. A separate subset of fish (Table 1) were exposed to a standardized 3 minute air exposure and then held in a 20 L container with 5 L of freshwater for 30 min prior to the collection of a post-stressor blood sample. Blood was sampled in the same manner as above at 30 ± 3 minutes post-air exposure corresponding with the approximate peak in GCs in brown trout (Pickering and Pottinger 1989). Blood samples were held in a water-ice slurry for no more than 2 hrs until centrifuged for 5 min (10,000 rpm) to isolate plasma. Plasma isolate was decanted, flash frozen and stored (-80°C) for later analysis of plasma cortisol concentrations. Total length and wet mass of each individual were then measured. Next, a uniquely coded 23 mm PIT tag (RI-TRP-RRHP, half duplex, 134 kHz, diameter 3.85 mm and a mass of 0.6 g in air; Texas Instruments, Plano, Texas, USA) was inserted into the body cavity through a small incision made with a scalpel. Previous tag retention studies on salmonids have found negligible effects on survival and growth (Larsen et al. 2013) and generally low PIT tag rejection rates ($< 4\%$ after seven months; Ombredane et al. 1998). Following tagging fish were

then placed in a 60 L container of fresh river water and monitored for a minimum of 30 minutes to ensure recovery before they were ultimately released back into the stream near their point of capture. All fish recovered. Length and mass were used to calculate Fulton's condition factor (K) for each sampled fish where $K = \text{mass (g)} \times 100 / \text{length}^3 \text{ (cm)}$ (Ricker 1975).

Cortisol Analysis

Plasma cortisol (ng mL^{-1}) was quantified using a commercially available enzyme linked immunosorbant assay (ELISA) (Enzo Life Sciences, Cortisol EIA Kit [901-071], BioAssay Systems) previously validated for use in fishes (Sink et al. 2008). This assay has a detection limit of $0.0567 \text{ ng mL}^{-1}$, therefore, the 5 samples with undetectable cortisol concentrations were assigned values equal to this detection limit (Haddy and Pankhurst 1999; Liss et al. 2013).

Tracking

To track movements of individuals from their point of release to the sea, two tracking stations were placed approximately 1 km upstream of Kolding Fjord on Gudsø Stream, which was the closest feasible place to the ocean. The first station (S1) was placed upstream of a millpond and the second station (S2) was placed downstream of the millpond, approximately 150 m from S1. Each station was comprised of a paired set of PIT gates spaced approximately 10-m apart. This allowed for an assessment of the direction of movement for individuals at each station. The same array was used by Midwood et al. (2015) and found to have a detection efficiency at S1 of 96.3%; detection efficiency could not be evaluated at S2, but missed-detections were assumed

to affect all trout equally and therefore should not bias conclusions related to baseline GC or post-stressor GC correlations with relative migration success to S1 and S2.

To estimate the proportion of individuals that stayed resident and did not migrate to the ocean, the stream was re-sampled using back-pack electrofishing on 18 and 19 June, 2013, at which point migratory individuals should have left the stream. Surveys started near the antenna and proceeded upstream through all areas where brown trout were initially captured (Figure 2). It was not possible to resample the entire system as it is greater than 16 km in length; however, given the shallow and narrow nature of the stream and the relatively high efficiency of capture for salmonid species using electrofishing (Kennedy and Strange 1981; Haynes and Baird 1994), efforts were likely sufficient to provide a relative estimate of residency.

Statistical Analyses

Multiple linear regression was used to quantify relationships between plasma cortisol concentrations (for both post-stressor and baseline groups) and timing of migration (time from sampling to S1 passage), while controlling for K , length or mass. Assumption of homogeneity of variance and normal distribution for the models were ensured by visual inspection of residual plots. If violated, the response variable was log or square root transformed. Multiple binomial logistic regression was used to test for the effect of plasma cortisol concentration (baseline and post-stressor) on the probability of successful migration to S1 (used as a proxy of migration strategy) and on the probability of successful migration to S2 given successful passage of S1 (used as a measure of migration success through the millpond for migrating individuals). Again,

separate tests were conducted using mass, length or K as a covariate in the model. Length, mass and K were also tested separately for effects on all migration response variables with the baseline and post-stressor fish subsets pooled. Two outliers were removed before analyses as they were outside the standard deviation of the mean: a baseline cortisol value of 44.5ng/mL and a peak cortisol value of 3.43ng/mL. All significant results found without the outliers were also found when the outliers were included in the analyses.

Variation in association with recorded mean values is given as standard error ($\pm$ SE) throughout. Statistical significance for all analyses was set at $\alpha = 0.05$ and all statistical analyses were conducted in R (v 3.2.2; R Core Team, 2016).

Results

Of the 198 fish tagged, 19 were recaptured in mid-June, 83 were detected passing S1 and 45 passing S2 (Table 2). All fish detected at S2 were also detected at S1, suggesting a detection efficiency of 100% at S1. Mean baseline cortisol was 5.6 ± 6.7 ng/mL and post-stressor cortisol was 69.2 ± 40.1 ng/mL (Table 1). Post-stressor (Multiple logistic regression: $Z_{2,96} = 1.44$, $p = 0.15$) and baseline (Multiple logistic regression: $Z_{2,96} = 0.15$, $p = 0.89$) plasma cortisol concentrations did not have a significant effect on the probability of successful passage at S1 when controlling for K , mass, or length ($p > 0.1$; Figure 2b and 2e). For fish that passed S1, the probability of successful passage to S2 increased with increasing individual baseline cortisol when controlling for mass (Multiple logistic regression: $Z_{2,27} = 1.96$, $p = 0.05$) and length (Multiple logistic regression: $Z_{2,27} = 2.01$, $p = 0.045$), but not for K ($p > 0.05$). For fish that passed

S1, the probability of successful passage to S2 showed no relationship with post stressor cortisol (All: $p>0.1$)

Mean migration time from blood sampling to S1 passage was 33.4 ± 19.1 days and mean time from S1 to S2 was 3.2 ± 7.6 days. Multiple regression showed no significant relationship between migration duration or date of passage, and post-stressor or baseline cortisol when controlling for mass, length or K (All: $p>0.1$). Migration time decreased with increasing fish length (Linear regression: $F_{1,79} = 6.82$, $p = 0.01$, $R^2 = 0.8$) and fish mass (Linear regression: $F_{1,79} = 4.4$, $p = 0.04$, $R^2 = 0.05$), but no relationship was found with K (Linear regression: $F_{1,79} = 3.14$, $p = 0.08$, $R^2 = 0.04$).

Multiple logistic regression revealed no effect of post-stressor and baseline cortisol on the probability of recapture in resident streams when controlling for K , length or mass (Figure 2; All: $p>0.1$). Length, mass and K also were found to have no effect on the probability of recapture in resident streams (All: Logistic regression: $p>0.1$). No relationship was found between K , mass or length and baseline or post-stressor cortisol (All: Linear regression: $p>0.1$). The probability of S1 passage and the probability of passage between S1 and S2 had no relationship with K , fish length and fish mass (All: $p>0.08$).

Discussion

As previous studies have found cortisol to be elevated during smoltification (Barton et al. 1985), we expected individual brown trout with higher baseline and post-stressor cortisol to have a

higher probability of migrating downstream to S1 and S2, coupled with a lower probability of recapture as resident. However, no such relationships were found.

We found no evidence suggesting that migration is correlated with endocrine regulation in juvenile brown trout, despite several studies having documented that cortisol levels rise during smoltification in anadromous fish (Barton et al. 1985; Redding et al. 1991; Björnsson et al. 2011), and also that GC regulation has the potential to play a role in determining alternative coping strategies (Zera and Harshman 2001; Landys et al. 2006; Williams 2008). Mass, length and body condition were also not found to have a relationship with plasma cortisol (baseline or post-stressor). Fish of greater mass and length (but not condition) were found to have shorter migration times similar to previous findings (Bohlin et al. 1996). Body condition, length and mass were not found to affect migration propensity to S1. the lack of correlation between our condition metrics and migration propensity is counter to previous findings of reduced condition leading to increased migration propensity (Peiman et al. 2017); however, it is possible that the comparatively limited recapture rate in the present study ($N=19$) prevented the detection of a size-based effect.

Baseline, but not post-stressor, cortisol concentration was found to be positively associated with passage through the millpond (i.e., from S1 to S2). Previous studies have found brown trout baseline cortisol without chronic stressors to be 0-5 ng/mL, while those exposed to chronic stressors such as confinement or crowding have shown baseline cortisol elevated to 10 ng/mL (Pickering and Pottinger 1989). The average baseline cortisol found in individuals

266 recaptured in resident streams, individuals that passed S1 and individuals that passed S2 was
267 5.6 ng/mL, 5.8 ng/mL and 8.4 ng/mL, respectively (Table 2). Thus, individuals that passed
268 through the millpond had baseline cortisol levels of individuals approaching chronically stressed
269 levels on average, while other fish had cortisol levels closer to what would be expected for fish
270 not showing chronic stress.

271
272 The distance between S1 and S2 is only 150 m, thus it is unlikely that S1 versus S2 passage
273 would be indicative of migration strategy. The millpond may therefore be an area of increased
274 mortality, suggesting S1 passage is more indicative of migration strategy (resident and
275 potamodromous versus anadromous) while passage between S1 and S2 is more indicative of
276 migration success of migrant individuals. Forty five percent of all brown trout that passed S1
277 were not detected passing S2, which is only 150 meters downstream, and those that did
278 migrate through the millpond, took much longer than when migrating through the stream,
279 taking an average of 3.2 days to move 150 meters. Similar patterns are found in other studies of
280 migrating fish at Gudso (Midwood et al. 2014, 2015). This indicates the millpond was
281 challenging for migrating trout. We suggest that many of these individuals likely suffered
282 mortality, as it is unlikely for such a large proportion of migrants to take up residency in such a
283 small area. Furthermore, there is anecdotal evidence of high concentration of foraging grey
284 heron (*Ardea cinerea* Linnaeus, 1758) and other birds at the outflow of millpond. Previous
285 studies on salmonid smolts have documented large losses resulting from predation in lakes
286 (Jepsen et al. 1998; Koed et al. 2006) and at weirs and related millponds (Aarestrup and Koed
287 2003). For example, Jepsen et al. (1998) found 90% of smolts died while passing through Lake

Tange, Denmark, with 56 % of the mortality cause by pike (*Esox lucius* (Linnaeus, 1758)) and 31% by birds. Another possibility for the reduced passage and long migration time through the millpond is that there is only one small exit and trout may have had a hard time getting out. While it is possible that overall lower passage at S2 is driven by reduced detection efficiency at this station, these missed-detections would affect all trout equally and thus would not affect the relationship found between baseline cortisol and passage between S1 and S2 other than by reducing statistical power. Our results, therefore, do not support the link between GC levels and migration strategy in brown trout, but they do suggest that baseline GC (and not post-stressor GC) is positively associated with migration success.

Generally it is hypothesized that increased baseline GC are associated with decreased survivorship because increased baseline GC are associated with increased chronic stressors (Cort-Fitness hypothesis: Bonier et al. 2009). Our findings appear to support the opposite trend as migratory fish with lower baseline cortisol appear to have lower successful passage through the millpond (Figure 2). Several other studies have also found higher baseline cortisol to be correlated with increased survivorship: Cabezas et al. (2007) found moderately elevated corticosterones were associated with decreased body condition but increased probability of survival in European wild rabbits (*Oryctolagus cuniculus* (Linnaeus, 1758)); Comendant et al. (2003) found higher corticosterone in female lizards (*Uta stansburiana* (Baird and Girard, 1852)) to be associated with increased survival to second ovulation; and Cote et al. (2006) found increased corticosterone to modify behaviour (locomotor and social activity), increasing survival in male adult common lizards (*Lacerta vivipara* (Jacquin, 1787)).

Another possible explanation for why our results are not consistent with the Cort-Fitness hypothesis is that it is difficult to discern whether higher baseline cortisol indicates exposure of the individual to greater chronic stress (which is assumed to be the case by the Cort-Fitness hypothesis; Bonier et al. 2009), or indicates a more competent ability to cope with chronic stressors. The first would likely lead to decreased survivorship with increasing baseline GC level, while the later would likely result in the opposite trend. In our study, all individuals may be subjected to similar chronic stressors as they were all reared in the same stream reach. Thus, baseline stress may be a better indicator of better ability to cope with chronic stressors than exposure to greater environmental stressors.

Post-stressor cortisol, a proxy for stress responsiveness, was not found to be a significant predictor of migration timing, survival or success. Potentially, the effect of stress responsiveness was masked, because we were not able to control for individual variability in baseline as fish were too small to obtain both a baseline and a post-stressor cortisol sample. However post-stressor cortisol was more variable ($SD=40.7$ ng/mL) than baseline cortisol ($SD=7.7$ ng/mL). Cockrem (2013) reviewed studies on baseline and stress-induced GCs and determined that there tended to be more variation in baseline GCs among individuals than in stress-induced levels. Although our findings are not consistent with that generalization, Cockrem (2013) did note that it was not ubiquitous with a number of exceptions. It is unclear why that pattern was observed here but reasons, among others, could include inter-individual differences in how the stressor was perceived or different stress histories. Despite the widely accepted belief that acute stress responses are adaptive, similar to our study several other

studies have failed to find a relationship between stress responsiveness and performance (Romero and Wikelski 2001; Blas et al. 2007), or may even reduce performance. These results may be an indication that different reactive scopes profiles may allow individuals to adequately cope with stressors. Additionally, the benefits of stress responsiveness are increasingly viewed as context dependent, as traits induced by a stress response can have variable consequences to an organisms' performance depending on the nature of the stressor (Breuner et al. 2008).

Similar to our findings in brown trout, the only other study we are aware of that examines individual variation in endocrine regulation in relation to migration in fish (anadromous semelparous sockeye salmon) found no correlation between GC levels and migration timing or rate. They also found individuals with higher stress responsiveness to have greater migration success, but that baseline cortisol not to be a significant predictor of migration success (Cook et al. 2014). Differences in the relationship between GC secretion and migration success found by our study may be attributed to differences in the stressors experienced. For example, salmon may have more chronic stressors such that their baseline levels are at the turning point of the baseline cortisol-survivorship curve hypothesized by Cabezas et al. (2007), where differences in baseline levels have negligible effects on survivorship. The inconsistency between findings may also be reflective of differences in physiology between species.

Conclusions

In summary, we found no evidence suggesting that cortisol regulates the migration life history trade off in juvenile brown trout. But, slightly higher levels of baseline GC (and not post-stressor GC) may favor migration performance. Further study is warranted to help determine the relationship between migration success and GC, with special focus on stress responsiveness within an individual.

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549 Table 1: The mean (with SD) and range of plasma cortisol, individual fish length, mass and
550 Fulton’s Condition factor (K), for juvenile brown trout (*Salmo trutta*) collected from the Gudsø
551 Stream, Denmark. Collected fish were blood sampled either within three minutes of capture
552 (baseline) or 30 minutes following a standardized three minute air exposure (post-stressor).

Treatment	Baseline	Post-Stressor
Sample size	99	99
Cortisol (ng/mL)	5.6 ± 6.7 (0.1 to 25.5)	69.2 ± 40.1 (7.2 to 197.6)
Length (mm)	146±126 (120 to 182)	147±13 (122 to 171)
Mass (g)	28.2±7.6 (15.8 to 48.5)	29.2±8.1 (17.1 to 45.8)
K	0.90±0.07 (0.66 to 1.06)	0.90±0.06 (0.72 to 1.09)

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554

555 Table 2. Number of individuals recaptured, number of sampled individuals that were detected
 556 at S1 and the mean cortisol concentration (with SD) of these individuals for baseline (sampled
 557 within three minutes of capture) and post-stressor (sampled 30 min minutes following a
 558 standardized three minute air exposure) treatment groups of juvenile brown trout (*Salmo*
 559 *trutta*) collected from the Gudsø Stream, Denmark.

Treatment	Baseline	Post-Stressor
Number recaptured	13	6
Cortisol of recaptured (ng/mL)	5.6±5.0	69.9±40.1
Number S1 passage	37	46
Cortisol Passing S1(ng/mL)	5.8±7.7	76.2±42.3
Number S2 passage	19	26
Cortisol Passing S2 (ng/mL)	8.4±9.0	79.2±42.4

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561

Figure 1. Location of Gudsø Stream in Denmark is shown as a black box in the bottom left panel (north-east of Kolding). The portions of the Stream where brown trout (*Salmo trutta*) were captured are shown as thick black lines and the areas re-sampled in June 2013 are shown as thick grey lines. The positions of the two PIT antennas, S1 and S2, are also shown in the larger map. Light gray areas represent water and the outflow location of Gudsø Stream into Kolding Fjord. Map data from the European Environment Agency.

Figure 2. The probability of recapture (a), successful S1 passage (b), and successful passage between S1 and S2 (c) as a function of individual baseline cortisol plasma concentration collected within 3 minutes of capture, for individual juvenile brown trout (*Salmo trutta*) on the Gudsø Stream, Denmark on the left, and on the right the probability of recapture (d), successful S1 passage (e), and successful passage between S1 and S2 (f), as a function of individual cortisol plasma concentration 30 minutes post 3 minute air exposure. Open circles represent actual values found for individuals and the line represents the values predicted using a binary logistic regression with no covariates. Binary logistic regression found no significant relationships except for (c): increased baseline cortisol was associated with greater probability of successful S2 passage for fish that had completed S1 passage, with marginal significance when mass ($p=0.05$) or length ($p=0.04$) was included in the model as a covariate, but not when Fulton's condition factor was included as a covariate. Significant relationships are indicated by three stars.

Figure 1

Figure 2