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Anti-protozoal activity and metabolomic analyses of *Cichorium intybus* L. against *Trypanosoma cruzi*

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**ARTICLE INFO**

Chagas disease, caused by the protozoa *Trypanosoma cruzi*, is a potentially life-threatening parasitic zoonosis infecting 6–7 million people worldwide, mainly in Latin America. Due to the limited numbers of drugs available against this neglected disease and their frequent adverse effects, novel anti-chagasic agents are urgently needed. *Cichorium intybus* L. (chicory) is a bioactive plant with potent activity against parasitic nematodes, but its effects on protozoans are poorly known and no studies have explored its trypanocidal potential. Here, we investigated the activity of *C. intybus* against extracellular and intracellular stages of *T. cruzi*, including the prediction of trypanocidal compounds by metabolomic analyses and bioactivity-based molecular networking. Purified *C. intybus* extracts were prepared from leaves and roots of five *C. intybus* cultivars (cv. ‘Benulite’, ‘Goldine’, ‘Larigot’, ‘Maestoso’ and ‘Spadona’). All *C. intybus* extracts induced concentration-dependent effects against *T. cruzi* trypomastigotes. *C. intybus* leaf extracts had higher trypanocidal selectivity and lower cytotoxicity on mammalian cells than root extracts. The leaf extract of *C. intybus* cv. Goldine also significantly reduced the number of mammalian cells infected with *T. cruzi* amastigotes. Metabolomic and bioactivity-based molecular networking analyses revealed 11 compounds in *C. intybus* leaves strongly linked with activity against trypanomastigotes, including the sesquiterpene lactone lactucin, and flavonoid- and fatty acid-derivatives. Furthermore, seven distinct *C. intybus* molecules (including two sesquiterpene lactone-derivatives) were predicted to be involved in reducing the number of mammalian cells infected with amastigotes. This is the first report of the anti-protozoal activity of *C. intybus* against trypanosomatid parasites and expands our understanding of the anti-parasitic effects of this plant and its bioactive metabolites. Further studies to elucidate the anti-protozoal compound(s) in *C. intybus* and their mode(s) of action will improve our knowledge of using this bioactive plant as a promising source of novel broad-spectrum anti-parasitic compounds with associated health benefits and biomedical potential.

**1. Introduction**

Chagas disease is a vector-borne zoonosis and potentially life-threatening disease caused by the protozoan parasite *Trypanosoma cruzi*. Approximately 6–7 million people are infected with *T. cruzi* worldwide, mainly in Latin America, and with an increasing number of cases also been reported in the US, Canada, Europe, Australia and Japan (Schmunis and Yadon, 2010; Pérez-Molina and Molina, 2018; Elkheir et al., 2021). Chagas disease is a neglected tropical disease, and only two effective anti-chagasic drugs are available; benznidazole and nifurtimox...
investigated for its activity against parasitic helminths in animals, which et al., 2021). During the past 20 years, 2016). Recently, the anthelmintic activity of pure SL from Pea et al., 2011; Pea Cichorium intybus is known to synthesize several natural bioactive metabolites with associated health benefits, such as sesquiterpene lactones (SL), polyphenols and fatty acids, which are detectable in leaves and roots (Bogdanovic et al., 2019; Peña-Espinoza et al., 2020; Janda et al., 2021). A growing scientific interest has focused on the study of C. intybus compounds and their biological activities, including anti-inflammatory and anti-parasitic effects (Bischoff et al., 2004; Pea et al., 2018; Baixinho et al., 2021; Perović et al., 2021). During the past 20 years, C. intybus has been intensively investigated for its activity against parasitic helminths in animals, which has been linked to its content of SL (Peña-Espinoza et al., 2018). Previous in vitro studies have reported that SL-rich C. intybus extracts can induce potent anthelmintic effects against parasitic nematodes (Foster et al., 2011; Peña-Espinoza et al., 2015, 2017, 2020; Williams et al., 2016). Recently, the anthelmintic activity of pure SL from C. intybus against parasitic nematodes has been confirmed by molecular networking and bioactivity-guided fractionation (Valente et al., 2021). Nevertheless, little is known about the potential of C. intybus and its bioactive compounds against other major pathogens of medical or veterinary importance, such as protozoan parasites.

To date, only two studies have explored the anti-protozoal activity of C. intybus. Bischoff et al. (2004) reported direct in vitro effects of purified SL (lactucin and lactucopicrin) from C. intybus roots against the malarial parasite Plasmodium falciparum, with total inhibition of parasite growth at 10 and 50 μg/mL of lactucin and lactucopicrin, respectively. More recently, Woolsey et al. (2019) described the in vitro activity of purified C. intybus leaf and root extracts against the zoonotic protozoa Cryptosporidium parvum, demonstrating a concentration-dependent inhibition of parasite growth. However, no studies have evaluated the anti-parasitic activity of C. intybus against T. cruzi or other trypanosomatid parasites. The objectives of the present study were to investigate the anti-protozoal activity of C. intybus against extracellular and intracellular stages of T. cruzi and their cytotoxicity in mammalian host cells, and to carry out metabolomic analyses and bioactivity-based molecular networking of the tested fractions to facilitate prediction and identification of trypanocidal compounds.

2. Materials and methods

2.1. Plant material and extraction

Leaf and root fresh material from five different C. intybus cultivars was used for preparation of purified extracts, evaluation of anti-protozoal and cytotoxicity activities and metabolomic analyses. The sources of plant material, sample collection and preparation of purified extracts are described in details in Valente et al. (2021). Briefly, fresh leaves and roots were collected from four industrial C. intybus cultivars grown by Sensus B.V. (Roosendaal, The Netherlands): cultivar (cv.) ‘Benulite’, cv. ‘Goldine’, cv. ‘Larigot’ and cv. ‘Maestoso’ (sampled in September 2016 from the test field of Sensus, Colijnsplaat, The Netherlands). Fresh leaves and roots were also sampled from C. intybus cv. ‘Spadona’, grown as a pure sward at the experimental farm of the University of Copenhagen (Taastrup, Denmark), in September 2017. Collected plant material (leaves and roots) from the five chiochy cultivars was subjected to an extraction method optimized for recovery of SL described by Valente et al. (2021). In brief, freeze-dried plant material was dissolved in methanol/H2O (4/1; v/v) with 2% (v/v) formic acid and extracted for 10 min in sonication at room temperature. Resulting crude extracts were recovered and dried under reduced pressure followed by freeze-drying. Dried crude extracts were incubated overnight with Viscozyme® L cellulolytic enzyme mixture (Sigma-Aldrich) for removal of bound sugars, and C. intybus metabolites were recovered with ethyl acetate. Resulting ethyl acetate extracts were dried under reduced pressure, dissolved in 14% methanol in dichloromethane and purified by solid-phase extraction (SPE). After SPE, collected liquid fractions were dried under nitrogen flux and the resulting purified extracts were weighed and stored at −20 °C until in vitro studies and metabolomic analyses. A total of 10 purified extracts (from root and leaf material of the five C. intybus cultivars described above) were selected and used in the present study. Separate aliquots from the same extracts were dissolved in either 100% dimethyl sulfoxide (DMSO) at a stock concentration of 100 mg dry extract/mL (for in vitro assays) or 100% methanol at a concentration of 10 mg dry extract/mL (for metabolite profiling).

2.2. Metabolite profiling of C. intybus extracts by untargeted metabolomics

The metabolite profile of all ten purified C. intybus extracts was analysed by an untargeted metabolomic platform as described in Valente et al. (2021). Briefly, purified C. intybus extracts (10 mg dry extract/mL in methanol) were analysed in a ultra-high performance liquid chromatography-high resolution mass spectrometry (UHPLC-HRMS) platform (Agilent Infinity 1290 UHPLC system QTOF, coupled with an Agilent 6545 QTOF MS; Agilent Technologies, Santa Clara, CA, USA). Resulting LC-MS/MS chromatograms from every purified C. intybus extract were converted to mzXML files and pre-processed using MZmine. For each extract, a table containing all extracted MS features (.csv file) associated with their MS fragmentation spectra (.mgf file) was obtained, and all data was submitted for compound identification and bioactivity-based molecular networking analyses in the Global Natural Products Social (GNPS)’s spectral libraries and Molecular Networking Platform (https://gnps.ucsd.edu/; Wang et al., 2016).

2.3. Parasite and mammalian cell cultures

Trypanosoma cruzi (Y strain, Discrete Type Unit II) cultures were maintained in infected Vero cells (Chlorocebus sabaeus; ATCC® CCL-81) following the method described by Cortes et al. (2015), with minor modifications. Vero cells were cultivated in culture flasks with RPMI 1640 supplemented with 2% (v/v) foetal bovine serum (FBS), penicillin 100 U/mL and streptomycin 100 μg/mL (37 °C, 5% CO2). Vero cells were then infected with T. cruzi trypomastigotes at 4:1 density (trypomastigote:cell), and 4–5 days post-infection, infected cells released T. cruzi trypomastigotes to the culture media that were collected and immediately used for in vitro assays (described in section 2.4) or to infect new Vero cells to maintain T. cruzi cultures. Uninfected Vero cells were cultivated separately for cytotoxicity studies (section 2.5).

2.4. Anti-protozoal activity of C. intybus against extracellular T. cruzi trypomastigotes

First, we assessed the direct anti-protozoal activity of all ten purified C. intybus extracts (tested individually) on isolated T. cruzi trypomastigotes obtained as described in section 2.3. Trypomastigotes (105 trypomastigotes/well) were placed in black 96-well plates and incubated with decreasing concentrations of each C. intybus extract (dissolved in DMSO) for 24 h in RPMI supplemented with antibiotics (37 °C, 5% CO2). Final concentrations tested in triplicates ranged from 100 to 6.3 μg C. intybus extract/mL (0.1% DMSO in all wells). Benznidazole (BNZ) was
used as positive control and tested at a single concentration of 260 μg/mL (i.e. 100 μM BNZ) to enable the complete inhibition of trypanomastigote viability, based on previous results with T. cruzi Y strain (Torchelsen et al., 2021). Positive controls (260 μg BNZ/mL, 0.1% DMSO in well) and negative controls (0.1% DMSO) were run in triplicates at similar conditions. After 24 h of incubation, the viability of exposed trypanomastigotes was evaluated using the resazurin reduction assay (Rolón et al., 2006), which measures the reduction of resazurin (non-fluorescent) to resorufin (highly fluorescent) by metabolically active cells and is directly proportional to the number of viable trypanomastigotes (O’Brien et al., 2006; Rolón et al., 2006). Briefly, 20 μL of resazurin (1 mM) were added to each well, and after 4 h of incubation (37 °C, 5% CO₂), the reduction of resazurin into resorufin by viable parasites was measured at 560 nm (excitation) and 590 nm (emission) in an automated Varioskan Flash reader (Thermo Fisher Scientific, USA).

To evaluate the potential confounding reduction of resazurin by the tested C. intybus extracts (and not by T. cruzi), blank wells containing the same tested concentrations of extracts and controls, but without trypanomastigotes, were incubated under the conditions as described above and evaluated with the resazurin reduction assay. The resulting relative fluorescent units (RFU) were normalized to their negative controls and expressed as parasite viability percentages. Three independent viability experiments with T. cruzi trypanomastigotes were performed.

2.5. Cytotoxicity of C. intybus on uninfected Vero cells

We then evaluated the potential cytotoxicity of C. intybus on mammalian cells by exposing uninfected Vero cells to decreasing concentrations of all ten purified C. intybus extracts (tested individually). Uninfected Vero cells were placed in black 96-well plates (5 × 10⁴ cells/well) and incubated in RPMI supplemented with antibiotics (37 °C, 5% CO₂). After overnight incubation to allow the attachment of Vero cells to the plates, the uninfected cells were exposed to decreasing concentrations of all C. intybus extracts in triplicates (100, 50 or 0 μg extract/mL; 0.1% DMSO in well). Vero cells exposed to BNZ (260 μg/mL) and DMSO (0.1%) in triplicates were run as positive and negative controls, respectively. After 24 h incubation, the viability of exposed Vero cells was measured by the resazurin reduction assay as described in section 2.4. The resulting RFU were normalized to the negative controls and expressed as cell viability percentages. Three independent viability experiments with uninfected Vero cells were performed.

2.6. Anti-protozoal activity of C. intybus against intracellular T. cruzi amastigotes

We further assessed the trypanocidal activity of C. intybus against T. cruzi amastigotes, the replicative intracellular stage of the parasite. The leaf extracts from the five C. intybus cultivars were selected for these experiments based on their higher selectivity towards T. cruzi (in comparison with the root extracts). We evaluated the effects of these five leaf C. intybus extracts on intracellular T. cruzi amastigotes using an in vitro infection model described by Cortes et al. (2015), with modifications. Briefly, sterile round coverslides (15 mm diameter) were added to each well of 24-well plates, followed by seeding of uninfected Vero cells (10⁴ cells/well) and T. cruzi trypanomastigotes (100:1 density, trypanomastigote: cell) in RPMI supplemented with 2% PBS and antibiotics. Parasite invasion of Vero cells was allowed for 48 h (37 °C, 5% CO₂), followed by removal of the supernatant and replacement by fresh medium containing C. intybus extracts (final concentration in well: 10 μg extract/mL, 0.1% DMSO). BNZ was used as positive control and tested at a single concentration of 26 μg/mL (i.e. 10 μM BNZ, 0.1% DMSO) to enable the complete inhibition of intracellular amastigotes, based on previous results with T. cruzi Y strain (Greco et al., 2017). Infected cells exposed to DMSO (0.1%) were incubated as negative controls. After 48 h incubation, exposed cells (attached to the upper side of the coverslides) were washed with sterile PBS and fixed in cold methanol (70%) for 12 h at 4 °C. Then, fixed cells were incubated with DAPI (DAPI:PBS, 1:50,000; NucBlue, Molecular Probes, USA) for 20 min in the dark (room temperature) to selectively stain the DNA of Vero cells and T. cruzi amastigotes. The coverslide containing stained cells were transferred into microscope slides and photographed using a fluorescence microscope (Leica DMI8) at 358 nm (excitation) and 461 nm (emission). Three images were randomly obtained for each cover slide (i.e. treatment) at 20× magnification, and the images were automatically analysed using the software Image J (1.52) to quantify the total number of Vero cells, the number of Vero cells infected with T. cruzi amastigotes and the number of T. cruzi amastigotes per infected cell.

2.7. Bioactivity-based molecular networking for prediction of trypanocidal compounds in C. intybus

The potential trypanocidal compounds in the tested purified C. intybus extracts were predicted by bioactivity-based molecular networking analyses (Wang et al., 2016; Nothias et al., 2018). Molecular networks represent the associations of molecules in related MS data by alignment of different compounds’ mass spectrum based on their similar fragmentation patterns (Wang et al., 2016). Bioactivity-based molecular networking integrates the molecular networks of these related MS data (e.g. different extracts from the same plant species) and their bioactivities (e.g. in vitro anti-parasitic activity of each extract) to predict “bioactivity scores” for each compound in all analysed extracts (Nothias et al., 2018; Peña-Espinoza et al., 2020). A bioactivity score is the correlation between the relative quantification of a single compound in an extract (expressed as individual peak area) and the bioactivity of this extract (e.g. EC₅₀ value), thus predicting the probability of a compound being active in a given biological system (Nothias et al., 2018). In the present study, bioactivity-based molecular networking analyses were performed as described by Valente et al. (2021). In brief, the LC-MS/MS chromatograms from the five C. intybus leaf extracts that were tested on T. cruzi trypanomastigotes and amastigotes (higher selectivity index), were pre-processed with MZmine. All extracted mass spectral features and aggregated MS² fragmentation spectra were submitted for compound identification and molecular networking at the GNPS platform using GNPS’s spectral libraries and feature-based mass spectral molecular networking online workflow (https://gnps.ucsd.edu/; Nothias et al., 2020). The resulting molecular network representing the relationship between all compounds detected in the tested purified C. intybus extracts is accessible at https://gnps.ucsd.edu/ProteoSAFe/status.jsp?task=6fb25560f4b948fa9ac0406fadb3b21.

The resulting molecular network was further processed in Cytoscape 3.8.2 (Shannon et al., 2003) to map bioactivity scores of all detected compounds in the tested extracts. Bioactivity scores were computed following the methodology and the R script provided by Nothias et al. (2018), with the modifications by Peña-Espinoza et al. (2020). Two distinct bioactivity scores were calculated for each compound, based on their predicted activity against extracellular T. cruzi trypomastigotes or towards intracellular T. cruzi amastigotes. Briefly, the individual peak area intensity of detected metabolites were normalized by adding 11 (× +1) to each peak intensity value and divided by the sum of all peak intensities in the same extract +1 (peak intensity +1)/(sum of all peak intensities +1), followed by scaling of the samples by normalizing the peak intensity to the total ion current. Next, bioactivity scores were calculated as the Pearson correlation coefficient (r) between the normalized individual peak areas and the bioactivity of each C. intybus leaf extract in the studies with T. cruzi trypomastigotes (1/[EC₅₀] × 10000) or in the assays with T. cruzi amastigotes (percentage reduction of infected Vero cells). Molecules with high bioactivity scores were defined as having a statistically significantly high positive correlation (r > 0.85, with significance of the correlation P < 0.03; Nothias et al., 2018). Resulting bioactivity scores (CVS format) were imported into the molecular network processed in Cytoscape (see above) to construct the
bioactivity-based molecular network for all compounds detected in the purified *C. intybus* extracts.

### 2.8. Statistical analyses

In the anti-protozoal studies of *C. intybus* against extracellular *T. cruzi* trypomastigotes, the effective concentrations of each tested extract able to inhibit the viability in 50% of exposed parasites (EC$_{50}$) were calculated. Tested extract concentrations were log-transformed, and parasite viability percentages obtained for each condition (nine replicates per concentration from three independent assays) were analysed by non-linear (least squares) regression using the model log (inhibitor) vs. response - variable slope in GraphPad Prism® 7.03 (GraphPad Software, San Diego, CA, USA). The R squared measure of goodness of fit ($R^2$) was calculated for each concentration-response curve. In the cytotoxicity assays with uninfected Vero cells, the cytotoxic concentrations of each *C. intybus* extract able to reduce the viability in 50% of exposed cells (CC$_{50}$) were calculated. Tested extract concentrations were log-transformed and cell viability percentages (nine replicates per concentration from three independent cytotoxicity studies) were analysed by

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**Fig. 1.** Metabolite profile and relative quantification of compounds from the three classes of compounds detected in purified *Cichorium intybus* extracts evaluated for trypanocidal activity. Leaf (L) and root (C) extracts from *C. intybus* (cv. Benulite, cv. Goldine, cv. Larigot, cv. Maestoso and cv. Spadona) were analysed by untargeted metabolomics using UHPLC-HRMS. Metabolites were identified based on the Global Natural Product Social Molecular Networking libraries. The relative quantification of the compounds is based on the peak area of the precursor ion and is illustrated in the heatmap as shades of green (with darker green representing more abundant compounds in the extracts). Identified hit compounds or unidentified derivatives (U; no hit in the libraries but grouped within a specific class of compounds by molecular networking) are presented with their molecular weights (in Da). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)
non-linear (least squares) regression as described above. The selectivity index (SI) of each *C. intybus* extract was calculated as the ratio of their EC50 values on uninfected Vero cells to their EC50 values on *T. cruzi* trypomastigotes (SI = EC50 Vero cells/EC50 T. cruzi trypomastigotes). Statistical differences between EC50 and CC50 obtained with leaf vs. root extracts from the same *C. intybus* cultivar were analysed by extra sum-of-squares F test with a null hypothesis of equal EC/CC50. In the anti-protozoal studies of *C. intybus* with intracellular *T. cruzi* amastigotes, the number of infected cells and the number of amastigotes per infected cell after exposure to purified leaf extracts or BNZ were compared with negative controls (DMSO) by one-way ANOVA with Dunnett’s post-test. The percentage reduction of infected Vero cells with *T. cruzi* amastigotes was also calculated for each purified *C. intybus* leaf extract. A value of P < 0.05 was considered significant.

3. Results

3.1. Metabolite profiling of purified *C. intybus* extracts by untargeted metabolomics

The metabolite profiling and relative quantification of the main compounds (according to the chemical class of compounds) identified in the ten *C. intybus* extracts are presented in Fig. 1. A summary of the total relative concentration of the main chemical classes detected in the purified extracts (based on the sum of individual peak areas) is described in Table 1. The LC/MS-MS chromatograms of each purified *C. intybus* extract are presented in Supplementary Figs. 1–10. The metabolomic analysis and compound identification in GNPS libraries revealed a different profile of identified compounds (library hits) and unidentified derivatives (no-hit in the libraries but grouped within a specific class of compounds) between the extracts. Molecules belonging to three distinct chemical classes were detected, with variations between *C. intybus* cultivars and plant parts: polyphenols, SL and fatty acids, with their derivatives (Fig. 1; Table 1). Based on the sum of peak areas of all detected compounds in the ten *C. intybus* extracts, polyphenols and their derivatives (including flavonoids) were the main identified compounds in all extracts (29.4–38.7% of total peak area), followed by SL and derivatives (8–17.5%) and fatty acids and derivatives (3.0–9.8%). Total polyphenol content was largely similar between *C. intybus* leaves and roots, except for single compounds such as chlorogenic acid, caffeic acid and esculetin (6-7-dihydroxycoumarin), which were almost only present in leaf extracts. Whereas the six major guaianolide SL (and their derivatives) synthesized by *C. intybus* were present in all extracts, and total SL was in average higher in leaves than in root material (Table 1). The individual SL lactucin, dihydro-lactucin, 8-deoxylactucin and dihydro-8-deoxylactucin were mainly detected in *C. intybus* leaf extracts but scarcely in root extracts, whereas luctucopenir and dihydro-lactucopenir were present at similar levels in both leaves and roots (Fig. 1). Not annotated molecules represented 44.6–54.1% of the total peak area in the ten chicory extracts (Table 1). These not annotated molecules correspond to compounds detected in the extracts by UHPLC-HRMS, but that could be neither identified in metabolomic databases (no library hit) nor grouped within a specific class of compounds by molecular networking. The relative quantification of these not annotated (unidentified) molecules in each purified *C. intybus* extract is presented in Supplementary Fig. 11.

3.2. Anti-protozoal activity of purified *C. intybus* extracts against extracellular *T. cruzi* trypomastigotes

The concentration-response curves of the leaf and root extracts from five chicory cultivars in the anti-protozoal assays with extracellular *T. cruzi* trypomastigotes are presented in Fig. 2. All tested extracts induced a concentration-dependent activity against *T. cruzi* trypomastigotes, but with differences between cultivars and leaf and root extracts. Root extracts induced a higher anti-protozoal activity than the leaf extracts from the same cultivar, with significantly lower EC50 (P < 0.001; Table 2), except for the Maestoso-Leaf extract that exerted a more potent trypanocidal effect in comparison with the Maestoso-Root extract (P < 0.0001; Table 2). The Larigot-Root and the Spadona-Leaf extracts induced the highest and lowest trypanocidal activity, respectively.

3.3. Cytotoxicity of purified *C. intybus* extracts on uninfected Vero cells

The cytotoxicity of *C. intybus* on uninfected mammalian Vero cells and the selectivity index (SI) of each tested extract against *T. cruzi* are presented in Table 2. The mean cell viability of Vero cells in the negative control (0.1% DMSO) at 24 h incubation was 99.33% (95% CI = 98.4–100). The leaf extracts were significantly less cytotoxic towards Vero cells (higher CC50) than the root extracts from all *C. intybus* cultivars (P < 0.0001). All the root extracts at the highest concentration tested (100 μg/mL) induced a marked cytotoxic effect and reduced the viability of exposed uninfected Vero cells below 50% (mean cell viability [95% CI] = 37.03% [33.1–40.9]). In contrast, the mean viability of Vero cells exposed to *C. intybus* leaf extracts at the highest concentration (100 μg/mL for 24 h) was ≥80% (95% CI = 77.8–81.9). This lower cytotoxicity resulted in a higher SI of purified *C. intybus* leaf extracts against *T. cruzi*, in comparison to the root extracts (Table 2). The Spadona-leaf extract had the highest trypanocidal SI (90.04), followed by the Maestoso (12.33) and the Goldine (6.00) leaf extracts.

3.4. Anti-protozoal activity of purified *C. intybus* extracts against intracellular *T. cruzi* amastigotes

The trypanocidal activity of the *C. intybus* leaf extracts (higher SI) was further tested on Vero cells infected with intracellular *T. cruzi* amastigotes (Fig. 3). Representative fluorescence microscopic images of Vero cells infected with *T. cruzi* amastigotes and exposed to *C. intybus* extracts at 10 μg/mL (or to negative/positive controls) are depicted in Fig. 3A and were further analysed for quantification of treatment effects.

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**Table 1**

Summary of the relative quantification by class of compounds of molecules detected in leaf (L) and root (R) purified extracts from five cultivars of *Cichorium intybus* (cv. Benulite, cv. Goldine, cv. Larigot, cv. Maestoso and cv. Spadona). Purified *C. intybus* extracts were analysed by untargeted metabolomics using UHPLC-HRMS and compounds were annotated based on the Global Natural Product Social Molecular Networking libraries. Peak areas of identified compounds (library hit) and not-identified derivatives (no hit in the libraries but grouped within a specific class of compounds) were calculated. Molecules belonging to three distinct chemical classes were detected, with variations between *C. intybus* cultivars and plant parts: polyphenols, SL and fatty acids, with their derivatives (Fig. 1; Table 1). Based on the sum of peak areas of all detected compounds in the ten *C. intybus* extracts, polyphenols and their derivatives (including flavonoids) were the main identified compounds in all extracts (29.4–38.7% of total peak area), followed by SL and derivatives (8–17.5%) and fatty acids and derivatives (3.0–9.8%). Total polyphenol content was largely similar between *C. intybus* leaves and roots, except for single compounds such as chlorogenic acid, caffeic acid and esculetin (6-7-dihydroxycoumarin), which were almost only present in leaf extracts. Whereas the six major guaianolide SL (and their derivatives) synthesized by *C. intybus* were present in all extracts, and total SL was in average higher in leaves than in root material (Table 1). The individual SL lactucin, dihydro-lactucin, 8-deoxylactucin and dihydro-8-deoxylactucin were mainly detected in *C. intybus* leaf extracts but scarcely in root extracts, whereas lactucopenir and dihydro-lactucopenir were present at similar levels in both leaves and roots (Fig. 1). Not annotated molecules represented 44.6–54.1% of the total peak area in the ten chicory extracts (Table 1). These not annotated molecules correspond to compounds detected in the extracts by UHPLC-HRMS, but that could be neither identified in metabolomic databases (no library hit) nor grouped within a specific class of compounds by molecular networking. The relative quantification of these not annotated (unidentified) molecules in each purified *C. intybus* extract is presented in Supplementary Fig. 11.

<table>
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<th>Plant material</th>
<th>Total peak area by class of compound (%)</th>
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<th>Fatty acids and derivatives</th>
<th>Not annotated</th>
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<td>Larigot-L</td>
<td>15.4</td>
<td>30.0</td>
<td>3.5</td>
<td>51.1</td>
</tr>
<tr>
<td>Larigot-R</td>
<td>9.6</td>
<td>32.5</td>
<td>5.9</td>
<td>52.0</td>
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<tr>
<td>Maestoso-L</td>
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<td>29.4</td>
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<td>51.3</td>
</tr>
<tr>
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<td>32.7</td>
<td>6.7</td>
<td>50.2</td>
</tr>
<tr>
<td>Spadona-L</td>
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<td>30.8</td>
<td>6.1</td>
<td>48.9</td>
</tr>
<tr>
<td>Spadona-R</td>
<td>8.0</td>
<td>37.5</td>
<td>9.8</td>
<td>44.6</td>
</tr>
</tbody>
</table>

* a Sum of peak areas (%) of all precursor ions identified as sesquiterpene lactones, polyphenols, fatty acids or their derivatives (no library hit but associated to a specific class of compounds) presented in details in Fig. 1.
* b Not annotated molecules (no library hit) and not grouped within any class of compound by molecular networking (presented in Supplementary Fig. 11).
The leaf extract from *C. intybus* cv. Goldine was the only able to significantly reduce the number of infected Vero cells by 35.2%, in comparison with the negative control (*P* = 0.0013; Fig. 3B). Leaf extracts from *C. intybus* cv. Larigot, Benulite and Maestoso also decreased the number of infected cells by 22.6%, 14.8% and 9.3%, respectively, in comparison with infected cells exposed to 0.1% DMSO (*P* > 0.06), while exposure to the Spadona-leaf extract did not affect the number of infected Vero cells (Fig. 3B). None of the tested leaf extracts significantly reduced the number of amastigotes per infected cell. The extracts from *C. intybus* cv. Goldine and cv. Benulite marginally decreased the number of amastigotes per cell by 14.3% and 13.6%, respectively, in comparison with infected cells exposed to 0.1% DMSO (*P* > 0.06), while exposure to the Spadona-leaf extract did not affect the number of infected Vero cells (Fig. 3B). In contrast, the positive control BNZ (tested at 26 μg/mL) induced a significant reduction in the number of infected cells and the number of amastigotes per infected cell by 94.4% and 86.2%, respectively (*P* < 0.0001; Fig. 3B).

### 3.5. Bioactivity-based molecular networking for prediction of trypanocidal compounds in purified *C. intybus* extracts

To further explore the relationship between the compounds present in the five *C. intybus* leaf extracts (higher selectivity towards *T. cruzi*), bioactivity-based molecular networking analyses were performed to facilitate the prediction and identification of trypanocidal molecules towards extracellular *Trypanosoma cruzi* trypomastigotes (Fig. 4). A bioactivity-based molecular network represents the relationship between structurally-related metabolites in the analysed extracts, and predicts the bioactivity of each molecule in those extracts. In Figs. 4 and 5, each node represents the spectra of one molecule, and pie charts inside nodes illustrate the relative concentration of each molecule among all *C. intybus* leaf extracts. The edges between nodes correspond to the spectrum-to-spectrum alignment between two compounds in relation to their fragmentation pattern. All identified compounds (library hit) and their derivatives were clustered within one of three classes of chemical compounds in the molecular network: SL, polyphenols or fatty acids.

First, we performed a molecular networking analysis by computing the bioactivity scores of each compound based on the bioactivities (EC50) of the purified leaf extracts against extracellular *T. cruzi* trypomastigotes (Fig. 4). The five *C. intybus* leaf extracts induced a concentration-dependent effect against *T. cruzi* trypomastigotes, but with different EC50 between extracts (Table 2). These EC50 values (normally distributed; Shapiro-Wilk normality test *P* > 0.9) were used for bioactivity score prediction by calculating the Pearson coefficient of correlation between the EC50 of the leaf extracts with the normalized peak intensity of each compound detected in the extracts (total 196 metabolites). In Fig. 4, the node sizes proportionally reflect the bioactivity score (Pearson correlation coefficient *r*) of each molecule, with yellow squares surrounding nodes indicating molecules with a significantly high bioactivity score, and thus a predicted high trypanocidal activity towards *T. cruzi* trypomastigotes (*r* > 0.85 with *P* < 0.03). The bioactivity-based molecular networking analysis resulted in 11 compounds with a significantly high bioactivity score, meaning that the increased concentration of these metabolites had a strong and significantly positive correlation with higher anti-protozoal activity against *T. cruzi* trypomastigotes. From the molecules with high bioactivity score, the only identified compound (library hit) was the SL lactucin (molecular weight of precursor ion [MW] = 277.107; *r* = 0.94; *P* = 0.017). Two other unidentified compounds (no library hit) clustered within the polyphenols (flavonoid compound with MW = 349.199; *r* = 0.94; *P* = 0.018) and fatty acids (molecule related with 9-Oxo-ODE with MW = 229.143; *r* = 0.96; *P* = 0.008) were also predicted to have significantly high bioactivity scores (Fig. 4). Moreover, eight not annotated compounds (not clustered in any of the mentioned chemical classes) also had significantly high bioactivity scores (Supplementary Fig. 12). Next, we performed a second molecular networking analysis by calculating the bioactivity scores of each compound based on the percentage reduction of...
C. intybus Maestoso-leaf vs. root extracts was also recently reported against the parasitic nematode Ascaris suum (Valente et al., 2021). In the present study, the enhanced anti-protozoal activity of the Maestoso-leaf extract, in comparison with the Maestoso-root extract, was likely related with their different metabolite profiles. In this regard, the Maestoso-leaf extract had the highest concentration of choric acid (MW = 163.039) among leaf and root extracts from all tested C. intybus cultivars, as well as the largest concentrations of an unidentified sesquiterpene lactone-derivative (MW = 205.158) and two fatty acid-derivatives (MW = 229.143, which had a significantly high bioactivity score against trypomastigotes, and MW = 425.377). These compounds may be related with the marked anti-T. cruzi of the Maestoso-leaf extract, and further studies should confirm their role in the apparent broad anti-parasitic effects elicited by leaves from this C. intybus cultivar.

In addition to their potent anti-trypomastigote activity, the C. intybus root extracts also induced a higher toxicity on mammalian cells, in comparison with the leaf extracts, irrespective of cultivar. The lower cytotoxicity of C. intybus leaf extracts towards uninfected cells could be explained by i) the presence of cytotoxic molecules in C. intybus roots that are absent in the leaves and/or ii) the synthesis of protective compounds in C. intybus leaves that reduced the cytotoxicity of other metabolites in the tested extracts. Additional bioactivity-based molecular networking analyses of C. intybus root and leaf extracts based on their cytotoxicity (CC50) on uninfected Vero cells (data not shown) predicted ten not annotated molecules only in the root extracts with significantly high bioactivity scores (i.e. higher cytotoxicity) towards mammalian cells (MWs = 763.552; 691.495; 647.469; 749.538; 705.512; 719.525; 661.485; 603.443; 837.590; 389.170). In comparison, merely one not annotated compound present in the leaf extracts was predicted to have a high bioactivity score against uninfected Vero cells (MW = 207.138). Thus, the presence of several of not annotated molecules with a predicted high cytotoxicity only in C. intybus roots may have been related with their increased cytotoxicity in our in vitro model. Furthermore, the leaf extracts from all C. intybus cultivars had markedly higher concentrations of chlorogenic acid, caffeic acid and esculetin, compared with root extracts. These polyphenols have been reported to elicit antioxidant and protective mechanisms in different biological systems exposed to toxic agents (Sato et al., 2011; Filipsky et al., 2015; Rashidi et al., 2022), and their enhanced concentration in C. intybus leaves may have contributed to their low toxicity by protecting the exposed cells from cytotoxic compounds.

Based on the low cytotoxicity of C. intybus leaves on mammalian cells, and their resulting higher anti-T. cruzi selectivity in comparison with roots, we tested all purified leaf extracts for their activity against intracellular T. cruzi amastigotes. Here, the leaf extract from C. intybus cv. Goldine was the only to significantly reduce the number of mammalian cells infected with T. cruzi amastigotes. However, none of the tested leaf extracts decreased the number of amastigotes per infected cell, which were in contrast significantly reduced by the positive control BNZ. Considering the marked trypanocidal activity and high SI of C. intybus leaf extracts against extracellular trypomastigotes, the lower activity against intracellular parasites may be explained by two causes. First, poor pharmacokinetics of C. intybus metabolites in the infected Vero cells linked to host cell factors (such as interferences in the uptake of the molecules by the plasma membrane) may have limited the cytoplasmic concentrations of the compounds that reached the intracellular parasites (Alcantara et al., 2018). Secondly, only one concentration of C. intybus leaf extracts was tested on infected Vero cells (10 μg extract/mL), and therefore, the poor activity on intracellular T. cruzi amastigotes may have been a consequence of the (low) concentration evaluated. Therefore, further studies could confirm the intracellular trypanocidal activity of C. intybus compounds, and study their uptake by host cells, at increased concentrations.

Bioactivity-based molecular networking analyses to infer potential trypanocidal molecules in C. intybus predicted 11 metabolites with a significantly high activity against T. cruzi trypomastigotes, and seven

### Table 2

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Plant part</th>
<th>T. cruzi trypomastigotes</th>
<th>R²</th>
<th>Vero cells</th>
<th>Selectivity Index</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>EC₅₀ (95% CI)</td>
<td>C₅₀ (95% CI)</td>
<td>(CC₅₀ Vero cells/EC₅₀ T. cruzi)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Benelite</td>
<td>Leaf</td>
<td>38.18 (36.4–40.3)</td>
<td>0.97</td>
<td>191.6 (160.2–245.0)</td>
<td>5.01</td>
</tr>
<tr>
<td>Root</td>
<td>32.53 (30.3–34.8)**</td>
<td>0.95</td>
<td>96.3 (94.8–97.7)</td>
<td>2.96</td>
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</tr>
<tr>
<td>Goldine</td>
<td>Leaf</td>
<td>36.18 (34.4–38.0)</td>
<td>0.96</td>
<td>217.1 (183.1–275.1)</td>
<td>6.00</td>
</tr>
<tr>
<td>Root</td>
<td>27.87 (26.7–29.1)**</td>
<td>0.98</td>
<td>73.24 (69.3–77.3)</td>
<td>2.63</td>
<td></td>
</tr>
<tr>
<td>Larigot</td>
<td>Leaf</td>
<td>34.3 (32.3–36.4)</td>
<td>0.96</td>
<td>180.3 (138.7–237.1)</td>
<td>5.26</td>
</tr>
<tr>
<td>Root</td>
<td>20.16 (18.8–21.6)**</td>
<td>0.94</td>
<td>89.47 (85.5–93.4)</td>
<td>4.44</td>
<td></td>
</tr>
<tr>
<td>Maestoso</td>
<td>Leaf</td>
<td>31 (30.1–32.0)</td>
<td>0.98</td>
<td>382.2 (191.3–9968)</td>
<td>12.33</td>
</tr>
<tr>
<td>Root</td>
<td>41.3 (38.7–43.9)***</td>
<td>0.95</td>
<td>102.1 (99.2–105.5)</td>
<td>2.47</td>
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</tr>
<tr>
<td>Spadona</td>
<td>Leaf</td>
<td>41.46 (38.3–44.7)</td>
<td>0.97</td>
<td>3733 (706.3–62041)</td>
<td>90.04</td>
</tr>
<tr>
<td>Root</td>
<td>29.79 (28.2–31.4)**</td>
<td>0.93</td>
<td>68.28 (65.3–71.5)</td>
<td>2.29</td>
<td></td>
</tr>
</tbody>
</table>

Statistical difference between purified leaf and root extract within the same cultivar: ***P < 0.0001; **P = 0.0002; CI = confidence interval; R² = goodness-of-fit of concentration-response curves for T. cruzi trypomastigotes.

* Effective concentration able to reduce the viability in 50% of the exposed T. cruzi trypomastigotes (EC₅₀) in μg extract/mL.

† Cytotoxic concentration able to reduce the viability in 50% of the exposed Vero cells (CC₅₀) in μg extract/mL.

infected Vero cells with T. cruzi amastigotes induced by the C. intybus leaf extracts (Fig. 5). This analysis resulted in seven compounds with a significantly high bioactivity score (r > 0.85; P < 0.03), including two unidentified SL-derivatives (MW = 483.129; r = 0.93; P = 0.021) and MW = 965.249; r = 0.91; P = 0.029; Fig. 5) and five not annotated compounds (Supplementary Fig. 13).

### 4. Discussion

In the present study, we have described anti-protozoal effects of C. intybus against T. cruzi trypomastigotes, with a higher trypanocidal selectivity and lower cytotoxicity on mammalian cells of purified leaf extracts. In addition, the leaf extract from C. intybus cv. Goldine significantly reduced the number of cells infected with T. cruzi amastigotes. Untargeted metabolomic and bioactivity-based molecular networking analyses revealed 11 compounds strongly linked with the anti-parasitic activity of C. intybus against trypomastigotes, including the SL lactucin, and flavonoid and fatty acid derivatives. Furthermore, seven distinct C. intybus molecules (including two SL-derivatives) were predicted to be involved in reducing the number of Vero cells infected with T. cruzi amastigotes.

All extracts from the five C. intybus cultivars induced a concentration-dependent activity against extracellular T. cruzi trypomastigotes, but with different potencies between leaf and root extracts and among cultivars. Root extracts were more potent than leaf extracts of the same cultivar against trypomastigotes, with one exception: the leaf extract of C. intybus cv. Maestoso. A higher anti-parasitic activity of C. intybus...
compounds with a significantly high activity in reducing the number of Vero cells infected with \textit{T. cruzi} amastigotes. Interestingly, none of the seven compounds with a high bioactivity towards intracellular amastigotes were predicted to have a significant activity against extracellular trypomastigotes, suggesting that distinct \textit{C. intybus} metabolites may act on different life stages of \textit{T. cruzi}. However, considering that in our study none of the purified \textit{C. intybus} extracts reduced the number of \textit{T. cruzi} amastigotes per infected cell, these computational predictions should be considered with caution. The SL lactucin was the only identified compound with a significantly high bioactivity score activity against trypomastigotes, whereas two unidentified SL-derivatives were predicted to have a significant high activity towards amastigotes. Previously, Bischoff et al. (2004) reported that pure lactucin and lactucopicrin isolated from \textit{C. intybus} exerted a concentration-dependent in vitro growth inhibition of the malarial parasite \textit{P. falciparum}, with complete parasite growth inhibition of lactucin and lactucopicrin at 10 and 50
Node sizes proportionally reflect the predicted bioactivity score of the molecule. The bioactivity score is the Pearson correlation coefficient ($r$) the more related the compounds are. Pie charts inside nodes describe the relative concentration (based on peak area) of each molecule among the different extracts. Edges (connections) between nodes represent the spectrum-to-spectrum alignment between two compounds in relation with their fragmentation pattern (i.e. the thicker the connection, the more related the compounds are). Each node represents one molecule detected by UHPLC-HRMS in five purified extracts of the tested purified extracts, either as single compounds or in combination. Additional experiments could also be performed to further test the most promising trypanocidal compounds suggested by our molecular networking analyses, such as the SL lactucin (high bioactivity score against Trypanosoma cruzi trypomastigotes), the two unidentified SL-derivatives with a high bioactivity in reducing the number of cells of C. intybus by molecular networking and bioguided-fractionation, confirming the SL 8-deoxylactucin as the main anti-nematodal metabolite (as single compound and in synergistic combination with other SL). In the present work, we tested similar purified C. intybus extracts as in Valente et al. (2021), and our analyses suggested that the compounds responsible for the anti-protozoal activity of C. intybus are different than those exerting anthelmintic effects, including various identified and not annotated molecules with high bioactivity scores towards T. cruzi. Therefore, further studies are needed to confirm the anti-protozoal metabolite(s) in C. intybus, for example by bioguided-fractionation of leaf material, followed by isolation and testing of purified compounds as single molecules and in combination. Additional experiments could also be performed to test the most promising trypanocidal compounds suggested by our molecular networking analyses, such as the SL lactucin (high bioactivity score against T. cruzi trypomastigotes), the two unidentified SL-derivatives with a high bioactivity in reducing the number of cells infected with T. cruzi amastigotes, as well as chicoric acid (possibly related with the higher trypanocidal activity of the Maestoso leaf-extract). Our present investigation thus provide a foundation for this further research that could lead to the identification of novel anti-protozoal compounds with therapeutic potential against Chagas disease and other trypanosomatid parasites, and which may also be used
as starting molecules for further chemical optimization. In addition, preliminary studies by our group showed a synergistic trypanocidal activity of purified C. intybus leaf extracts and BNZ (Peña-Espinoza et al., unpublished observations), and further research could also explore the potential combination of C. intybus compounds and established anti-chagasic drugs against T. cruzi.

In conclusion, we have described concentration-dependent trypanocidal effects of purified C. intybus extracts against extracellular T. cruzi trypanomastigotes, with C. intybus leaf extracts demonstrating higher antiparasitic selectivity and lower cytotoxicity on mammalian cells than root extracts. The leaf extract of C. intybus cv. Goldine was also able to significantly reduce the number of mammalian cells infected with T. cruzi amastigotes. Metabolomic and bioactivity-based molecular networking revealed distinct compounds in C. intybus leaves strongly linked with activity against T. cruzi trypanomastigotes and amastigotes, including the SL lactucin and SL-derivatives. Further studies elucidating the precise trypanocidal compound(s) and their mode(s) of anti-parasitic action will improve our knowledge on the use of C. intybus as a source of novel anti-trypanosomatid agents with associated health benefits and biomedical potential.

Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Matthew de Roode reports a relationship with Sensus B/V that includes: employment.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ijpjppdr.2022.08.002.

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