

Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world

Cardoso Alves, Samara; Díaz-Ruiz, Erick; Lisboa, Bruna; Sharma, Minaxi; Mussatto, Solange I.; Kumar Thakur, Vijay; Kalaskar, Deepak M.; Gupta, Vijai K.; Chandel, Anuj K.

Published in: Food Research International

Link to article, DOI: 10.1016/j.foodres.2023.112596

Publication date: 2023

Document Version Peer reviewed version

Link back to DTU Orbit

Citation (APA):

Cardoso Alves, S., Díaz-Ruiz, E., Lisboa, B., Sharma, M., Mussatto, S. I., Kumar Thakur, V., Kalaskar, D. M., Gupta, V. K., & Chandel, A. K. (2023). Microbial meat: A sustainable vegan protein source produced from agriwaste to feed the world. *Food Research International*, *166*, Article 112596. https://doi.org/10.1016/j.foodres.2023.112596

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from the public portal for the purpose of private study or research.

- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world

Samara Cardoso Alves, Erick Díaz-Ruiz, Bruna Lisboa, Minaxi Sharma, Solange I. Mussatto, Vijay Kumar Thakur, Deepak M. Kalaskar, Vijai K. Gupta, Anuj K. Chandel

PII:	S0963-9969(23)00141-2
DOI:	https://doi.org/10.1016/j.foodres.2023.112596
Reference:	FRIN 112596
To appear in:	Food Research International
Received Date:	6 November 2022
Revised Date:	27 January 2023
Accepted Date:	14 February 2023



Please cite this article as: Cardoso Alves, S., Díaz-Ruiz, E., Lisboa, B., Sharma, M., Mussatto, S.I., Kumar Thakur, V., Kalaskar, D.M., Gupta, V.K., Chandel, A.K., Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world, *Food Research International* (2023), doi: https://doi.org/10.1016/j.foodres.2023.112596

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2023 Elsevier Ltd. All rights reserved.

1	Microbial meat: A sustainable vegan protein source produced from agri-
2	waste to feed the world
3	Samara Cardoso Alves ^a , Erick Díaz-Ruiz ^a , Bruna Lisboa ^a , Minaxi Sharma ^b , Solange I.
4	Mussatto ^c , Vijay Kumar Thakur ^{d, e} , Deepak M. Kalaskar ^f , Vijai K. Gupta ^{d, e*} , Anuj K.
5 6	Chandel ^a *
7	^a Department of Biotechnology, Engineering School of Lorena, University of São Paulo,
8	Lorena, São Paulo 12.602.810, Brazil
9	^b Haute Ecole Provinciale de Hainaut- Condorcet, 7800 ATH, Belgium
10	^c Department of Biotechnology and Biomedicine, Technical University of Denmark, Søltofts
11	Plads, Building 223, 2800, Kongens Lyngby, Denmark
12	^d Biorefining and Advanced Materials Research Center, SRUC, Kings Buildings, West Mains
13	Road, Edinburgh EH9 3JG, UK
14	^e Center for Safe and Improved Food, SRUC, Kings Buildings, West Mains Road, Edinburgh
15	EH9 3JG, UK
16	^f UCL Institute of Musculoskeletal Sciences (IOMS), Division of Surgery and Interventional
17	Science, Royal National Orthopaedic Hospital-NHS Trust, Stanmore, Middlesex, HA7 4LP,
18	UK
19	
20	* Corresponding authors:
21	anuj10@usp.br, anuj.kumar.chandel@gmail.com (A.K.Chandel);
22	vijai.gupta@sruc.ac.uk, vijaifzd@gmail.com (V. K. Gupta)
23	
24	
25	

26

27 Abstract

In the modern world, animal and plant protein may not meet the sustainability criteria due to their high need for arable land and potable water consumption, among other practices. Considering the growing population and food shortage, finding alternative protein sources for human consumption is an urgent issue that needs to be solved, especially in developing countries.

33 In this context, microbial bioconversion of valuable materials in nutritious microbial cells 34 represent a sustainable alternative to the food chain. Microbial protein, also known as singlecell protein (SCP), consist of algae biomass, fungi or bacteria that are currently used as food 35 36 source for both humans and animals. Besides contributing as a sustainable source of protein to feed the world, producing SCP, is important to reduce waste disposal problems and production 37 costs meeting the sustainable development goals. However, for microbial protein as feed or 38 39 food to become an important and sustainable alternative, addressing the challenges of raising 40 awareness and achieving wider public regulatory acceptance is real and must be addressed with 41 care and convenience. In this work, we critically reviewed the potential technologies for 42 microbial protein production, its benefits, safety, and limitations associated with its uses, and perspectives for broader large-scale implementation. We argue that the information 43 44 documented in this manuscript will assist in developing microbial meat as a major protein 45 source for the vegan world.

46

47 Keywords: Single-cell protein, Microbial meat, Protein production, Sustainability

- 48
- 49 **1. Introduction**

50 The world population, which currently surpassed 8.0 billion inhabitants, is growing 51 exponentially and is expected to reach around 9.2 billion people by 2040. This rapid population

growth has led to global food insecurity. Furthermore, the COVID-19 pandemic has increased the risk of food insecurity and global hunger. According to the United Nations, hunger is about to reach millions of families worldwide due to factors such as geo-political conflicts, population rise, climate extremes, and the COVID-19. With this scenario, the agricultural sector needs a tremendous transformation to satisfy the growing global demand for food (UN, 2019; FAO, 2021; GLOBAL TRENDS, 2021).

58 The expansion of food production and the intensification of agriculture have caused a 59 high cost to the environment, contributing 31% of global anthropogenic GHG emissions and being responsible for strong changes in the composition and biodiversity of natural ecosystems, 60 61 such as soil erosion, acid rain, eutrophication, and climate change. Thus, international organizations, industries, governments and society have been called upon to provide 62 generalized responses to prevent the global food crisis (Hashempour-Baltork et al., 2020; 63 64 Kusmayadi et al., 2021; Tubiello et al., 2022). Nearly a billion people worldwide cannot afford 65 food that contains enough protein and calories required for their health. The lack of necessary protein sources causes serious health problems such as muscle weakness, defective immune 66 67 system, and growth deficiency (Berners-Lee et al., 2018). On the other hand, high consumption of meat products can cause health problems and seriously affect the environment/climate. 68 69 Therefore, it is crucial for the food businesses to provide viable alternatives to animal proteins, 70 particularly those derived from meat, into the market that are less expensive and consume less 71 natural resources. (Bonny et al., 2015; Hartmann & Siegrist, 2017; Prosekov & Ivanova, 2018). 72 Converting waste into valuable food or feed for humans and animals is not only an 73 environmentally friendly activity but also a healthy business work (Jurasz et al., 2018; Sharif et al., 2021). Currently, large part of waste pollutes the environment or is processed into low 74 75 value-added products such as biofuels and biogas. However, high-quality products such as

single cell oil, single cell proteins, chemicals, and enzymes, among others, can also be obtained
from wastes, and different production methods are being developed for that.

Microbial meat is one of the most relevant high-quality diet products that can be obtained 78 79 from agri-waste resources (El-Bakry et al., 2015; Finco et al., 2017; van der Spiegel et al., 80 2013). SCP is a protein of microbial origin produced from a pure or mixed culture of bacteria, 81 fungi, veasts or microalgae (Hashempour-Baltork et al., 2020). SCPs are dry cells that can be 82 used as protein supplements or as protein-rich ingredients in human and animal diets, providing 83 interesting benefits from a nutritional point of view (Anupama & Ravindra, 2000; Geada et al., 84 2021). Moreover, from an environmental perspective, SCP does not require a large area of land 85 or large reservoirs of water for its production, making it an excellent alternative to vegetable 86 protein sources. Its production also does not emit greenhouse gases into the environment as 87 animal protein sources do. Furthermore, the production of SCP is independent of seasonal and 88 climatic variations and can be carried out throughout the year (Mekonnen & Hoekstra, 2014; 89 Miller et al., 2019; Sharif et al., 2021). Cultured meat is guite different from microbial protein 90 as it is produced by cultivating animal cells in reactors and considered as pure animial meat 91 (Choudharay et al., 2020). While microbial protein seems a viable protein source for vegan 92 population, cultured meat is considered an ideal protein for non-vegeterians.

The selection of cheap and suitable substrates or biodegradable agro-industrial byproducts as a source of nutrients for microorganisms to grow and produce proteins is of fundamental importance to allow an incredible growth of the microorganism at a reduced production cost (Pogaku et al., 2009; Ravindra et al., 2009). In this sense, apple pomace, yam skins, potato skins, citrus pulp, pineapple residues, and papaya residues are some of the substrates used as a nutrition source in microbial cultivation (Diwan et al., 2018; Spalvins et al., 2019).

100 Microorganisms (algae and molds, 2-6h; bacteria and yeasts, 0.33-2h) generate protein more 101 efficiently than any animal or plant (1-2 years and a few months, respectively). In this way, the 102 production of protein biomass has several advantages in relation to livestock and conventional 103 crops. Furthermore, the microorganisms have a high protein rate based on dry mass (30-80%, 104 depending on the applied microorganism) and the protein has good nutritional value. In 105 addition, a wide variety of raw materials can be used as a substrate in the SCP, including low-106 value agro-industrial waste and by-products. Relatively small land areas can be used to carry 107 out continuous fermentation processes to cultivate microbial proteins in large quantities. The 108 generation of SCP does not depend on seasonal, weather and weather variations. In addition, 109 microorganisms are more easily genetically modified than plants and animals. In addition, the 110 SCP presents the essential amino acid requirements for human nutrition (Octasylva & Rurianto, 111 2020; Anupong et al., 2022).

Single-cell proteins will contribute to the greater popularization and wider availability of protein sources in foods. The greatest growth and demand for vegan meat was found in Europe. The growth rate of meat analogues is projected at 7.1% in 2025, growing sharply to 73% by 2050. European countries (51.5%), North America (26.8%), Asia-Pacific (11.8%), Latin America (6.3%), and the Middle East and Africa (3.6%) have the highest share of the global market for plant-based meat analogues (Kumar et al., 2022; Sheth & Patel, 2023).

The selection of cheap and suitable substrates or biodegradable agro-industrial byproducts as a source of nutrients for microorganisms to grow and produce proteins is of fundamental importance to allow an incredible growth of the microorganism at a reduced production cost (Pogaku et al., 2009; Ravindra et al., 2009). In this sense, apple pomace, yam skins, potato skins, citrus pulp, pineapple residues, and papaya residues are some of the substrates used as a nutrition source in microbial cultivation (Diwan et al., 2018; Spalvins et al., 2019). Finally, it is worth mentioning that growing concerns about the food crisis and/or

lack of healthy foods due to the increased population and environmental issues have resulted in more studies on the production of SCP as a potential meat substitute (Hashempour-Baltork et al., 2020). Therefore, this review summarizes and critically discusses different points related to the production of SCP in the form of microbial meat grown on agri-food waste resources as nutrient substrates, such as recent advancements in production technology, the protein recovery and purification processes, safety problems and the current industrial scenario.

131 **2. Single Cell Protein (SCP)**

Microbial protein is referred to as single-cell protein, although some of the producing microorganisms are multicellular, such as filamentous fungi or filamentous algae. In 1968, the term SCP was introduced for the first time, when scientists gathered to know the most appropriate terminologies in common practice, i.e. microbial protein at the Massachusetts Institute of Technology, United States (Matassa et al., 2020; Shharif et al., 2021). Microorganisms help with protein deficiencies when used to increase the amount of protein and improve the quality of fermented feeds (Bratosin et al., 2021).

The increase in the global search for protein will certainly make SCP more and more interesting, although protein of microbial origin has a low proportion of current human nutrition. The high speed of growth or the ability to apply substrates such as CO2 and methane, as carbon sources, makes the processes more efficient and sustainable compared to those employed in traditional agriculture (Balagurunathan et al., 2022; Yang et al., 2022).

Currently, SCP can be produced by a limited number of microbial species, especially when human demand is taken into account. The diversity of SCP sources applied in animal feed is greater than that certified for human consumption and is expanding (Thiviva et al., 2022). According to what will be brought forward, products derived from fungi, algae and bacteria are under development or being used. Typically, production processes proceed first with the preparation of the nutrient medium, then with the cultivation, then with the separation and

concentration of the SCP, in certain cases drying, and finally the final processing of the SCP
into ingredients and products (Jones et al., 2020; Nyyssölä et al., 2022).

152 High food grade substrates are generally used to produce SCP for human consumption. 153 However, there is belief in the development of processes to produce SCP from cheap waste 154 from the food and beverage processing industries, as well as from agricultural and forestry 155 sources. The SCP is composed of a high protein content, which varies between 60 to 82% based 156 on dry matter, in addition, carbohydrates, nucleic acids, vitamins, minerals and fats are also part 157 of its composition. Another benefit related to SCP is that it is rich in several essential amino 158 acids, such as methionine, lysine, which are not present in adequate proportions in most animal 159 and plant sources (Al-Mudhafr, 2019; Zha et al., 2021; Khan et al., 2022).

160

161 **2. Microorganisms as a protein source**

Different microorganisms, including microalgae, fungi, yeasts and bacteria, can be used as single cell proteins for food and feed applications due to their protein-rich composition. **Table 1** summarizes some examples of microorganisms used as SCP and their protein content. More details on their relevance as a source of protein and their production and utilization are discussed in the following sections.

167

Table 1

168

169 2.1. Microalgae

The consumption of algae dates thousands of years across different cultures. However, new developments are still needed today to boost the use of microalgae as a mainstream food option. Development of improved organoleptic traits, evaluation and increase of nutritional content, development of large-scale production units and also optimization of yields are some of the challenges for microalgae to be seen as a more common food source (Mourelle et al.,
2017; Torres-Tiji et al., 2020).

176 Microalgae have several features that make them attractive for large-scale production, 177 food, and feed applications. These features include high biomass yields per unit area, the ability 178 to grow on non-arable land, and the possibility of using non-potable water and even salt water 179 for its cultivation. Nevertheless, the scale up of the appropriate technologies and efficient 180 management of precision fermentation parameters and investments are necessary to develop 181 new microalgae-based products. First is selecting adequate species, which can be done by using 182 bioprospecting methods and searching from established Generally Recognized as Safe (GRAS) 183 species. The GRAS certification is needed for a new species, which is costly and time-184 consuming (FDA, 2017).

After the strain selection, it may be necessary to carry out genetic improvements to the 185 186 strain to enhance the desired traits, such as the yield, organoleptic trait or nutritional content. 187 Related to yield, high productivity, resistance factors and adaptation to outdoor growth are 188 examples of characteristics to be improved. Regarding the organoleptic traits, taste, aroma, 189 texture, palatability, color and appearance are some of the traits that can be improved. Finally, 190 in terms of nutritional content, the protein content and amino acid profile, the lipid content and 191 profile, and the aggregation of other nutritional molecules can be improved (Anderson et al., 192 2017).

Genetic improvements in microalgae can be done by random DNA alteration, UV mutagenesis, mating and genome shuffling, but these processes can be labor-intensive and timeconsuming. Controlled DNA manipulation can deliver faster and more precise results using techniques like targeted mutagenesis, synthetic genetic tools and recombinant protein expression systems. Finally, to obtain a high yield during the cultivation of the final species, it is necessary to work on bioprocess development, including medium optimization, growth systems adaptation, and developing a robust and cheap downstream process (Torres-Tiji et al.,200 2020).

Microalgae constitute a large market today since derived products like alginates and carrageenans are widely used in several industrial sectors, but specifically related to food and feed applications, there is still no precise market defined. Algae has several components of value for human nutrition, like, omega-3 fatty acids: DHA and EPA, natural pigments (betacarotene and astaxanthin) and glucans. Algal biomass can also be used as a nutritional complement (Gong & Bassi, 2016).

207 2.2. Fungi and yeast

208 Like algae, fungi are also not new to human diets. Mycoproteins, more specifically, were 209 first discovered in the early 1960's (Derbyshire & Delange, 2021). Since then, many studies 210 have been done to assess the safety and benefits of this type of proteins. When talking about 211 fungi protein, this type of food includes the fruiting bodies of edible mushrooms, as well as 212 several species of micro fungi such as molds and yeasts, and their derivatives. Recently, 213 research has been focused on the production and characterization of vegetative mycelia from 214 fungi to increase its protein content and further processing to obtain meat alternatives for human 215 consumption (Schweiggert-Weisz et al., 2020).

216 There are a number of advantages of using fungi as a food source, primarily the low land 217 requirements since they can grow in bioreactors with high metabolic rates, which avoids the 218 extensive use of land needed for growing and feeding animals for meat. Production of 219 mycoproteins in bioreactors is generally done based on submerged fermentation, with fungi 220 growing in liquid media containing its nutritional requirements. Another advantage of using 221 fungi as a food source is that their single-cell proteins can provide other nutrients to the human 222 diet, including different B-complex vitamins (Sharif et al., 2021). However, there are still some 223 challenges to be overcome to allow broader use of mycoprotein as a food source, mainly related

to the production costs. Further research is also needed to evaluate safety issues and also to
spread its benefits to the public (Schweiggert-Weisz et al., 2020). Recently, metabolic
engineering technologies have been proposed to modify microorganisms to obtain, for example,
an improved utilization of agro-industrial residues with simultaneous production of SCP
(Hülsen et al., 2018). However, the use of genetically modified organisms (GMO) does not
have public acceptance and is still a topic of discussion (Sharif et al., 2021).

Filamentous fungi are the preferred choice in SCP production at large scale. The fungus is grown in a synthetic medium and further mixed with egg albumin and other compounds to confer color and flavor, mimicking meat. In addition, using filamentous fungi has an advantage over plant-derived meat substitutes, as fungi produces filaments comparable to meat fibrils, conferring a similar meat texture to the product (Gmoser et al., 2020).

Yeasts, however, have been in the market for a long time. The production of SCP using 235 236 yeasts had an expressive significance in the war times. During the First World War, Germany 237 managed to substitute almost half of its imported protein sources by yeast. Initially, they used 238 brewer's yeast, but it was not enough to meet the demand as a protein source. In the beginning 239 of the Second World War, yeasts were used as a protein source in both army and civilian diets (Ugalde & Castrillo, 2002). Today, yeasts are often used as supplements in animal feed and in 240 241 vegetarian diets. Fungi, including the yeasts market, is the second largest single-cell protein 242 market after algae. Most of the SCP is still destined for the animal feed market, but human 243 consumption has been growing in recent years.

It is worth noting that yeasts have various benefits over bacteria in their manufacturing process, for example, they are larger than bacteria (cell size), and harvesting them from culture media is easy. Yeasts also have higher lysine and malic acid contents, although their protein content is usually lower, and they also have longer doubled times compared to bacteria (Raziq, 2020). 249

250 2.3. Bacteria

251 Bacteria have also been used as SCP for a long time, mainly for animal feed. Single-cell 252 protein derived from bacteria usually contains between 50 and 80% of protein (dry basis), and 253 the amino acid content is higher or similar to the FAO recommendations. Like fungi, bacteria 254 also have a high nucleic acid content, requiring previous processing (Strong et al., 2015). On 255 the other hand, bacteria have some advantages regarding the production process, such as faster 256 growth and shorter generation time when compared to fungi and yeasts. Additionally, they can 257 grow on several types of substrates, even in gaseous ones like hydrocarbons (Anupama & 258 Ravindra, 2000; Mussatto et al., 2021). In fact, using gases like CO₂, or diverse raw materials, 259 mainly waste/side streams from other industries, for bacteria cultivation may be appealing from 260 the perspectives of cost and sustainability. However, they are more difficult to harvest from the 261 culture medium due to their smaller size, requiring multiple unit operations for their recovery. 262 Some bacteria also have complex nutritional requirements (Nasseri et al., 2011).

263 For the selection of new strains for large-scale production of SCP, multiple criteria must 264 be considered, including the complexity of nutrients requirements, fermentation performance, 265 genetic stability during the cultivation process and growth morphology, the composition of the 266 final product generated by each strain in terms of protein and other components, and the 267 complexity of the downstream process required for purification (Razig, 2020). A significant 268 concern related to the utilization of bacteria as SCP is the possibility of producing toxins, which 269 can be extracellular (exotoxins) or intracellular (endotoxins). Toxins may cause adverse effects 270 in both animals and humans. Therefore, toxins' production must be carefully assessed to avoid 271 problems with regulatory bodies (Ritala et al., 2017).

272

273 3. Composition and safety issues of SCP obtained from different microorganisms

Table 2 shows the average composition of SCP obtained from algae, fungi/yeasts and bacteria, focusing on their nutritional value. Some important parameters/limitations to be considered for the SCP application in human and animal nutrition are also presented.

The acceptance and interest of a particular species for food or feed application greatly depend on its composition, growth rate, and associated toxin production. SCP for human consumption or animal feed must be free from all kinds of pathogens, toxins, and contaminants (heavy metals or other metal compounds, hydrocarbons). In addition, they should not cause food allergies, skin reactions, gastrointestinal reactions, diarrhea, vomiting, and other diseases (Ugbogu & Ugbogu, 2016). Therefore, it is essential to use toxicological studies to evaluate the safety of any produced SCP before marketing the products.

284 The main anti-nutritional factor in SCP is the presence of a high concentration of nucleic 285 acids, which is usually more abundant in microbial proteins than in other conventional protein 286 sources. This is one of the main factors limiting the SCP application in the food sector (Nalage 287 et al., 2015). Most nitrogen in SCP is in the form of amino acids, while the rest is in the form 288 of nucleic acids, which is a key property of fast-growing microorganisms. High nucleic acid 289 content is a problem because purine compounds derived from RNA break down and increase 290 uric acid concentration in the serum, ultimately leading to kidney stones and gout formation. It 291 has also been reported that living cells of microbes should be inactive before consumption. 292 Using an unprocessed product before killing the active microbes increases the incidence of skin 293 and gastrointestinal infections that can cause nausea and vomiting (Sharif et al., 2021). Anti-294 nutritional factors of SCP, like an elevated presence of nucleic acids, can be eliminated by 295 applying physical and/or chemical treatments during processing (Dantas et al., 2016). Different 296 techniques for nucleic acid reduction have been proposed to make SCP suitable for food applications. Chemical (sodium chloride, ammonium hydroxide, sodium hydroxide) and 297

enzymatic (ribonuclease, deoxyribonuclease) treatments can be used to treat biomass, obtaining
nucleic acid concentrations below 2% (w/w) (Yadav et al., 2016).

300

Table 2

301 Certain microorganisms can also produce toxic substances such as mycotoxins and 302 endotoxins during the production of SCP (Sharif et al., 2021). In addition, some carcinogenic 303 substances can be produced when microbes undergo mutations during the processing and 304 formation of the final product, which can be toxic to both humans and animals. However, all 305 these problems can be avoided by carefully selecting the microorganism and optimizing the 306 fermentation protocol for the production of SCP. The use of an appropriate substrate is also 307 useful to obtain SCP more beneficial to health. Recently, bacterial SCP obtained by culturing 308 bacteria in methanol as a carbon source was evaluated for mutagenicity in five in vivo tests in 309 various mammalian test systems, and the results showed no evidence of mutagenic activity due 310 to the substrate utilized for cultivation (Mahan et al., 2018; Spalvins et al., 2018).

311 Mycotoxin-generating fungi are undesirable sources of SCP as their toxins can cause 312 allergic reactions, carcinogenesis and even death in humans and animals. The fungus species 313 Aspergillus flavus, for example, produces aflatoxins of the B1, B2, G1 and G2 types, 314 Penicillium citrinum can produce citrine, trichothecenes and zearalenone, while Fusarium and 315 *Claviceps* species produce ergotamine. There is epidemiological evidence linking aflatoxins to 316 human liver cancer (Maiuolo et al., 2016). Recently, molecular biology techniques have been 317 explored to eliminate genes linked to mycotoxin synthesis. To isolate A. parasiticus and A. 318 *flavus* aflatoxin pathway clusters, the techniques of probing, cloning, expression libraries, 319 transcript mapping, and gene disruption have been applied. As an example, the aflR regulatory 320 gene, which controls the production of aflatoxins in Aspergillus, can become a target for 321 controlling the production of mycotoxins in this species. Although research in this field is still starting (Dubey et al., 2018; Xu et al., 2021), reliable and easily applicable techniques can be 322

expected in the near future. Some species of bacteria can also produce toxins, which limit their
use as SCP. *Methylomonas methanica* and *Pseudomonas* species, for example, produce
endotoxins that cause febrile reactions. However, heating can destroy these toxins (Mahan et
al., 2018; Ravindra et al., 2009).

According to the composition and potential limitations (**Table 2**), algal SCP has greater safety in terms of nucleic acid content and no toxin production compared to fungi and bacteria. In this way, the order of preference for food and feed application could be proposed as algae > fungi > bacteria (Anupama & Ravindra, 2000; Nasseri et al., 2011). However, this is a very general classification criteria, and studies should be done on each microorganism of interest to elucidate its potential to be used as SCP in food and feed applications.

333 4. Technology for SCP production

SCP can be produced by submerged, semi-solid, or solid-state fermentation. The process 334 335 for SCP production follows the steps shown in Figure 1. The first step consists of a screening 336 of potential microbial strains. This step is essential for an adequate selection of microorganisms 337 capable of producing a good amount of protein. Microbial strains can be isolated from different 338 habitats such as water, air, soil or other biological materials. The best strain can also be 339 optimized if necessary by mutation, selection and/or genetic protocols. The next step is the 340 choice of raw materials to be used for the bacterium cultivation, which is necessary to obtain 341 an appropriate composition of carbon, phosphorus, and nitrogen able to favor a high biomass 342 formation in a short time. The most desirable carbon sources are those containing 343 monosaccharides and disaccharides, as they are ready to use. The third step involves process 344 engineering and process optimization. At this stage, the best growing conditions for the selected 345 strain are determined and the metabolic pathways are elucidated (Nasseri et al., 2011; Ritala et 346 al., 2017; Singh et al., 2019; Ukaegbu-Obi, 2016). Then, the next step is developing the 347 technology, which consists of defining all the technical details and performance of the process

348 to make the production robust for large-scale application. Studies of economic factors make up 349 the next stage, where energy consumption and production costs are considered for the 350 implementation of the large-scale production process. Such analysis can also be done in parallel 351 or integrated with the technology development in the fourth step. Finally, attention should be 352 given to safety and environmental protection requirements. Since the single-cell protein will be 353 used for human or animal food, the product must have high safety, as some microorganisms 354 can also produce toxins that cause side effects to humans as well as to the environment. In this 355 way, the entire process must be properly monitored. Product authorizations for particular 356 applications and legal protection of innovative processes and strains of microorganisms, namely 357 exploration licenses, are the legal and controlled aspects that the innovation requires (Ritala et 358 al., 2017; Singh et al., 2019; Ukaegbu-Obi, 2016).

359

Figure 1

360 Process optimization for fermentation is a very important step in the processing of single 361 cell protein production since it should be able to result in high product yield. In this sense, an important aspect to consider is the medium's composition to be used for fermentation, which 362 363 must contain all the nutrients necessary for appropriate growth of the microorganism (Bellamy, 2009; Kadim et al., 2015). For fungi cultivation, for example, the fermentation medium must 364 365 include nutrients such as potassium, phosphorus, magnesium, trace elements, ammonium salts 366 and vitamins (like biotin), which are needed to develop mycelia (Gervasi et al., 2018). In 367 addition, the fermentation process requires a pure culture of the chosen organism, sterilization 368 of the growth medium, and a fermenter operated under suitable conditions to favor the microbial 369 growth (Nasseri et al., 2011). SCP can be produced by solid-state, semi-solid or submerged 370 fermentation. From these options, the submerged system usually results in higher production 371 efficiency (Hashempour-Baltork et al., 2020; Suman et al., 2015). At the end of the fermentation, the resulting biomass is collected by filtration or centrifugation and goes through
washing and drying steps to obtain the SCP (Figure 2).

374

Figure 2

375 Submerged fermentations are usually more expensive, leading to higher operational costs than 376 semi-solid or solid-state fermentations, giving lower protein yields. An alternative to reduce the 377 costs of submerged fermentations is to use food wastes, which have low value or no value at 378 all, to formulate the fermentation medium. Food wastes and a variety of cheap raw materials, 379 including agricultural wastes, have already been tested as substrates for the production of SCP 380 by different microorganisms, including orange peel, sugarcane bagasse, rice husk, wheat straw, 381 cassava residues, sawdust, corn cobs, coconut residues, mandarin residues, beet pulp, among 382 others (Bellamy, 2009; Diwan et al., 2018; Reihani & Khosravi-Darani, 2019; Ritala et al., 383 2017). Of course, the SCP production yields vary according to the substrate and microorganism 384 used. The use of banana peel as a substrate for the production of Aspergillus niger biomass, for 385 example, gave better yields (biomass yield of 2.29 and protein yield of 0.57 g/L) than crop 386 wastes like cucumber, orange, and pineapple waste (Oshoma & Eguakun-Owie, 2018). Also, 387 the use of pea processing by-products resulted in the production of A. oryzae var. oryzae CBS 388 819.72 mycobiomass with 38% more protein compared to the mycobiomass obtained from 389 synthetic medium (Souza Filho et al., 2018). Vinasse, the final residue obtained from the 390 production of bioethanol, can also be used for the production of SCP. Candida parapsilosis was 391 successfully cultivated in a medium containing 5 g/L of peptone and 70% v/v of vinasse (dos 392 Reis et al., 2019). The bioconversion of cheese whey is another appealing method for SCP 393 production since lactose can be used as a carbon source by different microorganisms, including 394 veasts of the genus Kluvveromvces (Coelho Sampaio et al., 2016; Dragone et al., 2009). It is 395 worth noting that the use of lignocellulosic materials, particularly, as substrates for the 396 production of SCP, is hindered by the complex structure of these materials, which is mainly

composed of cellulose, hemicellulose and lignin fractions. Thus, pre-treatment methods have
become essential to break the resistant lignin layer, reducing the crystallinity of cellulose and
increasing the availability of carbohydrates to be consumed by microorganisms (Mussatto &
Dragone, 2016; Sun et al., 2015; H. Zhang et al., 2021).

Tropea et al., (2022) utilized mixed food waste (fish, pineapple, banan, apple, citrus peel etc.) as a substrate to produce SCP by using *Saccharomyces cerevisiae* and observed the that the final protein concentration reached uo to 40.19% after 120 h of fermentation. While the true protein percentage was 10.86%. SCP production from pineapple waste, studied by Aruna (2019) and Tropea et al. (2015), observed the highest crude protein yields of 13.56% and 17.2%, respectively.

In a different study, certain food wastes (banana peel, citrus peel, carrot pomace, and potato
peel) were utilized for the production of SCP using yeast (isolated from durien fruit), and it was
reported that the crude protein yield was increased from 14.07 % (before fermentation) to
30.82% (after fermentation) (Chun et al., 2020).

It was investigated how well some industrially important microbes (kefir, *K. marxianus*, and *S. cerevisiae*) grew on substrates obtained from several common food industry wastes (whey, molasses, brewer's solid wastes, orange, and potato residues) during SSF (Aggelopoulos et al., 2014). Among all the three varieties, the highest protein content (38.5% w/w) of SCP was observed with *S. cerevisiae* AXAZ-1. The protein content observed in fermented biomasses of *S. cerevisiae* AXAZ-1 (38.5% w/w) and *K. marxianus* IMB3 (33.7%) were observed two times higher than their corresponding substrates before treatment (20.9-22.9%).

In summary, various materials and wastes can be used as a substrate for producing SCP. However, the material to be used for this application should meet some criteria, for example, they must be non-toxic, regenerable, abundant, and inexpensive. So, besides contributing to reducing production costs, the use of organic wastes as a carbon source for SCP production also helps with a more valuable solution for eliminating such residues (Reihani & Khosravi-Darani,
2019; Srivastava et al., 2011).

Table 3 summarizes different fermentation parameters related to the production of SCP from algae, fungi and bacteria. The growth rate, the substrate used for production (which will also impact the costs of the process), and risks of contamination are important aspects to consider for developing an SCP production technology. It is also of fundamental importance to optimize the fermentation process for each type of substrate and microorganism in order to maximize the production yield (Reihani & Khosravi-Darani, 2019; Sharif et al., 2021).

430

Table 3

431 **5. Protein recovery and purification**

The downstream process for protein recovery will have small variations according to the fermentation system and conditions used, microorganisms employed, and fermentation media used, among others.

435 5.1. Biomass recovery

436 Mechanical separation technologies allow the fermentation products to be recovered from 437 the fermentation broth. In submerged fermentation, for example, both the substrate to be 438 fermented and biomass produced are present in the liquid medium. The biomass can be 439 harvested continuously in this system, being then filtered or centrifuged and dried to obtain the 440 SCP. In the solid-state fermentation, biomass can be recovered by using less steps than in semi-441 solid-state fermentation. Because the later uses a higher amount of free water than solid-state 442 fermentation, it requires facile processing to obtain microbial biomass. The abysmal amount of 443 water in solid-state cultivation makes the downstream processing much more efficient, but heat dissipation from the biomass poses a critical problem. While the microbial biomass in semi-444 445 solid-state fermentation can be recovered by centrifugation, solid-state fermentation requires 446 special scrappers or mechanical systems to remove the biomass from the fermented slurry.

447 Single-cell from bacteria and yeasts are usually recovered by centrifugation, while 448 filamentous fungi are recovered by filtration (Ritala et al., 2017). Centrifugation is an energy-449 intensive process; however, such energy requirements can be offset by the high value of the 450 final product (Sheng et al., 2017). Membrane technology has been considered more favorable 451 than centrifugation for microalgae biomass collection due to its lower energy consumption, 452 being generally more cost-effective, and offering full biomass recovery (Zhang et al., 2019).

453

454 5.2. Cell disruption

To isolate an intracellular protein, cell membrane must be disrupted to release the cell 455 456 contents, which can be done by using a suitable lysis buffer to achieve high solubilization of 457 the target protein (Hernández et al., 2018). It is possible to use SCP as a complete cell preparation; however, breaking the cell makes the protein more accessible. Several methods 458 459 can be used to break the cell wall, including mechanical forces (crushing, crumbling, crushing, 460 pressure homogenization or ultrasound), hydrolytic enzymes (endogenous or exogenous), 461 chemical disruption with detergents, or even combinations of these methods (Nasseri et al., 462 2011).

463

464 5.3. Protein secretion

Several methodologies, such as precipitation, extraction, and filtration, have been developed to recover proteins from biological systems (McDonald et al., 2009). These methods have different advantages and disadvantages. Precipitation, for example, involves the adjustment of the physical properties (pH, salt and heat treatment) of the medium to improve the insolubility of proteins (Tovar Jiménez et al., 2012), being an easily scalable and low-cost method. However, thermal precipitation has the disadvantage of changing the structural characteristics of native proteins, which can affect their functional and nutritional value (Yadav

et al., 2014). Protein extraction is an important biological process to recover protein from the grown microorganism with desired purity. However, the extraction of protein from filamentous fungi is a complex and cumbersome process due to the presence of a chitinous cell wall. Several processing steps include the extreme conditions (pH, temperature, pressure, and requirement of solvents, among others) required to extract the protein. However, in the case of the use of microbial proteins in food and feed applications, extraction of protein employing these techniques is not required.

479 Pressure-driven membrane processes (microfiltration and ultrafiltration) and direct 480 osmosis membrane processes have also been explored for SCP collection due to their high 481 efficiency, ease of operation and scalability (Ye et al., 2018). Microfiltration membranes are 482 commonly used as a clarification step to ultrafiltration and nanofiltration processes, as well as 483 for cold sterilization of liquid foods and pharmaceuticals, and can also be used to fractionate 484 large macromolecules from smaller ones, such as casein from whey fractionation in the dairy 485 industry. In the biotech and pharmaceutical industries, microfiltration membranes offer the safety of a physical barrier to remove bacteria and other microbes (Tijing et al., 2020). On the 486 487 other hand, ultrafiltration has been more applied to eliminate organic substances from 488 wastewater treatment and in the textile industry. In ultrafiltration, the ultrafilter is supported 489 over a wire mesh, and the impure sample is poured over it. The impurity particles (electrolytes) 490 pass through the ultrafilter while the larger colloidal particles are retained (Tovar Jiménez et 491 al., 2012). This process is usually slow, but it can be accelerated by applying pressure or by 492 using a suction pump on the filtrate side (Tijing et al., 2020). Commercial whey protein is 493 purified through microfiltration and ultrafiltration processes, which avoid denaturation of the 494 protein. These methods are performed at low temperatures, removing the impurities in the whey 495 (fats and sugars) and producing a very high-quality protein.

It is worth noting that membrane contamination significantly decreases water 496 497 permeability and the overall system performance. In this sense, several techniques have been 498 reported to control/avoid membrane contamination, including feed pretreatment, system 499 operation below the critical flow, backwash, ultrasonic cleaning, chemical cleaning, and 500 mopping with air bubbles (Liao et al., 2018). In addition, most published studies use 501 commercial membranes, which are also used in the food industry. However, developing a 502 custom membrane to collect a specific type of microorganism would be very feasible 503 considering the diversity of species that can be used for SCP production, which have different 504 cell sizes (Lau et al., 2020). Moreover, coupling a good inlay control system with a suitable 505 membrane could also offer substantial performance advantages (Discart et al., 2015).

506

507 5.4. Purification

508 Chromatographic methods, including gel permeation, hydrophobic interaction, and 509 affinity chromatography, can be used for SCP purification. Such methods are easy to perform 510 but are complex to optimize due to the numerous parameters that need to be considered for this 511 process (Wingfield, 2015). For example, the choice of the column matrix, the buffer to be used, 512 the salt, the organic solvent, the reaction temperature, and the gradient are some of the 513 parameters to be considered for an efficient chromatography process. Chromatographic 514 methodologies are very popular for separating and purifying whey proteins, being used even 515 for large-scale protein separation (Bonnaillie et al., 2014).

516 5.5. Drying

517 Several techniques have been reported to dry microbial proteins and obtain powdered 518 proteins with desirable characteristics for application on an industrial scale. The mostly used 519 techniques are freezing-drying and spray-drying (Maltesen & van de Weert, 2008). Recently, 520 supercritical drying has emerged as another viable alternative for obtaining powdered proteins.

521 These techniques are based on three physical principles: sublimation, evaporation, and 522 precipitation (Son et al., 2020). Of course, different techniques use different stresses that can 523 compromise the final stability of the proteins. In addition, different techniques result in protein 524 powders with significantly different characteristics (Raziq, 2020). This fact should be 525 considered when focusing on the desired characteristics of the final product.

526

527

6. Industrial scenario and market of SCP

528 Currently, consumers demand healthy food and, at the same time, are concerned about 529 the environment. To meet a demanding and competitive market, manufacturers have 530 continuously innovated their production process using different raw materials and developing 531 more sustainable technologies to attract customers. Through iterative and incremental 532 methodologies, companies are creating innovative solutions to their existing products within a 533 constant innovation cycle to reduce toxic waste and the content of nucleic acids in their products 534 for a better human consumption, which, in the end, signals sales growth opportunities in the 535 food and beverage industries. Lallemand Inc., Montreal, Canada, for example, has been 536 developing innovative microbial products through external research partnerships with important universities and their own internal projects to create new and healthier products 537 538 (Hülsen et al., 2018; Matos, 2019).

539 The SCP production process can contribute to the environment's safety by reducing the 540 carbon footprint and using wastes/renewable resources as carbon sources for fermentation. Due 541 to this positive aspect, the high demand for healthy products and alternative protein sources, 542 the SCP market is expected to expand significantly from 2020 to 2030 (Banovic et al., 2018; 543 Hülsen et al., 2018; TMR, 2021). SCP has applications in food products as an important source 544 of proteins and vitamins and has also been used to improve the nutritional value of products 545 such as soups, baked goods, ready-to-serve meals, in diet recipes, among others. For animal

nutrition, SCP is used for fattening calves, pigs, poultry and fish. SCP is also used to increase
the nutritive value of soups, baked products and specialized diets. Besides the main applications
of SCP in food and feed products, it is also applied in the leather and paper processing industries
and as a foam stabilizer (Kumar et al., 2017; Zakaria et al., 2020).

550 The global SCP market is segmented into North America, Latin America, Western 551 Europe, and Pacific Asia (excluding Japan, the Middle East and Africa). A recent market study 552 indicated that the SCP market in Malaysia was at US\$ 9.7 million in 2020 and is expected to 553 reach US\$ 24.5 million in 2030 at an annual growth rate (CAGR) of 9.7%. Vietnam's SCP 554 market revenue was valued at over US\$ 26.7 million in 2020 and is expected to exceed US\$ 69.4 million by the end of 2030 (Khoshnevisan et al., 2020; TMR, 2021). The Asia region is 555 556 also made up of other prominent countries in the Association of Southeast Asian Nations 557 (ASEAN), such as India, China, Indonesia and Bangladesh. According to the Food and 558 Agriculture Organization of the United States, these countries produce about 50-60% of the 559 total aquaculture production. SCP is the most nutritious and cost-effective option for fishmeal, 560 and global growth is expected through the expansion of the aquaculture industry (Jones et al., 561 2020; Matos, 2017; Ritala et al., 2017).

562 North America is a global leader in the global SCP market due to the region's highly 563 developed food and feed industries. North America is favored by some prominent organizations 564 related to the food and feed industries, which allows a greater manufacturing and development 565 capacity for SCP production. Furthermore, the majority of individuals who adopt a high-protein 566 diet are in North America, which makes it possible to increase the market value share of the 567 one-time-only protein in the region (Nasseri et al., 2011; Spalvins et al., 2018). European 568 consumers are concerned about increasing the protein content in their food products as well as 569 in finding alternative protein sources (Banovic et al., 2018). Furthermore, the increased support 570 in sustainability and actions against animal cruelty in the region has also contributed to a better

571 market scenario for non-animal-based protein sources. SCP is also gaining significant attraction 572 in Latin American countries. However, the major share of the population relies on animal 573 protein because of the culture and availability of grazing lands, feeding crops for animals. 574 Nevertheless, the rising awareness about high-quality protein and the right nutrition, food -575 changing patterns, vegan proteins, and SCP is also getting sizeable attraction.

576 The main players in the SCP market, their respective countries and their segments are shown in Table 4. These companies are focused on business growth and innovation to 577 578 strengthen their positions in the global SCP market. Developing new products and strategic 579 collaborations are other approaches that the key players are considering to gain a competitive 580 advantage in the global SCP market. Angel Animal Nutrition, for example, launched an 581 innovative product called GroPro, which is a yeast-derived feed ingredient composed of 582 proteins necessary for the development of young animals. Afterwards, they launched a 583 completely innovative semi-dry yeast in the market in the form of a tetra pack, which is easy to 584 use and hygienic due to its reusable upper opening. This yeast product has about 20% moisture, 585 with characteristics of dry and fresh yeast (Ritala et al., 2017; TMR, 2021).

586

Table 4

587 Besides the big companies, startups are also increasingly investing in innovation, from 588 microalgae supplements for athletes to ice cream based on probiotic bacteria. For example, 589 Noko Foods, a French startup recently founded in 2021, develops herbal microalgae 590 supplements, drink shakes and food products for athletes. In addition, the startup Ninoko Labs, 591 founded in 2020 in Germany, produces alternative proteins from fungal mycelium in order to 592 compete with real meat in terms of cost and flavour. Another example of a biotechnology 593 company focused on innovation for human food is Bidifice Inc., Santiago, China which 594 develops ice cream rich in healthy probiotic bacteria to help people with chronic diseases and allergies (Bratosin et al., 2021; Sally Ho, 2021). 595

596

597 **7. Regulatory aspects**

598 SCP used as food or feed must be safe to produce and use. In most countries, there are 599 regulations to certify that food or feed is safe for human consumption. Generally, these 600 regulations differentiate between human food and animal feed, food that provides nutrition and 601 potentially flavor and aroma, and food additives such as colorants, preservatives, or feed 602 additives. In addition, although definitions differ among regions, international standards 603 regulated by the Joint FAO/WHO Expert Committee on Food Additives apply to internationally 604 traded products (Kannan et al., 2020; Ritala et al., 2017; Sharif et al., 2021).

605 Although the final SCP product is a protein and nutritional source, certain products may 606 enter the market as additives, providing, for example, color rather than SCP, which restricts the 607 extent to which they are added and their value as SCP. Therefore, the regulations differ 608 depending on the application (Zepka et al., 2010). Also, Smedley (2013) reported similarities 609 and differences among 7 jurisdictions (Canada, European Union, Brazil, China, Japan, United 610 States and South Africa) in terms of regulation of authorized food ingredients, as well as the 611 approval and management assessment process for feed components, and peculiarities between 612 regulations in these regions. In addition, as animals are not all the same in all regions, the 613 regulations for feeding pets differ in certain regions, requiring authorization before selling new 614 pet foods or additives.

It is worth noting that the final SCP product must not only be nutritious but also pass all toxicity tests to be marketed as a food product. In addition, the unwanted nucleic acid content, toxins and unwanted compounds that accumulate during the strain cultivation using substrates, such as hydrocarbons and petroleum contaminated with heavy metals, need to be removed (Gervasi et al., 2018). Decontamination and purification of the final product are essential for SCP to be used as a food source for consumption. 621

622 8. Role of microbial protein in the circular economy

623 Recently, the concept of circular economy, which implies the transformation of wastes 624 and industry side-streams to produce renewable energy and added-value compounds, has been 625 strongly encouraged to design a more sustainable economy (Dragone et al., 2020; Stiles et al., 626 2018). Microbial protein is becoming a potential product for incorporation in a circular 627 economy model due to its increased interest as an alternative protein source and numerous 628 applications in food, chemicals and pharmaceuticals (Lai et al., 2019). Microbial biomass, particularly microalgae, can also be recycled and used as a biofertilizer to sustainably improve 629 630 soil quality and crop nutrition (Abo et al., 2019).

631 In recent years, the global need to find alternative protein sources has driven the 632 development of new SCP processes. Using readily available raw materials and waste streams 633 as a substrate for SCP production is also a relevant driver to develop new processes. In this 634 sense, SCP production can fortify biorefineries' economic feasibility, besides being a 635 sustainable option for managing residual raw materials and wastes (Mahan et al., 2018). 636 Concerns about environmental pollution have also driven the development of new SCP 637 production processes. This can be seen especially in processes that have applied greenhouse 638 gases as a substrate, for example, the production of SCP using CO₂ or methane as a carbon 639 source. Although there are still important challenges to overcome for large-scale and 640 economically feasible implementation of these new processes using gases as substrate, they 641 have attracted great interest from a sustainability point of view (Puyol et al., 2017; Ritala et al., 642 2017; Ukaegbu-Obi, 2016).

Investment and profitability are key elements in estimating the economic viability of an
SCP production process. For large-scale SCP production, large bioreactors are required. Thus,
high oxygen transfer rates are needed to obtain a high amount of biomass during the cultivation,

646 which may cause an increased generation of heat from microbial metabolism that will lead to 647 the need for temperature control and reduction. In fact, operating costs, including labor, 648 consumables and energy, represent 45-55% of SCP manufacturing costs, while raw material 649 costs range from 35-55%. Using cheap raw materials and/or waste streams as carbon sources is 650 a good strategy to reduce substrate costs, as long as they do not compromise the quality of the 651 final product (Rodrigues, 2020). Finally, there is a relationship between cost and production 652 scale. Most of the SCP processes practiced on an industrial scale were set for a continuous 653 design, which proved to be the most profitable option (Poutanen et al., 2017; Ritala et al., 2017).

After everything that was presented and discussed in this work, the research brought here on microbial biomasses, showed that SCPs gained momentum, due to the increased demand for alternatives to proteins derived from plants and animals. In this way, certain products can become popular due to consumer acceptance and significant encouragement from government and regulatory authorities. The literature also made it possible to verify a diversity of possibilities of microorganisms capable of producing SCPs, including microalgae, fungi, bacteria and cyanobacteria.

661

662 9. Conclusions and future prospects

663 Microbial engineering has a relevant ability to enhance the competitiveness of the SCP product in terms of production cost, functionality and nutrition. The application of GRAS 664 665 microorganisms is considered safe and is always the right choice in microbial engineering for 666 SCP production, with the main objective of improving the production of intermediate raw 667 materials and the accumulation of biomass. Future research and promotion of meat-optional 668 protein sources is a major challenge. The judicious utilization of agri-residues and by-products 669 of agriculture and food processing for the cultivation of filamentous fungi, yeasts, bacteria and 670 microalgae, would allow in developing SCP in a sustainable manner. Furthermore, studies

671 aiming to correlate the consumption of alternative sources of protein and gains for human health 672 would very possibly increase consumers' attention to a more sustainable diet. In the near future, 673 it is expected that the development of new processes for SCP production using residual raw 674 materials, industry-side streams or even greenhouse gases (CO₂ or methane) as carbon sources 675 will become a reality to increase the protein market without affecting the environment and 676 potentially with a low production cost. This review can be useful for the start-ups to create new 677 products or processes by combining fermentation technologies and alternative meat protein 678 sources. Indeed, the sensory attributes and nutritional value of meat alternative foods can be 679 improved by fermentation with selected microorganisms.

680

ACKNOWLEDGEMENTS: A. K. Chandel gratefully acknowledges the financial support
from CNPq for scientific productivity program (Process number: 309214/2021-1). S. I.
Mussatto acknowledges the Novo Nordisk Foundation (NNF), Denmark, grant number
NNF20SA0066233. V. K. Gupta and V. K. Thakur would like to acknowledge the institutional
funding sources supported by the SRUC, UK for research and development.

686 AUTHOR CONTRIBUTIONS

Conceptualization, investigation, and writing—original draft: Samara Cardoso Alves, Erick
Díaz Ruiz and Bruna Lisboa. Writing— review and editing: Minaxi Sharma, Solange I.
Mussatto, Vijay Kumar Thakur, and Deepk M. Kalaskar. Conceptualization, writing— review
and editing: Vijai K. Gupta. Conceptualization, data curation, supervision, and writing—review
and editing: Anuj K. Chandel.

692 CONFLICTS OF INTEREST

693 The authors declare no conflicts of interest.

694 **REFERENCES:**

- 695 U. S. FDA. (2017). Regulatory Framework for Substances Intended for Use in Human Food
- 696 or Animal Food on the Basis of the Generally Recognized as Safe (GRAS) Provision of the
- 697 *Federal Food, Drug, and Cosmetic Act: Guidance for Industry. November,* 28.
- Abo, B. O., Odey, E. A., Bakayoko, M., & Kalakodio, L. (2019). Microalgae to biofuels
 production: A review on cultivation, application and renewable energy. *Reviews on Environmental Health*, 34(1), 91–99. https://doi.org/10.1515/reveh-2018-0052
- 701 Aggelopoulos, T., Katsieris, K., Bekatorou, A., Pandey, A., Banat, I. M., & Koutinas, A. A.
- (2014). Solid state fermentation of food waste mixtures for single cell protein, aroma
 volatiles and fat production. *Food chemistry*, *145*, 710-716.
- Al-Mudhafr, A. W. (2019). Microbiological sources and nutritional value of single cell protein
 (SCP). *International Journal for Research in Applied Sciences and Biotechnology*, 6(6).
- Anderson, M. S., Muff, T. J., Georgianna, D. R., & Mayfield, S. P. (2017). Towards a synthetic
- nuclear transcription system in green algae: Characterization of Chlamydomonas
 reinhardtii nuclear transcription factors and identification of targeted promoters. *Algal*
- 709 *Research*, 22, 47–55. https://doi.org/10.1016/j.algal.2016.12.002
- (o) 100000000, 22, 17 00. https://doi.org/10.1010/j.u.Bui.2010.12.002
- Anupama, & Ravindra, P. (2000). Value-added food: Single cell protein. *Biotechnology Advances*, 18(6), 459–479. https://doi.org/10.1016/S0734-9750(00)00045-8
- 712 W., Anupong, Jutamas. K., On-Uma, R., Sabour, A., Alshiekheid, М., 713 Karuppusamy, I.,...&Pugazhendhi, A. (2022). Sustainable bioremediation approach to treat 714 the sago industry effluents and evaluate the possibility of yielded biomass as a single cell 715 protein (SCP) using cyanide tolerant Streptomyces tritici D5. Chemosphere, 135248.
- Aruna, T.E. (2019). Production of value-added product from pineapple peels using solid state
 fermentation. *Innov. Food Sci. Emerg. Technol.*, *57*, 102193.
- 718 Banovic, M., Lähteenmäki, L., Arvola, A., Pennanen, K., Duta, D. E., Brückner-Gühmann, M.,

- & Grunert, K. G. (2018). Foods with increased protein content: A qualitative study on
 European consumer preferences and perceptions. *Appetite*, *125*, 233–243.
 https://doi.org/10.1016/j.appet.2018.01.034
- Balagurunathan, B., Ling, H., Choi, W. J., & Chang, M. W. (2022). Potential use of microbial
 engineering in single-cell protein production. *Current Opinion in Biotechnology*, 76,
 102740.
- Bellamy, W. D. (2009). Production of single-cell protein for animal feed from lignocellulose
 wastes. *World Animal Review*, 4–9.
- Berners-Lee, M., Kennelly, C., Watson, R., & Hewitt, C. N. (2018). Current global food
 production is sufficient to meet human nutritional needs in 2050 provided there is radical
 societal adaptation. *Elementa*, 6. https://doi.org/10.1525/elementa.310
- Bonnaillie, L. M., Qi, P., Wickham, E., & Tomasula, P. M. (2014). Enrichment and purification
 of casein glycomacropeptide from whey protein isolate using supercritical carbon dioxide
 processing and membrane ultrafiltration. *Foods*, 3(1), 94–109.
 https://doi.org/10.3390/foods3010094
- Bonny, S. P. F., Gardner, G. E., Pethick, D. W., & Hocquette, J. F. (2015). What is artificial
 meat and what does it mean for the future of the meat industry? *Journal of Integrative Agriculture*, *14*(2), 255–263. https://doi.org/10.1016/S2095-3119(14)60888-1
- Bratosin, B. C., Darjan, S., & Vodnar, D. C. (2021). Single cell protein: A potential substitute
 in human and animal nutrition. *Sustainability (Switzerland)*, *13*(16), 1–24.
 https://doi.org/10.3390/su13169284
- 740 Chun, N. C. W., Ismail, A. F., Makhatar, M. M. Z., Jamaluddin, F. M. N., Tajarudin, H. A.
- 741 (2020). Conversion of food waste via two-stage fermentation to controllable chicken feed
- nutrients by local isolated microorganism. Int. J. Recycl. Org. Waste Agric., 9, 33–47.
- 743 Choudhury, D., Tseng, T.W., Swartz E (2020). The business of cultured meat. Trends

744

Biotechnol. 38, 573-577.

- 745 Coelho Sampaio, F., da Conceição Saraiva, T. L., Dumont de Lima e Silva, G., Teles de Faria,
- 746 J., Grijó Pitangui, C., Aliakbarian, B., Perego, P., & Converti, A. (2016). Batch growth of
- 747 Kluyveromyces lactis cells from deproteinized whey: Response surface methodology
- 748 versus Artificial neural network-Genetic algorithm approach. *Biochemical Engineering*
- 749 *Journal*, 109, 305–311. https://doi.org/10.1016/j.bej.2016.01.026
- 750 Dantas, E. M., Valle, B. C. S., Brito, C. M. S., Calazans, N. K. F., Peixoto, S. R. M., & Soares,
- 751 R. B. (2016). Partial replacement of fishmeal with biofloc meal in the diet of postlarvae of
- the Pacific white shrimp Litopenaeus vannamei. *Aquaculture Nutrition*, 22(2), 335–342.
- 753 https://doi.org/10.1111/anu.12249
- De Oliveira, M. A. C. L., Monteiro, M. P. C., Robbs, P. G., & Leite, S. G. F. (1999). Growth
 and chemical composition of Spirulina maxima and Spirulina platensis biomass at
 different temperatures. *Aquaculture international*, 7(4), 261-275.
- Derbyshire, E. J., & Delange, J. (2021). Fungal Protein What Is It and What Is the Health
 Evidence? A Systematic Review Focusing on Mycoprotein. *Frontiers in Sustainable Food Systems*, 5(February). https://doi.org/10.3389/fsufs.2021.581682
- 760 Discart, V., Bilad, M. R., Moorkens, R., Arafat, H., & Vankelecom, I. F. J. (2015). Decreasing
- membrane fouling during Chlorella vulgaris broth filtration via membrane development
 and coagulant assisted filtration. *Algal Research*, 9, 55–64.
 https://doi.org/10.1016/j.algal.2015.02.029
- 764 Diwan, B., Parkhey, P., & Gupta, P. (2018). From agro-industrial wastes to single cell oils: a
- step towards prospective biorefinery. *Folia Microbiologica*, 63(5), 547–568.
 https://doi.org/10.1007/s12223-018-0602-7
- dos Reis, K. C., Coimbra, J. M., Duarte, W. F., Schwan, R. F., & Silva, C. F. (2019). Biological
- treatment of vinasse with yeast and simultaneous production of single-cell protein for feed

- supplementation. International Journal of Environmental Science and Technology, 16(2),
- 770 763–774. https://doi.org/10.1007/s13762-018-1709-8
- 771 Dragone, G., Kerssemakers, A. A. J., Driessen, J. L. S. P., Yamakawa, C. K., Brumano, L. P.,
- 8 Mussatto, S. I. (2020). Innovation and strategic orientations for the development of
- advanced biorefineries. *Bioresource Technology*, 302(January), 122847.
- 774 https://doi.org/10.1016/j.biortech.2020.122847
- Dragone, G., Mussatto, S. I., Oliveira, J. M., & Teixeira, J. A. (2009). Characterisation of
 volatile compounds in an alcoholic beverage produced by whey fermentation. *Food Chemistry*, *112*(4), 929–935. https://doi.org/10.1016/j.foodchem.2008.07.005.
- Duarte, L. C., Carvalheiro, F., Lopes, S., Neves, I., & Gírio, F. M. (2007). Yeast biomass
 production in brewery's spent grains hemicellulosic hydrolyzate. In *Biotechnology for Fuels and Chemicals* (pp. 637-647). Humana Press.
- 781 Dubey, M. K., Aamir, M., Kaushik, M. S., Khare, S., Meena, M., Singh, S., & Upadhyay, R. S.
- 782 (2018). PR Toxin biosynthesis, genetic regulation, toxicological potential, prevention
- and control measures: Overview and challenges. *Frontiers in Pharmacology*, 9(MAR), 1–
- 784 19. https://doi.org/10.3389/fphar.2018.00288
- El-Bakry, M., Abraham, J., Cerda, A., Barrena, R., Ponsá, S., Gea, T., & Sanchez, A. (2015).
- From wastes to high value added products: Novel aspects of SSF in the production of
- 787 enzymes. *Critical Reviews in Environmental Science and Technology*, *45*(18), 1999–2042.
- 788 https://doi.org/10.1080/10643389.2015.1010423
- Finco, A. M. de O., Mamani, L. D. G., Carvalho, J. C. de, de Melo Pereira, G. V., Thomaz-
- 790 Soccol, V., & Soccol, C. R. (2017). Technological trends and market perspectives for
- 791 production of microbial oils rich in omega-3. *Critical Reviews in Biotechnology*, 37(5),
- 792 656–671. https://doi.org/10.1080/07388551.2016.1213221.
- Gao, Y., Li, D., & Liu, Y. (2012). Production of single cell protein from soy molasses using

- 794 Candida tropicalis. Annals of microbiology, 62(3), 1165-1172.
- 795 Geada, P., Moreira, C., Silva, M., Nunes, R., Madureira, L., Rocha, C. M. R., Pereira, R. N.,
- 796 Vicente, A. A., & Teixeira, J. A. (2021). Algal proteins: Production strategies and 797 functional properties. Bioresource Technology, nutritional and *332*(March). https://doi.org/10.1016/j.biortech.2021.125125
- 798
- 799 Gervasi, T., Pellizzeri, V., Calabrese, G., Di Bella, G., Cicero, N., & Dugo, G. (2018).
- 800 Production of single cell protein (SCP) from food and agricultural waste by using
- 801 cerevisiae. Natural Product 32(6), 648-653. Saccharomyces Research,
- 802 https://doi.org/10.1080/14786419.2017.1332617
- 803 GLOBAL TRENDS. (2021). Demographics and human development. March, 2021.
- https://www.dni.gov/index.php/gt2040-home/gt2040-structural-forces/demographics-and-804
- 805 human-development
- 806 Gmoser, R., Fristedt, R., Larsson, K., Undeland, I., Taherzadeh, M. J., & Lennartsson, P. R.
- 807 (2020). From stale bread and brewers spent grain to a new food source using edible 808 filamentous Bioengineered, fungi. 11(1), 582-598.
- 809 https://doi.org/10.1080/21655979.2020.1768694
- 810 Gong, M., & Bassi, A. (2016). Carotenoids from microalgae: A review of recent developments.
- 811 Biotechnology 34(8), 1396–1412. Advances,
- 812 https://doi.org/10.1016/j.biotechadv.2016.10.005
- 813 Hartmann, C., & Siegrist, M. (2017). Consumer perception and behaviour regarding sustainable 814 protein consumption: A systematic review. Trends in Food Science and Technology, 61,
- 815 11-25. https://doi.org/10.1016/j.tifs.2016.12.006
- 816 Hashempour-Baltork, F., Khosravi-Darani, K., Hosseini, H., Farshi, P., & Reihani, S. F. S.
- 817 (2020). Mycoproteins as safe meat substitutes. Journal of Cleaner Production, 253,
- 818 119958. https://doi.org/10.1016/j.jclepro.2020.119958
- 819 Hernández, D., Molinuevo-Salces, B., Riaño, B., Larrán-García, A. M., Tomás-Almenar, C., &

820	García-González, M. C. (2018). Recovery of Protein Concentrates From Microalgal
821	Biomass Grown in Manure for Fish Feed and Valorization of the By-Products Through
822	Anaerobic Digestion. Frontiers in Sustainable Food Systems, 2(June), 1-11.
823	https://doi.org/10.3389/fsufs.2018.00028
824	Hülsen, T., Hsieh, K., Lu, Y., Tait, S., & Batstone, D. J. (2018). Simultaneous treatment and
825	single cell protein production from agri-industrial wastewaters using purple phototrophic
826	bacteria or microalgae – A comparison. Bioresource Technology, 254(January), 214–223.
827	https://doi.org/10.1016/j.biortech.2018.01.032
828	Jones, S. W., Karpol, A., Friedman, S., Maru, B. T., & Tracy, B. P. (2020). Recent advances in
829	single cell protein use as a feed ingredient in aquaculture. Current Opinion in
830	Biotechnology, 61(Table 1), 189–197. https://doi.org/10.1016/j.copbio.2019.12.026
831	Jurasz, J., Mikulik, J., Krzywda, M., Ciapała, B., & Janowski, M. (2018). Integrating a wind-
832	and solar-powered hybrid to the power system by coupling it with a hydroelectric power
833	station with pumping installation. <i>Energy</i> , 144, 549–563.
834	https://doi.org/10.1016/j.energy.2017.12.011
835	Kadim, I. T., Mahgoub, O., Baqir, S., Faye, B., & Purchas, R. (2015). Cultured meat from

muscle stem cells: A review of challenges and prospects. *Journal of Integrative Agriculture*, 14(2), 222–233. https://doi.org/10.1016/S2095-3119(14)60881-9.

- Kam, S., Kenari, A. A., & Younesi, H. (2012). Production of single cell protein in stickwater
 by Lactobacillus acidophilus and Aspergillus niger. *Journal of aquatic food product technology*, 21(5), 403-417.
- 841 Kannan, B., Rathipriya, A., Dhinakaran, A., & Hema, A. (2020). Single cell protein (scp)
- 842 production from Marine Lactobacillus spp. Indian Journal of Animal Research, 54(9),
- 843 1115-1119.https://doi.org/10.18805/ijar.B-3858.

844	Khan, M. K. I., Asif, M., Razzaq, Z. U., Nazir, A., & Maan, A. A. (2022). Sustainable food
845	industrial waste management through single cell protein production and characterization
846	of protein enriched bread. Food Bioscience, 46, 101406.

- Khoshnevisan, B., Tabatabaei, M., Tsapekos, P., Rafiee, S., Aghbashlo, M., Lindeneg, S., &
 Angelidaki, I. (2020). Environmental life cycle assessment of different biorefinery
 platforms valorizing municipal solid waste to bioenergy, microbial protein, lactic and
- succinic acid. Renewable and Sustainable Energy Reviews, 117(July 2019), 109493.
- 851 https://doi.org/10.1016/j.rser.2019.109493
- 852 Kornochalert, N., Kantachote, D., Chaiprapat, S., & Techkarnjanaruk, S. (2014). Use of
- 853 Rhodopseudomonas palustris P1 stimulated growth by fermented pineapple extract to treat
- latex rubber sheet wastewater to obtain single cell protein. *Annals of microbiology*, 64(3),
 1021-1032.
- Kumar, P., Chatli, M. K., Mehta, N., Singh, P., Malav, O. P., & Verma, A. K. (2017). Meat
 analogues: Health promising sustainable meat substitutes. *Critical Reviews in Food Science and Nutrition*, 57(5), 923–932. https://doi.org/10.1080/10408398.2014.939739.
- Kumar, P., Mehta, N., Abubakar, A. A., Verma, A. K., Kaka, U., Sharma, N., ... & Lorenzo, J.
- 860 M. (2022). Potential alternatives of animal proteins for sustainability in the food sector. *Food*
- 861 *Reviews International*, 1-26. https://doi.org/10.1080/87559129.2022.2094403.
- Kusmayadi, A., Leong, Y. K., Yen, H. W., Huang, C. Y., & Chang, J. S. (2021). Microalgae as
 sustainable food and feed sources for animals and humans Biotechnological and
 environmental aspects. *Chemosphere*, 271, 129800.
 https://doi.org/10.1016/j.chemosphere.2021.129800.
- Kurbanoglu, E. B., & Algur, O. F. (2002). Single-cell protein production from ram horn
 hydrolysate by bacteria. *Bioresource Technology*, *85*(2), 125-129.
- 868 Lai, Y. C., Chang, C. H., Chen, C. Y., Chang, J. S., & Ng, I. S. (2019). Towards protein

Journal Pre-proofs

- 869 production and application by using Chlorella species as circular economy. *Bioresource*
- 870 *Technology*, 289(May), 121625. https://doi.org/10.1016/j.biortech.2019.121625
- 871 Lau, A. K. S., Bilad, M. R., Nordin, N. A. H. M., Faungnawakij, K., Narkkun, T., Wang, D. K.,
- 872 Mahlia, T. M. I., & Jaafar, J. (2020). Effect of membrane properties on tilted panel
- 873 performance of microalgae biomass filtration for biofuel feedstock. *Renewable and*
- 874 Sustainable Energy Reviews, 120(March 2019), 109666.
- 875 https://doi.org/10.1016/j.rser.2019.109666
- 876 Liao, Y., Bokhary, A., Maleki, E., & Liao, B. (2018). A review of membrane fouling and its
- 877 control in algal-related membrane processes. *Bioresource Technology*, 264(April), 343–
- 878 358. https://doi.org/10.1016/j.biortech.2018.06.102
- 879 Mahan, K. M., Le, R. K., Wells, T., Anderson, S., Yuan, J. S., Stoklosa, R. J., Bhalla, A., Hodge,
- B., & Ragauskas, A. J. (2018). Production of single cell protein from agro-waste using
 Rhodococcus opacus. *Journal of Industrial Microbiology and Biotechnology*, 45(9), 795–
- 882 801. https://doi.org/10.1007/s10295-018-2043-3
- Maiuolo, J., Oppedisano, F., Gratteri, S., Muscoli, C., & Mollace, V. (2016). Regulation of uric
 acid metabolism and excretion. *International Journal of Cardiology*, *213*, 8–14.
 https://doi.org/10.1016/j.ijcard.2015.08.109
- Maltesen, M. J., & van de Weert, M. (2008). Drying methods for protein pharmaceuticals. *Drug Discovery Today: Technologies*, *5*(2–3), 81–88.
 https://doi.org/10.1016/j.ddtec.2008.11.001
- 889 Matassa, S., Papirio, S., Pikaar, I., Hülsen, T., Leijenhorst, E., Esposito, G., ... & Verstraete, W.
- (2020). Upcycling of biowaste carbon and nutrients in line with consumer confidence: the
 "full gas" route to single cell protein. *Green Chemistry*, 22(15), 4912-4929.
- Matos, Â. P. (2017). The Impact of Microalgae in Food Science and Technology. JAOCS,
 Journal of the American Oil Chemists' Society, 94(11), 1333–1350.

- 894 https://doi.org/10.1007/s11746-017-3050-7
- 895 Matos, Â. P. (2019). Microalgae as a Potential Source of Proteins. In *Proteins: Sustainable*
- 896 Source, Processing and Applications. https://doi.org/10.1016/b978-0-12-816695 897 6.00003-9
- 898 McDonald, C. A., Yang, J. Y., Marathe, V., Yen, T. Y., & Macher, B. A. (2009). Combining
- 899 results from lectin affinity chromatography and glycocapture approaches substantially
- 900 improves the coverage of the glycoproteome. *Molecular and Cellular Proteomics*, 8(2),
- 901 287–301. https://doi.org/10.1074/mcp.M800272-MCP200
- 902 Mekonnen, M. M., & Hoekstra, A. Y. (2014). Water footprint benchmarks for crop production:
- 903 A first global assessment. *Ecological Indicators*, 46, 214–223.
 904 https://doi.org/10.1016/j.ecolind.2014.06.013
- 905 Miller, R. G., Tangney, R., Enright, N. J., Fontaine, J. B., Merritt, D. J., Ooi, M. K. J., Ruthrof,
- K. X., & Miller, B. P. (2019). Mechanisms of Fire Seasonality Effects on Plant
 Populations. *Trends in Ecology and Evolution*, 34(12), 1104–1117.
 https://doi.org/10.1016/j.tree.2019.07.009
- 909 Mourelle, M. L., Gómez, C. P., & Legido, J. L. (2017). The potential use of marine microalgae
- 910 and cyanobacteria in cosmetics and thalassotherapy. *Cosmetics*, 4(4).
 911 https://doi.org/10.3390/cosmetics4040046
- Mussatto, S. I., & Dragone, G. M. (2016). Biomass Pretreatment, Biorefineries, and Potential
 Products for a Bioeconomy Development. In *Biomass Fractionation Technologies for a*
- 914 Lignocellulosic Feedstock Based Biorefinery. Elsevier Inc. https://doi.org/10.1016/B978915 0-12-802323-5.00001-3
- 916 Mussatto, S. I., Yamakawa, C. K., van der Maas, L., & Dragone, G. (2021). New trends in
- 917 bioprocesses for lignocellulosic biomass and CO2 utilization. *Renewable and Sustainable*
- 918 Energy Reviews, 152(June), 111620. https://doi.org/10.1016/j.rser.2021.111620

Journal Pre-proofs

- 919 Nalage, D. N., Khedkar, G. D., Kalyankar, A. D., Sarkate, A. P., Ghodke, S. R., Bedre, V. B.,
- 920 & Khedkar, C. D. (2015). Single Cell Proteins. In *Encyclopedia of Food and Health* (1st
- 921 ed., Issue January). Elsevier Ltd. https://doi.org/10.1016/B978-0-12-384947-2.00628-0
- 922 Nasseri, A. T., Rasoul-Amini, S., Morowvat, M. H., & Ghasemi, Y. (2011). Single cell protein:
- 923 Production and process. American Journal of Food Technology, 6(2), 103–116.
- 924 https://doi.org/10.3923/ajft.2011.103.116
- 925 Nyyssölä, A., Suhonen, A., Ritala, A., & Oksman-Caldentey, K. M. (2022). The role of single
 926 cell protein in cellular agriculture. *Current Opinion in Biotechnology*, 75, 102686.
- 927 Oshoma, C., & Eguakun-Owie, S. (2018). Conversion of Food waste to Single Cell Protein
- 928 using Aspergillus Niger. Journal of Applied Sciences and Environmental Management,
- 929 22(3), 350. https://doi.org/10.4314/jasem.v22i3.10
- 930 Octasylva, A. R. P., & Rurianto, J. (2020). Analisis industri telekomunikasi seluler di indonesia:
- 931 Pendekatan scp (structure conduct perfoemance). INOBIS: Jurnal Inovasi Bisnis dan
- 932 *Manajemen Indonesia*, 3(3), 391-408.
- 933 Pogaku, R., Rudravaram, R., Chandel, A. K., Linga, V. R., & Yim, Z. H. (2009). The effect of
- 934 de-oiled rice bran for single cell protein production using fungal cultures under solid state
- 935 fermentation. International Journal of Food Engineering, 5(2).
- 936 https://doi.org/10.2202/1556-3758.1502
- 937 Poutanen, K., Nordlund, E., Paasi, J., Vehmas, K., & Åkerman, M. (2017). Food economy 4.0:

938 VTT's vision towards intelligent, consumer-centric food production.

- Prosekov, A. Y., & Ivanova, S. A. (2018). Food security: The challenge of the present. *Geoforum*, 91(February), 73–77. https://doi.org/10.1016/j.geoforum.2018.02.030
- Puyol, D., Batstone, D. J., Hülsen, T., Astals, S., Peces, M., & Krömer, J. O. (2017). Resource
 recovery from wastewater by biological technologies: Opportunities, challenges, and
 prospects. *Frontiers in Microbiology*, 7(JAN), 1–23.
 https://doi.org/10.3389/fmicb.2016.02106

- Ravindra, P., Rudravaram, R., Chandel, A. K., Rao, L. V., & Hui, Y. Z. (2009). Bio (single
 cell) protein: Issues of production, toxins and commercialisation status. In *Food Science and Security*.
- Raziq, A. (2020). Single cell protein (SCP) production and potential substrates: A
 comprehensive review. *Pure and Applied Biology*, 9(3), 1743–1754.
 https://doi.org/10.19045/bspab.2020.90185
- Reihani, S. F. S., & Khosravi-Darani, K. (2019). Influencing factors on single-cell protein
 production by submerged fermentation: A review. *Electronic Journal of Biotechnology*,
- 953 *37*, 34–40. https://doi.org/10.1016/j.ejbt.2018.11.005
- 954 Ritala, A., Häkkinen, S. T., Toivari, M., & Wiebe, M. G. (2017). Single cell protein-state-of-
- the-art, industrial landscape and patents 2001-2016. *Frontiers in Microbiology*, 8(OCT).
 https://doi.org/10.3389/fmicb.2017.02009
- Rodrigues, A. G. (Ed.). (2020). New and Future Developments in Microbial Biotechnology and
 Bioengineering: Microbial Biomolecules: Properties, Relevance, and Their Translational *Applications*. Elsevier.
- 960 Rodríguez-Zavala, J. S., Ortiz-Cruz, M. A., Mendoza-Hernández, G., & Moreno-Sánchez, R.
- 961 (2010). Increased synthesis of α-tocopherol, paramylon and tyrosine by Euglena gracilis
 962 under conditions of high biomass production. *Journal of applied microbiology*, *109*(6),
 963 2160-2172.
- 964 Safafar, H., Uldall Nørregaard, P., Ljubic, A., Møller, P., Løvstad Holdt, S., & Jacobsen, C.
- 965 (2016). Enhancement of protein and pigment content in two Chlorella species cultivated
 966 on industrial process water. *Journal of Marine Science and Engineering*, 4(4), 84.
- 967 Sally Ho Green Queen (2021). Mushroom Meat & Probiotic Plant-Based Ice Cream? Meet
 968 These 9 Startups Disrupting Food. Available in:
 969 https://www.greenqueen.com.hk/mushroom-meat-probiotic-plant-based-ice-cream-meet-

- 970 these-9-startups-disrupting-food/.
- 971 Schweiggert-Weisz, U., Eisner, P., Bader-Mittermaier, S., & Osen, R. (2020). Food proteins
 972 from plants and fungi. *Current Opinion in Food Science*, 32, 156–162.
- 973 https://doi.org/10.1016/j.cofs.2020.08.003
- 974 Sharif, M., Zafar, M. H., Aqib, A. I., Saeed, M., Farag, M. R., & Alagawany, M. (2021). Single
- 975 cell protein: Sources, mechanism of production, nutritional value and its uses in
 976 aquaculture nutrition. *Aquaculture*, 531(August 2020), 735885.
 977 https://doi.org/10.1016/j.aquaculture.2020.735885
- 978 Sheng, A. L. K., Bilad, M. R., Osman, N. B., & Arahman, N. (2017). Sequencing batch
- 979 membrane photobioreactor for real secondary effluent polishing using native microalgae:
- 980 Process performance and full-scale projection. Journal of Cleaner Production, 168, 708–
- 981 715. https://doi.org/10.1016/j.jclepro.2017.09.083
- 982 Sheth, U., & Patel, S. (2023). Production, Economics, and Marketing of Yeast Single Cell
- 983 Protein. In Food Microbiology Based Entrepreneurship: Making Money From Microbes (pp.
- 984 133-152). Singapore: Springer Nature Singapore.

- 985 https://link.springer.com/chapter/10.1007/978-981-19-5041-4_8
- 987 Singh, D. P., Gupta, V. K., & Prabha, R. (2019). Microbial interventions in agriculture and
- 988 environment: Volume 1: Research trends, priorities and prospects. *Microbial Interventions*
- 989 in Agriculture and Environment: Volume 1 : Research Trends, Priorities and Prospects,
 990 1–596. https://doi.org/10.1007/978-981-13-8391-5
- 991 Smedley, K. O. (2013). Comparison of regulatory management of authorized ingredients,
- approval processes, and risk-assessment procedures for feed ingredients. *International Feed Industry Federation*, 102.
- 994 Son, W. S., Park, H. J., Lee, C. J., Kim, S. N., Song, S. U., Park, G., & Lee, Y. W. (2020).
- 995 Supercritical drying of vascular endothelial growth factor in mesenchymal stem cells

- 996 culture fluids. Journal of Supercritical Fluids, 157, 104710.
 997 https://doi.org/10.1016/j.supflu.2019.104710
- 998 Souza Filho, P. F., Nair, R. B., Andersson, D., Lennartsson, P. R., & Taherzadeh, M. J. (2018).
- 999 Vegan-mycoprotein concentrate from pea-processing industry byproduct using edible
- 1000 filamentous fungi. Fungal Biology and Biotechnology, 5(1), 1–10.
- 1001 https://doi.org/10.1186/s40694-018-0050-9
- Spalvins, K., Ivanovs, K., & Blumberga, D. (2018). Single cell protein production from waste
 biomass: Review of various agricultural by-products. *Agronomy Research*, *16*, 1493–
 1508. https://doi.org/10.15159/AR.18.129
- 1005 Spalvins, K., Vamza, I., & Blumberga, D. (2019). Single Cell Oil Production from Waste
- Biomass: Review of Applicable Industrial By-Products. *Environmental and Climate Technologies*, 23(2), 325–337. https://doi.org/10.2478/rtuect-2019-0071
- Srivastava, S., Pathak, N., & Srivastava, P. (2011). Identification of limiting factors for the
 optimum growth of Fusarium oxysporum in liquid medium. *Toxicology International*, *18*(2), 111–116. https://doi.org/10.4103/0971-6580.84262
- 1011 Stiles, W. A. V., Styles, D., Chapman, S. P., Esteves, S., Bywater, A., Melville, L., Silkina, A.,
- 1012 Lupatsch, I., Fuentes Grünewald, C., Lovitt, R., Chaloner, T., Bull, A., Morris, C., &
- 1013 Llewellyn, C. A. (2018). Using microalgae in the circular economy to valorise anaerobic
- 1014 digestate: challenges and opportunities. *Bioresource Technology*, 267(June), 732–742.
- 1015 https://doi.org/10.1016/j.biortech.2018.07.100
- 1016 Strong, P. J., Xie, S., & Clarke, W. P. (2015). Methane as a resource: Can the methanotrophs
- 1017 add value? Environmental Science and Technology, 49(7), 4001–4018.
 1018 https://doi.org/10.1021/es504242n
- 1019 Suman, G., Nupur, M., Anuradha, S., & Pradeep, B. (2015). Single Cell Protein Production: A
- 1020 Review. International Journal of Current Microbiology and Applied Sciences, 4(9), 251–

1021 262.

Sun, F. F., Hong, J., Hu, J., Saddler, J. N., Fang, X., Zhang, Z., & Shen, S. (2015). Accessory
enzymes influence cellulase hydrolysis of the model substrate and the realistic
lignocellulosic biomass. *Enzyme and Microbial Technology*, 79–80, 42–48.
https://doi.org/10.1016/j.enzmictec.2015.06.020.

- Taran, M., & Asadi, N. (2014). A novel approach for environmentally friendly production of
 single cell protein from petrochemical wastewater using a halophilic microorganism in
 different conditions. *Petroleum science and technology*, *32*(5), 625-630.
- 1029 Tijing, L. D., Ryan, J., Dizon, C., Ibrahim, I., Ray, A., Nisay, N., Kyong, H., & Advincula, R.
- 1030 C. (2020). 3D printing for membrane separation, desalination and water treatment.
 1031 Applied Materials Today, 18, 100486. https://doi.org/10.1016/j.apmt.2019.100486
- Thiviya, P., Gamage, A., Kapilan, R., Merah, O., & Madhujith, T. (2022). Single Cell Protein
 Production Using Different Fruit Waste: A Review. *Separations*, 9(7), 178.
- 1034 Torres-Tiji, Y., Fields, F. J., & Mayfield, S. P. (2020). Microalgae as a future food source.
- 1035
 Biotechnology
 Advances,
 41(August
 2019).

 1036
 https://doi.org/10.1016/j.biotechadv.2020.107536
 41(August
 2019).
- 1037 Tovar Jiménez, X., Arana Cuenca, A., Téllez Jurado, A., Abreu Corona, A., & Muro Urista, C.
- R. (2012). Traditional methods for whey protein isolation and concentration: Effects on
 nutritional properties and biological activity. *Journal of the Mexican Chemical Society*,
 56(4), 369–377. https://doi.org/10.29356/jmcs.v56i4.246
- 1041 Tropea, A., Ferracane, A., Albergamo, A., Potortì, A. G., Lo Turco, V., Di Bella, G. (2022).
- Single cell protein production through multi food-waste substrate fermentation.
 Fermentation, 8, 91. https://doi.org/10.3390/fermentation8030091.
- 1044 Tropea, A., Wilson, D., Lo Curto, R. B., Dugo, G., Saugman, P., Troy-Davies, P., Waldron, K.
- 1045 W. (2015). Simultaneous saccharification and fermentation of lignocellulosic waste

Journal Pre-proofs

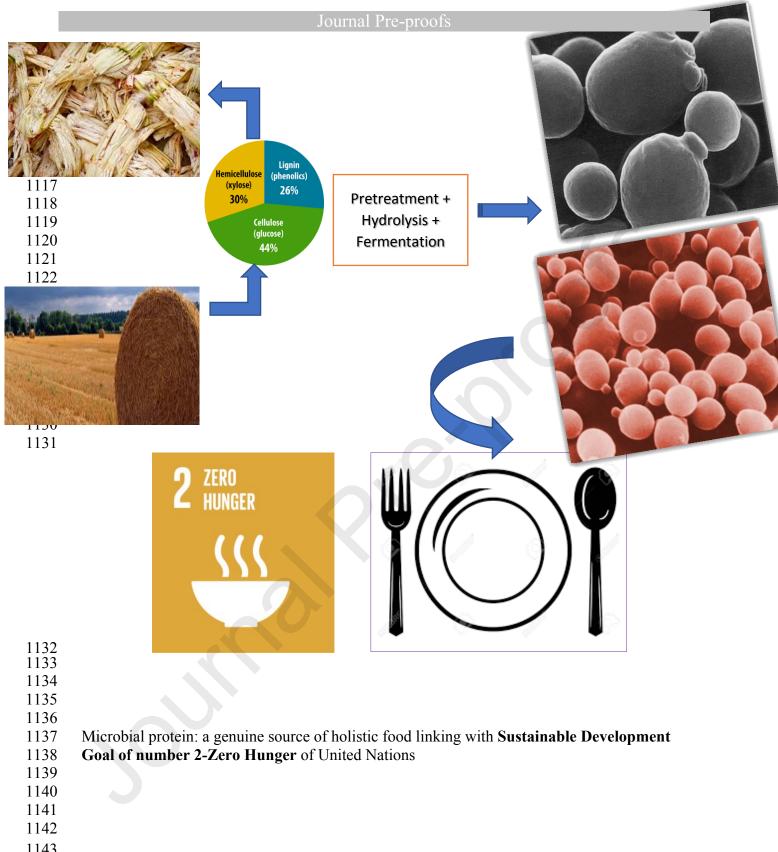
- 1046 material for second generation ethanol production. J. Biol. Res., 88, 142–143.
- 1047 Tubiello, F. N., Karl, K., Flammini, A., Gütschow, J., Conchedda, G., Pan, X., ... & Torero, M.
- 1048 (2022). Pre-and post-production processes increasingly dominate greenhouse gas
 1049 emissions from agri-food systems. *Earth System Science Data*, 14(4), 17951050 1809. https://doi.org/10.5194/essd-14-1795-
- 1051 Ugalde, U. O., & Castrillo, J. I. (2002). Agriculture and Food Production 123. Applied
 1052 Mycology and Biotechnology, 2, 123–149. http://www.bpfoods.com
- 1053 Ugbogu, E. A., & Ugbogu, O. C. (2016). a Review of Microbial Protein Production: Prospects
- and Challenges. FUW Trends in Science & Technology Journal Ftstjournal@gmail.Com
 April, 1(1), 20485170–20485182.
- 1056 Ukaegbu-Obi, K. (2016). Citation: Kelechi M. Ukaegbu-Obi. Single Cell Protein: A Resort to
- 1057 Global Protein Challenge and Waste Management Single Cell Protein: A Resort to Global
- 1058 Protein Challenge and Waste Management. Journal of Microbiology & Microbial
- *Technology*, *I*(1), 1–5. https://www.researchgate.net/profile/Kelechi-UkaegbuObi/publication/318947586 Citation Kelechi M Ukaegbu-
- 1061
 Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Manage
- 1062 ment_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_
- 1063 van der Spiegel, M., Noordam, M. Y., & van der Fels-Klerx, H. J. (2013). Safety of novel
 1064 protein sources (insects, microalgae, seaweed, duckweed, and rapeseed) and legislative
 1065 aspects for their application in food and feed production. *Comprehensive Reviews in Food*
- 1066 *Science and Food Safety*, *12*(6), 662–678. https://doi.org/10.1111/1541-4337.12032.
- 1067 Waghmare, A. G., Salve, M. K., LeBlanc, J. G., & Arya, S. S. (2016). Concentration and
- 1068 characterization of microalgae proteins from Chlorella pyrenoidosa. *Bioresources and* 1069 *Bioprocessing*, 3(1), 1-11.
- 1070 Wingfield, P. T. (2015). Overview of the purification of recombinant proteins. In Current

- 1071
 Protocols in Protein Science (Vol. 2015, Issue April).

 1072
 https://doi.org/10.1002/0471140864.ps0601s80
- 1073 Xu, H., Wang, L., Sun, J., Wang, L., Guo, H., Ye, Y., & Sun, X. (2021). Microbial
 1074 detoxification of mycotoxins in food and feed. *Critical Reviews in Food Science and*1075 *Nutrition*, 0(0), 1–19. https://doi.org/10.1080/10408398.2021.1879730
- 1076 Yadav, J. S. S., Bezawada, J., Ajila, C. M., Yan, S., Tyagi, R. D., & Surampalli, R. Y. (2014).
- 1077 Mixed culture of Kluyveromyces marxianus and Candida krusei for single-cell protein
- 1078 production and organic load removal from whey. *Bioresource Technology*, *164*, 119–127.
- 1079 https://doi.org/10.1016/j.biortech.2014.04.069
- 1080 Yadav, J. S. S., Yan, S., Ajila, C. M., Bezawada, J., Tyagi, R. D., & Surampalli, R. Y. (2016).
- 1081 Food-grade single-cell protein production, characterization and ultrafiltration recovery of
- residual fermented whey proteins from whey. *Food and Bioproducts Processing*, 99, 156–
- 1083 165. https://doi.org/10.1016/j.fbp.2016.04.012
- Yang, R., Chen, Z., Hu, P., Zhang, S., & Luo, G. (2022). Two-stage fermentation enhanced
 single-cell protein production by Yarrowia lipolytica from food waste. *Bioresource Technology*, 361, 127677.
- Ye, J., Zhou, Q., Zhang, X., & Hu, Q. (2018). Microalgal dewatering using a polyamide thin
 film composite forward osmosis membrane and fouling mitigation. *Algal Research*, *31*(January), 421–429. https://doi.org/10.1016/j.algal.2018.02.003
- Zakaria, Z. A., Boopathy, R., & Dib, J. R. (Eds.). (2020). Valorisation of Agro-industrial *residues-Volume I: Biological approaches. Springer International Publishing.*https://doi.org/10.1007/978-3-030-39137-9.
- 1093 Zepka, L. Q., Jacob-Lopes, E., Goldbeck, R., Souza-Soares, L. A., & Queiroz, M. I. (2010).
 1094 Nutritional evaluation of single-cell protein produced by Aphanothece microscopica
 1095 Nägeli. *Bioresource Technology*, 101(18), 7107–7111.

- 1096 https://doi.org/10.1016/j.biortech.2010.04.001
- 1097 Zha, X., Tsapekos, P., Zhu, X., Khoshnevisan, B., Lu, X., & Angelidaki, I. (2021).
 1098 Bioconversion of wastewater to single cell protein by methanotrophic
 1099 bacteria. *Bioresource Technology*, 320, 124351.
- 1100 Zhang, H., Han, L., & Dong, H. (2021). An insight to pretreatment, enzyme adsorption and
- 1101 enzymatic hydrolysis of lignocellulosic biomass: Experimental and modeling studies.
- 1102
 Renewable
 and
 Sustainable
 Energy
 Reviews,
 140(12),
 110758.

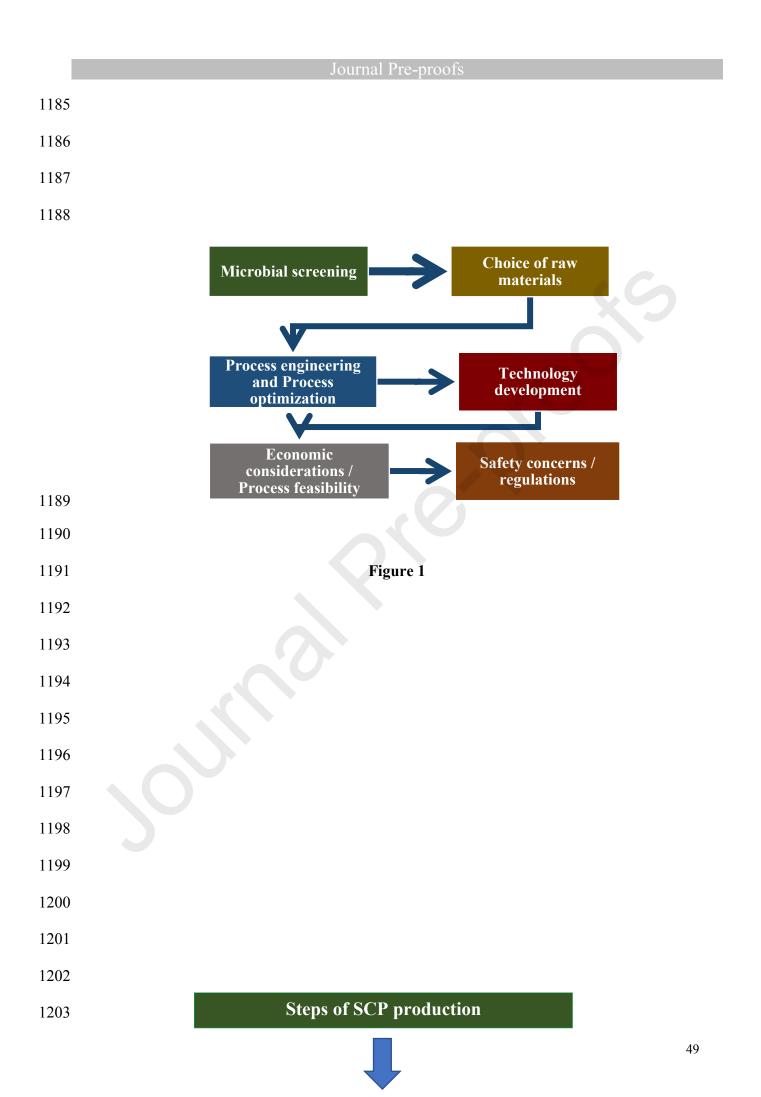
 1103
 https://doi.org/10.1016/j.rser.2021.110758
 https://doi.org/10.1016/j.rser.2021.110758
 https://doi.org/10.1016/j.rser.2021.110758
- 1104 Zhang, M., Yao, L., Maleki, E., Liao, B. Q., & Lin, H. (2019). Membrane technologies for
- 1105 microalgal cultivation and dewatering: Recent progress and challenges. *Algal Research*,
- 1106 *44*(April), 101686. https://doi.org/10.1016/j.algal.2019.101686.
- 1107 Zinjarde, S. S. (2014). Food-related applications of Yarrowia lipolytica. Food chemistry, 152,
- 1108 1-10.
- 1109

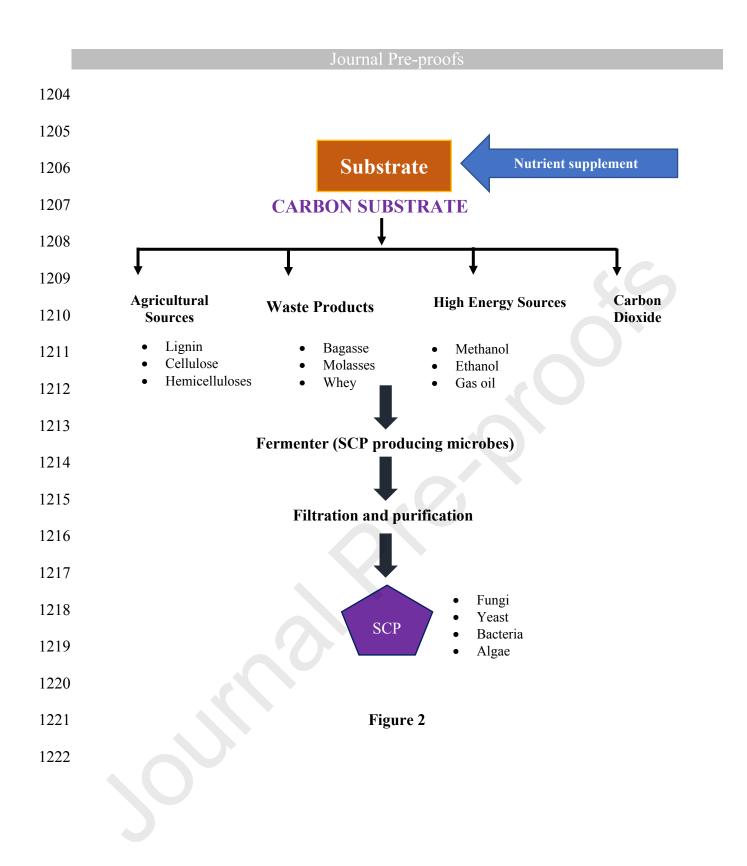


1144 Highlights:

- Microbial protein as a sustainable vegan protein source for the growing population
- Agro-industrial byproducts are renewable and surplus feedstock available round the
 year for microbial protein production
- Presence of high content of nucleic acid and toxins is a major concern of using microbial
 protein as food alternative
- Microbial protein can be produced with minimum carbon footprints and low water
 usage.
- Continuous cultivation of microorganisms employing semisolid state fermentation
 seems industrially viable strategy
- 1156
- 1157
- 1158
- 1159

	Journal Pre-proofs
1160	Figure captions
1161	
1162	Figure 1. Flowchart of the overall process for microbial protein production.
1163	
1164	Figure 2. Industrial production of microbial protein with a focus on the fermentation step
1165	
1166	
1167	
1168	
1169	
1170	
1171	
1172	
1173	
1174	
11751176	
1170	
1178	
1179	
1180	
1181	
1182	
1183	
1184	





1223	Table 1. Examp	oles of microo	rganisms use	d for single cell	protein i	production and	l their protein
1225	I able I. LAunip	Jies of microo	i gamomo use	a for single con	protoin	production and	i unon protom

1224 content

(Sp ChlChlChlEugFungi and yeastsCar Del	hospira platensis irulina maxima) lorella pyrenoidosa lorella sorokiana glena gracilis pergillus niger ndida tropicalis	content (%) 60-71 45 46-65 50-70 49	De Oliveira et al., 1999 Waghmare et al., 2016 Safafar et al., 2016 Rodríguez- Zavala et al., 2010 Kam et al., 2012
ChlChlEugFungi andyeastsCanDel	lorella pyrenoidosa lorella sorokiana glena gracilis pergillus niger ndida tropicalis	46-65 50-70 49	2016 Safafar et al., 2016 Rodríguez- Zavala et al., 2010
EugFungi andAspyeastsCanDel	glena gracilis pergillus niger ndida tropicalis	50-70 49	2016 Rodríguez- Zavala et al., 2010
Fungi and Asp yeasts Can Del	pergillus niger ndida tropicalis	49	Zavala et al., 2010
yeasts Can Deb	ndida tropicalis		Kam et al 2012
Del	<u> </u>		11uiii 00 ui., 2012
	· · · ·	56	Gao et al., 2012
Kla	baryomyces hansenii	32	Duarte et al., 2007
	yveromyces rxianus	59	Aggelopoulos et al., 2014
Yar	rowia lipolytica	54	Zinjarde, 2014
Bacteria Bac	cillus cereus	68	Kurbanoglu and Algur, 2002
Esc	herichia coli	66	Kurbanoglu and Algur, 2002
Hai	loarcula sp. IRU1	76	Taran and Asadi 2014
	odopseudomonas ustris	55–65	Kornochalert et al., 2014

- **Table 2.** Average composition of SCP obtained from different types of microorganisms and
- 1227 parameters / limitations to be considered for application in human and animal nutrition
- 1228 (Source: Anupma and Ravindra, 2000)

Algae 40–60	Fungi/yeasts	Bacteria
	30–70	50-83
45–65	35-50	60-80
4.6-7.0	6.5–7.8	4.3–5.8
1.4–2.6	1.5–1.8	2.2–3.0
5-10	5–13	8–10
9	n.a.	n.a.
6	n.a.	n.a.
4–6	9.70	15–16
7	6.6	8.6
n.a.	54	65
3	n.a.	n.a.
6.0	4.5-6.0	2.8
3	n.a.	n.a.
	1.4–2.6 5–10 9 6 4–6 7 n.a. 3 6.0 3	1.4–2.6 1.5–1.8 5–10 5–13 9 n.a. 6 n.a. 4–6 9.70 7 6.6 n.a. 54 3 n.a. 6.0 4.5–6.0

- 1240 **Table 3.** Fermentation parameters related to SCP production by different microorganisms
- 1241 (Source: Anupma and Ravindra, 2000).
- 1242

Parameter	Algae	Bacteria	Fungi	Fungi
			(Yeast)	(Filamentous)
Growth rate	Low	Highest	Quite high	Lower than
				bacteria and
				yeast
Substrate	Light, carbon	Wide range	Wide range	Mostly
	dioxide or		except carbon	lignocellulosic
	inorganic samples		dioxide	
pH range	Up to 11	5–7	5–7	3–8
Cultivation	Ponds, Bioreactors	Bioreactors	Bioreactors	Bioreactors
Contamination	High and serious	Precautions	Low	Least if pH is
risks		needed		less than 5

- **Table 4.** Single cell protein market players and their application segments (Source: Ritala et
- 1258 al., 2017).

Company	Country	Microorganism	Application segments
Amoco (BP)	United States	Candida utilis	Petrochemicals
Bega Cheese Ltd	Australia	Saccharomyces	Human food
Blue Green Foods		Aphanizomenon flos- aquae	Human food and animal feed
Cyanotech	United States	Spirulina platensis	Human food
BioProcess Algae LLC	United States	Desmodesmus sp.	Animal feed and nutrients
Calysta	United States	Methylococcus capsulatus	Fish feed
Algaeon Inc.	United States	Euglena gracillis	Human food and fish feed
Nucelis Inc.	United States	Yarrowia	Human food
Unibio A/S	United Kingdom	Methanotrophic bacteria	Dietary supplement and nutrients for animal
Euglena Co. Ltd.	Japan	Euglena	Human food
Biomin Holding GmbH	Austria	n.a	Animal feed
Evonik Industries AG	Germany	n.a	Animal feed
BlueBioTech Int. GmbH	Germany	Spirulina and Chlorella	Human food
Nutreco NV	Netherlands	n.a	Animal feed
Lallemand Inc	Canada	Saccharomyces cerevisiae and Torula	Human food and animal feed
Marlow Foods Ltd	United Kingdom	Fusarium venenatum	Human food
Vagan Pharma Ltd.	China	Bacterial	Animal feed
Angel Yeast Co. Ltd	China	Yeast	Animal feed
LeSaffre	France	Saccharomyces cerevisiae	Human food

1259 n. a: not available

1263 Credit author statement

Conceptualization, investigation, and writing—original draft: Samara Cardoso Alves, Erick
Díaz Ruiz and Bruna Lisboa. Writing— review and editing: Minaxi Sharma, Solange I.
Mussatto, Vijay Kumar Thakur, and Deepk M. Kalaskar. Conceptualization, writing— review
and editing: Vijai K. Gupta. Conceptualization, data curation, supervision, and writing—review
and editing: Anuj K. Chandel.

	Lours 1 Due une of					
	Journal Pre-proofs					
1271 1272	Conflict of Interest					
1273 1274 1275 1276 1277 1278 1279 1280 1281 1282 1283 1284	 All authors have participated in (a) conception and design, or analysis and interpretation of the data; (b) drafting the article or revising it critically for important intellectual content; and (c) approval of the final version. This manuscript has not been submitted to, nor is under review at, another journal or other publishing venue. The authors have no affiliation with any organization with a direct or indirect financial interest in the subject matter discussed in the manuscript The following authors have affiliations with organizations with direct or indirect 					
1285 1286 1287	financial interest in the subject matter discussed in the manuscript: Author's name:					
1287 1288 1289	Samara Cardoso Alves ^a , Erick Díaz-Ruiz ^a , Bruna Lisboa ^a , Minaxi Sharma ^b , Solange I.					
1290	Mussatto ^c , Vijay Kumar Thakur ^{d, e} , Deepak M. Kalaskar ^f , Vijai K. Gupta ^{d, e*} , Anuj K. Chandel					
1291	a*					
1292						
1293	Affiliation:					
1294	^a Department of Biotechnology, Engineering School of Lorena, University of São Paulo,					
1295	Lorena, São Paulo 12.602.810, Brazil					
1296	^b Laboratoire de "Chimie verte et Produits Biobasés", Haute Ecole Provinciale de Hainaut-					
1297	Département AgroBioscience et Chimie, 11, rue de la Sucrerie, 7800 Ath, Belgium					
1298	^c Department of Biotechnology and Biomedicine, Technical University of Denmark, Søltofts					
1299	Plads, Building 223, 2800, Kongens Lyngby, Denmark					
1300	^d Biorefining and Advanced Materials Research Center, Scotland's Rural College (SRUC),					
1301	Kings Buildings, West Mains Road, Edinburgh EH9 3JG, UK					
1302	^e Center for Safe and Improved Food, Scotland's Rural College (SRUC), Kings Buildings, West					
1303	Mains Road, Edinburgh EH9 3JG, UK					

- 1304 ^f UCL Institute of Musculoskeletal Sciences (IOMS), Division of Surgery and Interventional
- 1305 Science, Royal National Orthopaedic Hospital-NHS Trust, Stanmore, Middlesex, HA7 4LP,
- 1306 UK
- 1307
- 1308