



Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world

Cardoso Alves, Samara; Díaz-Ruiz, Erick; Lisboa, Bruna; Sharma, Minaxi; Mussatto, Solange I.; Kumar Thakur, Vijay; Kalaskar, Deepak M.; Gupta, Vijai K.; Chandel, Anuj K.

Published in:
Food Research International

Link to article, DOI:
[10.1016/j.foodres.2023.112596](https://doi.org/10.1016/j.foodres.2023.112596)

Publication date:
2023

Document Version
Peer reviewed version

[Link back to DTU Orbit](#)

Citation (APA):
Cardoso Alves, S., Díaz-Ruiz, E., Lisboa, B., Sharma, M., Mussatto, S. I., Kumar Thakur, V., Kalaskar, D. M., Gupta, V. K., & Chandel, A. K. (2023). Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world. *Food Research International*, 166, Article 112596.
<https://doi.org/10.1016/j.foodres.2023.112596>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Journal Pre-proofs

Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world

Samara Cardoso Alves, Erick Díaz-Ruiz, Bruna Lisboa, Minaxi Sharma, Solange I. Mussatto, Vijay Kumar Thakur, Deepak M. Kalaskar, Vijai K. Gupta, Anuj K. Chandel

PII: S0963-9969(23)00141-2
DOI: <https://doi.org/10.1016/j.foodres.2023.112596>
Reference: FRIN 112596

To appear in: *Food Research International*

Received Date: 6 November 2022
Revised Date: 27 January 2023
Accepted Date: 14 February 2023

Please cite this article as: Cardoso Alves, S., Díaz-Ruiz, E., Lisboa, B., Sharma, M., Mussatto, S.I., Kumar Thakur, V., Kalaskar, D.M., Gupta, V.K., Chandel, A.K., Microbial meat: A sustainable vegan protein source produced from agri-waste to feed the world, *Food Research International* (2023), doi: <https://doi.org/10.1016/j.foodres.2023.112596>

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2023 Elsevier Ltd. All rights reserved.



1 **Microbial meat: A sustainable vegan protein source produced from agri-**
2 **waste to feed the world**

3 Samara Cardoso Alves^a, Erick Díaz-Ruiz^a, Bruna Lisboa^a, Minaxi Sharma^b, Solange I.
4 Mussatto ^c, Vijay Kumar Thakur ^{d, e}, Deepak M. Kalaskar ^f, Vijai K. Gupta ^{d, e*}, Anuj K.
5 Chandel^{a*}

6
7 ^a *Department of Biotechnology, Engineering School of Lorena, University of São Paulo,*
8 *Lorena, São Paulo 12.602.810, Brazil*

9 ^b*Haute Ecole Provinciale de Hainaut- Condorcet, 7800 ATH, Belgium*

10 ^c *Department of Biotechnology and Biomedicine, Technical University of Denmark, Søtofts*
11 *Plads, Building 223, 2800, Kongens Lyngby, Denmark*

12 ^d*Biorefining and Advanced Materials Research Center, SRUC, Kings Buildings, West Mains*
13 *Road, Edinburgh EH9 3JG, UK*

14 ^e*Center for Safe and Improved Food, SRUC, Kings Buildings, West Mains Road, Edinburgh*
15 *EH9 3JG, UK*

16 ^f *UCL Institute of Musculoskeletal Sciences (IOMS), Division of Surgery and Interventional*
17 *Science, Royal National Orthopaedic Hospital-NHS Trust, Stanmore, Middlesex, HA7 4LP,*
18 *UK*

19
20 * Corresponding authors:

21 anuj10@usp.br, anuj.kumar.chandel@gmail.com (A.K.Chandel);

22 vijai.gupta@sruc.ac.uk, vijaifzd@gmail.com (V. K. Gupta)

23

24

25

26

Abstract

In the modern world, animal and plant protein may not meet the sustainability criteria due to their high need for arable land and potable water consumption, among other practices. Considering the growing population and food shortage, finding alternative protein sources for human consumption is an urgent issue that needs to be solved, especially in developing countries.

In this context, microbial bioconversion of valuable materials in nutritious microbial cells represent a sustainable alternative to the food chain. Microbial protein, also known as single-cell protein (SCP), consist of algae biomass, fungi or bacteria that are currently used as food source for both humans and animals. Besides contributing as a sustainable source of protein to feed the world, producing SCP, is important to reduce waste disposal problems and production costs meeting the sustainable development goals. However, for microbial protein as feed or food to become an important and sustainable alternative, addressing the challenges of raising awareness and achieving wider public regulatory acceptance is real and must be addressed with care and convenience. In this work, we critically reviewed the potential technologies for microbial protein production, its benefits, safety, and limitations associated with its uses, and perspectives for broader large-scale implementation. We argue that the information documented in this manuscript will assist in developing microbial meat as a major protein source for the vegan world.

Keywords: *Single-cell protein, Microbial meat, Protein production, Sustainability*

1. Introduction

The world population, which currently surpassed 8.0 billion inhabitants, is growing exponentially and is expected to reach around 9.2 billion people by 2040. This rapid population

52 growth has led to global food insecurity. Furthermore, the COVID-19 pandemic has increased
53 the risk of food insecurity and global hunger. According to the United Nations, hunger is about
54 to reach millions of families worldwide due to factors such as geo-political conflicts, population
55 rise, climate extremes, and the COVID-19. With this scenario, the agricultural sector needs a
56 tremendous transformation to satisfy the growing global demand for food (UN, 2019; FAO,
57 2021; GLOBAL TRENDS, 2021).

58 The expansion of food production and the intensification of agriculture have caused a
59 high cost to the environment, contributing 31% of global anthropogenic GHG emissions and
60 being responsible for strong changes in the composition and biodiversity of natural ecosystems,
61 such as soil erosion, acid rain, eutrophication, and climate change. Thus, international
62 organizations, industries, governments and society have been called upon to provide
63 generalized responses to prevent the global food crisis (Hashempour-Baltork et al., 2020;
64 Kusmayadi et al., 2021; Tubiello et al., 2022). Nearly a billion people worldwide cannot afford
65 food that contains enough protein and calories required for their health. The lack of necessary
66 protein sources causes serious health problems such as muscle weakness, defective immune
67 system, and growth deficiency (Berners-Lee et al., 2018). On the other hand, high consumption
68 of meat products can cause health problems and seriously affect the environment/climate.
69 Therefore, it is crucial for the food businesses to provide viable alternatives to animal proteins,
70 particularly those derived from meat, into the market that are less expensive and consume less
71 natural resources. (Bonny et al., 2015; Hartmann & Siegrist, 2017; Prosekov & Ivanova, 2018).

72 Converting waste into valuable food or feed for humans and animals is not only an
73 environmentally friendly activity but also a healthy business work (Jurasz et al., 2018; Sharif
74 et al., 2021). Currently, large part of waste pollutes the environment or is processed into low
75 value-added products such as biofuels and biogas. However, high-quality products such as

76 single cell oil, single cell proteins, chemicals, and enzymes, among others, can also be obtained
77 from wastes, and different production methods are being developed for that.

78 Microbial meat is one of the most relevant high-quality diet products that can be obtained
79 from agri-waste resources (El-Bakry et al., 2015; Finco et al., 2017; van der Spiegel et al.,
80 2013). SCP is a protein of microbial origin produced from a pure or mixed culture of bacteria,
81 fungi, yeasts or microalgae (Hashempour-Baltork et al., 2020). SCPs are dry cells that can be
82 used as protein supplements or as protein-rich ingredients in human and animal diets, providing
83 interesting benefits from a nutritional point of view (Anupama & Ravindra, 2000; Geada et al.,
84 2021). Moreover, from an environmental perspective, SCP does not require a large area of land
85 or large reservoirs of water for its production, making it an excellent alternative to vegetable
86 protein sources. Its production also does not emit greenhouse gases into the environment as
87 animal protein sources do. Furthermore, the production of SCP is independent of seasonal and
88 climatic variations and can be carried out throughout the year (Mekonnen & Hoekstra, 2014;
89 Miller et al., 2019; Sharif et al., 2021). Cultured meat is quite different from microbial protein
90 as it is produced by cultivating animal cells in reactors and considered as pure animal meat
91 (Choudharay et al., 2020). While microbial protein seems a viable protein source for vegan
92 population, cultured meat is considered an ideal protein for non-vegeterians.

93 The selection of cheap and suitable substrates or biodegradable agro-industrial by-
94 products as a source of nutrients for microorganisms to grow and produce proteins is of
95 fundamental importance to allow an incredible growth of the microorganism at a reduced
96 production cost (Pogaku et al., 2009; Ravindra et al., 2009). In this sense, apple pomace, yam
97 skins, potato skins, citrus pulp, pineapple residues, and papaya residues are some of the
98 substrates used as a nutrition source in microbial cultivation (Diwan et al., 2018; Spalvins et
99 al., 2019).

100 Microorganisms (algae and molds, 2-6h; bacteria and yeasts, 0.33-2h) generate protein more
101 efficiently than any animal or plant (1-2 years and a few months, respectively). In this way, the
102 production of protein biomass has several advantages in relation to livestock and conventional
103 crops. Furthermore, the microorganisms have a high protein rate based on dry mass (30-80%,
104 depending on the applied microorganism) and the protein has good nutritional value. In
105 addition, a wide variety of raw materials can be used as a substrate in the SCP, including low-
106 value agro-industrial waste and by-products. Relatively small land areas can be used to carry
107 out continuous fermentation processes to cultivate microbial proteins in large quantities. The
108 generation of SCP does not depend on seasonal, weather and weather variations. In addition,
109 microorganisms are more easily genetically modified than plants and animals. In addition, the
110 SCP presents the essential amino acid requirements for human nutrition (OctasyIva & Rurianto,
111 2020; Anupong et al., 2022).

112 Single-cell proteins will contribute to the greater popularization and wider availability of
113 protein sources in foods. The greatest growth and demand for vegan meat was found in Europe.
114 The growth rate of meat analogues is projected at 7.1% in 2025, growing sharply to 73% by
115 2050. European countries (51.5%), North America (26.8%), Asia-Pacific (11.8%), Latin
116 America (6.3%), and the Middle East and Africa (3.6%) have the highest share of the global
117 market for plant-based meat analogues (Kumar et al., 2022; Sheth & Patel, 2023).

118 The selection of cheap and suitable substrates or biodegradable agro-industrial by-
119 products as a source of nutrients for microorganisms to grow and produce proteins is of
120 fundamental importance to allow an incredible growth of the microorganism at a reduced
121 production cost (Pogaku et al., 2009; Ravindra et al., 2009). In this sense, apple pomace, yam
122 skins, potato skins, citrus pulp, pineapple residues, and papaya residues are some of the
123 substrates used as a nutrition source in microbial cultivation (Diwan et al., 2018; Spalvins et
124 al., 2019). Finally, it is worth mentioning that growing concerns about the food crisis and/or

125 lack of healthy foods due to the increased population and environmental issues have resulted in
126 more studies on the production of SCP as a potential meat substitute (Hashempour-Baltork et
127 al., 2020). Therefore, this review summarizes and critically discusses different points related to
128 the production of SCP in the form of microbial meat grown on agri-food waste resources as
129 nutrient substrates, such as recent advancements in production technology, the protein recovery
130 and purification processes, safety problems and the current industrial scenario.

131 **2. Single Cell Protein (SCP)**

132 Microbial protein is referred to as single-cell protein, although some of the producing
133 microorganisms are multicellular, such as filamentous fungi or filamentous algae. In 1968, the
134 term SCP was introduced for the first time, when scientists gathered to know the most
135 appropriate terminologies in common practice, i.e. microbial protein at the Massachusetts
136 Institute of Technology, United States (Matassa et al., 2020; Shharif et al., 2021).
137 Microorganisms help with protein deficiencies when used to increase the amount of protein and
138 improve the quality of fermented feeds (Bratosin et al., 2021).

139 The increase in the global search for protein will certainly make SCP more and more
140 interesting, although protein of microbial origin has a low proportion of current human
141 nutrition. The high speed of growth or the ability to apply substrates such as CO₂ and methane,
142 as carbon sources, makes the processes more efficient and sustainable compared to those
143 employed in traditional agriculture (Balagurunathan et al., 2022; Yang et al., 2022).

144 Currently, SCP can be produced by a limited number of microbial species, especially
145 when human demand is taken into account. The diversity of SCP sources applied in animal feed
146 is greater than that certified for human consumption and is expanding (Thiviva et al., 2022).
147 According to what will be brought forward, products derived from fungi, algae and bacteria are
148 under development or being used. Typically, production processes proceed first with the
149 preparation of the nutrient medium, then with the cultivation, then with the separation and

150 concentration of the SCP, in certain cases drying, and finally the final processing of the SCP
151 into ingredients and products (Jones et al., 2020; Nyssölä et al., 2022).

152 High food grade substrates are generally used to produce SCP for human consumption.
153 However, there is belief in the development of processes to produce SCP from cheap waste
154 from the food and beverage processing industries, as well as from agricultural and forestry
155 sources. The SCP is composed of a high protein content, which varies between 60 to 82% based
156 on dry matter, in addition, carbohydrates, nucleic acids, vitamins, minerals and fats are also part
157 of its composition. Another benefit related to SCP is that it is rich in several essential amino
158 acids, such as methionine, lysine, which are not present in adequate proportions in most animal
159 and plant sources (Al-Mudhafir, 2019; Zha et al., 2021; Khan et al., 2022).

160

161 **2. Microorganisms as a protein source**

162 Different microorganisms, including microalgae, fungi, yeasts and bacteria, can be used
163 as single cell proteins for food and feed applications due to their protein-rich composition.

164 **Table 1** summarizes some examples of microorganisms used as SCP and their protein content.
165 More details on their relevance as a source of protein and their production and utilization are
166 discussed in the following sections.

167

Table 1

168

169 **2.1. Microalgae**

170 The consumption of algae dates thousands of years across different cultures. However,
171 new developments are still needed today to boost the use of microalgae as a mainstream food
172 option. Development of improved organoleptic traits, evaluation and increase of nutritional
173 content, development of large-scale production units and also optimization of yields are some

174 of the challenges for microalgae to be seen as a more common food source (Mourelle et al.,
175 2017; Torres-Tiji et al., 2020).

176 Microalgae have several features that make them attractive for large-scale production,
177 food, and feed applications. These features include high biomass yields per unit area, the ability
178 to grow on non-arable land, and the possibility of using non-potable water and even salt water
179 for its cultivation. Nevertheless, the scale up of the appropriate technologies and efficient
180 management of precision fermentation parameters and investments are necessary to develop
181 new microalgae-based products. First is selecting adequate species, which can be done by using
182 bioprospecting methods and searching from established Generally Recognized as Safe (GRAS)
183 species. The GRAS certification is needed for a new species, which is costly and time-
184 consuming (FDA, 2017).

185 After the strain selection, it may be necessary to carry out genetic improvements to the
186 strain to enhance the desired traits, such as the yield, organoleptic trait or nutritional content.
187 Related to yield, high productivity, resistance factors and adaptation to outdoor growth are
188 examples of characteristics to be improved. Regarding the organoleptic traits, taste, aroma,
189 texture, palatability, color and appearance are some of the traits that can be improved. Finally,
190 in terms of nutritional content, the protein content and amino acid profile, the lipid content and
191 profile, and the aggregation of other nutritional molecules can be improved (Anderson et al.,
192 2017).

193 Genetic improvements in microalgae can be done by random DNA alteration, UV
194 mutagenesis, mating and genome shuffling, but these processes can be labor-intensive and time-
195 consuming. Controlled DNA manipulation can deliver faster and more precise results using
196 techniques like targeted mutagenesis, synthetic genetic tools and recombinant protein
197 expression systems. Finally, to obtain a high yield during the cultivation of the final species, it
198 is necessary to work on bioprocess development, including medium optimization, growth

199 systems adaptation, and developing a robust and cheap downstream process (Torres-Tiji et al.,
200 2020).

201 Microalgae constitute a large market today since derived products like alginates and
202 carrageenans are widely used in several industrial sectors, but specifically related to food and
203 feed applications, there is still no precise market defined. Algae has several components of
204 value for human nutrition, like, omega-3 fatty acids: DHA and EPA, natural pigments (beta-
205 carotene and astaxanthin) and glucans. Algal biomass can also be used as a nutritional
206 complement (Gong & Bassi, 2016).

207 **2.2. Fungi and yeast**

208 Like algae, fungi are also not new to human diets. Mycoproteins, more specifically, were
209 first discovered in the early 1960's (Derbyshire & Delange, 2021). Since then, many studies
210 have been done to assess the safety and benefits of this type of proteins. When talking about
211 fungi protein, this type of food includes the fruiting bodies of edible mushrooms, as well as
212 several species of micro fungi such as molds and yeasts, and their derivatives. Recently,
213 research has been focused on the production and characterization of vegetative mycelia from
214 fungi to increase its protein content and further processing to obtain meat alternatives for human
215 consumption (Schweiggert-Weisz et al., 2020).

216 There are a number of advantages of using fungi as a food source, primarily the low land
217 requirements since they can grow in bioreactors with high metabolic rates, which avoids the
218 extensive use of land needed for growing and feeding animals for meat. Production of
219 mycoproteins in bioreactors is generally done based on submerged fermentation, with fungi
220 growing in liquid media containing its nutritional requirements. Another advantage of using
221 fungi as a food source is that their single-cell proteins can provide other nutrients to the human
222 diet, including different B-complex vitamins (Sharif et al., 2021). However, there are still some
223 challenges to be overcome to allow broader use of mycoprotein as a food source, mainly related

224 to the production costs. Further research is also needed to evaluate safety issues and also to
225 spread its benefits to the public (Schweiggert-Weisz et al., 2020). Recently, metabolic
226 engineering technologies have been proposed to modify microorganisms to obtain, for example,
227 an improved utilization of agro-industrial residues with simultaneous production of SCP
228 (Hülsemann et al., 2018). However, the use of genetically modified organisms (GMO) does not
229 have public acceptance and is still a topic of discussion (Sharif et al., 2021).

230 Filamentous fungi are the preferred choice in SCP production at large scale. The fungus
231 is grown in a synthetic medium and further mixed with egg albumin and other compounds to
232 confer color and flavor, mimicking meat. In addition, using filamentous fungi has an advantage
233 over plant-derived meat substitutes, as fungi produces filaments comparable to meat fibrils,
234 conferring a similar meat texture to the product (Gmoser et al., 2020).

235 Yeasts, however, have been in the market for a long time. The production of SCP using
236 yeasts had an expressive significance in the war times. During the First World War, Germany
237 managed to substitute almost half of its imported protein sources by yeast. Initially, they used
238 brewer's yeast, but it was not enough to meet the demand as a protein source. In the beginning
239 of the Second World War, yeasts were used as a protein source in both army and civilian diets
240 (Ugalde & Castrillo, 2002). Today, yeasts are often used as supplements in animal feed and in
241 vegetarian diets. Fungi, including the yeasts market, is the second largest single-cell protein
242 market after algae. Most of the SCP is still destined for the animal feed market, but human
243 consumption has been growing in recent years.

244 It is worth noting that yeasts have various benefits over bacteria in their manufacturing
245 process, for example, they are larger than bacteria (cell size), and harvesting them from culture
246 media is easy. Yeasts also have higher lysine and malic acid contents, although their protein
247 content is usually lower, and they also have longer doubled times compared to bacteria (Raziq,
248 2020).

249

250 **2.3. Bacteria**

251 Bacteria have also been used as SCP for a long time, mainly for animal feed. Single-cell
252 protein derived from bacteria usually contains between 50 and 80% of protein (dry basis), and
253 the amino acid content is higher or similar to the FAO recommendations. Like fungi, bacteria
254 also have a high nucleic acid content, requiring previous processing (Strong et al., 2015). On
255 the other hand, bacteria have some advantages regarding the production process, such as faster
256 growth and shorter generation time when compared to fungi and yeasts. Additionally, they can
257 grow on several types of substrates, even in gaseous ones like hydrocarbons (Anupama &
258 Ravindra, 2000; Mussatto et al., 2021). In fact, using gases like CO₂, or diverse raw materials,
259 mainly waste/side streams from other industries, for bacteria cultivation may be appealing from
260 the perspectives of cost and sustainability. However, they are more difficult to harvest from the
261 culture medium due to their smaller size, requiring multiple unit operations for their recovery.
262 Some bacteria also have complex nutritional requirements (Nasseri et al., 2011).

263 For the selection of new strains for large-scale production of SCP, multiple criteria must
264 be considered, including the complexity of nutrients requirements, fermentation performance,
265 genetic stability during the cultivation process and growth morphology, the composition of the
266 final product generated by each strain in terms of protein and other components, and the
267 complexity of the downstream process required for purification (Raziq, 2020). A significant
268 concern related to the utilization of bacteria as SCP is the possibility of producing toxins, which
269 can be extracellular (exotoxins) or intracellular (endotoxins). Toxins may cause adverse effects
270 in both animals and humans. Therefore, toxins' production must be carefully assessed to avoid
271 problems with regulatory bodies (Ritala et al., 2017).

272

273 3. Composition and safety issues of SCP obtained from different microorganisms

274 **Table 2** shows the average composition of SCP obtained from algae, fungi/yeasts and
275 bacteria, focusing on their nutritional value. Some important parameters/limitations to be
276 considered for the SCP application in human and animal nutrition are also presented.

277 The acceptance and interest of a particular species for food or feed application greatly
278 depend on its composition, growth rate, and associated toxin production. SCP for human
279 consumption or animal feed must be free from all kinds of pathogens, toxins, and contaminants
280 (heavy metals or other metal compounds, hydrocarbons). In addition, they should not cause
281 food allergies, skin reactions, gastrointestinal reactions, diarrhea, vomiting, and other diseases
282 (Ugbogu & Ugbogu, 2016). Therefore, it is essential to use toxicological studies to evaluate the
283 safety of any produced SCP before marketing the products.

284 The main anti-nutritional factor in SCP is the presence of a high concentration of nucleic
285 acids, which is usually more abundant in microbial proteins than in other conventional protein
286 sources. This is one of the main factors limiting the SCP application in the food sector (Nalage
287 et al., 2015). Most nitrogen in SCP is in the form of amino acids, while the rest is in the form
288 of nucleic acids, which is a key property of fast-growing microorganisms. High nucleic acid
289 content is a problem because purine compounds derived from RNA break down and increase
290 uric acid concentration in the serum, ultimately leading to kidney stones and gout formation. It
291 has also been reported that living cells of microbes should be inactive before consumption.
292 Using an unprocessed product before killing the active microbes increases the incidence of skin
293 and gastrointestinal infections that can cause nausea and vomiting (Sharif et al., 2021). Anti-
294 nutritional factors of SCP, like an elevated presence of nucleic acids, can be eliminated by
295 applying physical and/or chemical treatments during processing (Dantas et al., 2016). Different
296 techniques for nucleic acid reduction have been proposed to make SCP suitable for food
297 applications. Chemical (sodium chloride, ammonium hydroxide, sodium hydroxide) and

298 enzymatic (ribonuclease, deoxyribonuclease) treatments can be used to treat biomass, obtaining
299 nucleic acid concentrations below 2% (w/w) (Yadav et al., 2016).

300 **Table 2**

301 Certain microorganisms can also produce toxic substances such as mycotoxins and
302 endotoxins during the production of SCP (Sharif et al., 2021). In addition, some carcinogenic
303 substances can be produced when microbes undergo mutations during the processing and
304 formation of the final product, which can be toxic to both humans and animals. However, all
305 these problems can be avoided by carefully selecting the microorganism and optimizing the
306 fermentation protocol for the production of SCP. The use of an appropriate substrate is also
307 useful to obtain SCP more beneficial to health. Recently, bacterial SCP obtained by culturing
308 bacteria in methanol as a carbon source was evaluated for mutagenicity in five in vivo tests in
309 various mammalian test systems, and the results showed no evidence of mutagenic activity due
310 to the substrate utilized for cultivation (Mahan et al., 2018; Spalvins et al., 2018).

311 Mycotoxin-generating fungi are undesirable sources of SCP as their toxins can cause
312 allergic reactions, carcinogenesis and even death in humans and animals. The fungus species
313 *Aspergillus flavus*, for example, produces aflatoxins of the B1, B2, G1 and G2 types,
314 *Penicillium citrinum* can produce citrine, trichothecenes and zearalenone, while *Fusarium* and
315 *Claviceps* species produce ergotamine. There is epidemiological evidence linking aflatoxins to
316 human liver cancer (Maiuolo et al., 2016). Recently, molecular biology techniques have been
317 explored to eliminate genes linked to mycotoxin synthesis. To isolate *A. parasiticus* and *A.*
318 *flavus* aflatoxin pathway clusters, the techniques of probing, cloning, expression libraries,
319 transcript mapping, and gene disruption have been applied. As an example, the aflR regulatory
320 gene, which controls the production of aflatoxins in *Aspergillus*, can become a target for
321 controlling the production of mycotoxins in this species. Although research in this field is still
322 starting (Dubey et al., 2018; Xu et al., 2021), reliable and easily applicable techniques can be

323 expected in the near future. Some species of bacteria can also produce toxins, which limit their
324 use as SCP. *Methylobacterium methanica* and *Pseudomonas* species, for example, produce
325 endotoxins that cause febrile reactions. However, heating can destroy these toxins (Mahan et
326 al., 2018; Ravindra et al., 2009).

327 According to the composition and potential limitations (**Table 2**), algal SCP has greater
328 safety in terms of nucleic acid content and no toxin production compared to fungi and bacteria.
329 In this way, the order of preference for food and feed application could be proposed as algae >
330 fungi > bacteria (Anupama & Ravindra, 2000; Nasserri et al., 2011). However, this is a very
331 general classification criteria, and studies should be done on each microorganism of interest to
332 elucidate its potential to be used as SCP in food and feed applications.

333 **4. Technology for SCP production**

334 SCP can be produced by submerged, semi-solid, or solid-state fermentation. The process
335 for SCP production follows the steps shown in **Figure 1**. The first step consists of a screening
336 of potential microbial strains. This step is essential for an adequate selection of microorganisms
337 capable of producing a good amount of protein. Microbial strains can be isolated from different
338 habitats such as water, air, soil or other biological materials. The best strain can also be
339 optimized if necessary by mutation, selection and/or genetic protocols. The next step is the
340 choice of raw materials to be used for the bacterium cultivation, which is necessary to obtain
341 an appropriate composition of carbon, phosphorus, and nitrogen able to favor a high biomass
342 formation in a short time. The most desirable carbon sources are those containing
343 monosaccharides and disaccharides, as they are ready to use. The third step involves process
344 engineering and process optimization. At this stage, the best growing conditions for the selected
345 strain are determined and the metabolic pathways are elucidated (Nasserri et al., 2011; Ritala et
346 al., 2017; Singh et al., 2019; Ukaegbu-Obi, 2016). Then, the next step is developing the
347 technology, which consists of defining all the technical details and performance of the process

348 to make the production robust for large-scale application. Studies of economic factors make up
349 the next stage, where energy consumption and production costs are considered for the
350 implementation of the large-scale production process. Such analysis can also be done in parallel
351 or integrated with the technology development in the fourth step. Finally, attention should be
352 given to safety and environmental protection requirements. Since the single-cell protein will be
353 used for human or animal food, the product must have high safety, as some microorganisms
354 can also produce toxins that cause side effects to humans as well as to the environment. In this
355 way, the entire process must be properly monitored. Product authorizations for particular
356 applications and legal protection of innovative processes and strains of microorganisms, namely
357 exploration licenses, are the legal and controlled aspects that the innovation requires (Ritala et
358 al., 2017; Singh et al., 2019; Ukaegbu-Obi, 2016).

359 **Figure 1**

360 Process optimization for fermentation is a very important step in the processing of single
361 cell protein production since it should be able to result in high product yield. In this sense, an
362 important aspect to consider is the medium's composition to be used for fermentation, which
363 must contain all the nutrients necessary for appropriate growth of the microorganism (Bellamy,
364 2009; Kadim et al., 2015). For fungi cultivation, for example, the fermentation medium must
365 include nutrients such as potassium, phosphorus, magnesium, trace elements, ammonium salts
366 and vitamins (like biotin), which are needed to develop mycelia (Gervasi et al., 2018). In
367 addition, the fermentation process requires a pure culture of the chosen organism, sterilization
368 of the growth medium, and a fermenter operated under suitable conditions to favor the microbial
369 growth (Nasseri et al., 2011). SCP can be produced by solid-state, semi-solid or submerged
370 fermentation. From these options, the submerged system usually results in higher production
371 efficiency (Hashempour-Baltork et al., 2020; Suman et al., 2015). At the end of the

372 fermentation, the resulting biomass is collected by filtration or centrifugation and goes through
373 washing and drying steps to obtain the SCP (**Figure 2**).

374 **Figure 2**

375 Submerged fermentations are usually more expensive, leading to higher operational costs than
376 semi-solid or solid-state fermentations, giving lower protein yields. An alternative to reduce the
377 costs of submerged fermentations is to use food wastes, which have low value or no value at
378 all, to formulate the fermentation medium. Food wastes and a variety of cheap raw materials,
379 including agricultural wastes, have already been tested as substrates for the production of SCP
380 by different microorganisms, including orange peel, sugarcane bagasse, rice husk, wheat straw,
381 cassava residues, sawdust, corn cobs, coconut residues, mandarin residues, beet pulp, among
382 others (Bellamy, 2009; Diwan et al., 2018; Reihani & Khosravi-Darani, 2019; Ritala et al.,
383 2017). Of course, the SCP production yields vary according to the substrate and microorganism
384 used. The use of banana peel as a substrate for the production of *Aspergillus niger* biomass, for
385 example, gave better yields (biomass yield of 2.29 and protein yield of 0.57 g/L) than crop
386 wastes like cucumber, orange, and pineapple waste (Oshoma & Eguakun-Owie, 2018). Also,
387 the use of pea processing by-products resulted in the production of *A. oryzae* var. *oryzae* CBS
388 819.72 mycobiomass with 38% more protein compared to the mycobiomass obtained from
389 synthetic medium (Souza Filho et al., 2018). Vinasse, the final residue obtained from the
390 production of bioethanol, can also be used for the production of SCP. *Candida parapsilosis* was
391 successfully cultivated in a medium containing 5 g/L of peptone and 70% v/v of vinasse (dos
392 Reis et al., 2019). The bioconversion of cheese whey is another appealing method for SCP
393 production since lactose can be used as a carbon source by different microorganisms, including
394 yeasts of the genus *Kluyveromyces* (Coelho Sampaio et al., 2016; Dragone et al., 2009). It is
395 worth noting that the use of lignocellulosic materials, particularly, as substrates for the
396 production of SCP, is hindered by the complex structure of these materials, which is mainly

397 composed of cellulose, hemicellulose and lignin fractions. Thus, pre-treatment methods have
398 become essential to break the resistant lignin layer, reducing the crystallinity of cellulose and
399 increasing the availability of carbohydrates to be consumed by microorganisms (Mussatto &
400 Dragone, 2016; Sun et al., 2015; H. Zhang et al., 2021).

401 Tropea et al., (2022) utilized mixed food waste (fish, pineapple, banan, apple, citrus peel etc.)
402 as a substrate to produce SCP by using *Saccharomyces cerevisiae* and observed the that the
403 final protein concentration reached up to 40.19% after 120 h of fermentation. While the true
404 protein percentage was 10.86%. SCP production from pineapple waste, studied by Aruna
405 (2019) and Tropea et al. (2015), observed the highest crude protein yields of 13.56% and 17.2%,
406 respectively.

407 In a different study, certain food wastes (banana peel, citrus peel, carrot pomace, and potato
408 peel) were utilized for the production of SCP using yeast (isolated from durien fruit), and it was
409 reported that the crude protein yield was increased from 14.07 % (before fermentation) to
410 30.82% (after fermentation) (Chun et al., 2020).

411 It was investigated how well some industrially important microbes (kefir, *K. marxianus*, and
412 *S. cerevisiae*) grew on substrates obtained from several common food industry wastes (whey,
413 molasses, brewer's solid wastes, orange, and potato residues) during SSF (Aggelopoulos et al.,
414 2014). Among all the three varieties, the highest protein content (38.5% w/w) of SCP was
415 observed with *S. cerevisiae* AXAZ-1. The protein content observed in fermented biomasses of
416 *S. cerevisiae* AXAZ-1 (38.5% w/w) and *K. marxianus* IMB3 (33.7%) were observed two times
417 higher than their corresponding substrates before treatment (20.9-22.9%).

418 In summary, various materials and wastes can be used as a substrate for producing SCP.
419 However, the material to be used for this application should meet some criteria, for example,
420 they must be non-toxic, regenerable, abundant, and inexpensive. So, besides contributing to
421 reducing production costs, the use of organic wastes as a carbon source for SCP production also

422 helps with a more valuable solution for eliminating such residues (Reihani & Khosravi-Darani,
423 2019; Srivastava et al., 2011).

424 **Table 3** summarizes different fermentation parameters related to the production of SCP
425 from algae, fungi and bacteria. The growth rate, the substrate used for production (which will
426 also impact the costs of the process), and risks of contamination are important aspects to
427 consider for developing an SCP production technology. It is also of fundamental importance to
428 optimize the fermentation process for each type of substrate and microorganism in order to
429 maximize the production yield (Reihani & Khosravi-Darani, 2019; Sharif et al., 2021).

430 **Table 3**

431 **5. Protein recovery and purification**

432 The downstream process for protein recovery will have small variations according to the
433 fermentation system and conditions used, microorganisms employed, and fermentation media
434 used, among others.

435 **5.1. Biomass recovery**

436 Mechanical separation technologies allow the fermentation products to be recovered from
437 the fermentation broth. In submerged fermentation, for example, both the substrate to be
438 fermented and biomass produced are present in the liquid medium. The biomass can be
439 harvested continuously in this system, being then filtered or centrifuged and dried to obtain the
440 SCP. In the solid-state fermentation, biomass can be recovered by using less steps than in semi-
441 solid-state fermentation. Because the later uses a higher amount of free water than solid-state
442 fermentation, it requires facile processing to obtain microbial biomass. The abysmal amount of
443 water in solid-state cultivation makes the downstream processing much more efficient, but heat
444 dissipation from the biomass poses a critical problem. While the microbial biomass in semi-
445 solid-state fermentation can be recovered by centrifugation, solid-state fermentation requires
446 special scrappers or mechanical systems to remove the biomass from the fermented slurry.

447 Single-cell from bacteria and yeasts are usually recovered by centrifugation, while
448 filamentous fungi are recovered by filtration (Ritala et al., 2017). Centrifugation is an energy-
449 intensive process; however, such energy requirements can be offset by the high value of the
450 final product (Sheng et al., 2017). Membrane technology has been considered more favorable
451 than centrifugation for microalgae biomass collection due to its lower energy consumption,
452 being generally more cost-effective, and offering full biomass recovery (Zhang et al., 2019).

453

454 **5.2. Cell disruption**

455 To isolate an intracellular protein, cell membrane must be disrupted to release the cell
456 contents, which can be done by using a suitable lysis buffer to achieve high solubilization of
457 the target protein (Hernández et al., 2018). It is possible to use SCP as a complete cell
458 preparation; however, breaking the cell makes the protein more accessible. Several methods
459 can be used to break the cell wall, including mechanical forces (crushing, crumbling, crushing,
460 pressure homogenization or ultrasound), hydrolytic enzymes (endogenous or exogenous),
461 chemical disruption with detergents, or even combinations of these methods (Nasseri et al.,
462 2011).

463

464 **5.3. Protein secretion**

465 Several methodologies, such as precipitation, extraction, and filtration, have been
466 developed to recover proteins from biological systems (McDonald et al., 2009). These methods
467 have different advantages and disadvantages. Precipitation, for example, involves the
468 adjustment of the physical properties (pH, salt and heat treatment) of the medium to improve
469 the insolubility of proteins (Tovar Jiménez et al., 2012), being an easily scalable and low-cost
470 method. However, thermal precipitation has the disadvantage of changing the structural
471 characteristics of native proteins, which can affect their functional and nutritional value (Yadav

472 et al., 2014). Protein extraction is an important biological process to recover protein from the
473 grown microorganism with desired purity. However, the extraction of protein from filamentous
474 fungi is a complex and cumbersome process due to the presence of a chitinous cell wall. Several
475 processing steps include the extreme conditions (pH, temperature, pressure, and requirement of
476 solvents, among others) required to extract the protein. However, in the case of the use of
477 microbial proteins in food and feed applications, extraction of protein employing these
478 techniques is not required.

479 Pressure-driven membrane processes (microfiltration and ultrafiltration) and direct
480 osmosis membrane processes have also been explored for SCP collection due to their high
481 efficiency, ease of operation and scalability (Ye et al., 2018). Microfiltration membranes are
482 commonly used as a clarification step to ultrafiltration and nanofiltration processes, as well as
483 for cold sterilization of liquid foods and pharmaceuticals, and can also be used to fractionate
484 large macromolecules from smaller ones, such as casein from whey fractionation in the dairy
485 industry. In the biotech and pharmaceutical industries, microfiltration membranes offer the
486 safety of a physical barrier to remove bacteria and other microbes (Tijing et al., 2020). On the
487 other hand, ultrafiltration has been more applied to eliminate organic substances from
488 wastewater treatment and in the textile industry. In ultrafiltration, the ultrafilter is supported
489 over a wire mesh, and the impure sample is poured over it. The impurity particles (electrolytes)
490 pass through the ultrafilter while the larger colloidal particles are retained (Tovar Jiménez et
491 al., 2012). This process is usually slow, but it can be accelerated by applying pressure or by
492 using a suction pump on the filtrate side (Tijing et al., 2020). Commercial whey protein is
493 purified through microfiltration and ultrafiltration processes, which avoid denaturation of the
494 protein. These methods are performed at low temperatures, removing the impurities in the whey
495 (fats and sugars) and producing a very high-quality protein.

496 It is worth noting that membrane contamination significantly decreases water
497 permeability and the overall system performance. In this sense, several techniques have been
498 reported to control/avoid membrane contamination, including feed pretreatment, system
499 operation below the critical flow, backwash, ultrasonic cleaning, chemical cleaning, and
500 mopping with air bubbles (Liao et al., 2018). In addition, most published studies use
501 commercial membranes, which are also used in the food industry. However, developing a
502 custom membrane to collect a specific type of microorganism would be very feasible
503 considering the diversity of species that can be used for SCP production, which have different
504 cell sizes (Lau et al., 2020). Moreover, coupling a good inlay control system with a suitable
505 membrane could also offer substantial performance advantages (Discart et al., 2015).

506

507 **5.4. Purification**

508 Chromatographic methods, including gel permeation, hydrophobic interaction, and
509 affinity chromatography, can be used for SCP purification. Such methods are easy to perform
510 but are complex to optimize due to the numerous parameters that need to be considered for this
511 process (Wingfield, 2015). For example, the choice of the column matrix, the buffer to be used,
512 the salt, the organic solvent, the reaction temperature, and the gradient are some of the
513 parameters to be considered for an efficient chromatography process. Chromatographic
514 methodologies are very popular for separating and purifying whey proteins, being used even
515 for large-scale protein separation (Bonnaillie et al., 2014).

516 **5.5. Drying**

517 Several techniques have been reported to dry microbial proteins and obtain powdered
518 proteins with desirable characteristics for application on an industrial scale. The mostly used
519 techniques are freezing-drying and spray-drying (Maltesen & van de Weert, 2008). Recently,
520 supercritical drying has emerged as another viable alternative for obtaining powdered proteins.

521 These techniques are based on three physical principles: sublimation, evaporation, and
522 precipitation (Son et al., 2020). Of course, different techniques use different stresses that can
523 compromise the final stability of the proteins. In addition, different techniques result in protein
524 powders with significantly different characteristics (Raziq, 2020). This fact should be
525 considered when focusing on the desired characteristics of the final product.

526

527 **6. Industrial scenario and market of SCP**

528 Currently, consumers demand healthy food and, at the same time, are concerned about
529 the environment. To meet a demanding and competitive market, manufacturers have
530 continuously innovated their production process using different raw materials and developing
531 more sustainable technologies to attract customers. Through iterative and incremental
532 methodologies, companies are creating innovative solutions to their existing products within a
533 constant innovation cycle to reduce toxic waste and the content of nucleic acids in their products
534 for a better human consumption, which, in the end, signals sales growth opportunities in the
535 food and beverage industries. Lallemand Inc., Montreal, Canada, for example, has been
536 developing innovative microbial products through external research partnerships with
537 important universities and their own internal projects to create new and healthier products
538 (Hülßen et al., 2018; Matos, 2019).

539 The SCP production process can contribute to the environment's safety by reducing the
540 carbon footprint and using wastes/renewable resources as carbon sources for fermentation. Due
541 to this positive aspect, the high demand for healthy products and alternative protein sources,
542 the SCP market is expected to expand significantly from 2020 to 2030 (Banovic et al., 2018;
543 Hülßen et al., 2018; TMR, 2021). SCP has applications in food products as an important source
544 of proteins and vitamins and has also been used to improve the nutritional value of products
545 such as soups, baked goods, ready-to-serve meals, in diet recipes, among others. For animal

546 nutrition, SCP is used for fattening calves, pigs, poultry and fish. SCP is also used to increase
547 the nutritive value of soups, baked products and specialized diets. Besides the main applications
548 of SCP in food and feed products, it is also applied in the leather and paper processing industries
549 and as a foam stabilizer (Kumar et al., 2017; Zakaria et al., 2020).

550 The global SCP market is segmented into North America, Latin America, Western
551 Europe, and Pacific Asia (excluding Japan, the Middle East and Africa). A recent market study
552 indicated that the SCP market in Malaysia was at US\$ 9.7 million in 2020 and is expected to
553 reach US\$ 24.5 million in 2030 at an annual growth rate (CAGR) of 9.7%. Vietnam's SCP
554 market revenue was valued at over US\$ 26.7 million in 2020 and is expected to exceed US\$
555 69.4 million by the end of 2030 (Khoshnevisan et al., 2020; TMR, 2021). The Asia region is
556 also made up of other prominent countries in the Association of Southeast Asian Nations
557 (ASEAN), such as India, China, Indonesia and Bangladesh. According to the Food and
558 Agriculture Organization of the United States, these countries produce about 50-60% of the
559 total aquaculture production. SCP is the most nutritious and cost-effective option for fishmeal,
560 and global growth is expected through the expansion of the aquaculture industry (Jones et al.,
561 2020; Matos, 2017; Ritala et al., 2017).

562 North America is a global leader in the global SCP market due to the region's highly
563 developed food and feed industries. North America is favored by some prominent organizations
564 related to the food and feed industries, which allows a greater manufacturing and development
565 capacity for SCP production. Furthermore, the majority of individuals who adopt a high-protein
566 diet are in North America, which makes it possible to increase the market value share of the
567 one-time-only protein in the region (Nasseri et al., 2011; Spalvins et al., 2018). European
568 consumers are concerned about increasing the protein content in their food products as well as
569 in finding alternative protein sources (Banovic et al., 2018). Furthermore, the increased support
570 in sustainability and actions against animal cruelty in the region has also contributed to a better

571 market scenario for non-animal-based protein sources. SCP is also gaining significant attraction
572 in Latin American countries. However, the major share of the population relies on animal
573 protein because of the culture and availability of grazing lands, feeding crops for animals.
574 Nevertheless, the rising awareness about high-quality protein and the right nutrition, food -
575 changing patterns, vegan proteins, and SCP is also getting sizeable attraction.

576 The main players in the SCP market, their respective countries and their segments are
577 shown in **Table 4**. These companies are focused on business growth and innovation to
578 strengthen their positions in the global SCP market. Developing new products and strategic
579 collaborations are other approaches that the key players are considering to gain a competitive
580 advantage in the global SCP market. Angel Animal Nutrition, for example, launched an
581 innovative product called GroPro, which is a yeast-derived feed ingredient composed of
582 proteins necessary for the development of young animals. Afterwards, they launched a
583 completely innovative semi-dry yeast in the market in the form of a tetra pack, which is easy to
584 use and hygienic due to its reusable upper opening. This yeast product has about 20% moisture,
585 with characteristics of dry and fresh yeast (Ritala et al., 2017; TMR, 2021).

586 **Table 4**

587 Besides the big companies, startups are also increasingly investing in innovation, from
588 microalgae supplements for athletes to ice cream based on probiotic bacteria. For example,
589 Noko Foods, a French startup recently founded in 2021, develops herbal microalgae
590 supplements, drink shakes and food products for athletes. In addition, the startup Ninoko Labs,
591 founded in 2020 in Germany, produces alternative proteins from fungal mycelium in order to
592 compete with real meat in terms of cost and flavour. Another example of a biotechnology
593 company focused on innovation for human food is Bidifice Inc., Santiago, China which
594 develops ice cream rich in healthy probiotic bacteria to help people with chronic diseases and
595 allergies (Bratosin et al., 2021; Sally Ho, 2021).

596

597 **7. Regulatory aspects**

598 SCP used as food or feed must be safe to produce and use. In most countries, there are
599 regulations to certify that food or feed is safe for human consumption. Generally, these
600 regulations differentiate between human food and animal feed, food that provides nutrition and
601 potentially flavor and aroma, and food additives such as colorants, preservatives, or feed
602 additives. In addition, although definitions differ among regions, international standards
603 regulated by the Joint FAO/WHO Expert Committee on Food Additives apply to internationally
604 traded products (Kannan et al., 2020; Ritala et al., 2017; Sharif et al., 2021).

605 Although the final SCP product is a protein and nutritional source, certain products may
606 enter the market as additives, providing, for example, color rather than SCP, which restricts the
607 extent to which they are added and their value as SCP. Therefore, the regulations differ
608 depending on the application (Zepka et al., 2010). Also, Smedley (2013) reported similarities
609 and differences among 7 jurisdictions (Canada, European Union, Brazil, China, Japan, United
610 States and South Africa) in terms of regulation of authorized food ingredients, as well as the
611 approval and management assessment process for feed components, and peculiarities between
612 regulations in these regions. In addition, as animals are not all the same in all regions, the
613 regulations for feeding pets differ in certain regions, requiring authorization before selling new
614 pet foods or additives.

615 It is worth noting that the final SCP product must not only be nutritious but also pass all
616 toxicity tests to be marketed as a food product. In addition, the unwanted nucleic acid content,
617 toxins and unwanted compounds that accumulate during the strain cultivation using substrates,
618 such as hydrocarbons and petroleum contaminated with heavy metals, need to be removed
619 (Gervasi et al., 2018). Decontamination and purification of the final product are essential for
620 SCP to be used as a food source for consumption.

621

622 8. Role of microbial protein in the circular economy

623 Recently, the concept of circular economy, which implies the transformation of wastes
624 and industry side-streams to produce renewable energy and added-value compounds, has been
625 strongly encouraged to design a more sustainable economy (Dragone et al., 2020; Stiles et al.,
626 2018). Microbial protein is becoming a potential product for incorporation in a circular
627 economy model due to its increased interest as an alternative protein source and numerous
628 applications in food, chemicals and pharmaceuticals (Lai et al., 2019). Microbial biomass,
629 particularly microalgae, can also be recycled and used as a biofertilizer to sustainably improve
630 soil quality and crop nutrition (Abo et al., 2019).

631 In recent years, the global need to find alternative protein sources has driven the
632 development of new SCP processes. Using readily available raw materials and waste streams
633 as a substrate for SCP production is also a relevant driver to develop new processes. In this
634 sense, SCP production can fortify biorefineries' economic feasibility, besides being a
635 sustainable option for managing residual raw materials and wastes (Mahan et al., 2018).
636 Concerns about environmental pollution have also driven the development of new SCP
637 production processes. This can be seen especially in processes that have applied greenhouse
638 gases as a substrate, for example, the production of SCP using CO₂ or methane as a carbon
639 source. Although there are still important challenges to overcome for large-scale and
640 economically feasible implementation of these new processes using gases as substrate, they
641 have attracted great interest from a sustainability point of view (Puyol et al., 2017; Ritala et al.,
642 2017; Ukaegbu-Obi, 2016).

643 Investment and profitability are key elements in estimating the economic viability of an
644 SCP production process. For large-scale SCP production, large bioreactors are required. Thus,
645 high oxygen transfer rates are needed to obtain a high amount of biomass during the cultivation,

646 which may cause an increased generation of heat from microbial metabolism that will lead to
647 the need for temperature control and reduction. In fact, operating costs, including labor,
648 consumables and energy, represent 45-55% of SCP manufacturing costs, while raw material
649 costs range from 35-55%. Using cheap raw materials and/or waste streams as carbon sources is
650 a good strategy to reduce substrate costs, as long as they do not compromise the quality of the
651 final product (Rodrigues, 2020). Finally, there is a relationship between cost and production
652 scale. Most of the SCP processes practiced on an industrial scale were set for a continuous
653 design, which proved to be the most profitable option (Poutanen et al., 2017; Ritala et al., 2017).

654 After everything that was presented and discussed in this work, the research brought
655 here on microbial biomasses, showed that SCPs gained momentum, due to the increased
656 demand for alternatives to proteins derived from plants and animals. In this way, certain
657 products can become popular due to consumer acceptance and significant encouragement from
658 government and regulatory authorities. The literature also made it possible to verify a diversity
659 of possibilities of microorganisms capable of producing SCPs, including microalgae, fungi,
660 bacteria and cyanobacteria.

661

662 **9. Conclusions and future prospects**

663 Microbial engineering has a relevant ability to enhance the competitiveness of the SCP
664 product in terms of production cost, functionality and nutrition. The application of GRAS
665 microorganisms is considered safe and is always the right choice in microbial engineering for
666 SCP production, with the main objective of improving the production of intermediate raw
667 materials and the accumulation of biomass. Future research and promotion of meat-optional
668 protein sources is a major challenge. The judicious utilization of agri-residues and by-products
669 of agriculture and food processing for the cultivation of filamentous fungi, yeasts, bacteria and
670 microalgae, would allow in developing SCP in a sustainable manner. Furthermore, studies

671 aiming to correlate the consumption of alternative sources of protein and gains for human health
672 would very possibly increase consumers' attention to a more sustainable diet. In the near future,
673 it is expected that the development of new processes for SCP production using residual raw
674 materials, industry-side streams or even greenhouse gases (CO₂ or methane) as carbon sources
675 will become a reality to increase the protein market without affecting the environment and
676 potentially with a low production cost. This review can be useful for the start-ups to create new
677 products or processes by combining fermentation technologies and alternative meat protein
678 sources. Indeed, the sensory attributes and nutritional value of meat alternative foods can be
679 improved by fermentation with selected microorganisms.

680

681 **ACKNOWLEDGEMENTS:** A. K. Chandel gratefully acknowledges the financial support
682 from CNPq for scientific productivity program (Process number: 309214/2021-1). S. I.
683 Mussatto acknowledges the Novo Nordisk Foundation (NNF), Denmark, grant number
684 NNF20SA0066233. V. K. Gupta and V. K. Thakur would like to acknowledge the institutional
685 funding sources supported by the SRUC, UK for research and development.

686 **AUTHOR CONTRIBUTIONS**

687 Conceptualization, investigation, and writing—original draft: Samara Cardoso Alves, Erick
688 Díaz Ruiz and Bruna Lisboa. Writing— review and editing: Minaxi Sharma, Solange I.
689 Mussatto, Vijay Kumar Thakur, and Deepk M. Kalaskar. Conceptualization, writing— review
690 and editing: Vijai K. Gupta. Conceptualization, data curation, supervision, and writing—review
691 and editing: Anuj K. Chandel.

692 **CONFLICTS OF INTEREST**

693 The authors declare no conflicts of interest.

694 **REFERENCES:**

- 695 U. S. FDA. (2017). *Regulatory Framework for Substances Intended for Use in Human Food*
696 *or Animal Food on the Basis of the Generally Recognized as Safe (GRAS) Provision of the*
697 *Federal Food, Drug, and Cosmetic Act: Guidance for Industry*. November, 28.
- 698 Abo, B. O., Odey, E. A., Bakayoko, M., & Kalakodio, L. (2019). Microalgae to biofuels
699 production: A review on cultivation, application and renewable energy. *Reviews on*
700 *Environmental Health*, 34(1), 91–99. <https://doi.org/10.1515/reveh-2018-0052>
- 701 Aggelopoulos, T., Katsieris, K., Bekatorou, A., Pandey, A., Banat, I. M., & Koutinas, A. A.
702 (2014). Solid state fermentation of food waste mixtures for single cell protein, aroma
703 volatiles and fat production. *Food chemistry*, 145, 710-716.
- 704 Al-Mudhafir, A. W. (2019). Microbiological sources and nutritional value of single cell protein
705 (SCP). *International Journal for Research in Applied Sciences and Biotechnology*, 6(6).
- 706 Anderson, M. S., Muff, T. J., Georgianna, D. R., & Mayfield, S. P. (2017). Towards a synthetic
707 nuclear transcription system in green algae: Characterization of *Chlamydomonas*
708 *reinhardtii* nuclear transcription factors and identification of targeted promoters. *Algal*
709 *Research*, 22, 47–55. <https://doi.org/10.1016/j.algal.2016.12.002>
- 710 Anupama, & Ravindra, P. (2000). Value-added food: Single cell protein. *Biotechnology*
711 *Advances*, 18(6), 459–479. [https://doi.org/10.1016/S0734-9750\(00\)00045-8](https://doi.org/10.1016/S0734-9750(00)00045-8)
- 712 Anupong, W., Jutamas, K., On-Uma, R., Sabour, A., Alshiekheid, M.,
713 Karuppusamy, I., ... & Pugazhendhi, A. (2022). Sustainable bioremediation approach to treat
714 the sago industry effluents and evaluate the possibility of yielded biomass as a single cell
715 protein (SCP) using cyanide tolerant *Streptomyces tritici* D5. *Chemosphere*, 135248.
- 716 Aruna, T.E. (2019). Production of value-added product from pineapple peels using solid state
717 fermentation. *Innov. Food Sci. Emerg. Technol.*, 57, 102193.
- 718 Banovic, M., Lähteenmäki, L., Arvola, A., Pennanen, K., Duta, D. E., Brückner-Gühmann, M.,

- 719 & Grunert, K. G. (2018). Foods with increased protein content: A qualitative study on
720 European consumer preferences and perceptions. *Appetite*, *125*, 233–243.
721 <https://doi.org/10.1016/j.appet.2018.01.034>
- 722 Balagurunathan, B., Ling, H., Choi, W. J., & Chang, M. W. (2022). Potential use of microbial
723 engineering in single-cell protein production. *Current Opinion in Biotechnology*, *76*,
724 102740.
- 725 Bellamy, W. D. (2009). Production of single-cell protein for animal feed from lignocellulose
726 wastes. *World Animal Review*, 4–9.
- 727 Berners-Lee, M., Kennelly, C., Watson, R., & Hewitt, C. N. (2018). Current global food
728 production is sufficient to meet human nutritional needs in 2050 provided there is radical
729 societal adaptation. *Elementa*, *6*. <https://doi.org/10.1525/elementa.310>
- 730 Bonnaillie, L. M., Qi, P., Wickham, E., & Tomasula, P. M. (2014). Enrichment and purification
731 of casein glycomacropeptide from whey protein isolate using supercritical carbon dioxide
732 processing and membrane ultrafiltration. *Foods*, *3*(1), 94–109.
733 <https://doi.org/10.3390/foods3010094>
- 734 Bonny, S. P. F., Gardner, G. E., Pethick, D. W., & Hocquette, J. F. (2015). What is artificial
735 meat and what does it mean for the future of the meat industry? *Journal of Integrative*
736 *Agriculture*, *14*(2), 255–263. [https://doi.org/10.1016/S2095-3119\(14\)60888-1](https://doi.org/10.1016/S2095-3119(14)60888-1)
- 737 Bratosin, B. C., Darjan, S., & Vodnar, D. C. (2021). Single cell protein: A potential substitute
738 in human and animal nutrition. *Sustainability (Switzerland)*, *13*(16), 1–24.
739 <https://doi.org/10.3390/su13169284>
- 740 Chun, N. C. W., Ismail, A. F., Makhatar, M. M. Z., Jamaluddin, F. M. N., Tajarudin, H. A.
741 (2020). Conversion of food waste via two-stage fermentation to controllable chicken feed
742 nutrients by local isolated microorganism. *Int. J. Recycl. Org. Waste Agric.*, *9*, 33–47.
- 743 Choudhury, D., Tseng, T.W., Swartz E (2020). The business of cultured meat. *Trends*

- 744 *Biotechnol.* 38, 573-577.
- 745 Coelho Sampaio, F., da Conceição Saraiva, T. L., Dumont de Lima e Silva, G., Teles de Faria,
746 J., Grijó Pitangui, C., Aliakbarian, B., Perego, P., & Converti, A. (2016). Batch growth of
747 *Kluyveromyces lactis* cells from deproteinized whey: Response surface methodology
748 versus Artificial neural network-Genetic algorithm approach. *Biochemical Engineering*
749 *Journal*, 109, 305–311. <https://doi.org/10.1016/j.bej.2016.01.026>
- 750 Dantas, E. M., Valle, B. C. S., Brito, C. M. S., Calazans, N. K. F., Peixoto, S. R. M., & Soares,
751 R. B. (2016). Partial replacement of fishmeal with biofloc meal in the diet of postlarvae of
752 the Pacific white shrimp *Litopenaeus vannamei*. *Aquaculture Nutrition*, 22(2), 335–342.
753 <https://doi.org/10.1111/anu.12249>
- 754 De Oliveira, M. A. C. L., Monteiro, M. P. C., Robbs, P. G., & Leite, S. G. F. (1999). Growth
755 and chemical composition of *Spirulina maxima* and *Spirulina platensis* biomass at
756 different temperatures. *Aquaculture international*, 7(4), 261-275.
- 757 Derbyshire, E. J., & Delange, J. (2021). Fungal Protein – What Is It and What Is the Health
758 Evidence? A Systematic Review Focusing on Mycoprotein. *Frontiers in Sustainable Food*
759 *Systems*, 5(February). <https://doi.org/10.3389/fsufs.2021.581682>
- 760 Discart, V., Bilad, M. R., Moorkens, R., Arafat, H., & Vankelecom, I. F. J. (2015). Decreasing
761 membrane fouling during *Chlorella vulgaris* broth filtration via membrane development
762 and coagulant assisted filtration. *Algal Research*, 9, 55–64.
763 <https://doi.org/10.1016/j.algal.2015.02.029>
- 764 Diwan, B., Parkhey, P., & Gupta, P. (2018). From agro-industrial wastes to single cell oils: a
765 step towards prospective biorefinery. *Folia Microbiologica*, 63(5), 547–568.
766 <https://doi.org/10.1007/s12223-018-0602-7>
- 767 dos Reis, K. C., Coimbra, J. M., Duarte, W. F., Schwan, R. F., & Silva, C. F. (2019). Biological
768 treatment of vinasse with yeast and simultaneous production of single-cell protein for feed

- 769 supplementation. *International Journal of Environmental Science and Technology*, 16(2),
770 763–774. <https://doi.org/10.1007/s13762-018-1709-8>
- 771 Dragone, G., Kerssemakers, A. A. J., Driessen, J. L. S. P., Yamakawa, C. K., Brumano, L. P.,
772 & Mussatto, S. I. (2020). Innovation and strategic orientations for the development of
773 advanced biorefineries. *Bioresource Technology*, 302(January), 122847.
774 <https://doi.org/10.1016/j.biortech.2020.122847>
- 775 Dragone, G., Mussatto, S. I., Oliveira, J. M., & Teixeira, J. A. (2009). Characterisation of
776 volatile compounds in an alcoholic beverage produced by whey fermentation. *Food*
777 *Chemistry*, 112(4), 929–935. <https://doi.org/10.1016/j.foodchem.2008.07.005>.
- 778 Duarte, L. C., Carvalheiro, F., Lopes, S., Neves, I., & Gírio, F. M. (2007). Yeast biomass
779 production in brewery's spent grains hemicellulosic hydrolyzate. In *Biotechnology for*
780 *Fuels and Chemicals* (pp. 637-647). Humana Press.
- 781 Dubey, M. K., Aamir, M., Kaushik, M. S., Khare, S., Meena, M., Singh, S., & Upadhyay, R. S.
782 (2018). PR Toxin - biosynthesis, genetic regulation, toxicological potential, prevention
783 and control measures: Overview and challenges. *Frontiers in Pharmacology*, 9(MAR), 1–
784 19. <https://doi.org/10.3389/fphar.2018.00288>
- 785 El-Bakry, M., Abraham, J., Cerda, A., Barrena, R., Ponsá, S., Gea, T., & Sanchez, A. (2015).
786 From wastes to high value added products: Novel aspects of SSF in the production of
787 enzymes. *Critical Reviews in Environmental Science and Technology*, 45(18), 1999–2042.
788 <https://doi.org/10.1080/10643389.2015.1010423>
- 789 Finco, A. M. de O., Mamani, L. D. G., Carvalho, J. C. de, de Melo Pereira, G. V., Thomaz-
790 Soccol, V., & Soccol, C. R. (2017). Technological trends and market perspectives for
791 production of microbial oils rich in omega-3. *Critical Reviews in Biotechnology*, 37(5),
792 656–671. <https://doi.org/10.1080/07388551.2016.1213221>.
- 793 Gao, Y., Li, D., & Liu, Y. (2012). Production of single cell protein from soy molasses using

- 794 *Candida tropicalis*. *Annals of microbiology*, 62(3), 1165-1172.
- 795 Geada, P., Moreira, C., Silva, M., Nunes, R., Madureira, L., Rocha, C. M. R., Pereira, R. N.,
796 Vicente, A. A., & Teixeira, J. A. (2021). Algal proteins: Production strategies and
797 nutritional and functional properties. *Bioresource Technology*, 332(March).
798 <https://doi.org/10.1016/j.biortech.2021.125125>
- 799 Gervasi, T., Pellizzeri, V., Calabrese, G., Di Bella, G., Cicero, N., & Dugo, G. (2018).
800 Production of single cell protein (SCP) from food and agricultural waste by using
801 *Saccharomyces cerevisiae*. *Natural Product Research*, 32(6), 648–653.
802 <https://doi.org/10.1080/14786419.2017.1332617>
- 803 GLOBAL TRENDS. (2021). Demographics and human development. March, 2021.
804 [https://www.dni.gov/index.php/gt2040-home/gt2040-structural-forces/demographics-and-](https://www.dni.gov/index.php/gt2040-home/gt2040-structural-forces/demographics-and-human-development)
805 [human-development](https://www.dni.gov/index.php/gt2040-home/gt2040-structural-forces/demographics-and-human-development)
- 806 Gmoser, R., Fristedt, R., Larsson, K., Undeland, I., Taherzadeh, M. J., & Lennartsson, P. R.
807 (2020). From stale bread and brewers spent grain to a new food source using edible
808 filamentous fungi. *Bioengineered*, 11(1), 582–598.
809 <https://doi.org/10.1080/21655979.2020.1768694>
- 810 Gong, M., & Bassi, A. (2016). Carotenoids from microalgae: A review of recent developments.
811 *Biotechnology Advances*, 34(8), 1396–1412.
812 <https://doi.org/10.1016/j.biotechadv.2016.10.005>
- 813 Hartmann, C., & Siegrist, M. (2017). Consumer perception and behaviour regarding sustainable
814 protein consumption: A systematic review. *Trends in Food Science and Technology*, 61,
815 11–25. <https://doi.org/10.1016/j.tifs.2016.12.006>
- 816 Hashempour-Baltork, F., Khosravi-Darani, K., Hosseini, H., Farshi, P., & Reihani, S. F. S.
817 (2020). Mycoproteins as safe meat substitutes. *Journal of Cleaner Production*, 253,
818 119958. <https://doi.org/10.1016/j.jclepro.2020.119958>
- 819 Hernández, D., Molinuevo-Salces, B., Riaño, B., Larrán-García, A. M., Tomás-Almenar, C., &

- 820 García-González, M. C. (2018). Recovery of Protein Concentrates From Microalgal
821 Biomass Grown in Manure for Fish Feed and Valorization of the By-Products Through
822 Anaerobic Digestion. *Frontiers in Sustainable Food Systems*, 2(June), 1–11.
823 <https://doi.org/10.3389/fsufs.2018.00028>
- 824 Hülsen, T., Hsieh, K., Lu, Y., Tait, S., & Batstone, D. J. (2018). Simultaneous treatment and
825 single cell protein production from agri-industrial wastewaters using purple phototrophic
826 bacteria or microalgae – A comparison. *Bioresource Technology*, 254(January), 214–223.
827 <https://doi.org/10.1016/j.biortech.2018.01.032>
- 828 Jones, S. W., Karpol, A., Friedman, S., Maru, B. T., & Tracy, B. P. (2020). Recent advances in
829 single cell protein use as a feed ingredient in aquaculture. *Current Opinion in*
830 *Biotechnology*, 61(Table 1), 189–197. <https://doi.org/10.1016/j.copbio.2019.12.026>
- 831 Jurasz, J., Mikulik, J., Krzywda, M., Ciapała, B., & Janowski, M. (2018). Integrating a wind-
832 and solar-powered hybrid to the power system by coupling it with a hydroelectric power
833 station with pumping installation. *Energy*, 144, 549–563.
834 <https://doi.org/10.1016/j.energy.2017.12.011>
- 835 Kadim, I. T., Mahgoub, O., Baqir, S., Faye, B., & Purchas, R. (2015). Cultured meat from
836 muscle stem cells: A review of challenges and prospects. *Journal of Integrative*
837 *Agriculture*, 14(2), 222–233. [https://doi.org/10.1016/S2095-3119\(14\)60881-9](https://doi.org/10.1016/S2095-3119(14)60881-9).
- 838 Kam, S., Kenari, A. A., & Younesi, H. (2012). Production of single cell protein in stickwater
839 by *Lactobacillus acidophilus* and *Aspergillus niger*. *Journal of aquatic food product*
840 *technology*, 21(5), 403-417.
- 841 Kannan, B., Rathipriya, A., Dhinakaran, A., & Hema, A. (2020). Single cell protein (scp)
842 production from Marine *Lactobacillus* spp. *Indian Journal of Animal Research*, 54(9),
843 1115-1119. <https://doi.org/10.18805/ijar.B-3858>.

- 844 Khan, M. K. I., Asif, M., Razzaq, Z. U., Nazir, A., & Maan, A. A. (2022). Sustainable food
845 industrial waste management through single cell protein production and characterization
846 of protein enriched bread. *Food Bioscience*, 46, 101406.
- 847 Khoshnevisan, B., Tabatabaei, M., Tsapekos, P., Rafiee, S., Aghbashlo, M., Lindeneg, S., &
848 Angelidaki, I. (2020). Environmental life cycle assessment of different biorefinery
849 platforms valorizing municipal solid waste to bioenergy, microbial protein, lactic and
850 succinic acid. *Renewable and Sustainable Energy Reviews*, 117(July 2019), 109493.
851 <https://doi.org/10.1016/j.rser.2019.109493>
- 852 Kornochalert, N., Kantachote, D., Chairapat, S., & Techkarnjanaruk, S. (2014). Use of
853 *Rhodospseudomonas palustris* P1 stimulated growth by fermented pineapple extract to treat
854 latex rubber sheet wastewater to obtain single cell protein. *Annals of microbiology*, 64(3),
855 1021-1032.
- 856 Kumar, P., Chatli, M. K., Mehta, N., Singh, P., Malav, O. P., & Verma, A. K. (2017). Meat
857 analogues: Health promising sustainable meat substitutes. *Critical Reviews in Food*
858 *Science and Nutrition*, 57(5), 923–932. <https://doi.org/10.1080/10408398.2014.939739>.
- 859 Kumar, P., Mehta, N., Abubakar, A. A., Verma, A. K., Kaka, U., Sharma, N., ... & Lorenzo, J.
860 M. (2022). Potential alternatives of animal proteins for sustainability in the food sector. *Food*
861 *Reviews International*, 1-26. <https://doi.org/10.1080/87559129.2022.2094403>.
- 862 Kusmayadi, A., Leong, Y. K., Yen, H. W., Huang, C. Y., & Chang, J. S. (2021). Microalgae as
863 sustainable food and feed sources for animals and humans – Biotechnological and
864 environmental aspects. *Chemosphere*, 271, 129800.
865 <https://doi.org/10.1016/j.chemosphere.2021.129800>.
- 866 Kurbanoglu, E. B., & Algur, O. F. (2002). Single-cell protein production from ram horn
867 hydrolysate by bacteria. *Bioresource Technology*, 85(2), 125-129.
- 868 Lai, Y. C., Chang, C. H., Chen, C. Y., Chang, J. S., & Ng, I. S. (2019). Towards protein

- 869 production and application by using *Chlorella* species as circular economy. *Bioresource*
870 *Technology*, 289(May), 121625. <https://doi.org/10.1016/j.biortech.2019.121625>
- 871 Lau, A. K. S., Bilad, M. R., Nordin, N. A. H. M., Faungnawakij, K., Narkkun, T., Wang, D. K.,
872 Mahlia, T. M. I., & Jaafar, J. (2020). Effect of membrane properties on tilted panel
873 performance of microalgae biomass filtration for biofuel feedstock. *Renewable and*
874 *Sustainable Energy Reviews*, 120(March 2019), 109666.
875 <https://doi.org/10.1016/j.rser.2019.109666>
- 876 Liao, Y., Bokhary, A., Maleki, E., & Liao, B. (2018). A review of membrane fouling and its
877 control in algal-related membrane processes. *Bioresource Technology*, 264(April), 343–
878 358. <https://doi.org/10.1016/j.biortech.2018.06.102>
- 879 Mahan, K. M., Le, R. K., Wells, T., Anderson, S., Yuan, J. S., Stoklosa, R. J., Bhalla, A., Hodge,
880 D. B., & Ragauskas, A. J. (2018). Production of single cell protein from agro-waste using
881 *Rhodococcus opacus*. *Journal of Industrial Microbiology and Biotechnology*, 45(9), 795–
882 801. <https://doi.org/10.1007/s10295-018-2043-3>
- 883 Maiuolo, J., Oppedisano, F., Gratteri, S., Muscoli, C., & Mollace, V. (2016). Regulation of uric
884 acid metabolism and excretion. *International Journal of Cardiology*, 213, 8–14.
885 <https://doi.org/10.1016/j.ijcard.2015.08.109>
- 886 Maltesen, M. J., & van de Weert, M. (2008). Drying methods for protein pharmaceuticals. *Drug*
887 *Discovery Today: Technologies*, 5(2–3), 81–88.
888 <https://doi.org/10.1016/j.ddtec.2008.11.001>
- 889 Matassa, S., Papirio, S., Pikaar, I., Hülsen, T., Leijenhurst, E., Esposito, G., ... & Verstraete, W.
890 (2020). Upcycling of biowaste carbon and nutrients in line with consumer confidence: the
891 “full gas” route to single cell protein. *Green Chemistry*, 22(15), 4912–4929.
- 892 Matos, Â. P. (2017). The Impact of Microalgae in Food Science and Technology. *JAOCs*,
893 *Journal of the American Oil Chemists' Society*, 94(11), 1333–1350.

- 894 <https://doi.org/10.1007/s11746-017-3050-7>
- 895 Matos, Â. P. (2019). Microalgae as a Potential Source of Proteins. In *Proteins: Sustainable*
896 *Source, Processing and Applications*. [https://doi.org/10.1016/b978-0-12-816695-](https://doi.org/10.1016/b978-0-12-816695-6.00003-9)
897 [6.00003-9](https://doi.org/10.1016/b978-0-12-816695-6.00003-9)
- 898 McDonald, C. A., Yang, J. Y., Marathe, V., Yen, T. Y., & Macher, B. A. (2009). Combining
899 results from lectin affinity chromatography and glycocapture approaches substantially
900 improves the coverage of the glycoproteome. *Molecular and Cellular Proteomics*, 8(2),
901 287–301. <https://doi.org/10.1074/mcp.M800272-MCP200>
- 902 Mekonnen, M. M., & Hoekstra, A. Y. (2014). Water footprint benchmarks for crop production:
903 A first global assessment. *Ecological Indicators*, 46, 214–223.
904 <https://doi.org/10.1016/j.ecolind.2014.06.013>
- 905 Miller, R. G., Tangney, R., Enright, N. J., Fontaine, J. B., Merritt, D. J., Ooi, M. K. J., Ruthrof,
906 K. X., & Miller, B. P. (2019). Mechanisms of Fire Seasonality Effects on Plant
907 Populations. *Trends in Ecology and Evolution*, 34(12), 1104–1117.
908 <https://doi.org/10.1016/j.tree.2019.07.009>
- 909 Mourelle, M. L., Gómez, C. P., & Legido, J. L. (2017). The potential use of marine microalgae
910 and cyanobacteria in cosmetics and thalassotherapy. *Cosmetics*, 4(4).
911 <https://doi.org/10.3390/cosmetics4040046>
- 912 Mussatto, S. I., & Dragone, G. M. (2016). Biomass Pretreatment, Biorefineries, and Potential
913 Products for a Bioeconomy Development. In *Biomass Fractionation Technologies for a*
914 *Lignocellulosic Feedstock Based Biorefinery*. Elsevier Inc. [https://doi.org/10.1016/B978-](https://doi.org/10.1016/B978-0-12-802323-5.00001-3)
915 [0-12-802323-5.00001-3](https://doi.org/10.1016/B978-0-12-802323-5.00001-3)
- 916 Mussatto, S. I., Yamakawa, C. K., van der Maas, L., & Dragone, G. (2021). New trends in
917 bioprocesses for lignocellulosic biomass and CO₂ utilization. *Renewable and Sustainable*
918 *Energy Reviews*, 152(June), 111620. <https://doi.org/10.1016/j.rser.2021.111620>

- 919 Nalage, D. N., Khedkar, G. D., Kalyankar, A. D., Sarkate, A. P., Ghodke, S. R., Bedre, V. B.,
920 & Khedkar, C. D. (2015). Single Cell Proteins. In *Encyclopedia of Food and Health* (1st
921 ed., Issue January). Elsevier Ltd. <https://doi.org/10.1016/B978-0-12-384947-2.00628-0>
- 922 Nasserri, A. T., Rasoul-Amini, S., Morowvat, M. H., & Ghasemi, Y. (2011). Single cell protein:
923 Production and process. *American Journal of Food Technology*, 6(2), 103–116.
924 <https://doi.org/10.3923/ajft.2011.103.116>
- 925 Nyssölä, A., Suhonen, A., Ritala, A., & Oksman-Caldentey, K. M. (2022). The role of single
926 cell protein in cellular agriculture. *Current Opinion in Biotechnology*, 75, 102686.
- 927 Oshoma, C., & Eguakun-Owie, S. (2018). Conversion of Food waste to Single Cell Protein
928 using *Aspergillus Niger*. *Journal of Applied Sciences and Environmental Management*,
929 22(3), 350. <https://doi.org/10.4314/jasem.v22i3.10>
- 930 Octasyiva, A. R. P., & Rurianto, J. (2020). Analisis industri telekomunikasi seluler di indonesia:
931 Pendekatan scp (structure conduct performance). *INOBI: Jurnal Inovasi Bisnis dan*
932 *Manajemen Indonesia*, 3(3), 391-408.
- 933 Pogaku, R., Rudravaram, R., Chandel, A. K., Linga, V. R., & Yim, Z. H. (2009). The effect of
934 de-oiled rice bran for single cell protein production using fungal cultures under solid state
935 fermentation. *International Journal of Food Engineering*, 5(2).
936 <https://doi.org/10.2202/1556-3758.1502>
- 937 Poutanen, K., Nordlund, E., Paasi, J., Vehmas, K., & Åkerman, M. (2017). Food economy 4.0:
938 VTT's vision towards intelligent, consumer-centric food production.
- 939 Prosekov, A. Y., & Ivanova, S. A. (2018). Food security: The challenge of the present.
940 *Geoforum*, 91(February), 73–77. <https://doi.org/10.1016/j.geoforum.2018.02.030>
- 941 Puyol, D., Batstone, D. J., Hülsen, T., Astals, S., Peces, M., & Krömer, J. O. (2017). Resource
942 recovery from wastewater by biological technologies: Opportunities, challenges, and
943 prospects. *Frontiers in Microbiology*, 7(JAN), 1–23.
944 <https://doi.org/10.3389/fmicb.2016.02106>

- 945 Ravindra, P., Rudravaram, R., Chandel, A. K., Rao, L. V., & Hui, Y. Z. (2009). Bio (single
946 cell) protein: Issues of production, toxins and commercialisation status. In *Food Science
947 and Security*.
- 948 Raziq, A. (2020). Single cell protein (SCP) production and potential substrates: A
949 comprehensive review. *Pure and Applied Biology*, 9(3), 1743–1754.
950 <https://doi.org/10.19045/bspab.2020.90185>
- 951 Reihani, S. F. S., & Khosravi-Darani, K. (2019). Influencing factors on single-cell protein
952 production by submerged fermentation: A review. *Electronic Journal of Biotechnology*,
953 37, 34–40. <https://doi.org/10.1016/j.ejbt.2018.11.005>
- 954 Ritala, A., Häkkinen, S. T., Toivari, M., & Wiebe, M. G. (2017). Single cell protein-state-of-
955 the-art, industrial landscape and patents 2001-2016. *Frontiers in Microbiology*, 8(OCT).
956 <https://doi.org/10.3389/fmicb.2017.02009>
- 957 Rodrigues, A. G. (Ed.). (2020). *New and Future Developments in Microbial Biotechnology and
958 Bioengineering: Microbial Biomolecules: Properties, Relevance, and Their Translational
959 Applications*. Elsevier.
- 960 Rodríguez-Zavala, J. S., Ortiz-Cruz, M. A., Mendoza-Hernández, G., & Moreno-Sánchez, R.
961 (2010). Increased synthesis of α -tocopherol, paramylon and tyrosine by *Euglena gracilis*
962 under conditions of high biomass production. *Journal of applied microbiology*, 109(6),
963 2160-2172.
- 964 Safafar, H., Uldall Nørregaard, P., Ljubic, A., Møller, P., Løvstad Holdt, S., & Jacobsen, C.
965 (2016). Enhancement of protein and pigment content in two *Chlorella* species cultivated
966 on industrial process water. *Journal of Marine Science and Engineering*, 4(4), 84.
- 967 Sally Ho – Green Queen (2021). Mushroom Meat & Probiotic Plant-Based Ice Cream? Meet
968 These 9 Startups Disrupting Food. Available in:
969 <https://www.greenqueen.com.hk/mushroom-meat-probiotic-plant-based-ice-cream-meet->

- 970 these-9-startups-disrupting-food/.
- 971 Schweiggert-Weisz, U., Eisner, P., Bader-Mittermaier, S., & Osen, R. (2020). Food proteins
972 from plants and fungi. *Current Opinion in Food Science*, 32, 156–162.
973 <https://doi.org/10.1016/j.cofs.2020.08.003>
- 974 Sharif, M., Zafar, M. H., Aqib, A. I., Saeed, M., Farag, M. R., & Alagawany, M. (2021). Single
975 cell protein: Sources, mechanism of production, nutritional value and its uses in
976 aquaculture nutrition. *Aquaculture*, 531(August 2020), 735885.
977 <https://doi.org/10.1016/j.aquaculture.2020.735885>
- 978 Sheng, A. L. K., Bilad, M. R., Osman, N. B., & Arahman, N. (2017). Sequencing batch
979 membrane photobioreactor for real secondary effluent polishing using native microalgae:
980 Process performance and full-scale projection. *Journal of Cleaner Production*, 168, 708–
981 715. <https://doi.org/10.1016/j.jclepro.2017.09.083>
- 982 Sheth, U., & Patel, S. (2023). Production, Economics, and Marketing of Yeast Single Cell
983 Protein. In *Food Microbiology Based Entrepreneurship: Making Money From Microbes* (pp.
984 133-152). Singapore: Springer Nature Singapore.
985 https://link.springer.com/chapter/10.1007/978-981-19-5041-4_8
- 986
- 987 Singh, D. P., Gupta, V. K., & Prabha, R. (2019). Microbial interventions in agriculture and
988 environment: Volume 1: Research trends, priorities and prospects. *Microbial Interventions
989 in Agriculture and Environment: Volume 1 : Research Trends, Priorities and Prospects*,
990 1–596. <https://doi.org/10.1007/978-981-13-8391-5>
- 991 Smedley, K. O. (2013). Comparison of regulatory management of authorized ingredients,
992 approval processes, and risk-assessment procedures for feed ingredients. *International
993 Feed Industry Federation*, 102.
- 994 Son, W. S., Park, H. J., Lee, C. J., Kim, S. N., Song, S. U., Park, G., & Lee, Y. W. (2020).
995 Supercritical drying of vascular endothelial growth factor in mesenchymal stem cells

- 996 culture fluids. *Journal of Supercritical Fluids*, 157, 104710.
997 <https://doi.org/10.1016/j.supflu.2019.104710>
- 998 Souza Filho, P. F., Nair, R. B., Andersson, D., Lennartsson, P. R., & Taherzadeh, M. J. (2018).
999 Vegan-mycoprotein concentrate from pea-processing industry byproduct using edible
1000 filamentous fungi. *Fungal Biology and Biotechnology*, 5(1), 1–10.
1001 <https://doi.org/10.1186/s40694-018-0050-9>
- 1002 Spalvins, K., Ivanovs, K., & Blumberga, D. (2018). Single cell protein production from waste
1003 biomass: Review of various agricultural by-products. *Agronomy Research*, 16, 1493–
1004 1508. <https://doi.org/10.15159/AR.18.129>
- 1005 Spalvins, K., Vamza, I., & Blumberga, D. (2019). Single Cell Oil Production from Waste
1006 Biomass: Review of Applicable Industrial By-Products. *Environmental and Climate
1007 Technologies*, 23(2), 325–337. <https://doi.org/10.2478/rtuct-2019-0071>
- 1008 Srivastava, S., Pathak, N., & Srivastava, P. (2011). Identification of limiting factors for the
1009 optimum growth of *Fusarium oxysporum* in liquid medium. *Toxicology International*,
1010 18(2), 111–116. <https://doi.org/10.4103/0971-6580.84262>
- 1011 Stiles, W. A. V., Styles, D., Chapman, S. P., Esteves, S., Bywater, A., Melville, L., Silkina, A.,
1012 Lupatsch, I., Fuentes Grünewald, C., Lovitt, R., Chaloner, T., Bull, A., Morris, C., &
1013 Llewellyn, C. A. (2018). Using microalgae in the circular economy to valorise anaerobic
1014 digestate: challenges and opportunities. *Bioresource Technology*, 267(June), 732–742.
1015 <https://doi.org/10.1016/j.biortech.2018.07.100>
- 1016 Strong, P. J., Xie, S., & Clarke, W. P. (2015). Methane as a resource: Can the methanotrophs
1017 add value? *Environmental Science and Technology*, 49(7), 4001–4018.
1018 <https://doi.org/10.1021/es504242n>
- 1019 Suman, G., Nupur, M., Anuradha, S., & Pradeep, B. (2015). Single Cell Protein Production: A
1020 Review. *International Journal of Current Microbiology and Applied Sciences*, 4(9), 251–

- 1021 262.
- 1022 Sun, F. F., Hong, J., Hu, J., Saddler, J. N., Fang, X., Zhang, Z., & Shen, S. (2015). Accessory
1023 enzymes influence cellulase hydrolysis of the model substrate and the realistic
1024 lignocellulosic biomass. *Enzyme and Microbial Technology*, 79–80, 42–48.
1025 <https://doi.org/10.1016/j.enzmictec.2015.06.020>.
- 1026 Taran, M., & Asadi, N. (2014). A novel approach for environmentally friendly production of
1027 single cell protein from petrochemical wastewater using a halophilic microorganism in
1028 different conditions. *Petroleum science and technology*, 32(5), 625–630.
- 1029 Tijjing, L. D., Ryan, J., Dizon, C., Ibrahim, I., Ray, A., Nisay, N., Kyong, H., & Advincula, R.
1030 C. (2020). 3D printing for membrane separation , desalination and water treatment.
1031 *Applied Materials Today*, 18, 100486. <https://doi.org/10.1016/j.apmt.2019.100486>
- 1032 Thiviya, P., Gamage, A., Kapilan, R., Merah, O., & Madhujith, T. (2022). Single Cell Protein
1033 Production Using Different Fruit Waste: A Review. *Separations*, 9(7), 178.
- 1034 Torres-Tijji, Y., Fields, F. J., & Mayfield, S. P. (2020). Microalgae as a future food source.
1035 *Biotechnology Advances*, 41(August 2019).
1036 <https://doi.org/10.1016/j.biotechadv.2020.107536>
- 1037 Tovar Jiménez, X., Arana Cuenca, A., Téllez Jurado, A., Abreu Corona, A., & Muro Urista, C.
1038 R. (2012). Traditional methods for whey protein isolation and concentration: Effects on
1039 nutritional properties and biological activity. *Journal of the Mexican Chemical Society*,
1040 56(4), 369–377. <https://doi.org/10.29356/jmcs.v56i4.246>
- 1041 Tropea, A., Ferracane, A., Albergamo, A., Potorti, A. G., Lo Turco, V., Di Bella, G. (2022).
1042 Single cell protein production through multi food-waste substrate fermentation.
1043 *Fermentation*, 8, 91. <https://doi.org/10.3390/fermentation8030091>.
- 1044 Tropea, A., Wilson, D., Lo Curto, R. B., Dugo, G., Saugman, P., Troy-Davies, P., Waldron, K.
1045 W. (2015). Simultaneous saccharification and fermentation of lignocellulosic waste

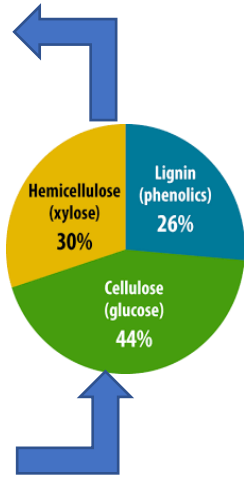
- 1046 material for second generation ethanol production. *J. Biol. Res.*, 88, 142–143.
- 1047 Tubiello, F. N., Karl, K., Flammini, A., Gütschow, J., Conchedda, G., Pan, X., ... & Torero, M.
1048 (2022). Pre-and post-production processes increasingly dominate greenhouse gas
1049 emissions from agri-food systems. *Earth System Science Data*, 14(4), 1795-
1050 1809. <https://doi.org/10.5194/essd-14-1795->
- 1051 Ugalde, U. O., & Castrillo, J. I. (2002). Agriculture and Food Production 123. *Applied*
1052 *Mycology and Biotechnology*, 2, 123–149. <http://www.bpfoods.com>
- 1053 Ugbogu, E. A., & Ugbogu, O. C. (2016). a Review of Microbial Protein Production: Prospects
1054 and Challenges. *FUW Trends in Science & Technology Journal Fstjournal@gmail.Com*
1055 *April*, 1(1), 20485170–20485182.
- 1056 Ukaegbu-Obi, K. (2016). Citation: Kelechi M. Ukaegbu-Obi. Single Cell Protein: A Resort to
1057 Global Protein Challenge and Waste Management Single Cell Protein: A Resort to Global
1058 Protein Challenge and Waste Management. *Journal of Microbiology & Microbial*
1059 *Technology*, 1(1), 1–5. [https://www.researchgate.net/profile/Kelechi-Ukaegbu-](https://www.researchgate.net/profile/Kelechi-Ukaegbu-Obi/publication/318947586_Citation_Kelechi_M_Ukaegbu-Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Management_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_)
1060 [Obi/publication/318947586_Citation_Kelechi_M_Ukaegbu-](https://www.researchgate.net/profile/Kelechi-Ukaegbu-Obi/publication/318947586_Citation_Kelechi_M_Ukaegbu-Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Management_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_)
1061 [Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Manage](https://www.researchgate.net/profile/Kelechi-Ukaegbu-Obi/publication/318947586_Citation_Kelechi_M_Ukaegbu-Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Management_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_)
1062 [ment_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_](https://www.researchgate.net/profile/Kelechi-Ukaegbu-Obi/publication/318947586_Citation_Kelechi_M_Ukaegbu-Obi_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_Management_Single_Cell_Protein_A_Resort_to_Global_Protein_Challenge_and_Waste_)
- 1063 van der Spiegel, M., Noordam, M. Y., & van der Fels-Klerx, H. J. (2013). Safety of novel
1064 protein sources (insects, microalgae, seaweed, duckweed, and rapeseed) and legislative
1065 aspects for their application in food and feed production. *Comprehensive Reviews in Food*
1066 *Science and Food Safety*, 12(6), 662–678. <https://doi.org/10.1111/1541-4337.12032>.
- 1067 Waghmare, A. G., Salve, M. K., LeBlanc, J. G., & Arya, S. S. (2016). Concentration and
1068 characterization of microalgae proteins from *Chlorella pyrenoidosa*. *Bioresources and*
1069 *Bioprocessing*, 3(1), 1-11.
- 1070 Wingfield, P. T. (2015). Overview of the purification of recombinant proteins. In *Current*

- 1071 *Protocols in Protein Science* (Vol. 2015, Issue April).
1072 <https://doi.org/10.1002/0471140864.ps0601s80>
- 1073 Xu, H., Wang, L., Sun, J., Wang, L., Guo, H., Ye, Y., & Sun, X. (2021). Microbial
1074 detoxification of mycotoxins in food and feed. *Critical Reviews in Food Science and*
1075 *Nutrition*, 0(0), 1–19. <https://doi.org/10.1080/10408398.2021.1879730>
- 1076 Yadav, J. S. S., Bezawada, J., Ajila, C. M., Yan, S., Tyagi, R. D., & Surampalli, R. Y. (2014).
1077 Mixed culture of *Kluyveromyces marxianus* and *Candida krusei* for single-cell protein
1078 production and organic load removal from whey. *Bioresource Technology*, 164, 119–127.
1079 <https://doi.org/10.1016/j.biortech.2014.04.069>
- 1080 Yadav, J. S. S., Yan, S., Ajila, C. M., Bezawada, J., Tyagi, R. D., & Surampalli, R. Y. (2016).
1081 Food-grade single-cell protein production, characterization and ultrafiltration recovery of
1082 residual fermented whey proteins from whey. *Food and Bioproducts Processing*, 99, 156–
1083 165. <https://doi.org/10.1016/j.fbp.2016.04.012>
- 1084 Yang, R., Chen, Z., Hu, P., Zhang, S., & Luo, G. (2022). Two-stage fermentation enhanced
1085 single-cell protein production by *Yarrowia lipolytica* from food waste. *Bioresource*
1086 *Technology*, 361, 127677.
- 1087 Ye, J., Zhou, Q., Zhang, X., & Hu, Q. (2018). Microalgal dewatering using a polyamide thin
1088 film composite forward osmosis membrane and fouling mitigation. *Algal Research*,
1089 31(January), 421–429. <https://doi.org/10.1016/j.algal.2018.02.003>
- 1090 Zakaria, Z. A., Boopathy, R., & Dib, J. R. (Eds.). (2020). *Valorisation of Agro-industrial*
1091 *residues-Volume I: Biological approaches*. Springer International Publishing.
1092 <https://doi.org/10.1007/978-3-030-39137-9>.
- 1093 Zepka, L. Q., Jacob-Lopes, E., Goldbeck, R., Souza-Soares, L. A., & Queiroz, M. I. (2010).
1094 Nutritional evaluation of single-cell protein produced by *Aphanothece microscopica*
1095 Nageli. *Bioresource Technology*, 101(18), 7107–7111.

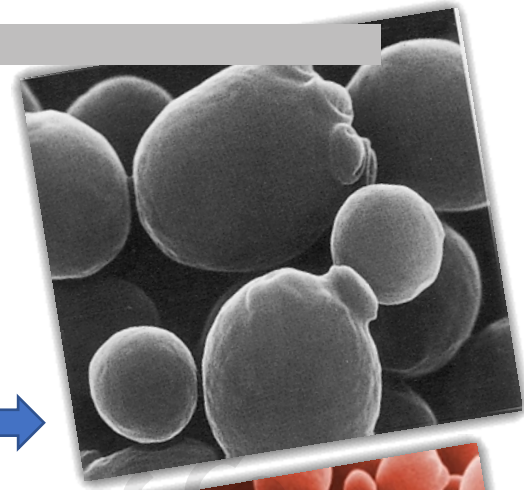
- 1096 <https://doi.org/10.1016/j.biortech.2010.04.001>
- 1097 Zha, X., Tsapekos, P., Zhu, X., Khoshnevisan, B., Lu, X., & Angelidaki, I. (2021).
1098 Bioconversion of wastewater to single cell protein by methanotrophic
1099 bacteria. *Bioresource Technology*, 320, 124351.
- 1100 Zhang, H., Han, L., & Dong, H. (2021). An insight to pretreatment, enzyme adsorption and
1101 enzymatic hydrolysis of lignocellulosic biomass: Experimental and modeling studies.
1102 *Renewable and Sustainable Energy Reviews*, 140(12), 110758.
1103 <https://doi.org/10.1016/j.rser.2021.110758>
- 1104 Zhang, M., Yao, L., Maleki, E., Liao, B. Q., & Lin, H. (2019). Membrane technologies for
1105 microalgal cultivation and dewatering: Recent progress and challenges. *Algal Research*,
1106 44(April), 101686. <https://doi.org/10.1016/j.algal.2019.101686>.
- 1107 Zinjarde, S. S. (2014). Food-related applications of *Yarrowia lipolytica*. *Food chemistry*, 152,
1108 1-10.
- 1109



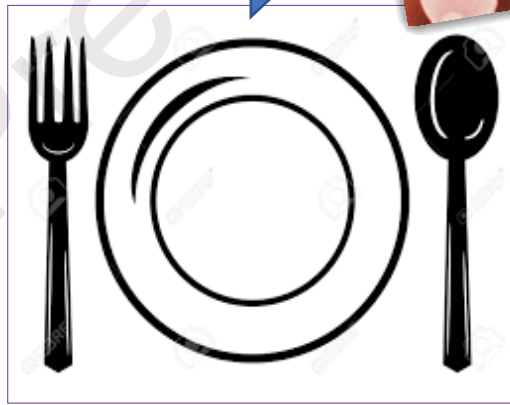
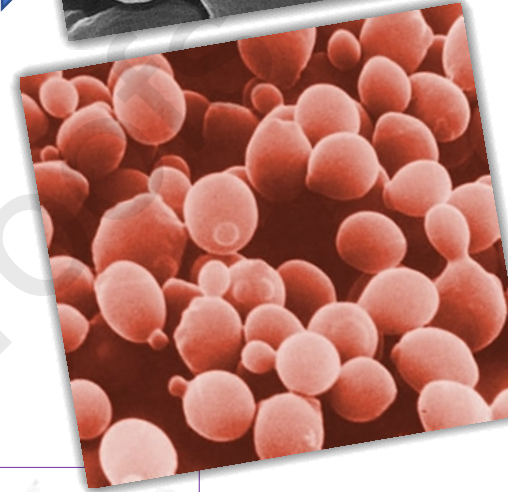
1117
1118
1119
1120
1121
1122



Pretreatment +
Hydrolysis +
Fermentation



1130
1131



1132
1133
1134
1135
1136
1137
1138
1139
1140
1141
1142
1143

Microbial protein: a genuine source of holistic food linking with **Sustainable Development Goal of number 2-Zero Hunger** of United Nations

1144 **Highlights:**

1145

1146

1147

- Microbial protein as a sustainable vegan protein source for the growing population

1148

- Agro-industrial byproducts are renewable and surplus feedstock available round the year for microbial protein production

1149

1150

- Presence of high content of nucleic acid and toxins is a major concern of using microbial protein as food alternative

1151

1152

- Microbial protein can be produced with minimum carbon footprints and low water usage.

1153

1154

- Continuous cultivation of microorganisms employing semisolid state fermentation seems industrially viable strategy

1155

1156

1157

1158

1159

1160 **Figure captions**

1161

1162 **Figure 1.** Flowchart of the overall process for microbial protein production.

1163

1164 **Figure 2.** Industrial production of microbial protein with a focus on the fermentation step

1165

1166

1167

1168

1169

1170

1171

1172

1173

1174

1175

1176

1177

1178

1179

1180

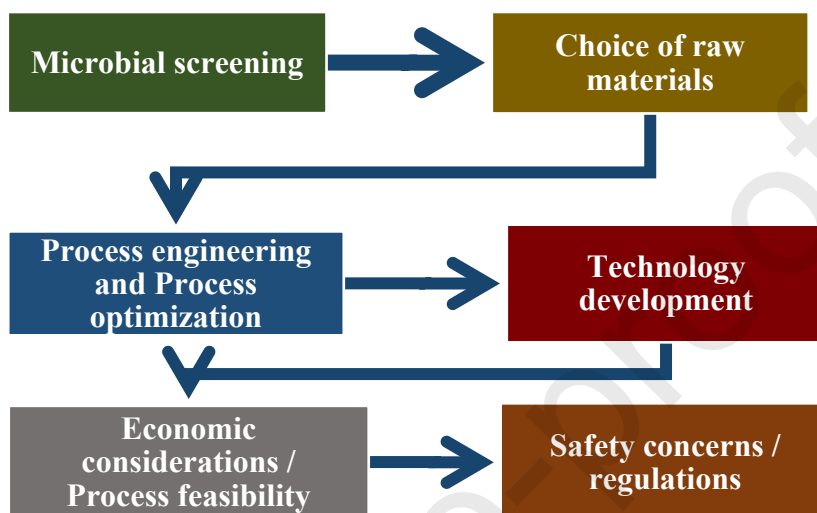
1181

1182

1183

1184

1185
1186
1187
1188



1189

1190

1191

Figure 1

1192

1193

1194

1195

1196

1197

1198

1199

1200

1201

1202

1203

Steps of SCP production



1204
1205
1206
1207
1208
1209
1210
1211
1212
1213
1214
1215
1216
1217
1218
1219
1220
1221
1222

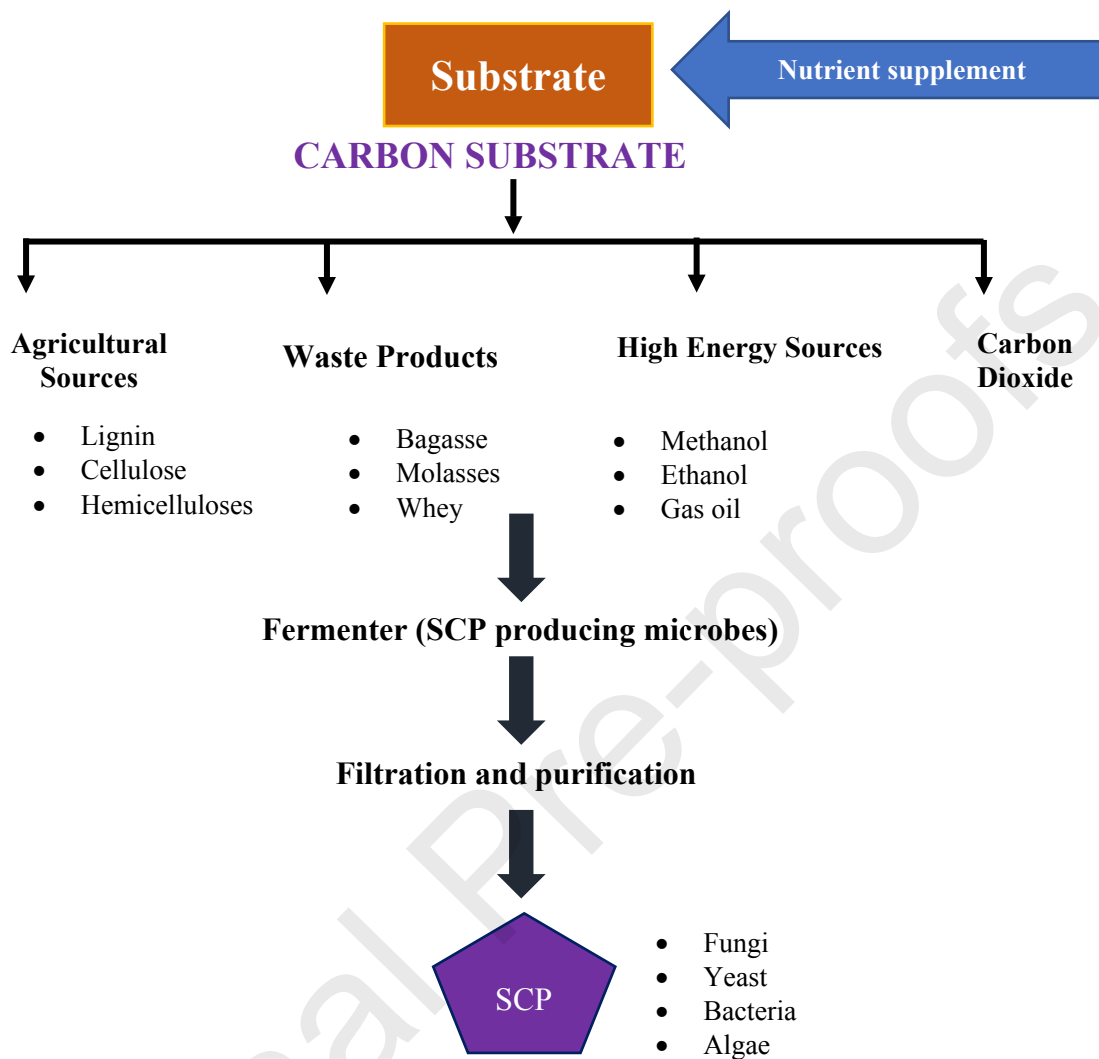


Figure 2

1223 **Table 1.** Examples of microorganisms used for single cell protein production and their protein
 1224 content

Microorganism	Protein content (%)	Reference
Microalgae	<i>Arthospira platensis</i> (<i>Spirulina maxima</i>)	60-71 De Oliveira et al., 1999
	<i>Chlorella pyrenoidosa</i>	45 Waghmare et al., 2016
	<i>Chlorella sorokiana</i>	46-65 Safafar et al., 2016
	<i>Euglena gracilis</i>	50-70 Rodríguez-Zavala et al., 2010
Fungi and yeasts	<i>Aspergillus niger</i>	49 Kam et al., 2012
	<i>Candida tropicalis</i>	56 Gao et al., 2012
	<i>Debaryomyces hansenii</i>	32 Duarte et al., 2007
	<i>Kluyveromyces marxianus</i>	59 Aggelopoulos et al., 2014
	<i>Yarrowia lipolytica</i>	54 Zinjarde, 2014
Bacteria	<i>Bacillus cereus</i>	68 Kurbanoglu and Algur, 2002
	<i>Escherichia coli</i>	66 Kurbanoglu and Algur, 2002
	<i>Haloarcula sp.</i> IRU1	76 Taran and Asadi, 2014
	<i>Rhodopseudomonas palustris</i>	55–65 Kornochalart et al., 2014

1226 **Table 2.** Average composition of SCP obtained from different types of microorganisms and
 1227 parameters / limitations to be considered for application in human and animal nutrition
 1228 (Source: Anupma and Ravindra, 2000)

Component	Composition (wt%)		
	Algae	Fungi/yeasts	Bacteria
Protein	40–60	30–70	50–83
Total nitrogen (Protein + nucleic acids)	45–65	35–50	60–80
Lysine	4.6–7.0	6.5–7.8	4.3–5.8
Methionine	1.4–2.6	1.5–1.8	2.2–3.0
Fats/Lipids	5–10	5–13	8–10
Carbohydrate	9	n.a.	n.a.
Bile pigment and Chlorophyll	6	n.a.	n.a.
Nucleic acids	4–6	9.70	15–16
Mineral salts	7	6.6	8.6
Amino acids	n.a.	54	65
Ash	3	n.a.	n.a.
Moisture	6.0	4.5–6.0	2.8
Fiber	3	n.a.	n.a.

1229 Range of values are due to the type of substrate, culture conditions and microorganism used.

1230 n.a.: not available.

1231

1232

1233

1234

1235

1236

1237

1238

1239

1240 **Table 3.** Fermentation parameters related to SCP production by different microorganisms
 1241 (Source: Anupma and Ravindra, 2000).

1242

Parameter	Algae	Bacteria	Fungi (Yeast)	Fungi (Filamentous)
Growth rate	Low	Highest	Quite high	Lower than bacteria and yeast
Substrate	Light, carbon dioxide or inorganic samples	Wide range	Wide range except carbon dioxide	Mostly lignocellulosic
pH range	Up to 11	5–7	5–7	3–8
Cultivation	Ponds, Bioreactors	Bioreactors	Bioreactors	Bioreactors
Contamination risks	High and serious	Precautions needed	Low	Least if pH is less than 5

1243

1244

1245

1246

1247

1248

1249

1250

1251

1252

1253

1254

1255

1256

1257 **Table 4.** Single cell protein market players and their application segments (Source: Ritala et
 1258 al., 2017).

Company	Country	Microorganism	Application segments
Amoco (BP)	United States	<i>Candida utilis</i>	Petrochemicals
Bega Cheese Ltd	Australia	<i>Saccharomyces</i>	Human food
Blue Green Foods	---	<i>Aphanizomenon flos-aquae</i>	Human food and animal feed
Cyanotech	United States	<i>Spirulina platensis</i>	Human food
BioProcess Algae LLC	United States	<i>Desmodesmus</i> sp.	Animal feed and nutrients
Calysta	United States	<i>Methylococcus capsulatus</i>	Fish feed
Algaeon Inc.	United States	<i>Euglena gracillis</i>	Human food and fish feed
Nucelis Inc.	United States	<i>Yarrowia</i>	Human food
Unibio A/S	United Kingdom	Methanotrophic bacteria	Dietary supplement and nutrients for animal
Euglena Co. Ltd.	Japan	<i>Euglena</i>	Human food
Biomin Holding GmbH	Austria	n.a	Animal feed
Evonik Industries AG	Germany	n.a	Animal feed
BlueBioTech Int. GmbH	Germany	<i>Spirulina and Chlorella</i>	Human food
Nutreco NV	Netherlands	n.a	Animal feed
Lallemand Inc	Canada	<i>Saccharomyces cerevisiae and Torula</i>	Human food and animal feed
Marlow Foods Ltd	United Kingdom	<i>Fusarium venenatum</i>	Human food
Vagan Pharma Ltd.	China	Bacterial	Animal feed
Angel Yeast Co. Ltd	China	Yeast	Animal feed
LeSaffre	France	<i>Saccharomyces cerevisiae</i>	Human food

1259 n. a: not available

1260

1261

1262

1263 **Credit author statement**

1264 Conceptualization, investigation, and writing—original draft: Samara Cardoso Alves, Erick
1265 Díaz Ruiz and Bruna Lisboa. Writing— review and editing: Minaxi Sharma, Solange I.
1266 Mussatto, Vijay Kumar Thakur, and Deepk M. Kalaskar. Conceptualization, writing— review
1267 and editing: Vijai K. Gupta. Conceptualization, data curation, supervision, and writing—review
1268 and editing: Anuj K. Chandel.

1269
1270

Journal Pre-proofs

Conflict of Interest

1271
1272
1273
1274
1275
1276
1277
1278
1279
1280
1281
1282
1283
1284
1285
1286
1287
1288
1289
1290
1291
1292

1293
1294
1295
1296
1297
1298
1299
1300
1301
1302
1303

- All authors have participated in (a) conception and design, or analysis and interpretation of the data; (b) drafting the article or revising it critically for important intellectual content; and (c) approval of the final version.
- This manuscript has not been submitted to, nor is under review at, another journal or other publishing venue.
- The authors have no affiliation with any organization with a direct or indirect financial interest in the subject matter discussed in the manuscript
- The following authors have affiliations with organizations with direct or indirect financial interest in the subject matter discussed in the manuscript:

Author's name:

Samara Cardoso Alves^a, Erick Díaz-Ruiz^a, Bruna Lisboa^a, Minaxi Sharma^b, Solange I. Mussatto^c, Vijay Kumar Thakur^{d,e}, Deepak M. Kalaskar^f, Vijai K. Gupta^{d,e*}, Anuj K. Chandel^{a*}

Affiliation:

^a *Department of Biotechnology, Engineering School of Lorena, University of São Paulo, Lorena, São Paulo 12.602.810, Brazil*

^b *Laboratoire de "Chimie verte et Produits Biobasés", Haute Ecole Provinciale de Hainaut-Département AgroBioscience et Chimie, 11, rue de la Sucrerie, 7800 Ath, Belgium*

^c *Department of Biotechnology and Biomedicine, Technical University of Denmark, Søtofts Plads, Building 223, 2800, Kongens Lyngby, Denmark*

^d *Biorefining and Advanced Materials Research Center, Scotland's Rural College (SRUC), Kings Buildings, West Mains Road, Edinburgh EH9 3JG, UK*

^e *Center for Safe and Improved Food, Scotland's Rural College (SRUC), Kings Buildings, West Mains Road, Edinburgh EH9 3JG, UK*

1304 ^f UCL Institute of Musculoskeletal Sciences (IOMS), Division of Surgery and Interventional
1305 Science, Royal National Orthopaedic Hospital-NHS Trust, Stanmore, Middlesex, HA7 4LP,
1306 UK
1307
1308

Journal Pre-proofs