

Effects of temperature and food availability on feeding and egg production of *Calanus hyperboreus* from Disko Bay, western Greenland

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ABSTRACT: The effects of temperature and food availability on feeding and egg production of the Arctic copepod *Calanus hyperboreus* were investigated in Disko Bay, western Greenland, from winter to spring 2009. The abundance of females in the near bottom layer and the egg production of *C. hyperboreus* prior to the spring bloom document that reproduction relies on lipid stores. The maximum *in situ* egg production (\pm SE) of 54 ± 8 eggs female⁻¹ d⁻¹ was recorded in mid-February at chlorophyll *a* concentrations below $0.1 \mu\text{g l}^{-1}$, whereas no egg production was observed in mid-April when the spring bloom developed. After reproduction, the females migrated to the surface layer to exploit the bloom and refill their lipid stores. In 2 laboratory experiments, initiated before and during the spring bloom, mature females were kept with and without food at 5 different temperatures ranging from 0 to 10°C and the fecal pellet and egg production were monitored. Food had a clear effect on fecal pellet production but no effect on egg production, while temperature did not have an effect on egg or fecal pellet production in any of the experiments. Analyses of carbon and lipid content of the females before and after the experiments did not reflect any effect of food or temperature in the pre-bloom experiment, whereas in the bloom experiment a clear positive effect of food was detected in female biochemical profiles. The lack of a temperature response suggests a future warmer ocean could be unfavorable for *C. hyperboreus* compared to smaller *Calanus* spp. which are reported to exploit minor temperature elevations for increased egg production.

KEY WORDS: *Calanus hyperboreus* · Egg production · Fecal pellet production · Effect of temperature

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INTRODUCTION

The annual productivity cycle in arctic ecosystems is greatly influenced by interannual variations in sea ice cover and solar irradiance as the breakup of the sea ice increases available light to the surface water

in the spring. In Disko Bay, western Greenland, the breakup of the sea ice varies greatly between years (Nielsen & Hansen 1995, S.D. Madsen et al. 2001, 2008, Hansen et al. 2006, S.J. Madsen et al. 2008, Dünweber et al. 2010). However, a general increase in mean air temperature of $0.4^\circ\text{C yr}^{-1}$ and a reduction

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in sea ice cover of 50% were observed from 1991 to 2004 (Hansen et al. 2006). This makes Disko Bay an ideal site for investigating the impact of climate-change-mediated variation in the ice cover on succession patterns in pelagic food webs.

Calanus hyperboreus, *C. glacialis*, and *C. finmarchicus* are key species in arctic marine food webs. With their ability to convert phytoplankton to high-energy wax esters, they provide a high-quality food source for fish, seabirds, and marine mammals (Heide-Jørgensen & Acquarone 2002, Karnovsky et al. 2003, Falk-Petersen et al. 2009). All 3 *Calanus* species are adapted to arctic conditions by having life cycles with seasonal ontogenetic migration and accumulation of lipids during spring and summer, as well as hibernation and arrested development during winter (Conover 1988, Madsen et al. 2001). *C. glacialis* and *C. hyperboreus* are true Arctic species, whereas *C. finmarchicus* have their main distribution in the Atlantic (Conover 1988). However, in Disko Bay, all 3 co-exist (Madsen et al. 2001).

In early spring, when the breakup of the sea ice triggers the formation of the spring bloom (Dünwaber et al. 2010), *Calanus* spp. ascend from deep waters (Madsen et al. 2001) and start feeding to support egg production and to refuel lipid reserves (Madsen et al. 2001, Swalethorp et al. 2011). When the bloom has ceased and the *Calanus* species have refilled their lipid stores, they stop eating and descend to the near-bottom layers where they slow down their metabolism and over-winter in a state of diapause (Lee et al. 2006).

Calanus hyperboreus differ from *C. glacialis* and *C. finmarchicus* in a number of traits, particularly lifecycle and reproductive strategies. *C. hyperboreus* have the longest life span of the 3 species, living typically between 2 and 5 yr (Scott et al. 2000, Madsen et al. 2001). In contrast to the 2 others, *C. hyperboreus* do not produce eggs after their ascent. They complete spawning during winter in the deep waters using their lipid stores to fuel egg production, and their eggs ascend to the photic zone (Hirche & Niehoff 1996, Melle & Skjoldal 1998). Winter spawning gives *C. hyperboreus* an advantage since the eggs have developed to the first feeding nauplii stage at the onset of the bloom. This enables nauplii of *C. hyperboreus* to undergo more developmental stages during the productive season and to better exploit even a short-lasting bloom. *C. hyperboreus* accumulate lipids more effectively than the 2 other *Calanus* species (Pasternak et al. 2001, Søreide et al. 2008) and can therefore descend to deeper waters earlier, sometime between June and August (Mad-

sen et al. 2001). Furthermore, the large lipid reserves increase the ability of *C. hyperboreus* to arrest development and thereby survive in areas with high variability in ice cover (Scott et al. 2000), e.g. the Disko Bay area.

The air temperatures in the Arctic have been predicted to increase by 4 to 7°C over the next 100 yr (ACIA 2004). Increasing temperatures reduce sea ice coverage and thickness. Furthermore, a warmer climate will increase meltwater runoff to the sea, and in combination, these factors can be expected to lead to an earlier stabilization of the water column and as a consequence earlier onset of the arctic spring bloom (Hansen et al. 2003). In Disko Bay, noteworthy hydrographic changes have occurred during the last decades (Hansen et al. in press). The duration of ice cover has decreased by approximately 50% (Hansen et al. 2006), and the date of ice break, and the subsequent spring bloom, now occurs earlier. In addition, there has been an acceleration in meltwater input from the Jakobshavn glacier to the bay due to increased submarine melting as a result of the increased presence of warmer oceanic waters from the Atlantic in the bay (Holland et al. 2008, Motyka et al. 2011).

An increase in temperature will not only prolong the productive season of the phytoplankton and the production of the *Calanus* community but may also impact the relative composition of the *Calanus* biomass. Kjellerup et al. (2012, this volume) showed a significant effect of temperature on egg production and feeding of *C. finmarchicus* and *C. glacialis*, including evidence that *C. finmarchicus* have a stronger positive response to increasing temperatures than *C. glacialis*. These results are supported by the models of Slagstad et al. (2011), which demonstrated that *C. finmarchicus* will take over the northern Barents Sea at the expense of *C. glacialis* in a warmer future. Several studies of temperature effects on production of arctic copepods have been conducted. Among these, the relationship between temperature, food concentration, and reproduction has been established for *C. finmarchicus* and *C. glacialis* (Hirche & Kwasniewski 1997, S.J. Madsen et al. 2008, Kjellerup et al. 2012). However, information on the effects of temperature on the functional biology of *C. hyperboreus* is limited (Hirche 1987, Plourde et al. 2003).

The aim of the present study was therefore to investigate the effects of temperature and food availability on feeding and egg production of *Calanus hyperboreus* in Disko Bay before and during the phytoplankton spring bloom. In parallel, bloom dynamics and *in situ* egg production of *C. hyperboreus* were followed in Disko Bay.

MATERIALS AND METHODS

Study site

Sampling was conducted from 10 February to 25 May 2009 about 1 nautical mile off the coast of Qeqertarsuaq in Disko Bay, western Greenland (69° 15' N, 53° 33' W, Fig. 1) The station is 250 to 300 m deep and was previously used in studies of the pelagic community of the bay (Nielsen & Hansen 1995, Madsen et al. 2001, S.J. Madsen et al. 2008, Dünweber et al. 2010). Sampling on 10 February and from 17 April to 25 May was carried out from a research vessel. On all other sampling dates, samples were taken through a hole made in the sea ice.

Hydrography and phytoplankton

Temperature and salinity in the water column were measured using a Seabird SBE25-01 CTD, and water samples from 1, 20, 50, 75, 100, 150, 200, and 250 m were taken with a 30 l Niskin water bottle. Water samples were kept cold and dark in 10 l plastic containers and transported back to the laboratory. Here, 500 ml triplicates from each depth were filtered onto GF/F filters and extracted overnight in 5 ml 96% ethanol (Jespersen & Christoffersen 1987) and fluorescence was measured on a Turner Design Model 700 fluorometer before and after acid addition. Salinity measurements were calibrated against salinity samples taken approximately once a month ($n = 4$) throughout the study phase, and analyzed on an 8410-Portasal salinometer (Guildline).

Depth distribution of *Calanus hyperboreus*

Female *Calanus hyperboreus* were sampled on 10 February and 17 April in 50 m depth intervals ($n = 5$) from 250 m to the surface. This was done using a Hydrobios Multinet (type Midi, opening 0.25 m²) with nets of 50 μ m in mesh size. The samples were immediately preserved in buffered formalin (2% final concentration), and in the laboratory we quantified the proportion of mature females ready to spawn (i.e. with visible orange gonads, stage GS4 according to Niehoff & Hirche 1996, Niehoff & Runge 2003).

In situ egg production

Calanus hyperboreus females were sampled from the bottom to the surface using a WP-2 net (200 μ m) equipped with a large non-filtering cod end. The samples were diluted and stored in a thermobox. In the laboratory, mature females (GS4 sensu Niehoff & Hirche 1996, Niehoff & Runge 2003; 13 to 20 replicates) were sorted out and placed individually in 600 ml polycarbonate bottles filled with 45 μ m screened surface water. The bottles were incubated at 5°C for 48 h, after which the content of each bottle was concentrated on a 45 μ m filter. The eggs were counted and the prosome length of the females measured. All eggs spawned within the 48 h were considered to belong to the same clutch (spawning interval at 4.5°C for *C. hyperboreus* is 5.6 d, Plourde et al. 2003). As only mature females with visible well-developed gonads (GS4) were incubated, the egg production (EP) rate measured would overestimate population EP. Therefore, EP rates were corrected for maturity of the female population by multiplying the observed EP by the proportion of mature females in the population based on the biomass samples (see Fig. 5a, Table 3). As carbon content of the females decreased by more than 50% over the period investigated, none of the previously established length-weight regressions could be used to estimate carbon content of females. An exponential decrease in dry weight (DW) over the spring has been demonstrated for *C. hyperboreus* (Conover & Siferd 1993). Therefore, average carbon content of females was esti-

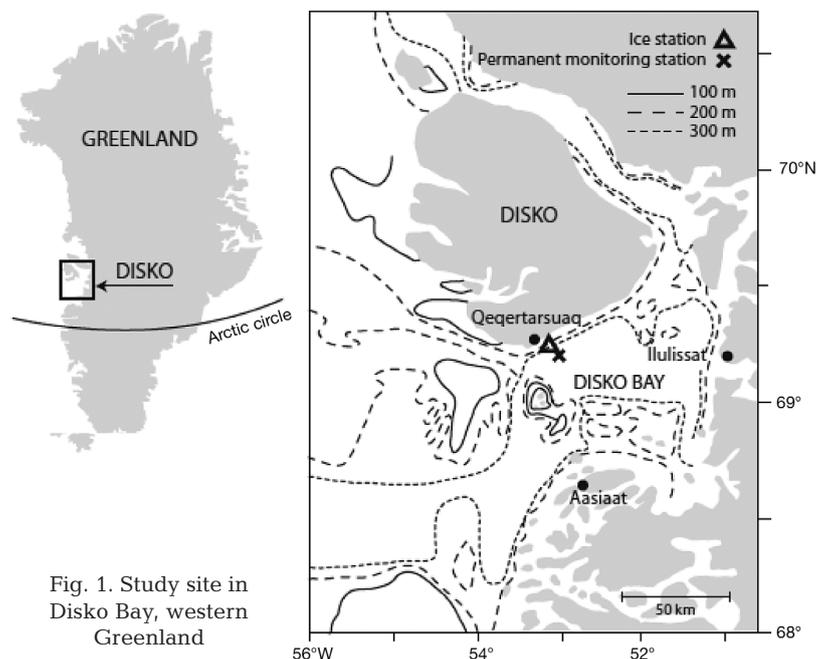


Fig. 1. Study site in Disko Bay, western Greenland

mated for each date using an exponential regression between *in situ* carbon content of females collected on 10 February and 17 April (see Table 4). Eggs from females sampled on 10 February were collected and immediately measured, and a mean egg volume was calculated assuming a spherical shape. The carbon content of eggs was calculated using a volume to carbon conversion factor for *C. glacialis* and *C. finmarchicus* of $1.10 \times 10^{-7} \mu\text{g C } \mu\text{m}^{-3}$ (Swalethorp et al. 2011). The carbon contents of females and eggs were then used to calculate specific EP (SEP). To estimate average total fecundity of females, an exponential regression was fitted to the observed EP. Using this regression, a new daily EP was estimated and summed over the period of investigation.

Laboratory experiment

The laboratory experiment was conducted twice, each time over a 2 wk period. The first experiment was set up on 10 February, before the spring bloom, and the second on 17 April, during the spring bloom. Females used in the experiments were collected in the same manner as for the *in situ* EP experiment, and kept on ice during handling.

Setup

Within 3 h after the mature females (GS4) were sampled, they were carefully sorted out and incubated at 5 different temperatures: 0, 2.5, 5, 7.5, and 10°C. Before starting each experiment, the copepods were acclimated to the temperature for 3 to 6 d, and individuals that were fed later, during the experiment, were acclimated to food concentration for 3 d. Thirty females were used at each temperature, half of which were kept starved in 0.2 μm filtered sea water and the other half kept under saturated food conditions in 0.2 μm filtered sea water with 15 μg chlorophyll (chl) a l^{-1} of the diatom *Thalassiosira weissflogii* (equal to 680 $\mu\text{g C l}^{-1}$; Reigstad et al. 2005). Cultures of *T. weissflogii* were grown in a 12:12 light:dark cycle (2 Osram L, 36 W/840, Lumilux cool white placed 40 cm away) in 0.2 μm filtered seawater at room temperature, and B₁ medium (1 ml l^{-1}) (Hansen 1989), silicate (0.9 ml l^{-1}), and vitamins (0.5 ml l^{-1}) were added every other day. The cultures were renewed every 1 to 2 wk and were constantly aerated.

Five thermo boxes filled with freshwater were used to keep the temperatures constant. Hobo thermo loggers were used throughout the experiment to log the

Table 1. Temperature in laboratory experiments

Intended temperature (°C)	Measured temperature \pm SD (°C)	
	Pre-bloom	Bloom
0	0.5 \pm 0.7	0.7 \pm 1.3
2.5	2.6 \pm 0.3	2.7 \pm 0.2
5	5.1 \pm 1.0	5.8 \pm 0.8
7.5	7.3 \pm 0.7	7.5 \pm 0.4
10	10.0 \pm 0.4	10.1 \pm 0.5

temperature every 15 min (Table 1). In each thermo-box, 2 buckets (10 l each) filled with 8.3 l filtered sea water (0.2 μm) were placed, and *Thalassiosira weissflogii* was added to one of these. In each bucket, 15 female *Calanus hyperboreus* were contained in a cylinder with false bottom (400 μm mesh). Every day the cylinders were carefully transferred to new buckets with 2.5 l filtered water at the corresponding temperature. The water from the old buckets was filtered with a 45 μm filter by reverse filtration, and the concentrated samples were collected and preserved in Lugol's solution (2% final concentration). Finally, 5.8 l of this filtered water were transferred to the new buckets and phytoplankton culture was added to adjust food concentration for the fed females. The eggs and pellets collected in the experiment were counted daily. Length and width of approximately 30 pellets from every temperature, both starved and fed, were measured on Days 2, 7, and 14 for both experiments in order to calculate an average fecal pellet volume. Only pellets with length at least 3 times their width were counted and measured.

Mortality in the 2 experiments averaged 1% d^{-1} . During the experiment, dead females were removed, their prosome length measured, and subsequently replaced with new individuals previously starved and kept at 5°C. The females were acclimated to the proper temperature for approximately half a day before being added to the buckets. At the end of both experiments, prosome length of every individual was measured and a mean female length in each treatment was calculated.

Fecal pellet production as a proxy for grazing

All fecal pellet measurements from the starved treatments were corrected for shrinkage due to fixation in Lugol's solution, as this reduces the volume of pellets from starved individuals by 21% (Kjellerup et al. 2012). Fecal pellet volumes for the fed and starved treatments in each experiment were then calculated

Table 2. *Calanus hyperboreus*. Mean \pm SD fecal pellet volume ($10^5 \mu\text{m}^3$) for fed and starved females in each experiment

	— Pre-bloom —		— Bloom —	
	n	Pellet volume	n	Pellet volume
Fed	460	48.1 \pm 23.7	456	32.9 \pm 13.0
Starved	425	36.6 \pm 20.7	174	12.3 \pm 6.5

from the length and width of pellets assuming that they were of a cylindrical shape. As no significant effect of temperature on pellet volume was detected, a mean volume for fed or starved females was calculated (Table 2). From these values, the carbon content was calculated using a conversion factor of $8.03 \times 10^{-8} \mu\text{g C } \mu\text{m}^{-3}$ (Reigstad et al. 2005) for the fed treatment and $4.75 \times 10^{-8} \mu\text{g C } \mu\text{m}^{-3}$ (Seuthe et al. 2007) for the starved treatment. These factors were measured in experiments with food concentrations comparable to this experiment using *Calanus finmarchicus* and *C. glacialis*. The carbon contents of females and fecal pellets were then used to calculate a cumulative carbon specific fecal pellet production (SPP_{cum}) for each treatment in each experiment (see Fig. 6).

Egg production

Together with the female carbon content, the mean carbon content of eggs (estimated as described for the *in situ* EP) was used to calculate the cumulative specific egg production (SEP_{cum}) for each treatment in each experiment (see Fig. 6).

Carbon measurements

Before each experiment, 24 of the females collected in the field were washed in filtered seawater ($0.2 \mu\text{m}$), their prosome length was measured, and they were placed in pre-weighed tin capsules. They were then dried for 24 h at 60°C and stored frozen (-30°C) for 8 to 10 mo. After re-drying the samples, the carbon content of each individual was measured on a CHNS Automatic Elemental Analyzer (EA 1110 CHNS, CE Instruments). This procedure was later repeated on approximately 7 females from each treatment after the experiments had ended. The carbon content was used to make a linear interpolation between the initial weight and the weight on the last day of each treatment for both experiments. These relationships were then used to estimate the carbon weight of females for each day of the experiments

and subsequently to calculate daily carbon specific egg and pellet production (SEP and SPP).

Lipid measurements

Approximately 20 females before the experiments and 5 females from each treatment after the experiments were placed individually in lipid test tubes with a Teflon cap. One ml chloroform:methanol solution (2:1 by volume) was added, and the samples were stored at -30°C for 2 to 4 mo and then at -80°C for 7 mo. Before analyses, an additional 2 ml chloroform:methanol solution was added. For a more detailed description, see Swalethorp et al. (2011).

Data analysis

The effects of temperature and food availability were tested with a general linear model (GLM, SAS Version 9.1) where the response (y) equals:

$$y = \text{intercept} + (k_{\text{food}} \times \text{food}) + (k_{\text{temp}} \times \text{temp}) \quad (1)$$

The model describes change in either SPP, SEP, carbon, nitrogen, or lipid content over the incubation period, where temp is the temperature in the experiment and food is a variable that has a value of 0 for starved females and 1 for fed females. On a few occasions (e.g. Eq. 5), the time of the year was included by adding a third term ($k_{\text{season}} \times \text{season}$), where the variable season has a value of 0 in the pre-bloom experiment and a value of 1 in the bloom experiment. The coefficients k_{food} , k_{temp} and k_{season} quantify the effects of temperature, food and season, respectively. During analysis of lipid content, the values for triacylglycerol (TAG) at 10°C were not included in the model, as they were unrealistically high and therefore considered as outliers (see Table 7).

SPP_{rate} and SEP_{rate} were estimated as the slopes in a 2-phase model using an iterative non-linear SAS procedure for each of the 10 different treatments to estimate the coefficients that best explained the observed rates for SPP_{cum} and SEP_{cum} . A visual inspection of the time course (see Fig. 6) clearly showed that the cumulative production increased linearly with time but also that a shift in the rate of production, both upward and downward, occurred during many of the experiments. In order to model this variability, a 2-phase model was constructed:

$$\begin{aligned} \text{if day} \leq l & \text{ then } p = \text{day} \times k_1 \\ \text{if day} > l & \text{ then } p = k_1 \times l + k_2 \times (\text{day} - l) \end{aligned} \quad (2)$$

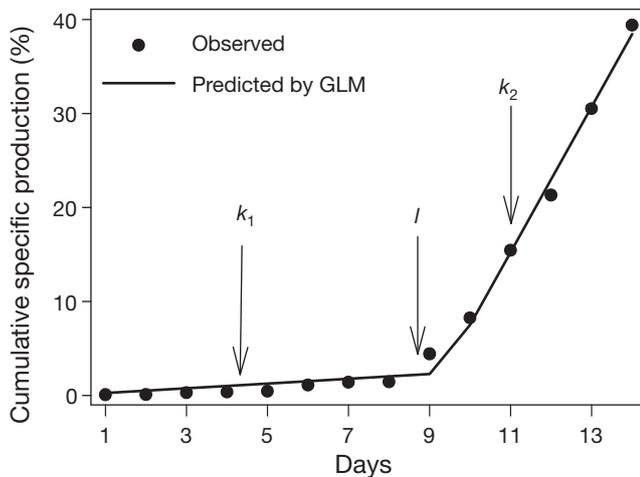


Fig. 2. Model used to establish specific fecal pellet production rate (SPP_{rate}) and specific egg production rate (SEP_{rate}) from the cumulative production. k_1 : slope of the line (% C d^{-1}) from Day 1 to l (the intercept between the fitted lines); k_2 : slope of the line (% C d^{-1}) from l to Day 14

where p is the cumulative production of pellets or eggs, k_1 and k_2 are the coefficients for the daily production, and l is the time where the shift from k_1 to k_2 occurs (Fig. 2). To avoid k_1 or k_2 being determined based on <3 data points, bounds were placed on l so that $3 \leq l \leq 13$. Tests were performed with a free estimate of l and with a constant value of $l = 6$, and they showed only minor deviations in the estimates of k_1 and k_2 . The parameters were estimated with SAS proc NLIN. Temperature coefficients for changes in k_1 and k_2 with temperature were estimated using a simple linear model followed by a t -test of the coefficient relative to 0. Carbon-specific values are given in % for SPP and SEP or as % d^{-1} for SPP_{rate} and SEP_{rate} ($\mu g C_{egg} \mu g C_{female}^{-1} d^{-1} \times 100$). Unless otherwise noted, all reported means are given \pm SE.

Energy budget for females

An energy budget was established following Auel et al. (2003) for the 2 experiments and for *in situ* development of EP, using the observed differences in total lipid content between the beginning and the end of the experiments, the number of eggs spawned, the lipid content of *Calanus hyperboreus* eggs ($0.54 \pm 0.01 \mu g$, i.e. $5.4 \times 10^{-4} mg egg^{-1}$; S. Jung-Madsen et al. unpubl.), an energy content of lipids of $42.7 J mg^{-1}$ (Conover 1964, Båmstedt 1986), and a respiration rate of females of $0.26 ml O_2 g^{-1} DW h^{-1}$ (Auel et al. 2003) converted to $10.4 ml O_2 g^{-1} C d^{-1}$ (assuming a carbon content of 60% of dry weight;

Omori 1969, Plourde et al. 2003). Finally, to convert respiration into daily energy requirements, an oxy-caloric equivalent of $19.64 J ml^{-1} O_2$ was assumed as typical for lipid-based metabolism (Ikeda et al. 2000). The energy budget for *in situ* EP was calculated by multiplying average female fecundity over the season (66 d) by lipid content of eggs and comparing it with the loss of female lipids occurring in the same period. Potential *in situ* EP (eggs $female^{-1} d^{-1}$) was calculated as:

$$EP_{potential} = \frac{(TL_{loss} \times 42.7 - 10.4 \times C_{females} \times 19.64 \times 66)/42.7}{5.4 \times 10^{-4}} \quad (3)$$

where TL_{loss} = loss of total lipids (mg) and $C_{females}$ = average carbon content of females (g) during the period. $C_{females}$ was estimated by averaging the carbon content of females calculated for each day over the period of 66 d assuming an exponential relationship between measurements on 10 February and 17 April.

RESULTS

Hydrography and phytoplankton

In February there was a clear pycnocline just below 100 m. The temperature increased from about $-1.6^\circ C$ in the surface layers to $3^\circ C$ in the bottom layers, and salinity increased from 32.9 in the surface to 34.2 at 250 m (Fig. 3A). The chl *a* concentration was very low throughout the water column, with values increasing toward the surface and reaching a maximum concentration at $0.05 \mu g l^{-1}$ at 20 m. Due to malfunction of the CTD, no CTD cast from April can be presented. Instead, Fig. 3B shows point measurements of temperature, salinity, and chl *a* made at 8 depths. In April, a weak pycnocline at about 50 m was present, but the main pycnocline was still situated just below 100 m. As in February, the temperature at the bottom was around $3^\circ C$. Chl *a* was found from the surface and down to 150 m, showing that the phytoplankton spring bloom was well underway. Highest chl *a* concentrations were found above 50 m, peaking at $1.6 \mu g l^{-1}$ at 1 m.

Depth distribution of *Calanus hyperboreus*

From February to mid-April, the majority of females were found in the deepest strata (Fig. 4) at

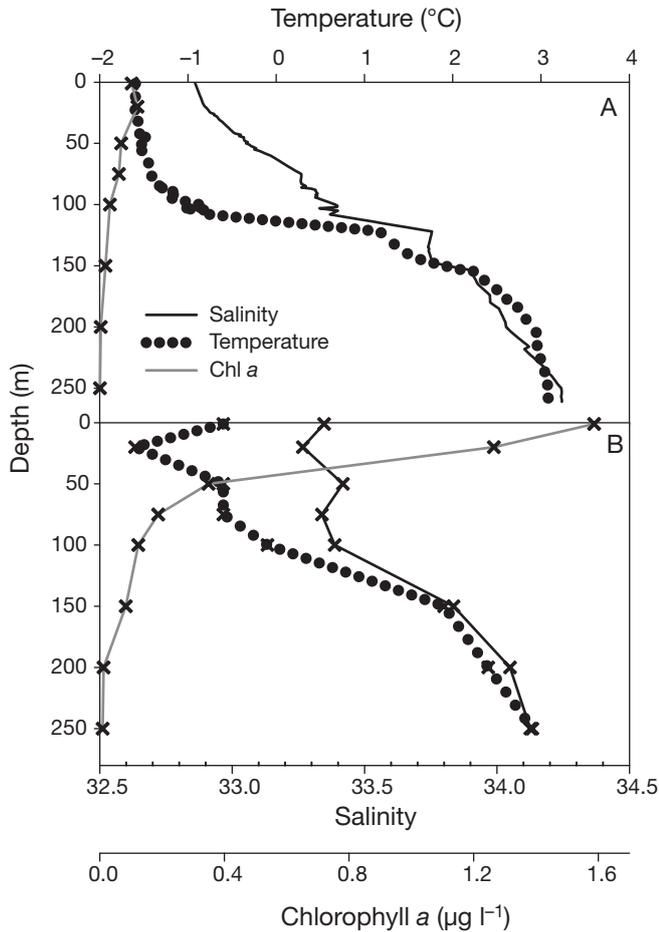


Fig. 3. Hydrography of Disko Bay on (A) 10 February and (B) 17 April 2009, showing salinity, temperature and chlorophyll concentration. (A) CTD data of salinity and temperature and point measurements of chl *a*. (B) Point measurements of all parameters at 8 depths (crosses) due to malfunction of CTD on 17 April

constant positive temperatures (3°C) and very low food concentration. At the beginning of the second experiment on 17 April, 10% of the females were found in the surface waters, indicating that the ascent towards the surface had just begun. By late April, the majority of females had ascended to surface waters.

In situ egg production

The proportion of ripe females and *in situ* EP were measured between 10 February and 17 April. Egg diameter was $198 \pm 7 \mu\text{m}$, giving an egg volume of $40.8 (\pm 5) \times 10^5 \mu\text{m}^3$ ($n = 110$, mean \pm SD). The measurements of *in situ* EP showed that EP was inversely correlated with the chl *a* concentration of the water

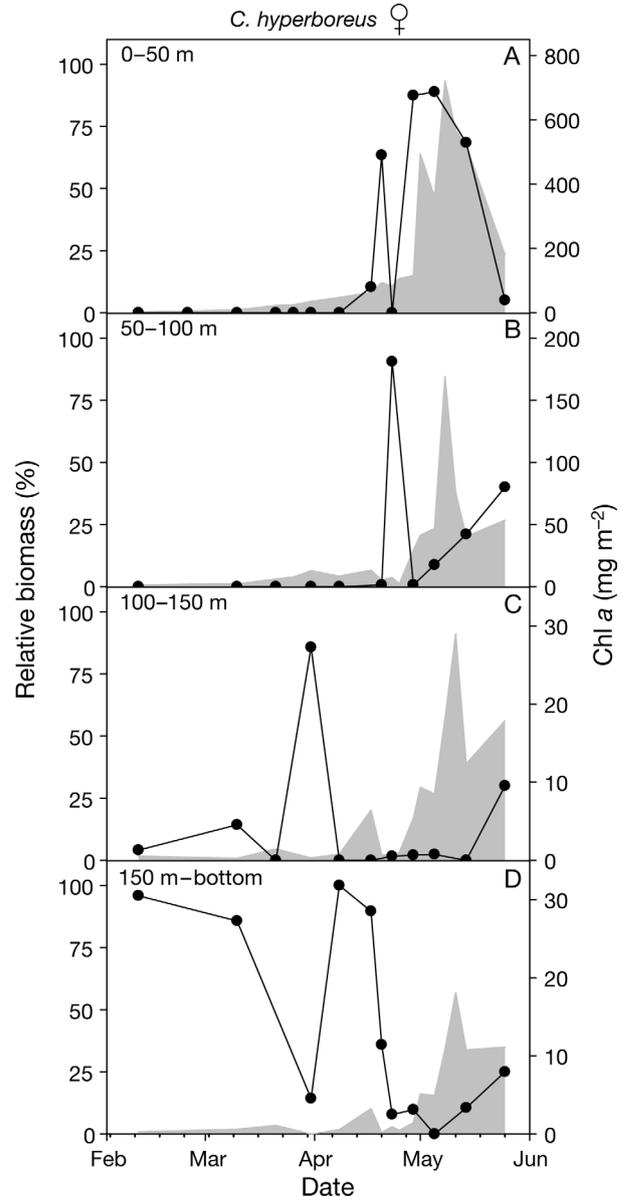


Fig. 4. *Calanus hyperboreus*. Relative depth distribution of females (left y-axis) and integrated chlorophyll (chl) *a* (shaded area; right y-axis: note different scale) in the different depths from 10 February to 25 May

(Fig. 5). Mean population EP was 54 ± 8 eggs female⁻¹ d⁻¹ before the spring bloom and ceased as the proportion of ripe females declined, until 17 April at the beginning of the spring bloom when spawning was terminated (Fig. 5, Table 3). Clutch size was quite variable, ranging between 9 and 416 eggs clutch⁻¹. During the main spawning event (February to March), average clutch size ranged between 107 ± 19 and 165 ± 52 eggs, whereas in early April when EP had ceased, clutch size averaged 30 ± 5 eggs (Table 3).

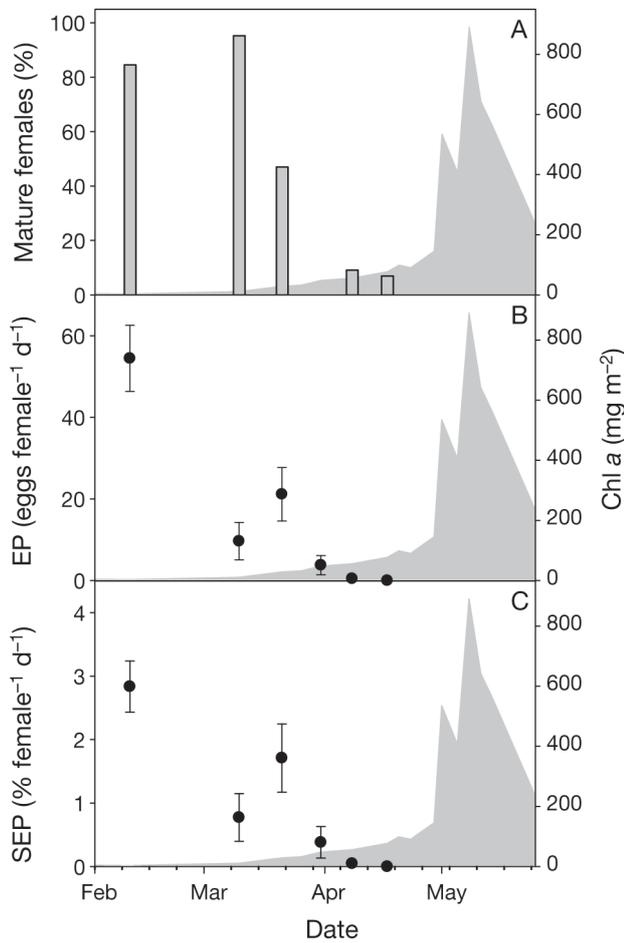


Fig. 5. *Calanus hyperboreus*. (A) Percentage of mature females, (B) *in situ* egg production (EP) \pm SE, and (C) specific *in situ* egg production (SEP) \pm SE, between February and April 2009. The shaded area is integrated chlorophyll in the upper 100 m

Mean SEP started at $3.5 \pm 0.5\% \text{ d}^{-1}$ and declined to $0.06 \pm 0.03\% \text{ d}^{-1}$ on 8 April until it reached 0 on 17 April. During the same period, the integrated chl *a* concentration down to 100 m increased from 3.2 to 76.9 mg m^{-2} .

Laboratory experiment

Surprisingly, we observed no positive effect of temperature on either egg or fecal pellet production in the pre-bloom or bloom period. However, food had a clear positive effect on fecal pellet production, whereas there was no effect on EP (Fig. 6).

Pellet production as a proxy for grazing

The mean SPP_{cum} after 2 wk varied from 0.1 to 7.9% in the 4 groups of experiments. The separate GLMs for the pre-bloom and bloom experiments showed a strong positive effect of food for both periods (Table 4). Also, pellet production was observed in the experiment without food, and even though the intercept in the GLM (estimated value at 0°C without food) was not significantly positive, the mean SPP_{cum} after 2 wk at higher temperatures was significantly different from 0 (Table 4). The pellets produced by starved females were clear and empty 'ghost type' pellets (Seuthe et al. 2007, Kjellerup et al. 2012). There was no significant effect of temperature on SPP_{cum} , but both coefficients were positive (with and without food, Table 4) and the temperature coefficient in a model for just the pre-bloom experiment without food was significantly positive:

$$\text{SPP}_{\text{cum}} = 10.3 \pm 2.9 \text{ (} p = 0.038 \text{)} + (1.57 \pm 0.47 \text{ (} p = 0.046 \text{)} \times \text{temp}), r^2 = 0.78 \quad (4)$$

The effects of temperature and season were also significant in a common GLM for all experiments:

$$\text{SPP}_{\text{cum}} = 8.9 \pm 5.1 \text{ (} p = 0.1 \text{)} + (60 \pm 4.6 \text{ (} p < 0.0001 \text{)} \times \text{food}) + (1.8 \pm 0.6 \text{ (} p = 0.01 \text{)} \times \text{temp}) - (17.0 \pm 4.6 \text{ (} p = 0.0019 \text{)} \times \text{season}), r^2 = 0.92 \quad (5)$$

Thus, overall there was a tendency for a positive effect of temperature on SPP_{cum} .

Fig. 7 shows a more detailed pattern for the relationship between SPP_{rate} , time, temperature, and food availability. For the pre-bloom experiment, there was an increase in the SPP_{rate} over time as k_2 was higher than k_1 , whereas the opposite was observed in the

Table 3. *Calanus hyperboreus*. *In situ* reproductive values for females used to correct the measured egg production (EP) for maturity of females and to calculate specific EP rates. Maturity is the percentage of mature females (stage GS4, see 'Materials and methods') found in the upper 200 m, and spawning frequency is the proportion of mature females that produced eggs during the 48 h incubation. Daily EP and clutch size are given \pm SE, n is the number of females incubated

Date (dd/mm/yy)	Maturity %	Spawning frequency %	Clutch size \pm SE (range): eggs clutch ⁻¹	EP \pm SE: eggs female ⁻¹ d ⁻¹	n
10/02/09	85	95	134 \pm 17 (17–279)	64 \pm 9	18
12/03/09	93	20	107 \pm 19 (73–148)	10 \pm 5	20
21/03/09	47	50	165 \pm 52 (40–416)	45 \pm 15	20
31/03/09	9	25	117 \pm 67 (19–203)	14 \pm 8	12
08/04/09	7	33	30 \pm 15 (9–45)	5 \pm 2	13
17/04/09	0	0	0	0	19

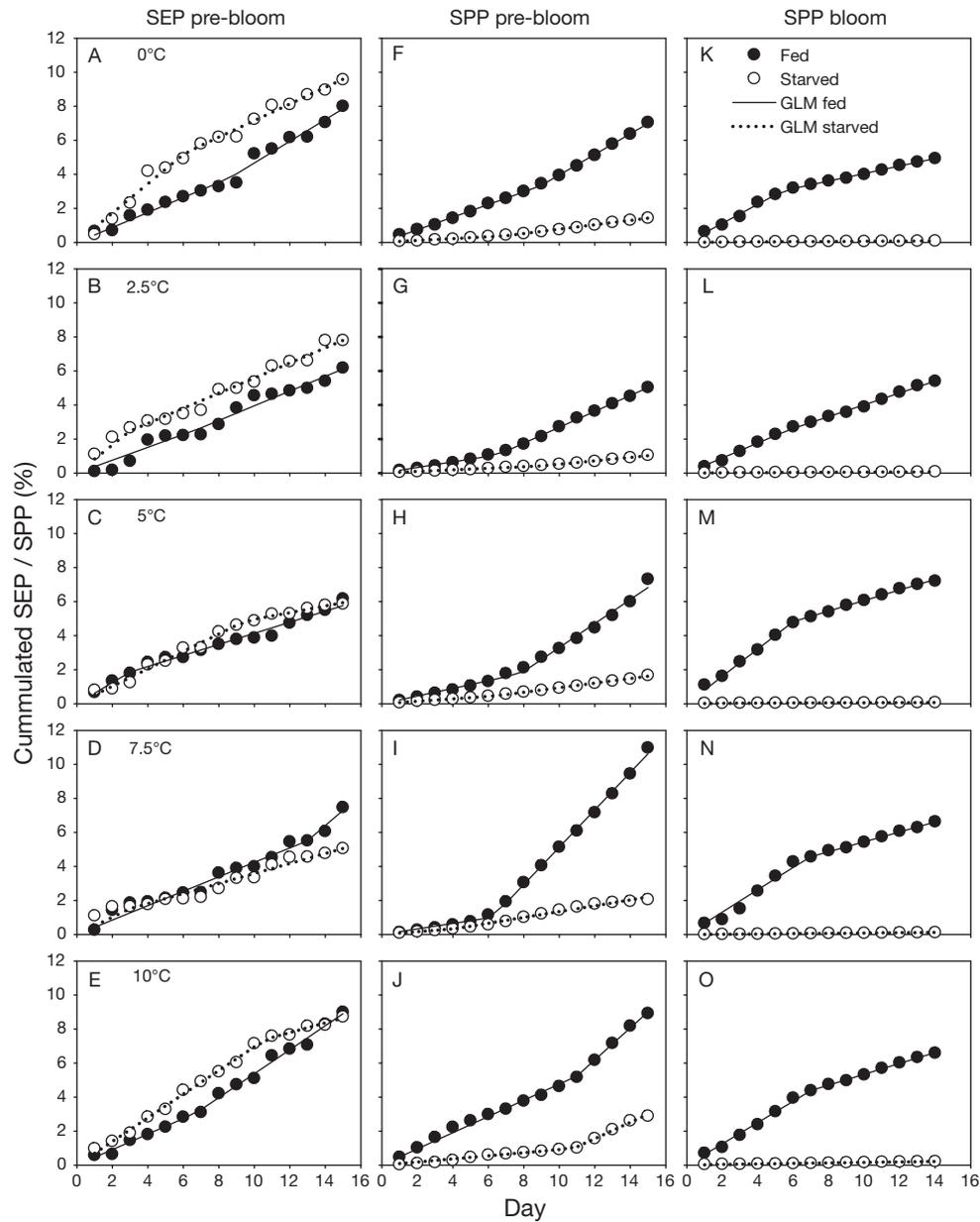


Fig. 6. *Calanus hyperboreus*. Cumulative specific egg production (SEP_{cum}) and cumulative specific fecal pellet production (SPP_{cum}) before and during spring bloom at 0, 2.5, 5, 7.5, and 10°C. Filled and empty symbols: fed and starved females, respectively. Lines: values of production predicted by the general linear model and used for estimating k_1 and k_2 (see Eq. 2 and Fig. 2)

Table 4. *Calanus hyperboreus*. Cumulative specific pellet and egg production (SPP_{cum} and SEP_{cum}) at the end of each experiment. Intercept and coefficients for general linear models (GLM, Eq. 1) as a function of temperature and food availability are given for the 2 periods of the season. Mean values are calculated across 5 experiments at temperatures from 0 to 10°C ($n = 5$). All values given \pm SE; p-values are given in parentheses, with significant values in **bold**

		Mean values (%)		Intercept (%)	GLM parameters	Food (%)
		Fed	Starved		Temp. ($^{\circ}\text{C}^{-1}$)	
SPP_{cum}	Pre-bloom	7.9 ± 1.0	1.8 ± 0.3	0.46 ± 0.84 (0.60)	0.27 ± 0.12 (0.057)	6.0 ± 0.84 (0.0002)
	Bloom	6.2 ± 0.4	0.1 ± 0.03	-0.37 ± 0.37 (0.35)	0.01 ± 0.5 (0.11)	6.0 ± 0.37 (<0.0001)
SEP_{cum}	Pre-bloom	7.4 ± 0.5	7.4 ± 0.9	7.5 ± 1 (0.002)	-0.03 ± 0.19 (0.88)	-0.05 ± 1.3 (0.97)

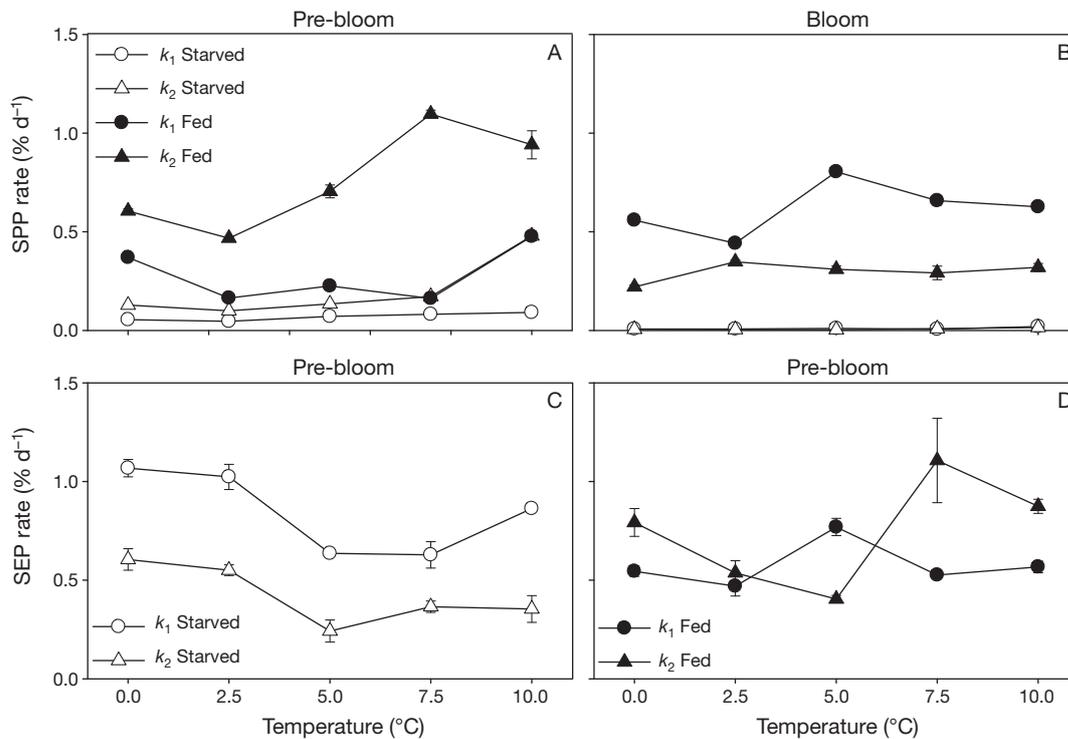


Fig. 7. *Calanus hyperboreus*. (A, B) Specific fecal pellet production rates (SPP_{rate}) \pm SE before and after the bloom, and (C, D) specific egg production rates (SEP_{rate}) \pm SE before the bloom as a function of temperature. k_1 (k_2) represents the first (last) part of the experiment. The filled (empty) symbols are fed (starved) females

bloom experiment with food (Fig. 7A,B). The SPP_{rate} in the pre-bloom experiment ranged from 0.16% d⁻¹ (k_1 at 7.5°C) to 1.1% d⁻¹ (k_2 at 7.5°C) for fed females and from 0.046% d⁻¹ (k_1 at 2.5°C) to 0.48% d⁻¹ (k_2 at 10°C) for starved females. During the bloom experiment, the SPP_{rate} for fed females ranged from 0.2 (k_2 at 0°C) to 0.8% d⁻¹ (k_1 at 5°C). In the starved treatments, almost no fecal pellets were produced, thus specific values were always lower than 0.019% d⁻¹ (k_1 at 10°C). Changes in k_1 and k_2 with temperature were analyzed with linear regression. The only experiment with a significant relationship between SPP_{rate} and temperature was the pre-bloom experiment without food. Here, the SPP_{rate} increased by $0.044 \pm 0.011\%$ °C⁻¹ ($p = 0.026$). For all other experiments, the relationship with temperature was positive but not significant (data not presented); however, as shown in Eqs. (4) & (5), the SPP_{cum} after 2 wk was significantly positively related with temperature.

Egg production

Values for EP only exist for the pre-bloom experiment, as the females had stopped spawning at the

beginning of the bloom experiment (Fig. 5). SEP_{cum} over 2 wk was independent of both temperature and food availability (Table 4). Although food availability had no effect on SEP_{cum} , it had a pronounced effect on the time course of EP (Fig. 7C,D). In general, fed females had a lower SEP_{rate} at all temperatures in the first part of the experiment (k_1) compared to starved females, whereas the rate values were reversed in the later part of the incubation (k_2), such that after 14 to 15 d there was no effect of food.

SEP rates varied from 0 to 1.1% d⁻¹. Maximal SEP rates were found at the lower temperatures for starved females (1.07 and 1.02% d⁻¹, k_1 at 0 and 2.5°C, respectively) and at high temperatures for fed females (1.11 and 0.87% d⁻¹, k_2 at 7.5 and 10°C, respectively). Nevertheless, there was no significant effect of temperature on SEP_{rate} for either fed or starved females (Table 4).

Carbon, nitrogen, and lipid content

Overall, *Calanus hyperboreus* lost carbon during the experiments (Table 5, Fig. 8). The loss was most pronounced in the pre-bloom experiment where the

Table 5. *Calanus hyperboreus*. Total carbon (C) and nitrogen (N) content at the end of each experiment. Initial values ($\mu\text{g female}^{-1}$) represent values on Day 0 ($n = 24$). Mean end values are followed by changes in percent of the initial value ($\Delta\%$) across 5 experiments at temperatures from 0 to 10°C ($n = 34$ to 36). Intercept (%) and coefficients for general linear models (GLM; Eq. 1) for the changes in percent of start values as a function of temperature ($\% ^\circ\text{C}^{-1}$) and food availability (%) are also given for the 2 periods of the season. All values given \pm SE; p-values are given in parentheses, with significant values in **bold**

		Initial value	Mean end values		Intercept (%)	GLM parameters	
		($\mu\text{g female}^{-1}$) <i>In situ</i>	Fed	Starved		Temp. ($\% ^\circ\text{C}^{-1}$)	Food (%)
C	Pre-bloom	1692 \pm 107	1091 \pm 77, -36 \pm 5	1140 \pm 91, -33 \pm 5	-25.8 \pm 7.0 (0.0005)	-1.4 \pm 1.0 (0.18)	2.9 \pm 7.0 (0.68)
	Bloom	716 \pm 41	746 \pm 34, 4 \pm 5	592 \pm 31, -17 \pm 4	-14.0 \pm 6.4 (0.03)	-0.6 \pm 0.9 (0.48)	21 \pm 6.4 (0.0013)
N	Pre-bloom	206 \pm 11	178 \pm 8, -14 \pm 4	165 \pm 10, -20 \pm 5	-14.2 \pm 6.2 (0.02)	-1.1 \pm 0.9 (0.20)	6.1 \pm 6.2 (0.33)
	bloom	127 \pm 4	155 \pm 5, 22 \pm 4	116 \pm 3, -8 \pm 2	-10.0 \pm 4.4 (0.03)	0.3 \pm 0.6 (0.61)	31 \pm 4.4 (<0.0001)

average loss for both fed and starved females after 2 wk was 34% of the initial carbon content. In the bloom experiment, the initial carbon content of the females had decreased by 58% compared to the pre-bloom experiment. After 2 wk of incubation, a significant difference between fed and starved treatments was observed (Table 5). Fed females were able to maintain their starting weight or even gain weight during the experiment, whereas starved females showed a net loss of 17% carbon. Food availability had a positive effect on the carbon content in the bloom experiment ($p = 0.0013$), whereas the effect was insignificant in the pre-bloom experiment ($p = 0.68$). The effect of temperature on final carbon weight was not significant in either of the 2 experiments when tested separately or when tested with a GLM across the 2 periods. There was, however, a tendency toward a negative effect of temperature of about $1\% ^\circ\text{C}^{-1}$ in both experiments (Table 5).

The overall pattern for changes in nitrogen content resembled that of carbon. There was a loss in nitrogen content at all temperatures of between 8 and 20% except for the bloom experiments with food, where the nitrogen content increased by 22%. There was no effect of temperature or food on the nitrogen loss in the pre-bloom experiment, whereas in the bloom experiment there was a clear positive effect of food availability (Table 5).

Similar to the pattern described for carbon and nitrogen content, there was an overall loss of total lipids during the experimental period. The lipid loss was generally not related to either food or temperature (Table 6, Fig. 9). The lipid content of the females was analyzed in 5 groups: total lipids (TL), wax esters (WE), triacylglycerol (TAG), phospholipids (PL), and sterols (STE). As STE constituted less than 2% of TL, and no significant change during the experiments were observed, results are not included in this section.

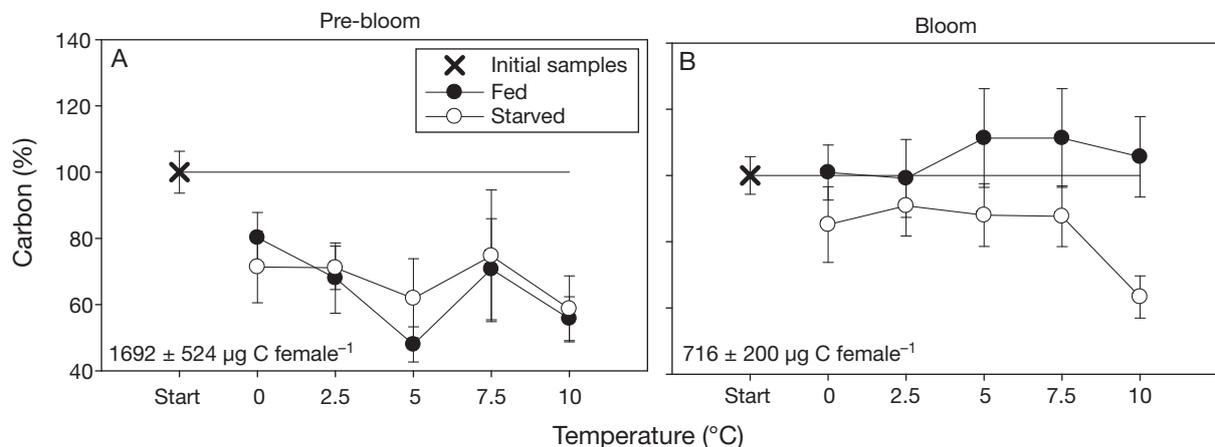


Fig. 8. *Calanus hyperboreus*. Carbon content at the end of the incubation period for the pre-bloom and bloom experiment at temperatures from 0 to 10°C . Values are given in % of start content \pm SE. Filled and empty symbols: fed and starved females, respectively, the straight line represents unchanged carbon content, and the cross is the carbon value at the beginning of each experiment. The initial carbon value is given at the bottom of each panel

Table 6. *Calanus hyperboreus*. Total lipids (TL), wax esters (WE), triacylglycerol (TAG), and phospholipids (PL) at the end of each experiment. Initial values ($\mu\text{g female}^{-1}$) represent values on Day 0 ($n = 15$). Mean end values are followed by changes in percent of the initial value ($\Delta\%$), across 5 experiments at temperatures from 0 to 10°C ($n = 22$ to 33). Intercept (%) and coefficients for general linear models (GLM; Eq. 1) for the changes in percent of start values as a function of temperature (°C) and food availability are also given for the 2 periods of the season. All values given \pm SE; p-values are given in parentheses, with significant values in **bold**

		Initial value ($\mu\text{g female}^{-1}$) <i>In situ</i>	Mean end values ($\mu\text{g female}^{-1}$, $\Delta\%$)		Intercept (%)	GLM parameters Temp. ($\% \text{ } ^\circ\text{C}^{-1}$)	Food (%)
			Fed	Starved			
TL	Pre-bloom	997 \pm 110	606 \pm 54, -39 \pm 5	478 \pm 59, -52 \pm 6	-47.9 \pm 7.6 (<0.0001)	-1.0 \pm 1.2 (0.43)	13.6 \pm 8.4 (0.11)
	Bloom	264 \pm 26	230 \pm 24, -16 \pm 9	221 \pm 29, -25 \pm 9	-2.4 \pm 14.6 (0.87)	-2.66 \pm 2.0 (0.19)	1.8 \pm 14.2 (0.90)
WE	Pre-bloom	919 \pm 101	518 \pm 50, -44 \pm 5	419 \pm 55, -54 \pm 6	-50.4 \pm 7.7 (0.0001)	-0.9 \pm 1.2 (0.45)	11.5 \pm 8.5 (0.18)
	Bloom	234 \pm 25	194 \pm 23, -21 \pm 9	194 \pm 28, -23 \pm 10	2.02 \pm 15.6 (0.90)	-3.6 \pm 2.2 (0.10)	2.5 \pm 15.2 (0.85)
TAG	Pre-bloom	9.6 \pm 1.3	12.4 \pm 1.1, 30 \pm 12	9.3 \pm 1.3, -3.2 \pm 14	-1.9 \pm 17.6 (0.91)	-0.3 \pm 2.8 (0.92)	33.3 \pm 19.6 (0.09)
	Bloom	9.6 \pm 0.4	2.5 \pm 0.2, -74 \pm 2.3	2.1 \pm 0.2, -78 \pm 2.1	-76.1 \pm 3.2 (<0.0001)	-0.5 \pm 0.6 (0.35)	3.7 \pm 3.2 (0.26)
PL	Pre-bloom	63 \pm 8	70 \pm 5, 11 \pm 8	47 \pm 5, -25 \pm 7	-19.9 \pm 9.9 (0.05)	-1.3 \pm 1.6 (0.43)	37.7 \pm 11.1 (0.0013)
	Bloom	19 \pm 3	29 \pm 2.5, 52 \pm 13	19 \pm 1.7, -0.2 \pm 9	-3.1 \pm 16.4 (0.85)	-0.6 \pm 2.3 (0.78)	52.3 \pm 16.0 (0.0026)

However, data for STE content are available in Table 7. In the pre-bloom experiment, TL content of the females decreased by 45 to 70% in starved treatments and by 30 to 52% in fed treatments (Fig. 9, Table 7). The lipid composition was dominated by WE, which on average constituted 78 to 92% of TL in all treatments. The trend in WE therefore clearly mimicked the trend in TL (Fig. 9A,B). TAG constituted less than 3% of TL. PL constituted on average 9 to 18% of TL in all treatments. There was a clear positive effect of food on PL content, where PL increased in fed females and decreased in starved females. There was no significant effect of temperature but overall coefficients were negative (Table 6).

From pre-bloom to bloom experiment, the *in situ* content of lipids decreased by 74%. Despite this large decrease in TL, the amount of TAG remained the same (Table 6). The lipid composition of the females at the end of each experiment was similar to what was found in the pre-bloom experiment. WE dominated with 72 to 89% of TL, followed by PL (8 to 24%) and with TAG constituting less than 2% (Table 7). Again, the trend in WE mimicked the trend in TL, where no significant trend related to either temperature or food was apparent (Fig. 9E,F, Table 6). The amount of TAG decreased significantly in all treatments, ranging from 82 to 75% loss. The decrease was independent of temperature and food. The amount of PL increased for both fed and starved females at low temperatures, but at temperatures $>5^\circ\text{C}$, PL of starved females decreased, whereas PL in fed females continued to increase to a maximum of 181% at 10°C (Fig. 9H). As in the pre-bloom, the effect of food was highly significant, whereas the effect of temperature was not (Table 6).

DISCUSSION

In situ condition

The spring bloom in 2009 was well underway in mid-April when spawning of *Calanus hyperboreus* had terminated (Fig. 5). This confirms that EP in *C. hyperboreus* is decoupled from the phytoplankton spring bloom, which has previously been shown in Disko Bay (Madsen et al. 2001), the Greenland Sea (Hirche & Niehoff 1996), and the Barents Sea (Melle & Skjoldal 1998). The relative distribution of *C. hyperboreus* females showed that they were at overwintering depths in February and had only just started their ascent in mid-April when the chlorophyll content of the water was rising.

The *in situ* EP showed a maximum of 54 eggs female $^{-1}$ d $^{-1}$ in February, after which EP decreased steadily until mid-April when spawning ended. Madsen et al. (2001) measured *in situ* EP of *Calanus hyperboreus* in Disko Bay on 1 occasion in the middle of March 1997 and found EP to be 33.3 \pm 3.4 eggs female $^{-1}$ d $^{-1}$. In our study, EP in March ranged between 10 and 21 eggs female $^{-1}$ d $^{-1}$. In the Greenland Sea, a maximum production of 23 eggs female $^{-1}$ d $^{-1}$ was found in February 1988 and 1989, whereas data from November and December showed an EP as high as 148 eggs female $^{-1}$ d $^{-1}$ (Hirche & Niehoff 1996). Generally, higher EP rates were found in November and December with values decreasing towards March. This corresponds well with what was shown in our study, viz. a clear reduction in EP as spring approached. When average female fecundity was estimated from February to April, a total number of

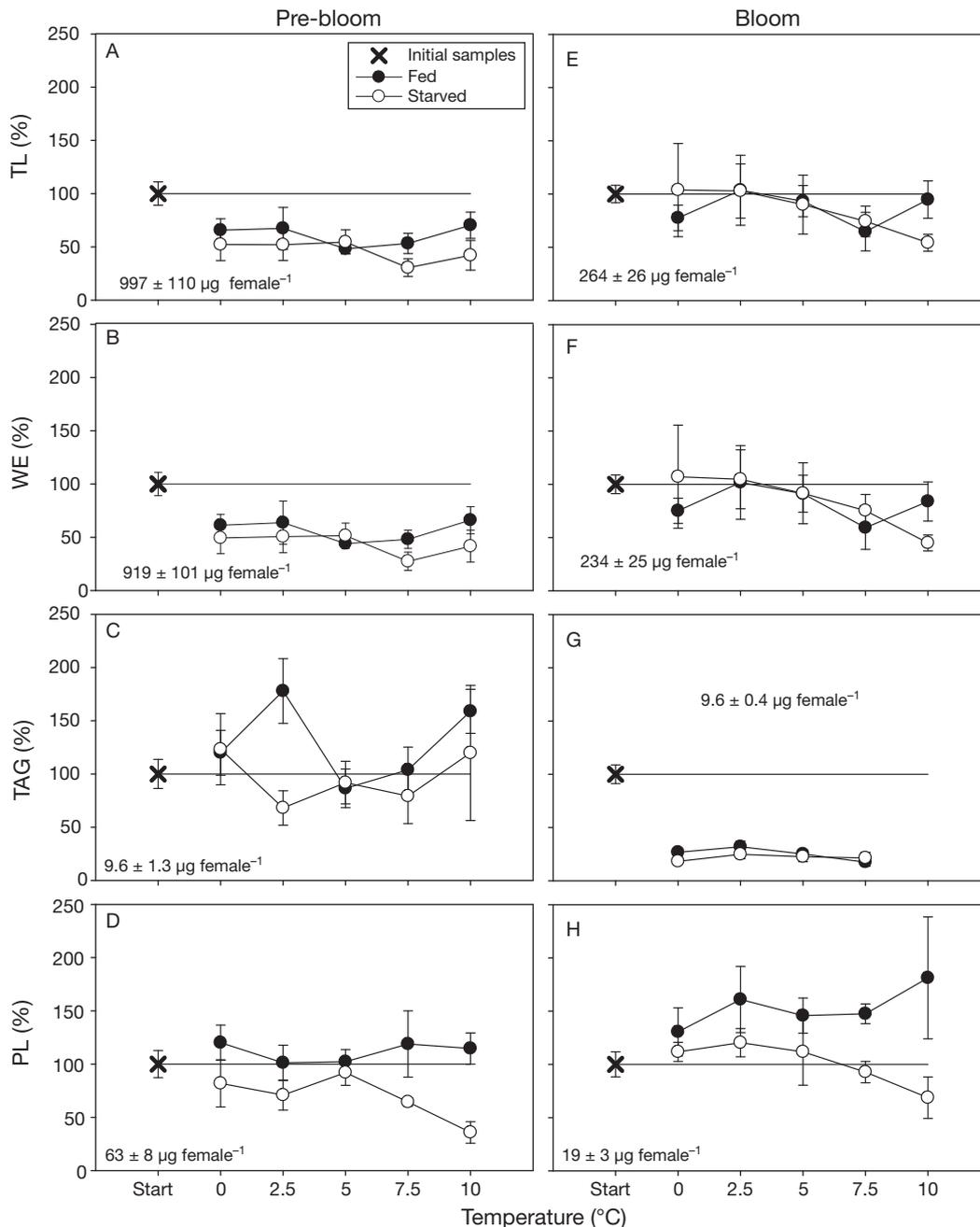


Fig. 9. *Calanus hyperboreus*. Total lipid (TL), wax ester (WE), triacylglycerol (TAG), and phospholipids (PL) at the end of the incubation period for the pre-bloom and bloom experiment at temperatures from 0 to 10°C. Values are given per female as % of start content ± SE. Filled and empty symbols: fed and starved females, respectively, the straight line represents unchanged lipid content, and the cross is the lipid value at the beginning of each experiment. The initial lipid value is given in each panel

1164 eggs female⁻¹ was found. During the same period, a decrease in lipid content of 74% was seen. This number compares well with previous studies where female fecundity was measured in the laboratory. Conover (1967) found female EP ranging from 429 to 3397 eggs female⁻¹ yr⁻¹, while other studies have observed average fecundity between 762 and

1500 eggs female⁻¹ (Conover & Siferd 1993, Hirche & Niehoff 1996, Plourde et al. 2003) and a carbon loss over the same period of 81% (Plourde et al. 2003). Comparing the number of eggs laid over the spawning period with the amount of TL lost in that same period and knowing the TL content of eggs, it was calculated that 86% of the lost lipids were converted

into eggs. However, this number leaves too little energy to cover metabolic costs. If instead a potential EP was calculated based on the lipid loss minus the energy needed for sustaining metabolism during the period (assuming a respiration rate of 0.26 ml O₂ gDW h⁻¹ and carbon content of 60% of DW), potential EP would be only 693 eggs female⁻¹, which equals 51% of the lost lipids and compares well with the assumption that 42% of an observed loss in *C. hyperboreus* female DW would be converted into reproductive products (Conover & Siferd 1993).

Laboratory experiments

Egg production

The SEP_{rate} in the laboratory experiment showed no significant temperature or food dependence, suggesting that EP is determined by the lipid content of the female and not by environmental conditions during spawning. As a positive effect of temperature was documented for arctic *Calanus glacialis* in similar laboratory experiments (Kjellerup et al. 2012), it was surprising not to observe any temperature response in *C. hyperboreus*. Kjellerup et al. (2012) showed that SEP_{rate} of *C. glacialis* in a pre-bloom situation peaked at 7.5°C. The SEP_{rate} would be expected to increase with temperatures until a certain limit where high temperatures would no longer be beneficial. The results indicate that *C. hyperboreus* is a strictly Arctic species that does not benefit from higher temperatures when considering fecal pellets and EP.

A lack of response to temperature in feeding activity does not necessarily mean that *Calanus hyperboreus* is not 'impacted' by elevated temperature. Respiration in *C. hyperboreus* is lower in winter even during reproduction than in spring (Conover 1962), suggesting that animals are in 'dormancy' and optimize EP from lipid reserves. Compared with the 2 smaller *Calanus* spp.,

Table 7. *Calanus hyperboreus*. Mean ± SE of carbon (C), nitrogen (N), and total lipids (TL) in µg female⁻¹, at the beginning and the end of each experiment in the pre-bloom and bloom period and mean ± SE of pellet and egg production in the different incubations. n = number of replicates, length = prosome length of females in mm. Wax esters (WE), triacylglycerol (TAG), phospholipids (PL), and sterols (STE) all in % of TL, and pellet production (PP) and egg production (EP) in pellet or egg female⁻¹ d⁻¹

	Carbon and nitrogen				Lipids				Pellet and egg production						
	n	Length (mm)	C (µg)	N (µg)	C/N	n	Length (mm)	TL (µg)	WE (%)	TAG (%)	PL (%)	STE (%)	n	PP	EP
Pre-bloom															
Initial	24	6.2±0.04	1692±107	206±11	8.1	18	6.2±0.05	997±110	92±0.4	1.0±0.1	6.4±0.4	0.7±0.1	-	-	-
0-	7	6.3±0.13	1207±183	174±21	6.8	8	6.2±0.08	521±152	86±0.6	2.6±0.5	10.4±0.8	0.7±0.2	15	7.0±0.8	18.1±4.2
0+	7	6.4±0.04	1356±129	210±13	6.4	4	6.4±0.20	656±105	86±0.7	1.8±0.3	11.7±0.5	0.9±0.1	16	16.1±1.6	15.2±3.5
2.5-	7	6.4±0.1	1203±111	179±9	6.7	7	6.0±0.09	518±149	88±1.6	1.3±0.2	10.0±1.4	0.8±0.4	15	4.9±0.5	15.2±3.6
2.5+	7	6.4±0.04	1151±179	167±10	6.7	5	6.4±0.20	671±197	85±2.6	3.1±0.6	11.2±1.8	1.0±0.4	15	10.9±1.3	11.8±3.2
5-	8	6.4±0.03	1047±202	147±14	6.8	8	6.3±0.09	544±113	85±2.1	1.6±0.1	12.4±2.1	0.9±0.2	16	7.3±0.6	12.3±3.1
5+	7	6.4±0.15	811±90	153±9	5.3	5	6.3±0.02	480±43	84±2.0	1.7±0.3	13.7±1.8	0.7±0.2	15	12.4±1.2	11.0±2.1
7.5-	7	6.4±0.14	1264±336	186±42	6.4	5	6.4±0.10	303±83	78±5.4	2.6±0.3	17.7±4.9	1.5±0.3	15	10.9±1.5	10.2±2.9
7.5+	7	6.5±0.11	1196±258	188±30	6.1	5	6.4±0.10	531±95	84±3.1	2.0±0.5	13.4±3.5	0.9±0.2	15	20.0±3.3	13.6±3.3
10-	7	6.2±0.08	993±168	141±13	6.9	5	6.3±0.08	420±140	88±2.8	2.3±0.6	8.7±2.9	1.0±0.4	14	12.2±3.0	16.9±3.1
10+	7	6.3±0.05	943±112	170±16	5.5	5	6.4±0.10	700±123	85±2.3	2.5±0.6	11.1±1.5	1.0±0.5	14	18.4±1.9	16.2±2.8
Bloom															
Initial	24	6.4±0.04	716±41	127±4	5.6	16	6.3±0.07	264±26	88±1.1	4.1±0.4	7.5±0.9	0.7±0.2	-	-	-
0-	7	6.3±0.06	610±82	113±9	5.3	4	6.4±0.07	274±116	88±3.3	0.9±0.3	11.3±3.1	0.1±0.1	13	0.7±0.1	-
0+	7	6.4±0.07	722±60	140±6	5.1	5	6.5±0.10	205±32	86±0.9	1.6±0.5	12.1±0.9	0.7±0.3	16	9.0±1.3	-
2.5-	6	6.5±0.08	651±65	124±4	5.2	4	6.5±0.20	272±67	88±3.3	1.0±0.2	10.5±3.4	0.5±0.3	14	0.6±0.1	-
2.5+	7	6.4±0.08	710±84	155±9	4.5	5	6.4±0.01	274±87	84±2.6	1.5±0.3	13.9±2.5	0.8±0.4	15	10.0±0.8	-
5-	7	6.4±0.05	630±68	120±7	5.2	5	6.3±0.10	238±73	89±1.8	1.1±0.2	10.0±1.8	0.1±0.1	17	0.5±0.1	-
5+	7	6.3±0.07	797±107	160±13	4.9	4	6.4±0.20	247±39	85±3.8	1.0±0.1	13.0±3.8	1.3±0.1	15	13.5±1.7	-
7.5-	7	6.4±0.12	628±66	121±6	5.1	5	6.3±0.05	197±38	89±1.0	1.0±0.2	9.7±1.1	0.6±0.4	15	0.9±0.1	-
7.5+	7	6.4±0.08	743±40	160±9	4.7	4	6.4±0.10	171±48	73±10	1.2±0.4	23.8±8.9	2.2±1.3	15	13.4±2.1	-
10-	7	6.3±0.12	453±46	103±6	4.3	5	6.3±0.08	143±21	72±2.9	17.9±8.3	8.5±2.1	2.1±0.8	15	1.6±0.4	-
10+	7	6.6±0.17	756±87	158±13	4.8	4	6.4±0.20	251±46	77±4.7	8.3±3.9	13.0±3.4	1.7±0.2	14	13.1±1.5	-

inactive or 'dormant' copepods such as *C. hyperboreus* could respond differently to temperature during short-term experiments, as 'dormant' animals could take a longer time to 'wake up' and adapt/react to changes in incubation conditions. Using an energy budget of EP based on long-term experiments, Plourde et al. (2003) suggested that the reproductive potential of *C. hyperboreus* would decrease between 0 and 4°C even with a similar daily carbon loss because of elevated respiration costs.

As *Calanus hyperboreus* spawns prior to the spring bloom when no food is available, the lack of a positive effect of food on EP is expected. The 2 other *Calanus* species in Disko Bay do not spawn until the beginning of the bloom (Madsen et al. 2001, S.J. Madsen et al. 2008) and therefore show a completely different food response. A significantly lower EP has been found in starving females for both *C. glacialis* and *C. finmarchicus* (S.J. Madsen et al. 2008, Kjellerup et al. 2012). Even though no significant effect of food was found in our study after the 2 wk period, differences in the course of production were observed, where SEP_{rate} increased for fed females and decreased for starved females in the last part of the experiment (k_2 , Fig. 7C,D). Therefore, there might have been a positive effect of food if the experiment had continued for a longer time. A possible explanation for the initial lower EP of fed females is that the copepods need to adjust their metabolism to feeding, and that this takes resources away from EP. Hence, the effect of food on EP rate may depend on the pre-feeding history of the animals. This may explain the opposing results on the effects of food on EP of *C. hyperboreus* that have been found previously. Some studies have found EP to be independent of food (Conover 1967, Plourde et al. 2003), whereas other studies conducted later in the season have found *C. hyperboreus* females to produce more eggs when food was available as a supplement to internal lipids (Sømme et al. 1934, Niehoff 2007; Fig. 9). In general, EP rates measured in the laboratory experiment were lower than the *in situ* rates measured at the same time. As different incubation methods were used, the values found should not be compared directly. The handling method was rougher in the laboratory experiment, where a large amount of water was concentrated on a small sieve which increased the risk of breaking and disintegrating eggs. Because of the large lipid content, eggs of *C. hyperboreus* are rather fragile. Furthermore, neither of the methods prevented cannibalism of eggs, as eggs of *C. hyperboreus* are positively buoyant and hence do not sink through the sieve. Therefore, EP in this study may be underestimated. Although average

SEP in the laboratory experiments was rather low (ranging between 0.3 and 0.6 % d⁻¹ in the 15 d period), it is still comparable to what was reported in another laboratory study where SEP of 0.7 % d⁻¹ was measured over a 9 d period (Hirche & Niehoff 1996).

Fecal pellet production

As expected, the fecal pellet production of fed females was significantly higher both before and during the spring bloom. In the first experiment, there seemed to be a lag phase in SPP. This could be because females were collected before the spring bloom and needed some time to reach the maximal intake of food. Consequently, the highest production was not reached before Day 6 or 7 of the experiment. The opposite tendency was observed in the second experiment, where pellet production started out high and then leveled off. The reason for this opposite tendency is unknown. Kjellerup et al. (2012) found a lag phase for both *Calanus glacialis* and *C. finmarchicus* not only before the bloom but also during the bloom. SPP_{rate} was higher before the spring bloom than during the bloom at all temperatures for both fed and starved females. One explanation for this decrease over the spring could be differences in assimilation efficiency related to the lifecycle of the females. In the first experiment, the females may not have been ready to feed, as they had been dwelling in deep waters where the ambient food concentration is very low. Normally they would not encounter food at this time of year, so they may not have been ready to assimilate the ingested food effectively. This might also explain the difference in food response observed between the pre-bloom and bloom experiment. Even though fed females seemed to be grazing in both periods, an effect of food on body weight was only obvious in the pre-bloom experiment (Fig. 8).

In the second experiment, the bloom was underway, and ingested food provided energy for the regeneration of gonads and lipid stores, which had been exhausted by spawning. Indeed, initial carbon and lipid content had decreased 2 and 4 times, respectively, between the 2 experiments (Figs. 8 & 9). These stores would need to be refilled for the females to reproduce again in the following year. Furthermore, *in situ* investigations from Disko Bay in 2008 showed a 3.5-fold increase in carbon content and a 4.7-fold increase in lipid content of *Calanus hyperboreus* females over the spring bloom, indicating such a refueling process (Swailethorp et al. 2011). As the second experiment was conducted very early in

the bloom and only a slight increase in carbon and lipid content was observed for fed females, it is likely that the animals had just started feeding *in situ* and the rebuilding of lipid stores had not yet begun.

Another explanation for the lower SPP_{rate} in the bloom experiment could be that spent females are about to die (Head & Harris 1985). This could also explain why k_2 was consistently lower than k_1 in the second experiment, as dying females would slowly stop all feeding. The feeding of the females in the first experiment could in such a scenario be explained by a need to attain some additional energy for EP (Takahashi et al. 2002, Niehoff 2007). Even though no effect of food on EP was seen in this experiment, the finding of a higher EP rate in the last part of the experiment for fed females makes this a likely explanation. Further studies of the fate of spawning females should be made to confirm such a hypothesis. In general, we would expect to see the same temperature dependency in pellet production as in EP, i.e. low production at low temperatures, a temperature optimum, and a decline at temperatures that are too high. As was the case for the SEP_{rate} , no convincing effect of temperature on SPP_{rate} was observed before or during the spring bloom in the temperature range investigated here.

The measured SPP_{rate} ranging from 0.003 to 1.1 % d^{-1} was low compared to values obtained for *Calanus finmarchicus* and *C. glacialis* in a similarly designed experiment from 2008 where values ranged from 0.006 to 20.4 % d^{-1} (Kjellerup et al. 2012) but comparable to *in situ* values measured for *C. hyperboreus* in the area during the same year, which ranged from 0.01 to 0.46 % d^{-1} (Swailethorp et al. 2011). The fecal pellets produced in the starved treatments are not due to grazing but due to forced elimination of the intestinal epithelium (Besiktepe & Dam 2002) fueled by the stored lipids as shown for the smaller *Calanus* species by Kjellerup et al. (2012).

Carbon and lipid content over the course of the experiment

The loss of carbon and lipids in females during the pre-bloom experiment, as well as the loss observed *in situ* between the pre-bloom and the bloom experiment, is partly due to the production of eggs during this period. Comparing mean lipid loss (462 μg) and mean number of eggs laid (211), and knowing the lipid content of an egg (0.54 μg ; S. Jung-Madsen et al. unpubl.), on average 26% of the lipid lost during the incubation was channeled directly into EP.

However, this is most likely underestimated because of the underestimated EP rate (see earlier discussion). On the other hand, if assuming an EP rate equal to the *in situ* rate (54 eggs female $^{-1}$ d^{-1}) over the same period (15 d), then 96% of the lipids should have gone into reproduction, leaving too little energy to cover metabolic costs. The 26% fits better with what was calculated for the *in situ* situation and what was estimated by Conover & Siferd (1993). As a consequence of the experimental design, sample sizes of lipid and carbon measurements done at the end of each experiment were quite small. The low number of animals analyzed might limit the ability to detect minor changes in biochemical composition over the relative short duration of the experiment. The variation between individuals (SE) was quite high, and higher at the termination of each experiment than at the beginning. This could be a consequence of the smaller sample size, but it might also reflect plasticity in the individual response.

Temperature effects on *Calanus hyperboreus*

The temperature interval of 0 to 10°C to which the females were exposed in this study did not reveal a temperature response in the monitored rates. Comparable studies of temperature effects on both SPP and SEP for *Calanus hyperboreus* are not available, although temperature-related studies investigating EP and lifecycle patterns exist. Conover (1962) investigated the respiration of *C. hyperboreus* over a range of 2 to 8°C and found the species to regulate their metabolism well over this interval if previously acclimatized to the temperature. Ringuette et al. (2002) found that chl *a* concentration and not temperature had the greatest impact on recruitment of *C. hyperboreus* copepodites, whereas they found that the recruitment of *C. glacialis* was more temperature dependent. On the other hand, Plourde et al. (2003) investigated EP at a temperature interval of 0 and 8°C for *C. hyperboreus* and concluded that high temperatures could reduce the reproductive output of *C. hyperboreus* by 30% and shorten the spawning period significantly. Hirche (1987) studied respiration and mortality at increasing temperatures (−0.8 to 17°C) and found *C. hyperboreus* to be the least temperature tolerant of the 3 *Calanus* species.

Both *Calanus glacialis* and *C. finmarchicus* show a positive response to higher temperatures in terms of pellet production and EP rates (Kjellerup et al. 2012). Thus, the finding that *C. hyperboreus* shows no temperature response suggests potential future changes

in composition of the *Calanus* community in Disko Bay. In a warmer climate, the fact that *C. finmarchicus* has a clear advantage of temperatures up to at least 10°C while *C. glacialis* increases production rates up to 7.5°C could give these 2 species a competitive advantage over *C. hyperboreus*.

Other opposing and more indirect effects of a warmer climate will also influence the future biomass composition. Ringuette et al. (2002) suggested that a longer productive season in the Arctic as a consequence of a warmer climate could result in earlier recruitment of *Calanus glacialis* and *C. hyperboreus* and a possibility for them to complete their lifecycles faster and thereby increase their population sizes. However, Plourde et al. (2003) showed that a warmer climate would lead to a shorter winter-spawning season for *C. hyperboreus* and a subsequent mismatch between the development from egg to the first feeding nauplii stage and the spring bloom which could lead to a decrease in population size. Because a temperature increase will impact developmental stages and phases of *Calanus* spp. life cycles differently, it is difficult to predict exactly how the composition of the *Calanus* community will change with increasing temperature in the future. A warmer climate may also change the relative composition of the copepod community in general, with replacement of the arctic species adapted to sea ice by subarctic and boreal generalist species (Darnis et al. 2008)

In conclusion, this study demonstrates the winter-spawning strategy of *Calanus hyperboreus* in which reproduction is coupled to the spring bloom with a time lag of 1 yr. Furthermore, it was documented that temperature had no positive effect on either egg or pellet production of *C. hyperboreus*. This finding suggests that this high-energy *Calanus* species will lose in competition with the 2 smaller *Calanus* species in a future warmer climate because of the ability of the smaller species to exploit the higher temperature to increase their grazing and EP rates (Kjellerup et al. 2012). However, to fully elucidate the changes in the future *Calanus* community, the growth response of the smaller non-reproductive stages have to be investigated and considered.

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