



## Application of WGS data for O-specific antigen analysis and *in silico* serotyping of *Pseudomonas aeruginosa* isolates

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1 Application of WGS data for O-specific antigen analysis and *in*  
2 *silico* serotyping of *Pseudomonas aeruginosa* isolates

3

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17 **Abstract**

18

19 Accurate typing methods are required for efficient infection control. The emergence  
20 of whole genome sequencing (WGS) technologies has enabled the development of  
21 genomics-based methods applicable for routine typing and surveillance of bacterial  
22 pathogens. In this study, we developed the *Pseudomonas aeruginosa* serotyper  
23 (PAst) program, which enabled *in silico* serotyping of *P. aeruginosa* isolates using  
24 WGS data. PAst has been made publically available as a web-service, and aptly  
25 facilitate high-throughput serotyping analysis. The program overcomes critical issues  
26 such as the loss of *in vitro* typeability often associated with *P. aeruginosa* isolates  
27 from chronic infections, and quickly determines the serogroup of an isolate based on  
28 the sequence of the O-specific antigen (OSA) gene cluster. Here, PAst analysis of  
29 1649 genomes resulted in successful serogroup assignments in 99.27% of the cases.  
30 This frequency is rarely achievable by conventional serotyping methods. The limited  
31 number of non-typeable isolates found using PAst was the result of either complete  
32 absence of OSA genes in the genomes or the artifact of genomic misassembly. With  
33 PAst, *P. aeruginosa* serotype data can be obtained from WGS information alone.  
34 PAst is a highly efficient alternative to conventional serotyping methods in relation  
35 to outbreak surveillance of serotype O12 and other high-risk clones, while  
36 maintaining backward compatibility to historical serotype data.

37

38 **Introduction**

39

40 *Pseudomonas aeruginosa* is a Gram-negative opportunistic pathogen and a major  
41 cause of mortality and morbidity among hospitalized and compromised patients  
42 including those with cystic fibrosis (CF). *P. aeruginosa* is well known for its ability to  
43 cause chronic and extensively drug resistant infections (1). The outer membrane  
44 lipopolysaccharide (LPS) layer is a major virulence factor of *P. aeruginosa* (2). LPS has  
45 been linked to antibiotic resistance and immune evasion. Furthermore, LPS is one of  
46 the receptors that determines susceptibility of the bacterium to bacteriophages and  
47 pyocins (2–4). Our ability to control *P. aeruginosa* infections depends on the  
48 availability of accurate typing methods. Previously, serotyping was a benchmark  
49 typing method for *P. aeruginosa*. In the 1980's the International Antigenic Typing  
50 Scheme (IATS) was established to classify the species *P. aeruginosa* into 20 serotypes  
51 (O1-O20) (5–7). Today, serotyping is infrequently used in the clinic for typing  
52 purposes, mainly because of the time consuming protocol, the need for a continuous  
53 supply of serotype-specific antisera, and a high prevalence of polyagglutinating or  
54 non-typeable isolates.

55

56 The loss of *P. aeruginosa* typeability has been known for decades, and has often  
57 been linked to bacteria isolated from chronic infections, where typeability is lost  
58 over time during the course of infection (8, 9). A study performed by Pirnay *et al* (10)  
59 showed that 65% of all *P. aeruginosa* isolates examined were either non- or multi-  
60 typeable and therefore assigning a particular serotype to these strains would be  
61 difficult. The occurrence of these non- or multi-typeable isolates was higher when  
62 evaluating isolates sampled exclusively from CF infections (10). Multi-typeability has  
63 been associated with poor prognosis for CF patients, and is a trait of persistent or  
64 chronic infection. This correlates with the observation that *P. aeruginosa* isolates  
65 from chronic CF infections are initially resistant to human serum but evolve to  
66 becoming serum sensitive over time. This is likely due to the loss of production of O-  
67 antigen, which protects the bacterial cell from the human serum (8). The mechanism  
68 underlying loss of typeability over time is not fully understood, but is most likely due

69 to modifications of LPS structures over extended periods of bacteria-host  
70 interactions as a means to improve fitness in the host and to evade host immune  
71 system, bacteriophages and antibiotic therapy.  
72  
73 The knowledge concerning the serotype of an isolate is important for monitoring  
74 outbreaks and for understanding the structures of the LPS expressed on the surface  
75 of these bacteria. O11 and O12 are more predominant than other serotypes in the  
76 clinic, and intriguingly, these serotypes have been associated with multi-drug  
77 resistance (MDR) (10–13). This implies that these particular LPS structures improve  
78 fitness within the hosts and the hospital environments in ways that we currently do  
79 not understand. Specifically for the O12 serotype, it has been shown that horizontal  
80 gene transfer of LPS genes has resulted in MDR isolates and the switching of a  
81 certain serotype to O12 (14). To continuously monitor LPS structure and evolution,  
82 serotyping can help to improve our understanding of the isolates that successfully  
83 infect patients. The continued collection of these data will also enable retrospective  
84 population analysis, as serotype has been recorded for decades also prior to the  
85 emergence of other DNA-based typing methods such as MLST and PCR.  
86  
87 *P. aeruginosa* LPS is comprised of three domains: lipid A, core oligosaccharide, and  
88 O-antigen (2). Most *P. aeruginosa* isolates produce two forms of O-antigen  
89 simultaneously: common polysaccharide antigen (CPA) and O-specific antigen (OSA).  
90 While CPA is relatively conserved, OSA is variable and defines the serotype of an  
91 isolate (2, 15). OSA is encoded in a gene cluster varying in size from just under 15 kb  
92 to over 25 kb. The OSA gene cluster is flanked by the genes *ihfB/himD* and *wbpM*.  
93 The 20 serotypes harbor 11 distinct OSA gene clusters, each with a high number of  
94 unique genes (16). With the emergence of whole genome sequencing (WGS)  
95 methods it is now possible to assign an isolate into one of 11 serogroups based on  
96 the sequence and structure of the OSA gene cluster (11, 14, 17).  
97  
98 The present study presents a program that our group has developed for fast and  
99 reliable *in silico* serotyping of *P. aeruginosa* isolates using WGS data – the  
100 *Pseudomonas aeruginosa* serotyper (PAst). The program has been made publically

101 available as a web-service, and can enable high throughput serotyping analysis based  
102 on analysis of the OSA gene cluster. Using PAST, issues with typeability of clinical  
103 isolates can be overcome, and serotyping can be performed in a rapid and cost-  
104 effective way in the clinic as whole genome sequencing of isolates become  
105 accessible.  
106

107 **Materials and Methods**

108

109 **PAst verification and isolates included in the study**

110 To evaluate the efficiency of the *in silico* serotyping using PAst, all available *P.*  
111 *aeruginosa* genomes were acquired and analyzed. These *P. aeruginosa* genomes  
112 were downloaded from NCBI and included 1120 genome assemblies (Supplementary  
113 Table 1, extracted 18.08.2015). An exclusively CF-related *P. aeruginosa* dataset was  
114 constructed, due mainly to the documented high level of non-typeability in  
115 persistent infecting clones. The isolates described by Marvig *et al.* 2015 (475  
116 genomes) (18) were used as the initial dataset. These were assembled using SPAdes  
117 (19) prior to analysis. Additional CF isolates were recovered by searching for *P.*  
118 *aeruginosa* genome assemblies related to CF in PATRIC (54 genomes) (20). It was  
119 verified that frequently observed CF-specific strains such as DK2 and LES were part of  
120 the dataset. The final dataset included 529 CF-related *P. aeruginosa* genome  
121 assemblies. *In silico* serotyping of both datasets was performed using PAst in order  
122 to evaluate typeability of the program. Non-typeable isolates (i.e., isolates in which  
123 %coverage of reference OSA was < 95%) were manually examined for either  
124 biological or technical explanations of the lack of typeability.

125

126 **PAst specifications**

127 The PAst program is developed using the programming language Perl for *in silico*  
128 serotyping of *P. aeruginosa* isolates using WGS data. It is based on a BLASTn analysis  
129 of the assembled input genome, against an OSA cluster database. OSA clusters with  
130 > 95% coverage in the query genome represents a positive hit for a serogroup. Since  
131 *P. aeruginosa* isolates have been described which either harbor multiple OSA  
132 clusters or no clusters at all, the program accommodates multi-, mono- and non-  
133 typeability based on analysis of the number of positive OSA hits and coverage (Figure  
134 1). Compared to other studies (11, 14, 17) PAst optimizes *in silico* serotyping further  
135 by distinguishing members of the O2 serogroup through identification of the  
136 acquired phage-related *wzy<sub>β</sub>* within serotypes O2 and O16 (21, 22). This enables  
137 typing into 12 serogroups as opposed to the 11 described by Raymond *et al.* (16).

138 Together with a summary of the best hit(s) from the analysis and the BLAST report,  
139 the user receives a multi fasta file containing the sequence(s) of the OSA cluster  
140 from the analyzed isolate for use in future analysis.

141

#### 142 **The *P. aeruginosa* OSA cluster database**

143 The database was constructed using the WGS data of the 20 *P. aeruginosa* IATS  
144 serotype reference isolates (14). The genomes were assembled using SPAdes (19)  
145 and the OSA clusters extracted via identification of the *ihfB/himD* gene flanking the  
146 cluster upstream and the *wbpM* gene flanking the cluster downstream. The clusters  
147 were aligned within their serotypes, described by Raymond *et al.* 2002 and their  
148 shared structure confirmed (16). A representative cluster of each serotype was  
149 selected for the database (Table 1). Also included in the database was the *wzy<sub>β</sub>* gene  
150 for distinguishing the O2 and O5 serotypes, as the two serogroups share OSA cluster  
151 organization, but only the O2 and O16 serotype harbor the *wzy<sub>β</sub>* gene present on a  
152 prophage.

153

#### 154 ***In silico* serotyping of *P. aeruginosa* isolates using PAst**

155 PAst has been implemented as a simple and user-friendly web-tool available on the  
156 Center for Genomic Epidemiology (CGE) service platform  
157 (<https://cge.cbs.dtu.dk/services/PAst-1.0/>). The tool accommodates raw reads, draft  
158 assemblies (contigs or scaffolds) and complete genomes from all WGS platforms.  
159 Raw read data are processed and assembled as previously described for other CGE  
160 tools (23). Following analysis of the input data, the web-tool outputs the predicted  
161 serogroup of the query genome, the %coverage of the reference OSA cluster, as well  
162 as the OSA cluster sequence in multi fasta format, for the user to continue exploring  
163 the OSA genes (Fig. 1). If multiple positive hits are found (multi-typeability), all the  
164 identified OSA clusters are written for the user (Fig. 1). In the case of a non-typeable  
165 query genome (where no OSA cluster has >95% coverage) the best hit identified is  
166 written for the user together with the sequence of this hit (Fig. 1).  
167 For batch analysis of larger datasets (only applicable for assembled genomes) the  
168 PAst Perl program has been made available on Github:  
169 <https://github.com/Sandramses/PAst>



170 **Results**

171 The PAst web server tool identifies and analyzes the nucleotide sequence of the O-  
172 specific antigen (OSA) gene cluster within the provided genomes and place them into  
173 one of twelve serogroups defined in Table 1. These serogroups are defined by  
174 sequence similarities between the 20 IATS serotypes (16) as well as  
175 absence/presence of the discriminatory *wzy*<sub>β</sub> gene (21, 22) and are as such different  
176 from previously groupings of serotypes on the basis of *in vitro* serotyping data (11,  
177 14, 17). All serogroups contained three or less of the 20 IATS serotypes (Table 1).

178

179 **More than 97% of the *P. aeruginosa* dataset is typeable using PAst**

180 To evaluate the typeability efficiency of PAst all *P. aeruginosa* genome assemblies  
181 available in NCBI (1120 genomes on date of extraction) were analyzed. A total of  
182 97.68% (1094) of the 1120 genomes were typed unambiguously to a single  
183 serogroup by PAst (Fig. 2). This means that each genome assembly had a single  
184 BLAST hit of >95% OSA coverage to one sequence in our reference OSA database  
185 (Fig. 2). No isolates were found to be multi-typeable and 2.32% (26 genomes) of the  
186 1120 genomes were found to be non-typeable (Fig. 2). In these cases, no significant  
187 BLAST hit of >95% OSA coverage to one of the sequence in the reference OSA  
188 database was identified. PAst correctly determined the serogroup of the 20 IATS  
189 strains as well as PAO1 (serotype O5), PA14 (serotype O10), and PAK (serotype O6).

190

191 The analysis showed that all serogroups were represented in the 1120 genomes (Fig.  
192 2). Four of the 12 serogroups represented 70% of the genomes analyzed; these were  
193 O3, O6, O11 and O12 (Fig. 2). The smallest serogroup was O13, which contained  
194 only four genomes. We note that the same clone type could be present multiple  
195 times in the dataset, and that a substantial sampling bias would therefore be  
196 expected. The distribution of serotypes in our analysis thus describes what has been  
197 chosen for sequencing and does not necessarily match the distribution of serotypes  
198 in the actual *P. aeruginosa* population. This does not affect the high confidence of  
199 PAst, as it shows that un-ambiguous typing of multiple isolates from the same  
200 lineage is possible.

201

202 **PAst overcomes non-typeability issues from *in vitro* typing of CF lineages**

203 *P. aeruginosa* isolates from CF infections are often non-typeable with conventional  
204 serotyping assays. To explore if our genomics-based method could enable  
205 acquisition of serotype information in such isolates, we analyzed 529 genome  
206 assemblies of *P. aeruginosa* isolates sampled from CF infections. This dataset  
207 contained multiple examples of isolates of the same lineage that had been sampled  
208 during the course of infection. This enabled us to investigate whether *in silico*  
209 typeability might be lost over time as has frequently been observed for *in vitro*  
210 serotyping of isolates from chronic CF infections. Interestingly, 99.81% of the  
211 genomes in the CF-specific dataset could be typed to single serogroups. More  
212 importantly, no multi-typeable isolates were observed and only one isolate was  
213 deemed non-typeable (Fig. 3). All serogroups were represented in the dataset,  
214 except for O12. The absence of O12 serotypes among CF isolates has previously been  
215 reported (10). Serotypes O1, O6 and O7/O8 represented ~65% of the CF-specific  
216 dataset and the smallest representation of serotypes was the O9 serogroups with  
217 only two isolates from these samples (Fig. 3).

218

219 Well-known transmissible CF-specific clone types such as *P. aeruginosa* DK1 (24),  
220 DK2 (25), and LES (26) are represented in the dataset due to multiple isolates being  
221 sampled from various patients over several decades. Using our PAst tool, the typing  
222 problems documented from *in vitro* typing of such lineages were not observed, and  
223 the DK1, DK2 and LES isolates were consistently *in silico* serotyped with PAst. DK1  
224 and DK2 were found to belong to the O3 serogroup, while the LES lineage belonged  
225 to the O6 serogroup.

226

227 **Complete loss of O-specific antigen defining genes is a rare event**

228 Out of two WGS-based datasets (n = 1649) that were *in silico* typed with PAst, our  
229 results yielded a total of 27 non-typeable isolates. The lack of typeability in these 27  
230 genome assemblies was further investigated to resolve whether non-typeability in  
231 these cases was due to technical or biological reasons. We found that the %OSA  
232 coverage of the non-typeable isolates ranged from a minimum of 1.91% to a

233 maximum 93.96% OSA coverage (Supplementary Table 2). Of the 27 isolates  
234 classified as non-typeable, thirteen were found to have OSA coverage of 0-20%,  
235 whereas seven isolates had OSA coverage of 80-95% (Fig. 4). The best hit (serogroup)  
236 for each of the non-typeable isolates was then examined to evaluate if certain  
237 serogroups were more prone to be problematic in the PAST analysis and why. The 27  
238 isolates were found to distribute across 6 serogroups (O1, O2, O6, O7, O11 and O13),  
239 while 15/27 isolates showed a best hit to be typed as the O11 serogroup (Fig. 4).

240

241 The group of non-typeable isolates with a best hit to the O11 serogroup were  
242 analyzed separately to identify the reason for the lack of typeability. Of the 15 O11  
243 serogroup isolates, nine had an OSA coverage of 14.94-15.84% (Supplementary  
244 Table 2); these corresponded to the presence of only the two flanking genes  
245 *himD/ihfB* and *wbpM*. This observation shows that a best hit of a non-typeable  
246 isolate to the O11 OSA cluster with a coverage of ~15% is the result of a complete  
247 absence of an OSA cluster but the presence of the flanking genes. Two other isolates  
248 had an OSA coverage of <2%, and corresponded to the absence of the entire OSA  
249 cluster as well as the flanking genes (Supplementary table 2). In summary, a total of  
250 11 of the 27 non-typeable isolates (or 11 of 1649 isolates analyzed in total) were  
251 non-typeable due to a lack of the OSA cluster sequences.

252

### 253 **Genome mis-assembly accounts for false non-typeability**

254 Since the seven non-typeable isolates with the highest OSA coverage (80-95%) in  
255 Figure 4 were all candidates for harboring complete and functioning OSA clusters,  
256 we analyzed the cause of non-typeability in this group of isolates. For each of the  
257 isolates, we examined whether there were mis-assembly or assembly gaps within  
258 the OSA gene cluster; we also looked for the occurrence of insertion sequence (IS)  
259 elements, which often cause gaps in *de novo* assembly. Indeed, five of the seven  
260 isolates contained assembly gaps within their OSA cluster, which account for the  
261 observed lowered OSA coverage (Table 2). The remaining two isolates had no gaps  
262 within their OSA sequence (Table 2). However, both of these isolates had a best type  
263 hit to the O11 serogroup, which is known to contain OSA sequences of both the O11  
264 and the O17 serotypes (16) (Table 1). Interestingly, the OSA cluster in these two

265 serotypes differ only by the presence of two IS elements and a deletion in the O17  
266 serotype OSA sequence (16). Alignment of the OSA sequence from the two non-  
267 typable isolates to the O11 and O17 reference OSA sequences, respectively,  
268 contained an O17 OSA gene cluster, which had been misassembled into  
269 concatenated O11 serotype OSA clusters because of the O17 IS elements.

270 **Discussion**

271

272 The serotyping technique has been one of the standard tools for epidemiological  
273 studies and infection controls for many decades. The available historical records of *P.*  
274 *aeruginosa* serotypes offer a vast amount of information about *P. aeruginosa*  
275 epidemiology and population structures (27–30). Although problems with non-  
276 typeable isolates have been described since the implementation of the method, the  
277 serotype information is still applicable today for outbreak tracking, strain typing, and  
278 studies of LPS structure and evolution. The present study presents a newly  
279 developed Web Server tool called PAsT, which is user friendly, reliable, and high-  
280 throughput for *in silico* serotyping of *P. aeruginosa* isolates.

281

282 In contrast to conventional serology-based *in vitro* serotyping, PAsT *in silico*  
283 serotyping has a very low occurrence of non-typeability. Of the 1649 analyzed  
284 genomes, only 27 non-typeable isolates were detected across two separate *P.*  
285 *aeruginosa* datasets. One dataset represents all available whole genome assemblies  
286 of *P. aeruginosa*, while the other specifically represents genomes from CF infections,  
287 which are known to contain high occurrences of non-typeability due to adaptability  
288 of the bacteria into a biofilm life-style associated with chronicity of the infection (Fig.  
289 1 and 2). Importantly, since the frequency of non-typeability of *in vitro* serotyped *P.*  
290 *aeruginosa* isolates may amount to over 65% (10), analysis with PAsT is clearly  
291 advantageous and superior compared to conventional *in vitro* serotyping.

292 Importantly, the superiority of the PAsT tool as a reliable and fast typing method is  
293 consistent with other published tools for *in silico* serotyping (31–35). Similar to both  
294 the SerotypeFinder (*in silico* serotyping of *E. coli* (31)), LisSero (*in silico* serotyping of  
295 *Listeria monocytogenes* (34, 35)) and SeqSero (*in silico* serotyping of *Salmonella* (32))  
296 PAsT resolves the OSA cluster information to the most accurate typing possible as a  
297 serogroup representing 1-3 serotypes.

298

299 Interestingly, we observed a high level of conservation of the OSA gene cluster  
300 within the *P. aeruginosa* genome. In contrast to certain well-documented difficulties

301 in serology-based *in vitro* serotyping, PAST identified complete OSA clusters (with  
302 >95% sequence being present) in 99.27% of the analyzed genomes. As such only 12  
303 of the 1649 isolates examined were found to be devoid of the OSA cluster and an  
304 additional 8 isolates were found to contain only a partial OSA cluster in their  
305 genomes (<80% OSA sequence compared to the reference). These findings indicate  
306 that the loss of typeability of *P. aeruginosa* isolates during the course of infection is  
307 either due to mutations (rather than larger deletions) or is linked to other parts of  
308 the LPS biosynthesis, such as regulatory genes or transport of the structure to the  
309 cell surface. A study by Bélanger *et al.* reported that mutation in any of the four *wbp*  
310 genes (*wbpO*, *wbpP*, *wbpV* and *wbpM*) in the OSA gene cluster could disrupt the *P.*  
311 *aeruginosa* O6 OSA biosynthesis (36). Furthermore, key genes involved in the OSA  
312 assembly and translocation through the Wzx/Wzy-dependent pathway not localized  
313 within the OSA cluster, for instance, *waal*, are essential for O-antigen expression  
314 (37, 38). It is possible that more OSA-related genes might be present in the *P.*  
315 *aeruginosa* genomes, which have not been discovered yet. Overall, our study  
316 demonstrates that a complete lack of an OSA gene cluster is a rarely observed  
317 phenomenon in *P. aeruginosa*.

318

319 PAST will enable further investigations of the diversity, evolution and variability of  
320 the OSA clusters. For example, the sequence of the cluster is part of the output  
321 material from the *in silico* serotyping which can then be readily analyzed for  
322 sequence variations to provide new knowledge on the mechanisms behind loss of  
323 typeability *in vitro* and *in silico*. Furthermore, PAST will enable systematic analysis of  
324 serotype switching by horizontal gene transfer and genetic recombination of the  
325 OSA gene cluster among different clone types. This recently described phenomenon  
326 has contributed to the evolution of the multi-drug resistant *P. aeruginosa* serotype  
327 O12 population that has successfully disseminated across hospitals worldwide (14).  
328 It is currently unknown if there are additional cases of such serotype switching by  
329 recombination.

330

331 The new PAST Web Server tool makes *in silico* serotyping of *P. aeruginosa* using WGS  
332 data a fast and reliable method. The use of PAST can play an important role in future

333 surveillance of LPS evolution and possible outbreak detection. With the emergence  
334 of rapidly disseminating, high-risk clones of *P. aeruginosa*, such as the O12 ST111  
335 clone, new and reliable typing techniques for improved monitoring and tracking of  
336 such outbreaks are becoming increasingly important (13). With the lowered cost of  
337 sequencing and the increased focus on WGS of pathogens in clinics and hospital  
338 settings, genomics-based tools can assist in designing future treatments and  
339 containment of outbreaks.

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486 **Figure legends**

487 **FIG 1** Workflow illustrating the *in silico* serotyping of the *Pseudomonas aeruginosa*  
488 serotyper (PAst).

489

490 **FIG 2** The distribution of the different serogroups (in %) identified via *in silico*  
491 serotyping of the *P. aeruginosa* dataset using PAst. The analysis is based on all  
492 available *P. aeruginosa* genomes assemblies (n = 1120).

493

494 **FIG 3** The distribution of the different serogroups (in %) identified via *in silico*  
495 serotyping of CF specific *P. aeruginosa* isolates (n = 529) using PAst.

496

497 **FIG 4** Best-hit serotype distribution of the 27 non-typeable isolates as a function of  
498 the OSA coverage.

499

500 **Tables**

**TABLE 1** Serogroup definition in the PAST OSA database.

| Serogroup | Reference OSA cluster | Ref. gene              | Size (bp) | Serotypes within serogroup |
|-----------|-----------------------|------------------------|-----------|----------------------------|
| O1        | O1                    |                        | 18.368    | O1                         |
| O2        | O2                    | <i>wzy<sub>6</sub></i> | 23.303    | O2, O16                    |
| O3        | O3                    |                        | 20.210    | O3, O15                    |
| O5        | O2                    |                        | 23.303    | O5, O18, O20               |
| O4        | O4                    |                        | 15.279    | O4                         |
| O6        | O6                    |                        | 15.649    | O6                         |
| O7        | O7                    |                        | 19.617    | O7, O8                     |
| O9        | O9                    |                        | 17.263    | O9                         |
| O10       | O10                   |                        | 17.635    | O10, O19                   |
| O11       | O11                   |                        | 13.868    | O11, O17                   |
| O12       | O12                   |                        | 25.864    | O12                        |
| O13       | O13                   |                        | 14.316    | O13, O14                   |

501

**TABLE 2** Non-typeable *P. aeruginosa* isolates with %OSA coverage of 80-95% with specification of assemblies.

| Strain                       | Size (Mb) | Scaffolds | %GC  | Best hit | %OSA  | <i>wbpM</i> | <i>himD</i> | Gap |
|------------------------------|-----------|-----------|------|----------|-------|-------------|-------------|-----|
| <i>P. aeruginosa</i> E2      | 635.733   | 196       | 66.4 | O7       | 83.31 | +           | +           | +   |
| <i>P. aeruginosa</i> IGB83   | 648.065   | 249       | 66.4 | O2       | 84.46 | +           | +           | +   |
| <i>P. aeruginosa</i> VRFPA04 | 681.803   | 1         | 66.5 | O11      | 86.96 | +           | +           | -   |
| <i>P. aeruginosa</i>         | 627.851   | 176       | 66.1 | O6       | 90.54 | +           | +           | +   |
| <i>P. aeruginosa</i> 148     | 664.374   | 128       | 66.1 | O11      | 90.93 | +           | +           | -   |
| <i>P. aeruginosa</i> ID4365  | 677.663   | 172       | 66.1 | O7       | 91.74 | +           | +           | +   |
| <i>P. aeruginosa</i> C2773C  | 671.772   | 200       | 65.9 | O6       | 93.96 | +           | +           | +   |

502









