



Concurrent elevation of CO₂, O₃ and temperature severely affects oil quality and quantity in rapeseed

Namazkar, Shahla; Stockmarr, Anders; Frenck, Georg ; Egsgaard, Helge; Terkelsen, Thilde ; Mikkelsen, Teis Nørgaard; Ingvordsen, Cathrine Heinz; Bagger Jørgensen, Rikke

Published in:
Journal of Experimental Botany

Link to article, DOI:
[10.1093/jxb/erw180](https://doi.org/10.1093/jxb/erw180)

Publication date:
2016

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):
Namazkar, S., Stockmarr, A., Frenck, G., Egsgaard, H., Terkelsen, T., Mikkelsen, T. N., Ingvordsen, C. H., & Bagger Jørgensen, R. (2016). Concurrent elevation of CO₂, O₃ and temperature severely affects oil quality and quantity in rapeseed. *Journal of Experimental Botany*, 67(24), 4117-4125. <https://doi.org/10.1093/jxb/erw180>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



RESEARCH PAPER

Concurrent elevation of CO₂, O₃ and temperature severely affects oil quality and quantity in rapeseed

Shahla Namazkar¹, Anders Stockmarr², Georg Frenck³, Helge Egsgaard¹, Thilde Terkelsen⁴,
Teis Mikkelsen¹, Cathrine Heinz Ingvordsen^{1,5} and Rikke Bagger Jørgensen^{1*}

¹ Technical University of Denmark, DTU Environment, Risø Campus, Building 763, Frederiksborgvej 399, 4000 Roskilde, Denmark

² Technical University of Denmark, Department of Applied Mathematics and Computer Science, Richard Petersens Plads, Building 324, 2800 Kongens Lyngby, Denmark

³ University of Innsbruck, Institute of Ecology, Sternwartestraße 15, 6020 Innsbruck, Austria

⁴ Danish Cancer Society, Statistics, Bioinformatics and Registry, Strandboulevarden 49, 2100 København Ø, Denmark

⁵ CSIRO, Plant Industry, Black Mountain Laboratories, GPO Box 1600, Canberra, ACT 2601, Australia

* Correspondence: rijq@env.dtu.dk

Received 6 December 2015; Accepted 15 April 2016

Editor: Elizabeth Ainsworth, USDA/ARS

Abstract

Plant oil is an essential dietary and bio-energy resource. Despite this, the effects of climate change on plant oil quality remain to be elucidated. The present study is the first to show changes in oil quality and quantity of four rapeseed cultivars in climate scenarios with elevated [CO₂], [O₃] and temperature (T) combined and as single factors. The combination of environmental factors resembled IPCC's 'business as usual' emission scenario predicted for late this century. Generally, the climate scenarios reduced the average amounts of the six fatty acids (FAs) analysed, though in some treatments single FAs remained unchanged or even increased. Most reduced was the FA essential for human nutrition, C18:3- ω 3, which decreased by 39% and 45% in the combined scenarios with elevated [CO₂]+T+[O₃] and [CO₂]+T, respectively. Average oil content decreased 3–17%. When [CO₂] and T were elevated concurrently, the seed biomass was reduced by half, doubling the losses in FAs and oil content. This corresponded to a 58% reduction in the oil yield per hectare, and C18:3- ω 3 decreased by 77%. Furthermore, the polyunsaturated FAs were significantly decreased. The results indicate undesirable consequences for production and health benefits of rapeseed oil with future climate change. The results also showed strong interactive effects of CO₂, T and O₃ on oil quality, demonstrating why prediction of climate effects requires experiments with combined factors and should not be based on extrapolation from single factor experiments.

Key words: *Brassica napus* L., greenhouse gases, model, multifactor experiment, oilseed rape, vegetable oil quality.

Introduction

Current trends in greenhouse gas emissions suggest that the climate will change substantially in the 21st century (IPCC, 2013). Recent reports (Le Quéré *et al.*, 2014) indicate that the concentrations of greenhouse gases (CO₂ equivalents) are still

increasing – 2.3% in 2013 – and in the first half of 2015 the global average concentration of CO₂ exceeded 400 ppm (<http://www.esrl.noaa.gov/gmd/ccgg/trends/>). Current emissions are just above the RCP8.5 scenario, demanding substantial and

likely unrealistic reductions in emissions by 2020 in order to keep the average global warming below 2 °C (IPCC, 2013).

The changing climate will affect global food security (IPCC, 2014). A number of studies have investigated crop productivity, e.g. in rapeseed (Supit *et al.*, 2010; Frenck *et al.*, 2011; Frenck *et al.*, 2013), and these show that production is likely to be seriously impeded if greenhouse gas emissions continue at the current pace. We previously showed that when [CO₂] (~650 ± 50 ppm) and temperature (+5 °C above ambient) were elevated simultaneously, seed production was only 50% of that in rapeseed grown at ambient conditions (Frenck *et al.*, 2013). Though climate change effects on crop production are well researched, this is not the case for food quality. The recent IPCC report (AR5, Porter *et al.*, 2014) points out these uncertainties: 'Little is known about combined effects of climate change factors on food quality or the economic and behavioral changes that will occur. Thus, there is little confidence regarding effects of climate change on human health through changes in nutrient composition'.

Brassica napus L., rapeseed or oilseed rape, is the second largest source of vegetable oil after soybean worldwide (USDA, 2016) and the predominant oil crop in Europe (European Commission, 2013). Rapeseed oil is being used in many parts of the world for food (canola oil) or bio-fuel, and additionally rapeseed meal is used for animal feed (McKevith, 2005; Wittkop *et al.*, 2009; Hoekman *et al.*, 2012). Despite its importance only a few studies have investigated the oil content and oil quality of rapeseed and related species under predicted future climate change conditions (Triboi-Blondel and Renard, 1999; Tripathi and Agrawal, 2012; Baux *et al.*, 2013), and treatments with combined climate factors have not been explored experimentally. The present study reports the effects of predicted future climate conditions on the quantity and quality of the seed oil from rapeseed. The results on changes in oil quantity and quality are viewed in the perspective of the decrease in seed yield observed under future changed climate conditions; seed yield was accounted for previously by Frenck *et al.* (2013). We applied treatments where [CO₂], [O₃] and temperature (T) were elevated one by one, and treatments where the factors were elevated concurrently. The combined scenarios mostly resembled conditions of RCP8.5 (IPCC, 2013), or A1FI (IPCC, 2007) around the years 2075–2100. With a continued increase in greenhouse gas emissions, the combination of 650 ppm [CO₂] and +5 °C applied in this study resembles a realistic future scenario.

Our study is the first to document the effects on the crop quality – specifically oil quality – from multifactor climate scenarios. The results suggests that reliable projections of the future quality of rapeseed oil should be derived from such multifactor studies and not made by assuming additivity of results from single factor experiments.

Materials and methods

Plant material

The fatty acids (FA) and total oil (lipid) content were analysed in mature seeds from rapeseeds cultivated during their entire life cycle

in a climate phytotron under six different climate treatments. Four cultivars, 'Bolero', 'Mary', 'Mozart' and 'Tanto', were included in the study. The four cultivars were assumed to be representative for rapeseed, as they differed in year of release and breeding organization and covered different cultivation areas. All four cultivars belong to the 'double low' type (developed since the late 1970s), which was bred for low content of glucosinolates and erucic acid. Frenck *et al.* (2011) described the cultivars in more detail. Each cultivar was represented by an experimental population of 36 plants in each of the six climate treatments (216 plants per cultivar in total). Plants were cultivated at a density of 64 plants m⁻² corresponding to a normal field density of 40–80 plants m⁻². At maturity seeds were harvested and yield was determined. The quantity and quality of the seed oil were analysed from five replicate samples of the pooled seeds per cultivar-population and treatment. In total 120 seed samples were analysed (4 cultivars × 6 treatments × 5 replicates) for their content of FAs and total oil. After harvest the seeds were stored at 7 °C until sample preparation.

Climate treatments in the phytotron

In the different treatments the cultivars were grown together but on separate wheeled cultivation tables. The six different treatments corresponded to six phytotron chambers each 24 m² located at the Technical University of Denmark, DTU Risø Campus. Set points for the six different environments were:

- ambient – 390 ppm [CO₂], 19/12 °C (day/night), 20/20 ppb [O₃] (day/night)
- all factors elevated – 650 ppm [CO₂], 24/17 °C, 60/20 ppb [O₃]
- [CO₂] and T elevated in combination – 650 ppm [CO₂], 24/17 °C, 20/20 ppb [O₃]
- [CO₂] elevated as single factor – 650 ppm [CO₂], 19/12 °C, 20/20 ppb [O₃]
- T elevated as single factor – 390 ppm [CO₂], 24/17 °C, 20/20 ppb [O₃]
- [O₃] elevated as single factor – 390 ppm [CO₂], 19/12 °C, 60/20 ppb [O₃].

Over the experiment, the average values for the manipulated environmental factors were as follows: ambient [CO₂] 390 ± 17 ppm, elevated [CO₂] 654 ± 46 ppm, ambient [O₃] 18 ± 16 ppb, elevated [O₃] 52 ± 11 ppb, ambient T 18.5 ± 0.9/12.8 ± 0.9 °C (day/night) and elevated T 23.3 ± 0.8/17.9 ± 0.8 °C (day/night). More information about growth conditions and phytotron treatments is given by Frenck *et al.* (2011, 2013).

Due to a limited number of the large phytotron chambers available, the two-factor combinations with elevated [O₃] and elevated [CO₂] or elevated T (O₃+CO₂ and O₃+T) could not be executed; however, the modeling (see below) partly compensated for the incomplete design. Plants and their corresponding treatments were rotated both between and within the six chambers of the phytotron every week to minimize chamber effects (Frenck *et al.*, 2013).

During daytime (16/8 h day/night light regime), photosynthetically active radiation (PAR) just above the canopy was 520 μmol photons m⁻² s⁻¹. Plants were watered by a drip irrigation system providing 4.4 L m⁻² day⁻¹ delivered to the individual pots at the beginning of the daytime period. A relative humidity of 55/70% (day/night) was established in the growth chambers. All treatments received the same amount of water, and it could therefore be anticipated that treatments with elevated temperature experienced limitation of water to a higher degree than those with ambient temperature due to higher evapotranspiration. In treatments with elevated [CO₂], the reduced stomatal conduction might partly mitigate the transpiration loss. However, the water deficit was not quantified.

Analysis of FA composition and lipid content

Preparation of samples Five replicates of 100 mg seeds of each of the four rapeseed cultivars, times six treatments, were prepared and

analysed in duplicate by gas chromatograph flame ionization chromatography using flame ionization detection (GC-FID) (Agilent Technologies 7890A). The seeds were ball-milled to obtain homogeneous samples (Retsch MM301 mixermill, Germany, stainless steel balls, $d=20$ mm, 35 mL grinding jar) for 10 s. The sample size of 100 mg seeds corresponded to approximately 26 seeds (average seed weight 3.9 mg) randomly collected from the pooled seeds per cultivar and treatment. An amount of 5 mL of chloroform/methanol mixture (2:1 v/v) together with 10 mg internal standard (IS, C19:0) were added to each sample of crushed seeds. After evaporating the solvents, diethyl ether (4 mL) was added to each extracted seed sample followed by addition of methanol (15 mL). FA methyl esters (FAMES) were prepared based on the methodology according to [Craig and Murty \(1959\)](#) by adding 25% sodium methoxide solution in methanol (1 mL) to the extracted oil and maintaining this mixture at room temperature for 1 h. After the reaction with sodium methoxide, cyclohexane (3 mL) and distilled water (2 mL) were added. The diluted FAMES sample (1 μ L) was introduced into the gas chromatograph using a CTC auto-sampler (Agilent, Denmark) in split mode (1:25). The FAMES were separated using a 0.32 mm i.d. \times 30 m WCOT fused silica column coated with VF-23ms at a thickness of 0.25 μ m (Agilent, Denmark). The carrier gas was helium at a flow rate of 1.0 ml min⁻¹. Separation of the FAMES was achieved using a temperature gradient from 70 to 250 °C at 10 °C min⁻¹.

Calibration A concentration of 1 mg mL⁻¹ of C19:0 (the internal standard, IS) in cyclohexane was prepared as well as a set of calibration standards containing 50 mg of C16:0, C18:0, C18:1, C18:2, C18:3 and C20:1 in 50 mL cyclohexane. The stock standard solutions were prepared by adding 1, 0.75, 0.5 and 0.25 mL of each FA standard solution to 1 mL of the IS and the solutions were stored in darkness at 4 °C. These standards were analysed by GC-FID in order to establish the retention time and response factors of each compound. The method was validated using certified sunflower oil (Supelco 4-7123) and oil extracted from a rapeseed standard BCR-447 (COLZA, EC-JRC-IRMM, Geel Belgium).

Statistical analysis of effects under the climate treatments

To investigate potential interaction between the individual factors in the combined treatments, lipid content and individual FAs were analysed with a mixed effects model. The fixed effects were modeled with an incomplete three-way factorial design on the treatment types (ambient, [CO₂], [O₃] and T). The effect of cultivar was considered random; cultivar-level random effects were applied in order to obtain a general predictive model. To investigate cultivar specific responses, the mixed effects model was modified to a multiple regression model such that the cultivar effects were considered as fixed, and allowed to interact with the treatment types. Prior to analysis, content of the six different FAs and the total lipid content were ln transformed. The transformed content of each FA and the total lipid content were modeled separately. The applied mixed effects model may be described as:

$$\ln(\text{lipid content}_i) = \alpha + \beta_{\text{CO}_2} \text{CO}_2 + \beta_T T + \beta_{\text{O}_3} \text{O}_3 + \beta_{\text{CO}_2:T} \text{CO}_2 : T + \beta_{\text{CO}_2:\text{T}:\text{O}_3} \text{CO}_2 : T : \text{O}_3 + \eta Y_{\text{Accession}} + \varepsilon_i, \quad i = 1, \dots, 120 \quad (1)$$

where the treatment symbols are indicators of whether the given treatment is part of the total treatment for the i^{th} trial. So, for example, if the treatment in the i^{th} trial consisted of CO₂ alone, the indicator CO₂ would be 1, while the remaining indicators for T and O₃ and interactions would be equal to 0. The reference treatment is ambient, so that an ambient treatment will have all indicators equal to 0. Y indicates the random effect of *accession*. Because of the incompleteness of the factorial design, caused by the fact that combination treatments CO₂+O₃ and T+O₃ were not included in the study, the third order interaction CO₂:T:O₃, and the two two-way interactions CO₂:O₃ and T:O₃ could not be distinguished from each other. For this reason, an additive effect of the O₃ treatment could only be tested versus the

interaction effect with the simultaneous effect of CO₂ and T. Thus, if an additive effect of the O₃ treatment was rejected through testing the three-way interaction term CO₂:T:O₃, we could not say which of the factors – CO₂ and/or T – interacted with O₃. An additive effect of the CO₂ and T treatments relative to each other was tested through the standard two-way interaction term CO₂:T, as Eqn (1) without this term conforms with the additive effects.

Furthermore, a confirmatory analysis was carried out, where the content was analysed simultaneously for all FAs, with a model similar to Eqn (1). In this model, FA type was included as an explanatory variable, with main effects and interaction terms with the three basic treatment types. Confidence intervals are reported asymmetric, to reflect the non-linear ln transformation.

Model validation was performed through graphical procedures and normality tests of residuals. Parameter estimation was carried out using restricted maximum likelihood (REML), and tests were evaluated with T-distributions. All model analysis and testing were carried out using the software R, version 3.1.0 ([R Core Team, 2014](#)).

Lipid content (oil content) was calculated as the sum of the six individual FAs. Cultivar-specific variability of total oil and FA content to the different treatments is shown in [Fig. 3](#). The figure was constructed by keeping the effect of cultivar fixed in Eqn (1), and allowing for interaction between cultivar, treatment types and fatty acid type. Standard deviations and significances are derived from Eqn (1). The total content of polyunsaturated fatty acids was also estimated with Eqn (1). [Figure 4](#) depicts estimated changes relative to the ambient condition for total contents of polyunsaturated fatty acids, i.e. C18:2 and C18:3, according to Eqn (1).

Results

The contents of total lipid and FAs are given as percentage (%) of seed weight. Raw data can be found in [Supplementary Table S1](#) at *JXB* online, and summaries of the close to 750 individual measurements of FAs and total oil are presented in [Figs 1](#) and [2](#). The results show that the spectrum of FAs detected in the rapeseed samples was similar for all cultivars, and in agreement with previous reports on this crop ([Omidi et al., 2010](#); [Tan et al., 2011](#)). Oleic acid (C18:1) was the predominant FA, while linoleic acid (C18:2) and linolenic acid (C18:3) were the second and the third most abundant FAs. Furthermore, palmitic (C16:0), stearic (C18:0), and eicosenoic acids (C20:1) were present in trace amounts in all rapeseed varieties. The quantification method of the FAs was specifically designed to detect the six most common FAs in rapeseed. However, other unexpected FAs could also be detected due to the high resolution of the chromatographic method.

The oil composition exhibited significant shifts in the amount and proportion of the individual FAs in the different treatments compared with ambient. The changes in FAs and lipid content averaged over the four cultivars are shown in [Figs 1](#) and [2](#) and compared with ambient. The modeling ([Table 1](#)) revealed significant differences between treatments. It should be noted that the test statistics in [Table 1](#) accounts for intracorrelation between treatment effect estimates. Therefore, overlapping confidence intervals for parameters do not necessarily translate to statistically insignificant differences.

Fatty acids

All applied climate scenarios affected FA content, though in some treatments not all FAs were affected, e.g. the treatment

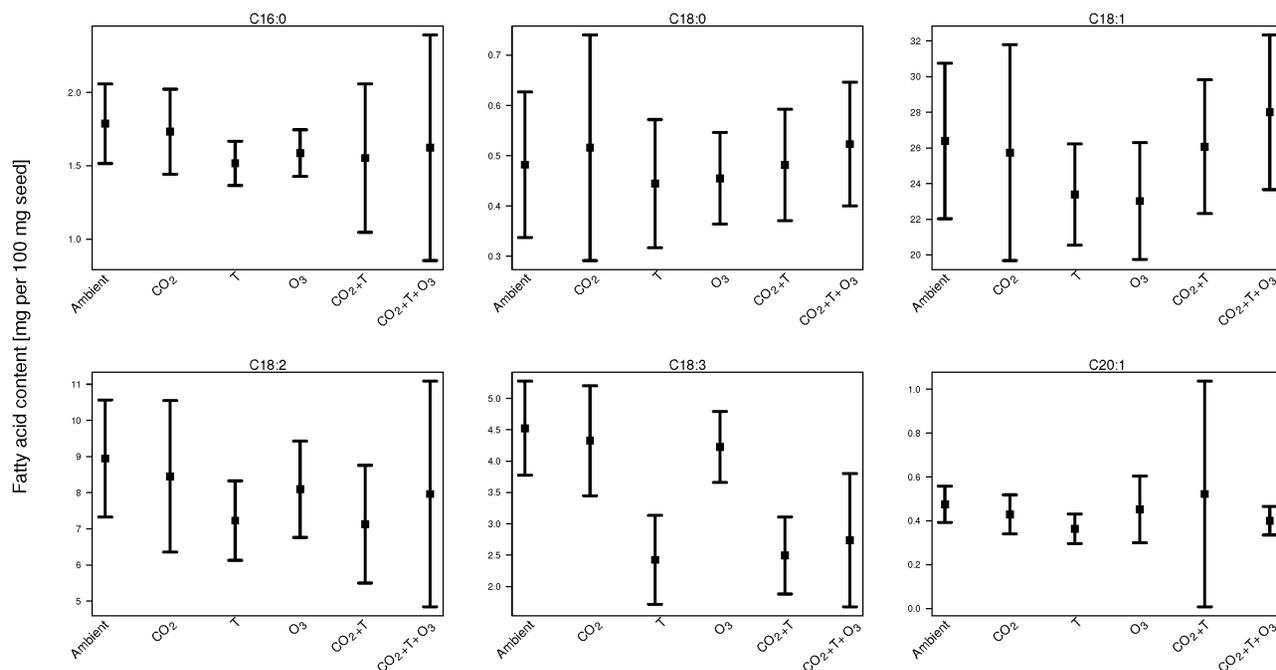


Fig. 1. Summary of the effects of future climate scenarios, CO₂, T, O₃, CO₂+T and CO₂+T+O₃, on the fatty acid content of rapeseed oil (mg fatty acid/100 mg seeds). Mean±standard error for six fatty acids (average of the four cultivars).

Table 1. Estimated levels of FAs and oil content and interaction between the factors CO₂, T and O₃ (all according to Eqn (1)). A P-value less than 0.05 for interaction between CO₂ and T, and between (CO₂,T) and O₃ indicates that factors interact and additivity cannot be assumed.

FA	Treatment						P-value for interaction between CO ₂ and T	P-value for interaction between (CO ₂ ,T) and O ₃
	mg (100mg) ⁻¹ seeds, (95% confidence intervals), P-values for difference from ambient treatment							
	Ambient	CO ₂	T	O ₃	CO ₂ +T	CO ₂ +T+O ₃		
C16:0	1.78 (1.69, 1.88)	1.73 (1.63, 1.82)	1.51 (1.43, 1.60)	1.58 (1.50, 1.67)	1.61 (1.52, 1.70)	1.70 (1.65, 1.75)	0.0008	<0.0001
	—	P=0.09	P<0.0001	P<0.0001	P<0.0001	P=0.01		
C18:0	0.47 (0.42, 0.53)	0.51 (0.45, 0.57)	0.44 (0.39, 0.50)	0.45 (0.40, 0.51)	0.48 (0.42, 0.54)	0.52 (0.46, 0.59)	0.58	0.009
	—	P=0.005	P=0.008	P=0.18	P=0.91	P=0.005		
C18:1	26.31 (24.52, 28.19)	25.57 (23.83, 27.40)	23.35 (21.76, 25.02)	22.70 (20.92, 24.58)	26.01 (24.25, 27.88)	27.93 (26.02, 29.93)	<0.0001	<0.0001
	—	P=0.17	P<0.0001	P<0.0001	P=0.59	P=0.004		
C18:2	8.82 (7.90, 9.83)	8.49 (7.60, 9.46)	7.30 (6.53, 8.13)	8.08 (7.22, 9.01)	7.02 (6.28, 7.82)	7.82 (6.99, 8.73)	0.22	<0.0001
	—	P=0.03	P<0.0001	P=0.0002	P<0.0001	P<0.0001		
C18:3	4.42 (4.05, 4.83)	4.39 (4.02, 4.79)	2.45 (2.24, 2.67)	4.22 (3.85, 4.62)	2.43 (2.22, 2.65)	2.69 (2.45, 2.94)	0.11	0.004
	—	P=0.76	P<0.0001	P=0.15	P<0.0001	P<0.0001		
C20:1	0.47 (0.44, 0.50)	0.43 (0.40, 0.46)	0.36 (0.34, 0.39)	0.42 (0.40, 0.46)	0.41 (0.38, 0.44)	0.40 (0.37, 0.43)	<0.0001	0.02
	—	P<0.0001	P<0.0001	P<0.0001	P<0.0001	P<0.0001		
Total oil (lipid sum)	42.49 (39.39, 45.78)	41.26 (38.23, 44.45)	34.51 (31.96, 37.21)	37.78 (35.02, 40.70)	38.22 (35.43, 41.17)	41.06 (38.06, 44.23)	<0.0001	<0.0001
	—	P=0.08	P<0.0001	P<0.0001	P<0.0001	P=0.03		

with elevated [CO₂] only had effects on C18:0, C18:2 and C20:1. The T increase of +5 °C day/night was the single factor with the largest detrimental effect on FA content as well as the quantity of the seed oil (Table 1). All six FAs were significantly decreased by this treatment, 8–46% (on average; Fig. 1

and Supplementary Table S1). The ozone treatment affected C16:0, C18:1, C18:2 and C20:1 significantly (Table 1) with decreases of 5–13% (Fig. 1 and Supplementary Table S1).

For the climate treatments combining elevated [CO₂], T and [O₃], the FA changes recorded were predominantly

unfavorable. In the scenario with all three factors elevated simultaneously, decreases were significant for the FAs C16:0 (9%), C18:2 (11%), C18:3 (39%) and C20:1 (16%), but the content of C18:0 and C18:1 increased (9% and 6%, respectively) (Table 1, Fig. 1 and Supplementary Table S1). When [CO₂] and T were elevated concurrently but [O₃] kept constant and low, reductions were significant for the FAs C16:0 (13%), C18:2 (20%) and C18:3 (45%) (Table 1, Fig. 1 and Supplementary Table S1).

The interactive effects between factors were modeled for the treatments with combination of factors. For the individual FAs, the interaction between elevated [CO₂] and elevated T in their two-factor treatment was significant for C16:0, C18:1 and C20:1 (Table 1). The interaction between elevated [O₃] and elevated [CO₂]+T, (CO₂,T), was found to be significant for all six FAs. For example, the estimated effect of the three-factor treatment on C18:3 was larger than could be expected from additivity of the three single factor treatment effects for this FA ($P=0.004$, Table 1).

Total lipid

The modeling revealed that the future climate scenarios decreased the oil content (Table 1). The treatments with elevated [CO₂] and T combined produced about 10% less lipid, and when all three factors were elevated simultaneously, the lipid sum decreased by 3%. In the single factor treatments with elevated T or O₃, the lipid content was reduced by approximately 17% and 11%, respectively (Fig. 2 and Supplementary Table S1).

There was a significant interaction between [CO₂] and T in their effect on total lipid ($P<0.0001$), and the effects from single factor treatments were not additive (Table 1). All three factors combined also had a greater effect than expected additive effects from single factor treatments. Assuming additivity of single factors, the expected lipid content would be $42.49-13.92=28.57$. However, the expected lipid content according to Table 1 when combining all factors was 41.06, which is greater than the expectation when assuming additivity of the single factor effects ($P<0.0001$).

Cultivar responses

The response of individual cultivars to the climate treatments are illustrated in Fig. 3 for total oil and two of the most important FAs from rapeseed, C18:3 and C18:1. Significant differences in the response of cultivars were noted for total oil across all treatments – the general trend was a decrease in oil content. Cultivars also showed different responses in C18:1 content under different climate treatments. Significant reductions in content were most common, but also significant increases in content were observed for the cultivars Bolero and Tanto, when they were grown in elevated [CO₂] or the three-factor treatment. Also for FA C18:3 differences among cultivars were observed in all treatments. All cultivars presented a significant reduction in C18:3 except Bolero, which showed no change in C18:3 content in elevated [CO₂]. Also for the other FAs (data not shown) variation among cultivars was the rule.

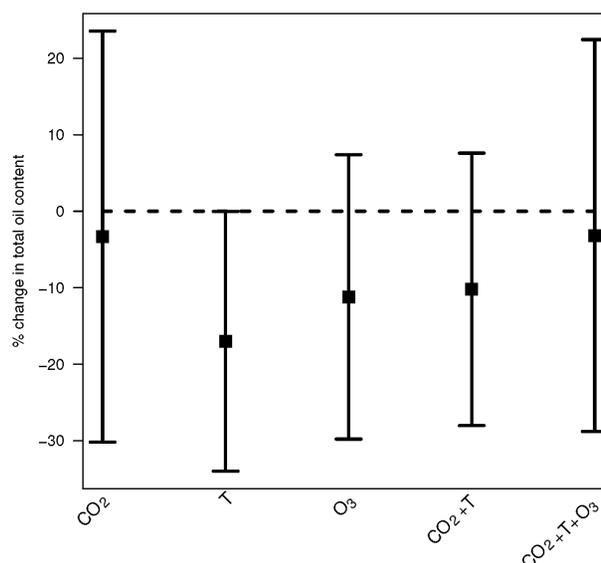


Fig. 2. Change in total rapeseed oil content (average of four cultivars) in the climate treatment, CO₂, T, O₃, CO₂+T and CO₂+T+O₃, relative to the ambient value (% change±standard error).

Change in fraction of the polyunsaturated FAs in the seed oil

In the present study the polyunsaturated FAs – considered an especially healthy component of the diet – were found to be significantly decreased by the climate treatments. The fraction of the two polyunsaturated FAs, C18:2 and C18:3 (ω 3, omega 3) of the total seed oil was reduced in all treatments relative to ambient (Fig. 4).

Discussion

The effect of multiple aspects of climate change on food quality has only been documented by a few experimental studies. Little is known about the effects of combining climate factors, making multi-treatment experiments and modeling pivotal; however, the single factor treatments are also needed for a fundamental understanding of the mechanisms of response.

Effects of the single factor treatments

Elevated [CO₂] Several authors (e.g. Long *et al.*, 2006; Hatfield *et al.*, 2011; IPCC 2014) have reviewed effects of the changing climate on crop production. It is generally assumed that elevated [CO₂] will increase crop production (Hatfield *et al.*, 2011). Many studies have reported that plants benefit from elevated [CO₂] due to a boost in photosynthesis, decreased stomatal conductance and improved leaf water status resulting in increased productivity (Leakey *et al.*, 2009; Högy *et al.*, 2010; Wang *et al.*, 2012). When it comes to the quality of oil crops, the limited research available indicates that elevated [CO₂] may have a positive effect, but certainly there are also negative impacts on oil quality by changing the composition of FAs on a dry weight basis. Illustrated by the handful of studies available on a very limited number

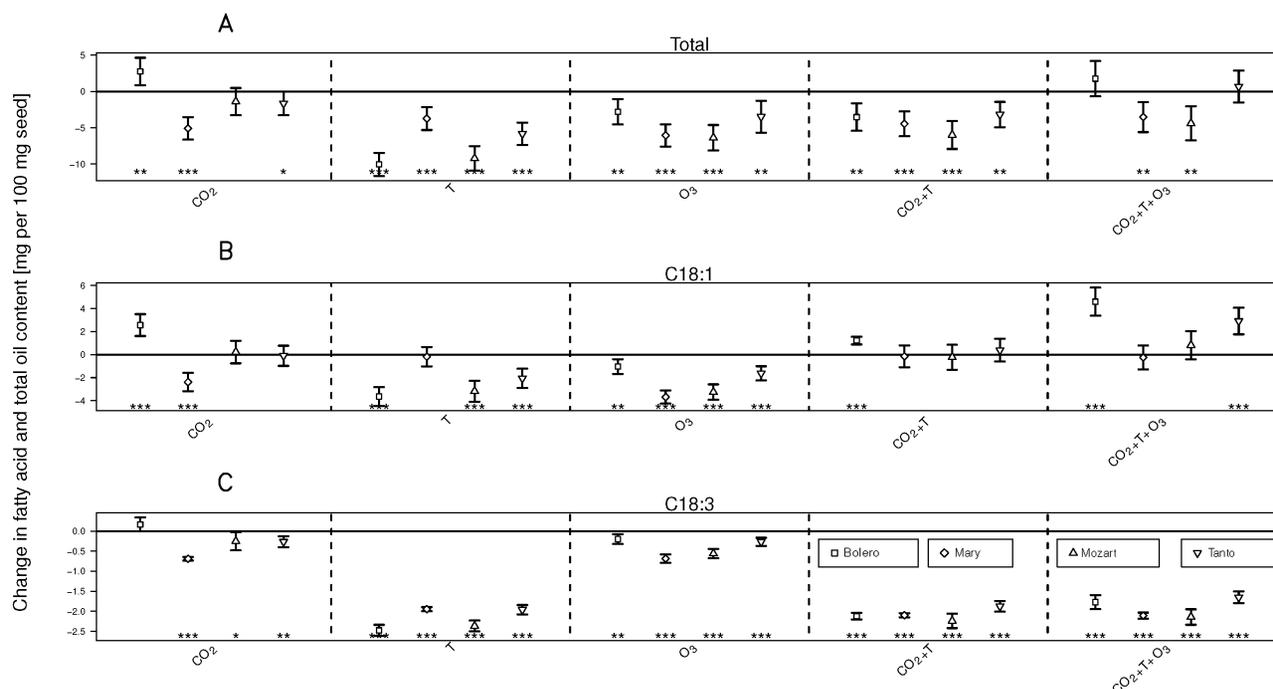


Fig. 3. Cultivar-specific changes in total rapeseed oil (A), and the fatty acids C18:1 (B) and C18:3 (C) content in the climate treatments, CO₂, T, O₃, CO₂+T and CO₂+T+O₃, relative to the ambient value (absolute change±standard error). Derived from Eqn (1) with cultivar effects kept fixed. Asterisks (***)*P*<0.001, (**)*P*<0.01, (**P*<0.05) indicate cultivar-specific responses to the corresponding treatment.

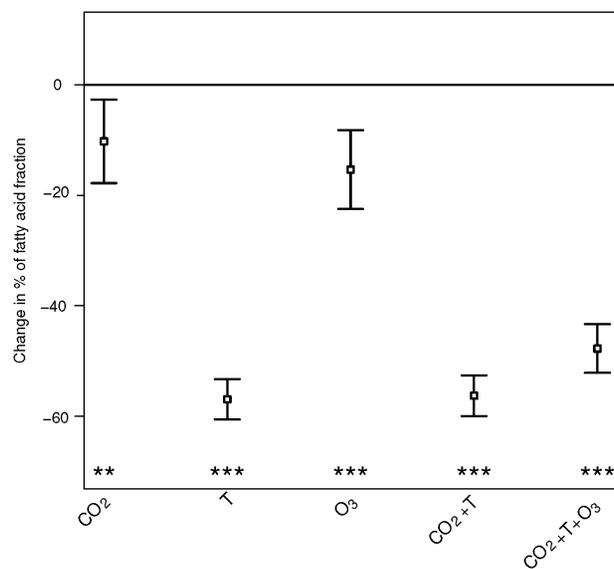


Fig. 4. Change in polyunsaturated fatty acid fraction of rapeseed oil in the climate treatments, CO₂, T, O₃, CO₂+T and CO₂+T+O₃, relative to the ambient value (% change±standard error). The polyunsaturated fatty acids were C18:2 and C18:3. Derived from Eqn (1). (***)*P*<0.0001, (**)*P*<0.01, (**P*<0.05).

of cultivars in sunflower (Pal *et al.*, 2014), castor bean (Vanaja *et al.*, 2008) and rapeseed (Högy *et al.*, 2010; Tian *et al.*, 2014), the crop quality response to elevated [CO₂] is not unequivocal. A growth chamber study of the rapeseed cultivar ‘Mozart’ – likely the same cultivar also analysed in the present study – demonstrated decreased seed oil% as well as seed yield, while vegetative biomass increased (Franzaring *et al.*, 2011; [CO₂] 550 ppm). Averaged over all four cultivars

of rapeseed our results showed that [CO₂] enrichment did not affect the oil% significantly; C18:0 increased and C18:2 and C20:1 were reduced (Table 1, Figs 1 and 2 and Supplementary Table S1). When comparing the response of individual cultivars, significant differences were noted for elevated [CO₂] (Fig. 3); however, ‘Mozart’ did not have changed oil% (Fig. 3), seed yield and aboveground biomass (Franck *et al.*, 2011), results that are contrary to the observations by Franzaring *et al.* (2011). The fact that the results on oil quality from elevated [CO₂] are not in agreement very likely reflects both inter- and intraspecific variation as well as experimental conditions. For example, the level of [CO₂] in the experiments will affect the photosynthetic response, transpiration, as well as the nutrients available (Franzaring *et al.*, 2011). This variability among studies underlines the need for analysis of more and diverse genotypes of a crop species grown in different environmental settings to create a broad spectrum of responses.

One suggested mechanism responsible for changes in FA content under elevated [CO₂] is that by increasing the availability of carbon, the production of malonyl CoA (the precursor required for synthesis of long- and short-chain FAs) changes, which possibly causes imbalance in FA proportions (Chakraborty and Uprety, 2012).

Elevated temperature In the present study elevated temperature was the factor affecting the oil composition and total oil the most. The lipid sum as well as all FAs were significantly decreased at a temperature increase of 5 °C (Table 1), and particularly the change in polyunsaturated FAs was large in treatments where temperature was elevated (Fig. 4). The effects of elevated T will certainly depend on

the magnitude of the treatment and the cardinal temperature of the species. In the RCP8.5 scenario with a possible 4–5 °C elevation of temperature, global crop yields are expected to decrease (Porter *et al.*, 2014). However, few experimental studies have analysed the quality of crops from scenarios where temperature is increased by 4 °C or more over the entire growing period (Porter *et al.*, 2014). Werteker *et al.* (2010) studied the effect of temperature on oil quality in rapeseed, soybean and sunflower in the last month before harvest. They found that in general, there was a negative correlation between temperature and the proportion of C18:3 and C18:2 in the crop oil. The same negative effect of elevated temperature on C18:3 has been reported in other studies (Thomas *et al.*, 2003; Kumar *et al.*, 2006). Byfield and Upchurch (2007) reported that higher temperatures increased expression of ω -3 FA desaturase in soybean, possibly resulting in lower C18:3 and higher C18:0 concentrations. In accordance with this, our treatments with elevated T significantly reduced concentrations of C18:3. In contrast C18:0 was found to be reduced in our study. The correct ratio of FAs in the human diet is important, especially because C18:3 is an essential FA that humans cannot synthesize.

Elevated ozone Effects of elevated ozone will depend on the regional tropospheric concentration and also on the tolerance of crop species to the oxidative stress (Mills *et al.*, 2007). The yield losses due to elevated [O₃] are estimated to be 10% for wheat and soybean and 3–5% for maize and rice (Van Dingenen *et al.*, 2009) since preindustrial times. In the present study it was found that the general effect of elevated [O₃] was a decrease in total lipid content, and a significant reduction in the content of the FAs C16:0, C18:1, C18:2 and C20:1, whereas C18:0 and C18:3 were unaffected (Table 1, Figs 1 and 2 and Supplementary Table S1). Also a previous report (De Bock *et al.*, 2011) found a reduction in lipid content of rapeseed, when ozone was increased 8 hours per day by 20 or 40 ppb above an ambient ozone concentration of 29–33 ppb. The authors suggested that this observation was explained by a change in carbon partitioning with an increased allocation of carbon to leaf injury repair. Tripathi and Agrawal (2013) exposed two cultivars of linseed to elevated [O₃] alone (27.7–59.0 ppb (ambient) + 10 ppb) and in combination with UV-B. They found that both as an individual factor and in combination with UV-B, ozone reduced oil content and oil quality of linseed; rancid oil of low nutritional value was favored. The same authors (Tripathi and Agrawal, 2012) analysed the effects of elevated [O₃] (26.3–69.5 ppb (ambient) + 10 ppb) in two cultivars of *Brassica campestris*, one of the progenitor species of oilseed rape. In this species, they found an increase in C18:1 and C18:3.

Interactive effects of factors in two- and three-factor treatments

The plant phenotypes resulting from an environment with a simultaneous increase of multiple factors like elevated [CO₂], T and [O₃] are little studied and not well understood. Generally, the effect of elevated T and elevated [O₃] on

plant production are considered counteractive to the effect of elevated [CO₂] (Tianhong *et al.*, 2005; Ainsworth *et al.*, 2008; Taub *et al.*, 2008; Gillespie *et al.*, 2012, Long, 2012). Our results on crop quality are consistent with this finding (Table 1), but additionally our analysis indicates that the effects of factors in combination cannot be estimated from combining results of single factor experiments. For example, in the scenario where T and CO₂ interacted, it was found that the content of C16:0, C18:1 and C20:1 could not have been predicted from the results of the individual single factor treatments (Table 1, second to last column). Also for the lipid content the outcome from combining T and CO₂ was influenced by their interaction (Table 1, second to last column). If [O₃] was increased together with T and [CO₂] in the three-factor combination, O₃ interacted with (CO₂,T) and influenced all FAs and the collective lipid sum (Table 1, last column). Undoubtedly the more interacting factors present in the environment, the more uncertainty there will be in extrapolating results from single factor experiments. Likewise, with respect to the response of individual cultivars in climate scenarios with combined factors, results from single factor treatments might not be additive (Fig. 3). The interactive effect of O₃ and CO₂ has been studied previously in regard to yield and oil content of *Brassica juncea* (Indian mustard), and the oil content was found to decrease at elevated [O₃] (25–35 ppb). This reduction was only partly compensated by exposing the plant to elevated [CO₂] (550 ppm [CO₂]; Singh *et al.*, 2013). Unfortunately, we were not able to resolve if O₃ interacted with CO₂, T or both factors at the same time, as the limitation of growth chambers prevented experimentation of the two-way interactions CO₂:O₃ and T:O₃.

Combined effects of reductions in seed yield, oil quantity and quality

Besides the unfavorable effects on oil content and quality of rapeseed, a reduction in seed production of this crop can be foreseen in a changed climate (Frenck *et al.*, 2011, 2013; Supplementary Table S2). Compared with ambient conditions, the treatment with elevated [CO₂] did not increase seed production significantly, while under elevated T and the combination of elevated [CO₂] and T, the seed yield was decreased by approximately 50% (Frenck *et al.*, 2013; Supplementary Table S2). This trend in seed production is supported by previous studies on wheat and rapeseed, where growth was studied in the two-factor treatments with elevated [CO₂] and T (Qaderi *et al.*, 2006; Manoj-Kumar *et al.*, 2012). In the two-factor treatment with elevated [CO₂] and T in our study, the essential FA, C18:3, decreased by 45% in addition to the 50% reduction of the seed set, and the oil content decreased by 10%. We acknowledge that our results were obtained in a phytotron, and therefore comparable experiments with combinations of elevated [CO₂] and T in the field might not have the same outcome. However, such experiments have not yet been performed because of technical and economic challenges. Modeling of European crop yields under climate change conditions has been performed (Supit *et al.*, 2012), and the results indicated that in northern Europe yields of spring crops either stagnate

or decline depending on the magnitude of increase in T and [CO₂]. If the results obtained in our study reflect a future worst case scenario, climate change effects on spring oilseed rape could be profound. Currently the average seed yield of spring rapeseed in Denmark is 2.48 ton ha⁻¹ (StatBank Denmark, 2016). However, assuming a yield reduction of 50%, when [CO₂] and T are elevated simultaneously to levels applied in the present study, the seed harvest will be reduced to 1.24 ton ha⁻¹. The oil yield will drop from the present 1.12 ton ha⁻¹ (45 wt% oil/seed; Wittkop *et al.*, 2009) to 0.47 ton ha⁻¹ (38 wt% oil/seed in our two-factor treatment), and the content of C18:3 will fall from 0.11 ton ha⁻¹ (today C18:3 constitutes approximately 10wt% of the oil (Omidi *et al.*, 2010); in the future the content will be reduced by 45%) to 0.03 ton ha⁻¹, when reductions in seed weight, oil content and C18:3 are added up. If this worst case scenario becomes a reality, the harvested oil per hectare will be reduced by 58%, and content of C18:3 will decrease by 77%. For the top three rapeseed producing countries, Canada (rapeseed crop worth approximately 8 billion CAD), India and China (each with a rapeseed area of 7–8 Mha) such a reduction would have significant economic consequences.

Unsaturated FAs

Unsaturated FAs are preferred in the human diet (McKevith, 2005; Gebauer *et al.*, 2006), since they are assumed to reduce the risk of cardiovascular diseases, while for bio-fuel storage saturated FAs are favored, as a decline in saturation causes an increase in oxidation of FA and therefore less stability (Hoekman *et al.*, 2012). Especially the polyunsaturated FAs are assumed to have health benefits (Kris-Etherton *et al.*, 2004; Reed *et al.*, 2007). Therefore, the consequences for human nutritional value of the observed dramatic decrease in C18:2 plus C18:3 (Fig. 4) in combinations of elevated [CO₂], T and [O₃] are profound, and could be a challenge for the future quality of rapeseed oil for consumption.

Though C18:1 is a monounsaturated FA, it is the most abundant and preferred FA in vegetable oil for bio-fuel production (Pinzi *et al.*, 2013). This FA was not reduced by the two-factor treatment, and even somewhat increased in the three-factor treatment (Table 1, Fig. 1). Therefore, bio-fuel quality may not be negatively affected by the future climate changes. However, oil quantity per hectare will decrease and affect the harvested amount of any FA.

Outlook

The overall decrease in quality and quantity of rapeseed oil found in the present study depicts a pessimistic future for seed oil production from rapeseed and for the health benefits of rapeseed oil. However, the difference in cultivar responses indicates that cultivars holding specific beneficial genes may be identified and exploited to retain oil quality and quantity despite comprehensive changes in the climate. At this point it seems advisable and profitable to initiate screening programs for genetic resources of crop plants in order to optimize food quality under future climate conditions.

Supplementary data

Supplementary data are available at *JXB* online.

Table S1. Data set on contents of fatty acids and total lipid.

Table S2. Detailed data on seed yield.

Acknowledgements

This work was supported in part by Nordic Council of Ministers (NordForsk) through the network 'Sustainable primary production in a changing climate' (grant number 18800).

References

- Ainsworth EA, Leakey A, Ort DR, Long SP. 2008. FACE-ing the facts: inconsistencies and interdependence among field, chamber and modeling studies of elevated CO₂ impacts on crop yield and food supply. *New Phytologist* **179**, 5–9.
- Baux A, Colbach N, Allirand JM, Julien A, Ney B, Pellet D. 2013. Insights into temperature effects on the fatty acid composition of rapeseed varieties. *European Journal of Agronomy* **49**, 12–19.
- Byfield G, Upchurch RG. 2007. Effect of temperature on microsomal omega-3 linoleate desaturase gene expression and linolenic acid content in developing soybean seeds. *Crop Science* **47**, 2445–2452.
- Chakraborty K, Upreti DC. 2012. Elevated CO₂ alters seed composition and quality in Brassica. *Indian Journal of Plant Physiology* **17**, 84–87.
- Craig BM, Murty NL. 1959. Quantitative fatty acid analysis of vegetable oils by gas-liquid chromatography. *Journal of the American Oil Chemist's Society* **36**, 549–552.
- De Bock M, Op de Beeck M, De Temmerman L, Guisez Y, Ceulemans R, Vandermeiren K. 2011. Ozone dose-response relationships for spring rapeseed and broccoli. *Atmospheric Environment* **45**, 1759–1765.
- European Commission. 2013. Agriculture in the European union: Statistical and Economic Information, Report 2013. http://ec.europa.eu/agriculture/statistics/agricultural/2013/pdf/full-report_en.pdf (14 April 2016, date last accessed).
- Franzaring J, Weller S, Schmid I, Fangmeier A. 2011. Growth, senescence and water use efficiency of spring rapeseed (*Brassica napus* L. cv. Mozart) grown in a factorial combination of nitrogen supply and elevated CO₂. *Environmental and Experimental Botany* **72**, 284–296.
- Frenck G, Linden LD, Mikkelsen TN, Brix H, Jørgensen RB. 2011. Increased [CO₂] does not compensate for negative effects on yield caused by higher temperature and [O₃] in *Brassica napus* L. *European Journal of Agronomy* **35**, 127–134.
- Frenck G, Linden L, Mikkelsen TN, Brix H, Jørgensen RB. 2013. Response to multi-generational selection under elevated [CO₂] in two temperature regimes suggests enhanced carbon assimilation and increased reproductive output in *Brassica napus* L. *Ecology and Evolution* **3**, 1163–1172.
- Gebauer SK, Psota TL, Harris WS, Kris-Etherton PM. 2006. n-3 fatty acid dietary recommendations and food sources to achieve essentiality and cardiovascular benefits. *American Journal of Clinical Nutrition* **83**, 1526S–1535S.
- Gillespie KM, Xu F, Richter KT, McGrath JM, Markelz RJC, Ort DR, Leakey ADB, Ainsworth EA. 2012. Greater antioxidant and respiratory metabolism in field-grown soybean exposed to elevated O₃ under both ambient and elevated CO₂. *Plant Cell and Environment* **35**, 169–184.
- Hatfield JL, Boote KJ, Kimball BA, Ziska LH, Izaurralde RC, Ort D, Thomson AM, Wolfe D. 2011. Climate impacts on agriculture: implications for crop production. *Agronomy Journal* **103**, 351–370.
- Hoekman SK, Broch A, Robbins C, Ceniceros E, Natarajan M. 2012. Review of biodiesel composition, properties, and specifications. *Renewable and Sustainable Energy Reviews* **16**, 143–169.
- Högy P, Franzaring J, Schwadorf K, Breuer J, Schütze W, Fangmeier A. 2010. Effects of free-air CO₂ enrichment on energy traits and seed quality of rapeseed. *Agriculture Ecosystem and Environment* **139**, 239–244.

- IPCC.** 2007. Summary for policymakers. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL, eds. *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge and New York: Cambridge University Press, 1–18.
- IPCC.** 2013. Summary for policymakers. In: Midgley PM, Stocker TF, Qin D, Plattner, GK, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex V, eds. *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge and New York: Cambridge University Press, 3–29.
- IPCC.** 2014. Summary for policymakers. In: White LL, Field CB, Barros VR, Dokken DJ, Mach KJ, Mastrandrea MD, Bilir TE, Chatterjee M, Ebi KL, Estrada YO, Genova RC, Girma B, Kissel ES, Levy AN, MacCracken S, Mastrandrea PR, eds. *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge and New York: Cambridge University Press, 2–31.
- Kris-Etherton PM, Binkoski AE, Hecker KD.** 2004. Polyunsaturated fatty acids and cardiovascular health. *Nutrition Reviews* **62**, 414–426.
- Kumar V, Rani A, Solanki S, Hussain SM.** 2006. Influence of growing environment on the biochemical composition and physical characteristics of soybean seed. *Journal of Food Composition and Analysis* **19**, 188–195.
- Leakey ADB, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP, Ort DR.** 2009. Elevated CO₂ effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. *Journal of Experimental Botany* **60**, 2859–2876.
- Le Quéré C, Peters GP, Andres RJ, et al.** 2014. Global carbon budget 2013. *Earth System Science Data Discussions* **6**, 689–760.
- Long SP.** 2012. Virtual Special Issue on food security – greater than anticipated impacts of near-term global atmospheric change on rice and wheat. *Global Change Biology* **18**, 1489–1490.
- Long SP, Ainsworth EA, Leakey ADB, Nösberger J, Ort DR.** 2006. Food for thought: lower-than-expected crop yield stimulation with rising CO₂ concentrations. *Science* **312**, 1918–1921.
- McKevith B.** 2005. Nutritional aspects of oilseeds. *Nutrition Bulletin* **30**, 13–26.
- Manoj-Kumar, Swarup A, Patra AK, Chandrakala JU, Manjaiah KM.** 2012. Effect of elevated CO₂ and temperature on phosphorus efficiency of wheat grown in an inceptisol of subtropical India. *Plant Soil and Environment* **58**, 230–235.
- Mills G, Buse A, Gimeno B, Bermejo V, Holland M, Emberson D, Pleijel H.** 2007. A synthesis of AOT40-based response functions and critical levels of ozone for agricultural and horticultural crops. *Atmospheric Environment* **41**, 2630–2643.
- Omid H, Tahmasebi Z, Naghdi Badi HA, Torabi H, Miransari M.** 2010. Fatty acid composition of canola (*Brassica napus* L.) as affected by agronomical, genotypic and environmental parameters. *Comptes Rendus Biologies* **333**, 248–254.
- Pal M, Chaturvedi AK, Pandey SK, Bahuguna RN, Khetarpal S, Anand A.** 2014. Rising atmospheric CO₂ may affect oil quality and seed yield of sunflower (*Helianthus annuus* L.). *Acta Physiologiae Plantarum* **36**, 2853–2861.
- Pinzi S, Rounce P, Herreros JM, Tsolakis A, Dorado MP.** 2013. The effect of biodiesel fatty acid composition on combustion and diesel engine exhaust emissions. *Fuel* **104**, 170–182.
- Porter JR, Xie L, Challinor AJ, Cochrane K, Howden SM, Iqbal MM, Lobell DB, Travasso MI.** 2014. Food security and food production systems. In: Field CB., Barros VR, Dokken DJ, Mach KJ, Mastrandrea MD, Bilir TE, Chatterjee M, Ebi KL, Estrada YO, Genova RC, Girma B, Kissel ES, Levy AN, MacCracken S, Mastrandrea PR, White LL, eds. *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge and New York: Cambridge University Press, 485–533.
- Qaderi MM, Kurepin LV, Reid DM.** 2006. Growth and physiological responses of canola (*Brassica napus*) to three components of global climate change: temperature, carbon dioxide and drought. *Physiologia Plantarum* **128**, 710–721.
- R Core Team.** 2014. R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. <http://www.R-project.org/>.
- Reed SC, Simpson MJ, Millington KJ, Ruxton CH.** 2007. The health benefits of omega-3 polyunsaturated fatty acids: a review of the evidence. *Journal of Human Nutrition and Dietetics* **20**, 275–285.
- Singh S, Bhatia A, Tomer R, Kumar V, Singh B, Singh SD.** 2013. Synergistic action of tropospheric ozone and carbon dioxide on yield and nutritional quality of Indian mustard (*Brassica juncea* (L.) Czern.). *Environmental Monitoring and Assessment* **185**, 6517–6529.
- StatBank Denmark.** 2016. Business sectors, agriculture, horticulture and forestry, crop production, Harvest by crop and unit 2009–2013. <http://www.statistikbanken.dk/statbank5a/default.asp?w=1680> (14 April 2016, date last accessed).
- Supit I, Diepen CA, Wit AJW, Kabat P, Baruth B, Ludwig F.** 2010. Recent changes in the climatic yield potential of various crops in Europe. *Agricultural Systems* **103**, 683–694.
- Supit I, Diepen CA, Wit AJW, Wolf J, Kabat P, Baruth B, Ludwig F.** 2012. Assessing climate change effects on European crop yields using the Crop Growth Monitoring System and a weather generator. *Agricultural and Forest Meteorology* **164**, 96–111.
- Tan H, Yang X, Zhang F.** 2011. Enhanced seed oil production in canola by conditional expression of *Brassica napus* LEAFY COTYLEDON1 and LEC1-LIKE in developing seeds. *Plant Physiology* **156**, 1577–1588.
- Taub DR, Miller B, Allen H.** 2008. Effects of elevated CO₂ on the protein concentration of food crops: a meta-analysis. *Global Change Biology* **14**, 565–575.
- Thomas JMG, Boote KJ, Allen LH, Gallo-Meagher M, Davis JM.** 2003. Elevated temperature and carbon dioxide effects on soybean seed composition and transcript abundance. *Crop Science* **43**, 1548–1557.
- Tian S, Jia Y, Ding Y, Wang R, Feng R, Song Z, Guo J, Zhou L.** 2014. Elevated atmospheric CO₂ enhances copper uptake in crops and pasture species grown in copper-contaminated soils in a micro-plot study. *Clean – Soil, Air, Water* **42**, 347–354.
- Tianhong Z, Yi S, Guohong H, Yan W, Bei S.** 2005. Respective and interactive effects of doubled CO₂ and O₃ concentration on membrane lipid peroxidation and anti-oxidative ability of soybean. *Science in China Series C: Life Sciences* **48**, 136–141.
- Triboi-Blondel AM, Renard M.** 1999. Effects of temperature and water stress on fatty acid composition of rapeseed oil. In: *Proceedings of the 10th International Rapeseed Congress*, 26–29 September 1999, Canberra, Australia.
- Tripathi R, Agrawal SB.** 2012. Effects of ambient and elevated level of ozone on *Brassica campestris* L. with special reference to yield and oil quality parameters. *Ecotoxicology and Environmental Safety* **85**, 1–12.
- Tripathi R, Agrawal SB.** 2013. Interactive effect of supplemental ultraviolet B and elevated ozone on seed yield and oil quality of two cultivars of linseed (*Linum usitatissimum* L.) carried out in open top chambers. *Journal of the Science of Food and Agriculture* **93**, 1016–1025.
- USDA.** 2016. Oilseeds. United States Department of Agriculture, Foreign Agricultural Service. <http://apps.fas.usda.gov/psdonline/psdHome.aspx> (14 April 2016, date last accessed).
- Vanaja M, Jyothi M, Ratnakumar P, Vagheera P, Reddy PR, Lakshmi NJ, Yadav SK, Maheshwari M, Venkateswarlu B.** 2008. Growth and yield responses of castor bean (*Ricinus communis* L.) to two enhanced CO₂ levels. *Plant, Soil and Environment* **54**, 38–46.
- Van Dingenen R, Dentener FJ, Raes F, Krol MC, Emberson L, Cofala J.** 2009. The global impact of ozone on agricultural crop yields under current and future air quality legislation. *Atmospheric Environment* **43**, 604–618.
- Wang D, Heckathorn SA, Wang X, Philpott SAA.** 2012. Meta-analysis of plant physiological and growth responses to temperature and elevated CO₂. *Oecologia* **169**, 1–13.
- Werteker M, Lorenz A, Johannes H, Berghofer E, Findlay CS.** 2010. Environmental and varietal influences on the fatty acid composition of rapeseed, soybeans and sunflowers. *Journal of Agronomy and Crop Science* **196**, 20–27.
- Wittkop B, Snowdon RJ, Friedt W.** 2009. Status and perspectives of breeding for enhanced yield and quality of oilseed crops for Europe. *Euphytica* **170**, 131–140.