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Abstract

Mitigation activities to improve water quality and quantity in streams as well as stream management and restoration efforts are conducted in the European Union aiming to improve the chemical, physical and ecological status of streams. Headwater streams are often characterised by impairment of hydro-morphological, chemical, and ecological conditions due to multiple anthropogenic impacts. However, they are generally disregarded as water bodies for mitigation activities in the European Water Framework Directive despite their importance for supporting a higher ecological quality in higher order streams.

We studied 11 headwater streams in the Hove catchment in the Copenhagen region. All sites had substantial physical habitat and water quality impairments due to anthropogenic influence (intensive agriculture, urban settlements, contaminated sites and low base-flow due to water abstraction activities in the catchment). We aimed to identify the dominating anthropogenic stressors at the catchment scale causing ecological impairment of benthic macroinvertebrate communities and provide a rank-order of importance that could help prioritising mitigation activities. We identified numerous chemical and hydromorphological impacts of which several were probably causing major ecological impairment, but we were unable to provide a robust rank-ordering of importance suggesting that targeted mitigation efforts on single anthropogenic stressors in the catchment are unlikely to have substantial effects on the ecological quality in these streams.

The SPEcies At Risk (SPEAR) index explained most of the variability in the macroinvertebrate community structure, and notably, SPEAR index scores were often very low (< 10 % SPEAR abundance). An extensive re-sampling of a subset of the streams provided evidence that especially insecticides were probably essential contributors to the overall ecological impairment of these streams.

Our results suggest that headwater streams should be considered in future management and mitigation plans. Catchment-based management is necessary because several anthropogenic stressors exceeded problematic thresholds, suggesting that more holistic approaches should be preferred.

1. Introduction

The conversion of land to agriculture and urban settlements are global phenomena that characterise the general intensification of land use. This intensification is considered one of the most important drivers for the increasing rate of biodiversity loss and impairment of ecosystem functions (Matson et al., 1997; MEA, 2005). Surface water ecosystems have been identified as being particularly vulnerable to these changes, and direct and indirect effects of increasing land use intensity on their physical condition, water chemistry, and aquatic biota have been extensively discussed in the literature (e.g. Allan, 2004; Ormerod et al., 2010; Pedersen, 2009). Nevertheless, human exploitation of water resources and the resultant impairment of freshwater ecosystems outpace our attempts to mitigate and manage these impacts (Ormerod et al., 2010).

In the European Union (EU), the implementation of the Water Framework Directive (WFD) obliges member states to obtain "good ecological and chemical status" of surface waters through improved catchment scale management (River Basin Management Plans, RBMPs). However, the WFD objectives are only applied to discrete and significant surface water bodies. Due to their small size, headwater streams (1st and 2nd order) are often not qualified as being significant surface water bodies and are therefore excluded from the RBMPs. The exclusion of headwater streams in the WFD is controversial, since (i) a major part of stream networks is low order streams (Freeman et al., 2007), (ii) due to the tight connection between land and streams in headwater systems, the majority of diffuse source chemicals enter stream networks in the headwater sections (e.g. Dodds and Oakes, 2008), (iii) headwater streams add important biodiversity to the overall stream system because of high habitat diversity (Meyer et al., 2007), and (iv) headwater streams are essential for the successful dispersal of species among stream networks due to their function as migration corridors (Freeman et al., 2007; Meyer et al., 2007). Consequently, the management of headwater

streams is crucial for maintaining ecosystem health in higher order streams that are targeted by the WFD aims.

The management of headwater streams almost always involves multiple and interrelated factors, with human activities resulting in many different types of environmental impact. For example, conventional agricultural practices often result in chemical pollution (nutrients and pesticides) and habitat degradation of streams (channelisation, weed-cutting, dredging and mowing of riparian vegetation). Moreover, several anthropogenic stressors may act concurrently with differing intensity; for example chemical pollution may be constituted of agricultural pesticides, macro- and micronutrients originating from agriculture, pesticides originating from urban area activities and scattered settlements, petroleum hydrocarbons and chlorinated solvents discharging to streams from contaminated sites (e.g. industrial sites, old landfills and former gasoline stations), and waste water treatment. Each of these sources discharges compounds with differing physicochemical properties. Moreover, the different sources act differently in time, for example diffuse source pollution with agricultural pesticides is primarily restricted to periods with heavy rainfall during pesticide spraying seasons, while the compounds discharging into streams from contaminated sites through groundwater are chronic sources of pollution that are likely to act with the highest intensity during summer low flow conditions due to a lower dilution factor. The effects of combinations of stressors often provide unpredictable results compared to what would be expected from the single stressors alone (Townsend et al., 2008). As a consequence, the studies of complex mixtures of anthropogenic stressors are challenging, but essential for the improvement of mitigation efforts and management strategies for these systems.

The need for ecological approaches to disentangle the effects of co-existing anthropogenic stressors has been urged by several authors (e.g. Beketov and Liess, 2012; Segner, 2011; Statzner and Bêche, 2010). Equally important is the characterisation of sum effects of stressor combinations.

Due to direct and/or indirect interactions of stressors, the knowledge of single stressor effects is often not sufficient to provide reliable predictions of multiple stressor effects (Rasmussen et al., 2012; Townsend et al., 2008). Applying ecological traits of macroinvertebrates has promising potential as a tool for disentangling effects of different anthropogenic stressors (see review by Statzner and Bêche, 2010). For example, sediment transport and diffuse source pesticide pollution has been shown to cause trait changes that may partly overrule effects of other anthropogenic stressors (e.g. Liess and von der Ohe, 2005; Matthaei et al., 2010; Schäfer et al, 2012a). However, comprehensive field studies aiming to distinguish the effects of pesticides and groundwater based contaminants (e.g. chlorinated solvents and petroleum hydrocarbons) from other anthropogenic stressors are still scarce (but see McKnight et al., 2012; Rasmussen et al., 2012). Moreover, stream ecosystems that are severely impacted by a multitude of anthropogenic stressors complicate the use of ecological traits for disentangling multiple-stressor effects, because several stressors may *a priory* be expected to cause similar changes in trait categories (Statzner and Bêche, 2010). Nevertheless, this may often be the scenario that managers of headwater streams will encounter.

We studied 11 headwater streams (1st and 2nd order) in the Hove catchment west of Copenhagen, Denmark. All streams were subject to multiple stressors of various origins, including contaminated sites, urban discharges, pesticide pollution from agriculture and habitat degradation. The purpose of this study was threefold: (1) to identify the main stressors at the catchment scale; (2) evaluate the total impact on the benthic macroinvertebrate communities, and if possible rank-order the importance of the dominating stressor types by the use of correlation analyses; (3) give guidance for prioritising focus areas for mitigation activities in headwater streams at the catchment level. We used a series of macroinvertebrate metrics for the interpretation of responses in the macroinvertebrate communities including the trait based metrics: the Danish Stream Fauna Index (DSFI; directed for organic pollution); the Lotic Invertebrate index for Flow Evaluation (LIFE;

directed for antecedent flow characteristics and substrate types); and the SPEcies At Risk index (SPEAR; directed for the effects of pesticides).

2. Materials and methods

2.1 Study area

The study catchment (195 km^2) is located west of Copenhagen on Sjaelland, Denmark (Fig. 1), where the catchment is characterised by a low elevation, clayey/loamy soils, a temperate climate and an average regional precipitation of 500 mm yr^{-1}. The catchment contains two dominant streams and their tributaries: the Hove stream and the Nybølle stream, the latter of which joins with the Hove stream in the north-central part of the catchment and eventually flows into Roskilde Fjord. Approximately 80 % of the total catchment is used for agriculture; the rest is comprised of ca. 15 % natural area (i.e. forest and wetlands), and 5 % urban area (i.e. settlements and industry). The dominant arable crop types are wheat and rapeseed, with large portions of the catchment used either as grassland or left uncultivated. There is significant groundwater abstraction for drinking water supply at the junction of Hove and Nybølle streams, i.e. along 3rd and 4th order stream sections.

The 11 study stream reaches were identified in order to represent the dominating anthropogenic stressors in the catchment: contaminated sites, urban discharges and diffuse pollution from agriculture. For each dominating stressor type, we selected 3-4 streams for the study. Groundwater abstraction and waste water treatment were present only at one 1st order stream each in the catchment, and so were not considered further. Specifically, sub-catchments for the study streams were constructed using ESRI ArcGIS$^®$ 9.3, in order to facilitate calculations for the specific land use percentages potentially influencing each sampling location (Table 1). The sub-catchments were produced from a soil and groundwater topography map with a 100 m x 100 m resolution and they

represent the total area that is drained by the stream from the sampling point and upstream. Site three was situated just outside the Hove catchment.

For the identification of stream reaches that could potentially be impacted by adjacent contaminated sites, we used model simulations to determine the locations where groundwater-surface water interactions would most likely to take place. For this, a MIKE SHE model of the Hove catchment was extracted and refined from a larger model of the island of Sjaelland, Denmark (Kürstein et al., 2009). Particle tracking simulations were then performed to determine the direction of groundwater flow from contaminated sites. Here, one particle was added for each cell where the contaminated sites are located. Possible particle sinks (or end locations) include: (i) rivers, by way of groundwater base-flow, (ii) rivers, by way of tile drains, (iii) pumping wells, (iv) an unsaturated zone cell, or (v) another saturated groundwater zone cell.
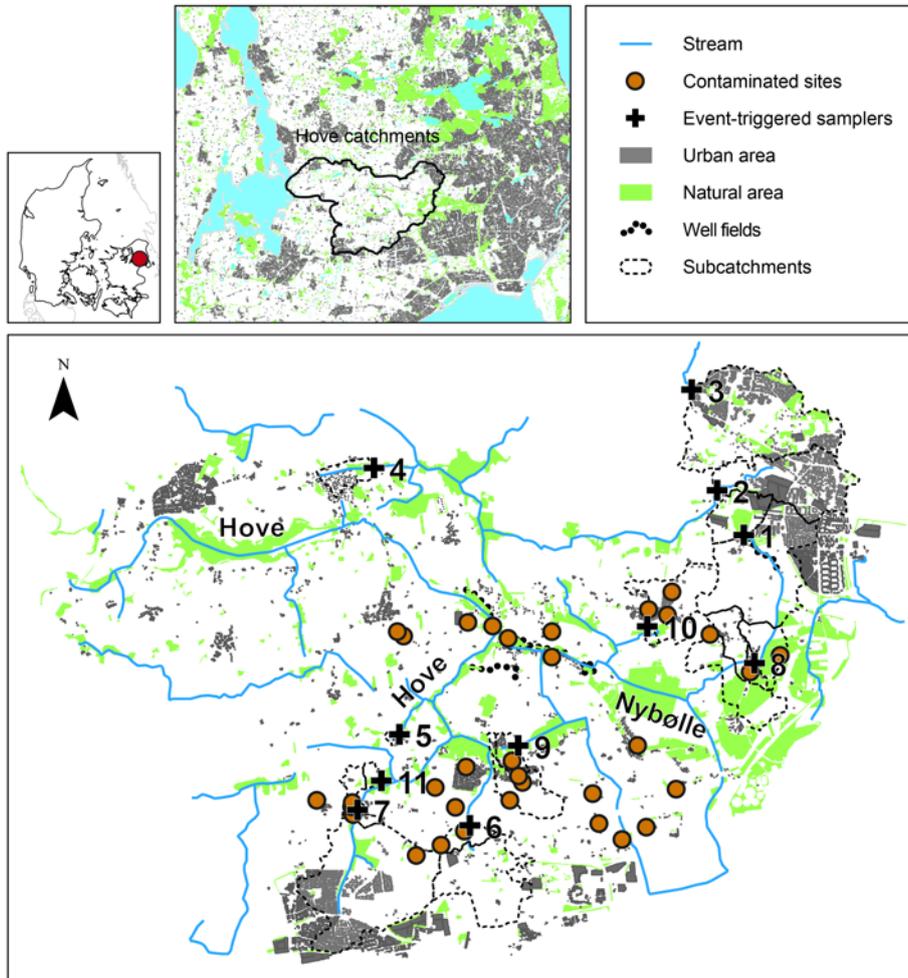
Figure 1: Overview of the field study area on Zealand, Denmark, including sampling locations (crosses), contaminated sites (circles) and well fields. Also shown is an overview of the sub-catchments (broken lines) draining to each sampling location, as well as the urban and natural areas; areas without color represent agricultural land (including grassland).

2.1.1 Control sites

We searched for control sites with "least disturbed conditions" (Stoddard et al., 2006) in the study catchment using data from the NOVANA program (the Danish stream monitoring program). We were, however, unable to find suitable sites and we thus expanded the search to the Copenhagen region. Available data (general water chemistry, hydromorphology and macroinvertebrate samples) for the identified sites were extracted from the ODA database (https://oda.dk) (Supplementary Material, Tables A.3 and C.4). In total, we included six NOVANA sites that qualified as having

"least disturbed conditions", considered by McKnight et al. (2012) for a nearby catchment. Consult McKnight et al. (2012) for selection criteria and sampling details.

2.2 Overall water sampling strategy and study set-up

In 2010, we conducted stream water chemistry sampling using different techniques and at different times in order to capture the expected maximum concentrations of chemicals from the identified potential anthropogenic sources in the 11 study streams (Fig. 1). Diffuse source pesticides were sampled with event-triggered water samplers (Liess and von der Ohe, 2005) during May-June; groundwater inflow of pesticides, chlorinated solvents and petroleum hydrocarbons were sampled manually in August during base-flow conditions. General water chemistry parameters were collected twice using manual grab sampling. Groundwater contaminants were one main focus area in the 2010 field campaign, and the selected list of pesticides for analysis was therefore restricted to water soluble compounds with long half-lives (only herbicides and degradation products of herbicides) (Table 2). Due to the comprehensive detections of currently used pesticides and historically low SPEAR values (see the Results section), we conducted a follow-up field campaign in 2011. Sites that were characterised by summer drought in 2010 or dissolved oxygen concentration < 4 mg L^{-1} were excluded. In 2011, we additionally sampled two of the control streams with the same sampling program as in 2010, but using the extended list of pesticide analyses for the event-triggered water samples. Moreover, we deployed passive *in situ* suspended particle samplers (SPS) for the detection of the most lipophilic pesticides. Due to limited quantity of sampling equipment, the SPS samplers were only deployed in the three streams with highest sum concentrations of pesticides during storm flow. Below, the sampling methods are described in detail.

2.2.1   General water chemistry

The biological oxygen demand (BOD_5) and micro- and macro-nutrient concentrations were measured in June and August, 2010. Conductivity, oxygen concentrations and pH were measured before collecting water samples for water chemistry. Water samples for cation were analysed according to the methods given in Milosevic et al. (2012). The general water chemistry is summarised in Table 3 and is furthermore provided in full detail (i.e. per sampling location) in Supplementary Material, Table A.2.

2.2.2 Pesticide sampling

Pesticides were sampled with two different sampling strategies in 2010; event-triggered samplers (May and June) (Liess and von der Ohe, 2005) and grab sampling (August) to differentiate between the compounds that are transported to streams via surface runoff and flow through tile drains during heavy precipitation events, and the compounds that mainly originate from groundwater inflow, respectively.

During May and June, 2010, two precipitation episodes triggered the sampling system in some or all study streams on the 15th and the 30th of May. The precipitation depths were 17.5 mm and 13 mm, respectively.

In 2011, a subset of the streams (sites 2, 3, 5, 6 and 7) were re-sampled during May and June using event-triggered water samplers. Additionally, we deployed event-triggered sampling systems in two of the control streams (C1 and C2). In 2011, the sampling systems were triggered twice on May 22nd and June 8th, where the precipitation depths were 11 mm and 12 mm, respectively. Furthermore, we collected suspended particles samples with a suspended particle sampler in three of the study streams (Sites 3, 6 and 7) (Laubel et al., 2001).

The selection of analysed pesticides for the 2010 field campaign was based on application frequency and total applied amounts in 2009 (Danish EPA, 2010). This list was augmented with a series of banned pesticides that have been previously found in groundwater monitoring surveys

(Malaguerra et al., 2012). In total, the water samples were analysed for 27 different herbicides (Table 2). The selection of analysed pesticides for the water samples collected in 2011 was further augmented with 4 fungicides and 4 insecticides. Moreover, the suspended sediment samples additionally collected in 2011 were analysed for a selected list of particularly lipophilic insecticides, fungicides and herbicides (Table 2).

Pesticide analyses for the event-triggered samplers and grab sampling were conducted by Eurofins Miljoe A/S Laboratories, and the suspended particle samples were analysed at the Swedish University of Agriculture (Uppsala, Sweden). Analyses of all the samples were based on solid phase extraction, and the final extracts were analysed by GC–MS or LC–MS. The minimum detection limit was 0.01 μg L^{-1} for all pesticide compounds in water samples and 1 ng g^{-1} for particles (dry weight).

2.2.3   Sampling for xenobiotics

Samples for benzene, toluene, ethylbenzene, m-/p- and o-xylene (BTEX), naphthalene, and the chlorinated solvents PCE, TCE, trans- and cis-1,2-DCE, 1,1-DCE, 1,1-DCA and 1,1,1-TCA, were collected at all 11 sampling locations at base-flow conditions. The sampling locations at the contaminated sites (sites 7 - 11, Fig. 1) were placed in the expected groundwater to surface water discharge zones for the contaminated site, as determined by additional up-gradient and down-gradient sampling (data not shown). Samples were collected in 40 mL glass vials and immediately preserved using 4 M H_2SO_4 and stored at 4 °C. The analytes were separated and identified by GC/MS using an Agilent 7980 gas chromatograph system equipped with an Agilent 5975 C electron impact (70 eV) triple-axis mass-selective detector. Detection and quantification limits were determined as described by Winslow et al. (2006), and are given in McKnight et al. (2012).

2.3 Hydromorphology

A habitat survey was conducted on a 50 metre reach in each of the study streams spanning the sites for macroinvertebrate sampling and physicochemical measurements. The characterisation of the physical stream dimensions and habitat properties was performed according to the Danish Habitat Quality Index (DHQI) (Pedersen et al., 2006). The DHQI ranks the physical habitat quality from -12 to 63 using a series of hydromorphological and riparian parameters, and the threshold level for good physical conditions is 26 (Dunbar et al., 2010a). Discharge was quantified using four current velocity measurements (using a Höentzsch µP-TAD flow-meter) across each of five transects.

2.4 Macroinvertebrate sampling and metrics

Benthic macroinvertebrates were sampled in May 2010 in all study streams, and again in early May and late June in 2011 in a subset of the study streams (sites 2, 3, 5, 6 and 7). Moreover, macroinvertebrates were sampled at two control sites (C1 and C2) in early May and late June, 2011. All macroinvertebrate sampling was conducted using a standardized kick-sampling procedure (25 cm x 25 cm net with 0.5 mm mesh size) (Skriver et al., 2000). The samples were preserved in 96% ethanol in the field. Macroinvertebrates were identified to the species or genus level with only few exceptions (Oigochaeta (order), Chironomidae (sub-family), Ostracoda (order), Heteroptera (family) and Simuliidae (family)) (see Supplementary Material, Tables C1-C4, for the complete species lists).

We used a series of macroinvertebrate community descriptors for the interpretation of response in the macroinvertebrate communities, including total species richness, total macroinvertebrate abundance, species richness and densities of Ephemeroptera, Plecoptera and Trichoptera (EPT) and Simpson diversity. Moreover, we applied the Danish Stream Fauna Index (DSFI; currently the only ecological quality assessment method for running waters in Denmark), which is directed at detecting the effects of organic pollution (e.g. high BOD_5 concentrations or high concentrations of

macro-nutrients, especially ammonia-N) (Skriver et al., 2000). The Lotic Invertebrate index for

Flow Evaluation (LIFE) was used to relate the macroinvertebrate community to antecedent flow

characteristics of each site (Extence et al., 1999). Each taxon is assigned to a flow group (ranging

from I to VI) indicating its preference for higher current velocities and clean coarse substrate types.

The LIFE score for one sample is calculated as the mean flow score for all taxa scored in the

sample. The SPEcies At Risk index was used to relate the macroinvertebrate community to the toxic

loads from pesticides. The SPEAR index aims at detecting impacts of diffuse source pesticide

pollution by applying physiological and ecological traits (Liess and von der Ohe, 2005).

2.5 Data treatment

We applied toxic units (TU) as a measure for the toxicity of xenobiotic compounds and

pesticides to benthic macroinvertebrates (using *Daphnia magna* as a surrogate species). The TU

was calculated for all compounds according to Tomlin (2000):

$$\log \text{TU}_{D.magna} = \log (C_i / LC50_i) \tag{1}$$

where $\log \text{TU}_{D.magna}$ is the log toxic unit for compound i, C_i is the measured concentration of

compound i and $LC50_i$ is the corresponding 48 h LC50 value for *Daphnia magna* exposed to

compound i. The log maximum $\text{TU}_{D.magna}$ (log max $\text{TU}_{D.magna}$) and the log summed $\text{TU}_{D.magna}$ (log

sum $\text{TU}_{D.magna}$) were calculated for all samples (Supplementary Material, Tables B.3-B.5). The

summation of all TUs is based on the principle of toxic additivity, i.e. an increase in the number of

mixture components has been suggested to lead to a decrease in the range of deviation from toxic

additivity (the Funnel hypothesis) (Warne and Hawker, 1995). For log max $\text{TU}_{D.magna}$ the suggested

threshold for effects of pesticides in the field is at least -3 (Schäfer et al., 2012b).

For interpretation of the potential toxicity of the pesticides detected in the suspended particle

sampler, the log max $\text{TU}_{D.magna}$ and the log sum $\text{TU}_{D.magna}$ were calculated. For these calculations, it

is thought to be necessary to account for the potential reduction in toxicity that is a result of the

sorption to (especially organic) micro-particles. The reduction in toxicity for pesticides that are sorbed to particles is proposed to be within the range of 100 to 600 (e.g. Ding et al., 2011; Maund et al., 2002; Yang et al., 2006b). We therefore calculated the log max $TU_{D.magna}$ and the log sum $TU_{D.magna}$ using this range for safety factors (Supplementary Material, Table B.6).

We derived eight response variables from the macroinvertebrate community data; taxonomic richness, total macroinvertebrate abundance, EPT abundance, Simpson diversity, DSFI scores, LIFE scores and %SPEAR$_{pesticides}$ abundance. Moreover, a Detrended Correspondence Analysis (DCA) of the macroinvertebrate community composition abundance data was used to reduce the species abundance matrix of the aquatic macroinvertebrate community from the 11 sites to two orthogonal axes for use as additional response variables in later analyses. We down-weighted rare species and removed all taxa that only occurred in one sample in order to improve the signal of the most overall trends. We performed Pearson correlations between the DCA axis scores and the macroinvertebrate metrics for each sample in order to evaluate the amount of variability in the macroinvertebrate communities that was explained by the used macroinvertebrate metrics. The EPT abundance and the LIFE scores were log transformed before analysis in order to meet assumptions of normality. The DCA analysis was performed in PC-ORD 6.0, and the Pearson correlations were performed in SAS Enterprise Guide 4.3.

While the sub-catchments were selected so that they each ideally contained one particular type of anthropogenic stress (conventional agriculture, urban settlements and contaminated sites), the results from the DCA and a cluster analysis (performed in PC-ORD 6.0, data not shown) showed that the macroinvertebrate community structure did not group according to the type of anthropogenic stress in the sub-catchment. In the further statistical analyses we therefore treated the study sites as one group.

A Principal Component Analysis (PCA) was performed to reduce 30 environmental parameters from each site to three orthogonal axes. This was done because the stream environment is a product of all environmental parameters acting in concert, and it is therefore more meaningful to provide an integrated measure of them all. Pairwise co-linearity ($r > 0.9$) was assessed prior to the analysis in order to remove correlated parameters, and substrate characteristics were collapsed into one group (proportion of soft substrate; consisting of sand and silt). Thus, the PCA was performed on 21 environmental variables. In order to interpret statistically significant contributions ($\alpha < 0.05$) of the different measured environmental parameters to the three PCA axes, we performed a Spearman rank correlation on PCA axis scores and environmental variables. The Spearman rank test was preferred since the majority of environmental data did not meet the criteria for normality, and common data transformations (log, inverse, square root and inverse square root) did not provide normally distributed data. The PCA analysis was performed in PC-ORD 6.0, and the subsequent Spearman rank test was performed in SAS Enterprise Guide 4.3. Moreover, we performed a Spearman rank correlation analysis on the macroinvertebrate metrics (including DCA axis scores for the first two axes of the DCA) and the set of environmental variables (including PCA axis scores for the first three axes of the PCA) in SAS Enterprise Guide 4.3.

An additional (multivariate) interpretation of the macroinvertebrate taxa and their relationship to the measured environmental variables was conducted using a Canonical Correspondence Analysis (CCA) in PC-ORD 6.0. The CCA analysis was chosen for analysing the combined species-environment dataset, since we aimed to determine whether some proportion of the community structure is more strongly related to one environmental variable than expected by chance (ter Braak and Prentice, 1988). Since the interpretation of results from a CCA becomes increasingly dubious when the number of environmental variables increases relative to the number of observations (here, $n = 11$), we only included the axis scores for the three PCA axes as environmental variables in this

analysis. This is meaningful because the PCA axes incorporate some proportion of all measured environmental variables. We used a Monte Carlo test (999 permutations) for the CCA to test for linear relationships between the two matrices ($\alpha = 0.05$).

We compared the relation between the log $mTU_{D.magna}$ for water and suspended particle samples taken in 2011 and the %SPEAR$_{pesticides}$ abundance with the relation obtained in a previous study in northern Germany (for stream sites without upstream forested sections) with similar geological and climatic conditions (Liess and von der Ohe, 2005). We performed this comparison by calculating the differences in Euclidean distances (difference between measured and predicted %SPEAR$_{pesticides}$ abundance using the regression line from the study of Liess and von der Ohe, 2005 as a prediction tool) for the log mTUs obtained from water samples and suspended particle samples. The average Euclidean distances using water samples and suspended particle samples were subsequently compared with a t-test ($\alpha = 0.05$), and the tested data fulfilled the requirements for normality and equal variances ($\alpha = 0.05$).

The control sites were not included in any of the statistical analyses, as the control sites and the study sites constitute two clusters of sites that have only a very limited overlap in terms of hydromorphological properties and water chemistry. Therefore, these clusters mainly constitute the two extremes on the summed environmental gradients and thus do not provide much information on biotic responses in between the environmental extremes. We therefore only interpreted the more restricted summed environmental gradients among the study sites, and the control sites were mainly included as evidence for an existing colonisation pool of sensitive species in the region.

3. Results

3.1 Characterisation of environmental stressors

In general, the study streams were characterised by being channelised and with grass dominating riparian vegetation, poor habitat conditions with the dominating substrate type being silt (73%) and sand (11%) and the DHQI score showing generally bad physical conditions (average DHQI = 9.8) (Table 3). The streams were characterised by moderate to high organic pollution ($BOD_5 > 2$ mg L^{-1}) detected at least once in each of the study streams in June and August, 2010. Moreover, oxygen concentrations were often low (< 5-6 mg L^{-1}), and ammonium and total P concentrations were moderate to high (Table3).

The results of the field campaigns in 2010 and 2011 disclosed a total of 21 herbicides, 5 fungicides and 2 insecticides (Table 4). Twenty of the herbicides and three fungicides were detected in stream water, whereas two herbicides, two fungicides and two insecticides were detected in the suspended particle samplers (n = 3). Presently banned pesticides were detected in storm-flow water as well as base-flow water, whereas currently used pesticides were mainly detected in storm-flow water and suspended sediment samples. In total, 7 of the 33 EU priority pollutants were detected, including atrazine, diuron, isoproturon, simazine, hexachlorobenzene, chlorpyriphos and gamma-hexachlorocyclohexane (lindane). The number of pesticides found per stream site ranged from 5 to 14 for storm-flow water, and from 1 to 10 for base-flow water. Notably, 4 to 5 herbicides were also detected at the control sites, sampled in 2011.

Converting pesticide concentrations to TU (Supplementary Material, Tables B.1-3), we found that the obtained log $mTU_{D.magna}$ were comparatively similar during storm-flow and base-flow in 2010 (water samples only analysed for herbicides), whereas the log $mTU_{D.magna}$ were several orders of magnitude higher when the insecticide chlorpyrifos was detected in the particle sampler (Supplementary Material, Table B.4). In fact, all log $mTU_{D.magna}$ values for the particle samples were above -3, regardless of the factor value applied.

The first axis of the Principal Component Analysis (PCA) of the measured physical and chemical variables explained 32 % of the variation in the dataset (n = 11) (Fig. 2) and was significantly correlated with log mTU$_{D.magna}$ (based on base-flow measurements), DHQI scores, proportion of soft substrate and total coverage of macrophytes (Supplementary Material, Table D.2). Moreover, PCA axis 1 was strongly correlated with a series of micro-nutrient concentrations (Mn, Ca, K, Na and Cl). The PCA axis 2 explained an additional 19% of the variation in the dataset, and this axis particularly reflected ammonium-N, oxygen and pH. The PCA axis 3 explained 15% of the variation in the dataset, and this axis was significantly correlated with log mTU$_{D.magna}$ (based on storm-flow measurements), total P and plant species richness. Thus, the three PCA axes cumulatively explained 66% of the variation in the dataset.
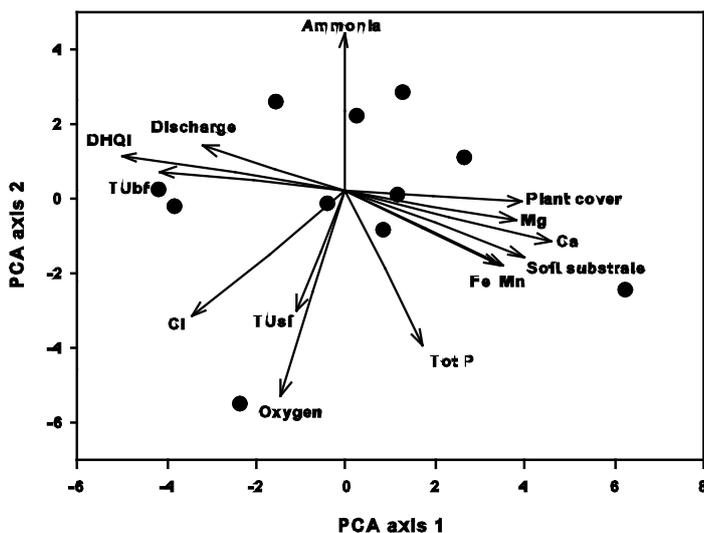
Figure 2: Principal Component Analysis (PCA) of measured environmental variables. Sites are represented as filled circles and the eigenvectors are presented for environmental variables that were significant components of the three PCA axes. However, data is only shown for PCA axis 1 and 2. PCA axes 1, 2 and 3 explained 32%, 19% and 15% of the variation in the dataset, respectively. TUsf and TUbf represent the log mTUD.magna based on storm flow and log sumTUD.magna based on base flow water samples, respectively.

3.2 Characterisation of the macroinvertebrate communities

The macroinvertebrate communities were generally characterised by few species and specimens of Ephemeroptera, Plecoptera and Trichoptera and all applied macroinvertebrate metrics (DSFI, LIFE and SPEAR) were characterised by relatively pronounced low scores (Table 5).

The first two axes of the macroinvertebrate community composition ordination (DCA) cumulatively explained 63% of the variability in the macroinvertebrate community composition, with DCA axis 1 explaining 52.5% and DCA axis 2 explaining an additional 10.6 % of the variability (Fig. 3). DCA axis 1 was significantly and negatively correlated to % SPEAR abundance (Fig. 4; $r = -0.86$, $P = 0.0008$), but there was no clear separation of SPEAR taxa from SPEnotAR taxa in the DCA ordination. The DCA axis 2 was significantly and positively correlated to total macroinvertebrate abundance (Fig. 4; $r = 0.70$, $P = 0.017$). Moreover, taxon richness and DSFI index scores were significantly correlated to DCA axis 2 scores ($r = 0.63$, $P = 0.036$ and $r = -0.58$, $P = 0.05$, respectively) (Supplementary Material, Table D.1).
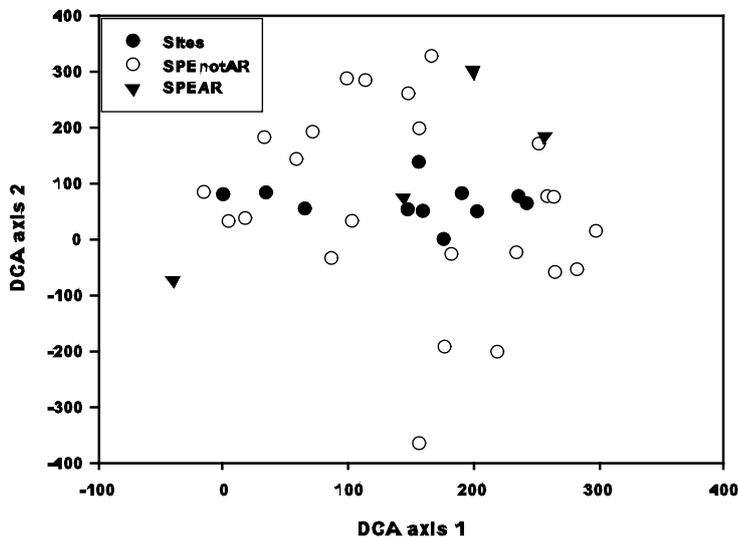
Figure 3: Detrended Correspondence Analysis (DCA) of community composition abundance data based on macroinvertebrate communities at the 11 sampling sites. Sites and macroinvertebrate community data are shown, and the taxa are additionally grouped into Species at Risk (SPEAR) and not at risk (SPEnotAR) for being affected by pesticide pollution. DCA axes 1 and 2 explained 55.2% and 10.6% of the total variation in the dataset, respectively.

3.3 Linking macroinvertebrate communities and environmental stressors

From the correlation matrix, presenting Spearman rank correlation coefficients on the correlation strengths between macroinvertebrate metrics and environmental parameters (Supplementary Material, Table D.3), it is seen that the flow-based LIFE index scores significantly decrease with increasing proportions of silt and macrophytes coverage, increasing axis scores of PCA 1 (primarily representing habitat quality with negative axis scores representing high habitat quality) and increasing concentrations of Mn. Moreover, LIFE index scores increased with increasing log sumTU$_{D.magna}$ for base-flow conditions, Nitrate-N and chloride concentrations. Increasing axis scores for PCA axis 2 (high values represent high concentrations of ammonium-N, low pH and low oxygen concentrations) correspond to significantly decreasing % SPEAR$_{pesticides}$ abundance. DCA axis 1 (primarily representing % SPEAR$_{pesticides}$ abundance) was not significantly correlated with any environmental variables. Axis scores for DCA axis 2 (high values indicate low DSFI score and high abundance and taxa richness) were negatively correlated with axis scores for PCA axis 3 (high values represent high log mTU$_{D.magna}$ during storm-flow, high total-P concentrations and low plant species richness).

In the Canonical Correspondence Analysis (CCA) of the macroinvertebrate species and axis scores of the three PCA axes representing all measured environmental variables, the total inertia was 1.328 and the proportion of variability explained was 20.0 % for CCA axis 1 and 8.4 % for CCA axis 2 (Fig. 5). The eigenvalue of the first axis was 0.265, and for the second axis 0.112 (Table 6). Several taxa that do not have strong requirements for well-mixed water and high current velocities are clustered to the right of the CCA ordination. In contrast, most of the taxa in the dataset that have specific requirements for moving and well-mixed water (*Baetis rhodani*, Limoniidae (e.g. Eriopterinae and *Dicranota* sp.) and to some extent *Gammarus pulex*) are clustered to the left of the ordination. This clustering is also reflected by eigenvectors for the strongest

environmental variables where the right part of the ordination is positively correlated with PCA axis 1 (reflecting high proportions of soft substrate and plant coverage) and PCA axis 2 (reflecting low oxygen concentrations). We cannot, however, reject the hypothesis of no relationship between the macroinvertebrate community matrix and the environmental variables matrix (P = 0.448; Table 6).

The two different sampling techniques for pesticides, used in 2011 (event-triggered water sampling and suspended particle samplers), clearly provided different results in terms of estimated toxicity to benthic macroinvertebrates (log $TU_{D.magna}$) (Fig. 6). In order to determine which sampling technique provides the better estimate for toxic impact on the benthic macroinvertebrates (%$SPEAR_{pesticides}$ abundance), we compared the average Euclidean distance from measured data points to predicted data points (the relation between log $mTU_{D.magna}$ and %$SPEAR_{pesticides}$ abundance previously established for streams without upstream forested sections in northern Germany (Liess and von der Ohe, 2005). The average Euclidean distance between predicted and measured data points was significantly higher when using the log $mTU_{D.magna}$ based on event-triggered water samplers compared to the log $mTU_{D.magna}$ based on suspended particle samplers (df = 6, t = 5.06, P = 0.002). In other words, the high log $mTU_{D.magna}$ that was thought to cause the low %$SPEAR_{pesticides}$ abundance was detected only when using suspended particle samplers and not when using event-triggered water samplers.
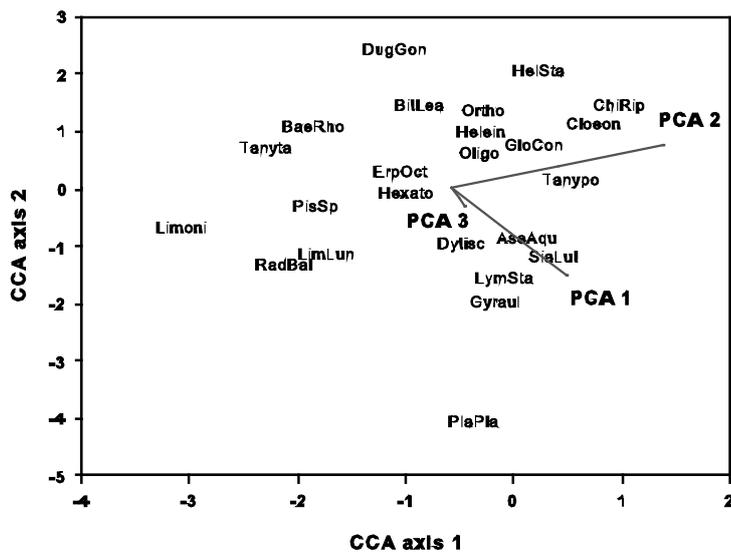
Figure 5: CCA with macroinvertebrates on the Hove catchment streams as dependent variables (indicated with taxa abbreviations) and environmental factors (represented by PCA axis scores) as independent variables, using LC scores. Eigenvectors (in red) depict the correlation between the environmental factors and the community matrix. The lengths of the vectors represent correlation strengths, and the vectors are extended by a factor 5 in order to promote the visibility. Only taxa occurring in more than one sample are presented in the ordination. AseAqu = *Asellus aquaticus*, BaeRho = *Baet is rhodani*, *ChiRip = Chironomus riparius*, Cloeon = *Cloeon dipterum*, Dytisc = Dytiscidae, ErpOct = *Erpobdella octaculata*, GamPul = *Gammarus pulex*, GloCon = *Glossiphonia concolor*, Gyraul = *Gyraulus* sp., Helein = Heleinae, HelSta = *Helobdella stagnalis*, Hexato = Hexatominae, LimLun = *Limnephilus lunatus*, Limoni = Limoniidae, LymSta = *Lymnaea* stagnalis, Oligo = Oligochaeta, Ortho = Orchocladinae, PisSp = *Pissidium* sp., PlaPla = *Planorbis planorbis*, RadBal = *Radix baltica*, SiaLut = *Sialis lutaria*, Simul = Simuliidae, SphaSp = *Sphaerium* sp., Tanypo = Tanypodinae, Tanyta = Tanytarsini.

Figure 6: The % SPEAR$_{pesticides}$ abundance as a function of log max TU$_{D.magna}$ for pesticides. The solid line indicates the relation for sites without upstream forested stream sections in Liess and von der Ohe (2005). The da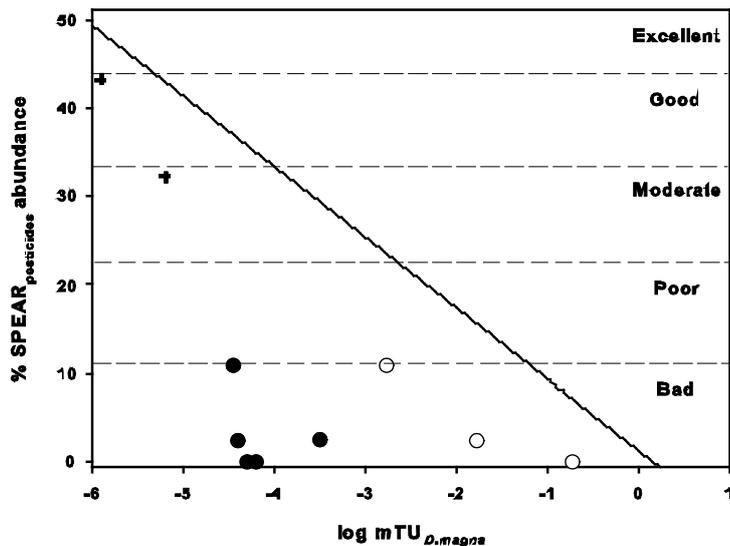shed horizontal lines indicate water quality categories based on the SPEAR index (Schletterer et al. 2010). All SPEAR values for the actual data points were collected in June, 2011. Crosses represent control sites without agriculture in the catchment (sites C1 and C2) and circles represent streams with multiple anthropogenic impacts. Filled circles represent data from storm flow water samples (n = 5), and open circles represent data from suspended sediment samplers (n = 3). The log mTU$_{D.magna}$ for suspended particles were calculated using LC50 values for 48 h acute mortality tests that were reduced by two orders of magnitudes (see supplementary material G, Table G4).

4. Discussion

4.1 Characterisation of environmental stressors

The headwater streams studied in the Hove catchment were heavily impacted by multiple anthropogenic sources of stress as shown by traditional water quality parameters (BOD$_5$, oxygen and ammonia-N), hydromorphological characteristics (flow conditions and physical habitat quality) and the high occurrence of pesticides (including EU priority pollutants). These indicators demonstrate that the study area has a worse condition than other low-order Danish streams previously studied (e.g. Dunbar et al., 2010a; Friberg et al., 2009; Rasmussen et al., 2012). Also, the total anthropogenic impact is comprised of a multitude of constituents; all being consequences of the human activities and intensive land-use that characterises the Hove catchment. Hence, the

biological communities must be interpreted in the context of the total amount of stress that is imposed to the system.

A detailed review of the chemical data collected in the 11 study streams in May-June and August, 2010, show that all study sites were characterised by at least one environmental parameter exceeding threshold values where sensitive macroinvertebrate species would be expected to disappear. In particular, BOD_5 (> 2 mg L^{-1}), oxygen concentrations ($< 5\text{-}6$ mg L^{-1}) and ammonium-N concentrations (> 0.2 mg L^{-1}) were often measured at critical levels which are likely caused by anthropogenic sources such as agriculture, untreated wastewater from scattered dwellings (Friberg et al., 2010) and discharge from landfills (Milosevic et al., 2012). Moreover, nitrate-N, phosphate-P and potassium were often found at high concentrations documenting the heavy use of agricultural fertilisers for crops (Table 3).

Twenty-one different herbicides were detected in stream water with concentrations up to 3.4 µg L^{-1} for the commonly applied MCPA (Danish EPA, 2010). Interestingly, the log sumTU$_{D.magna}$ for herbicides obtained during base-flow and storm flow conditions were within one order of magnitude of each other in all but one case (Supplementary Material, Tables B.1-3), suggesting that chronic exposure due to the inflow of pesticide contaminated groundwater is potentially an important stressor. However, the herbicides are probably not significantly impacting resident fauna (as discussed below). Importantly, however, several of the detected herbicides are characterised as EU priority pollutants e.g. atrazine, diuron, isoproturon and simazine. Toluene and TCE (pollutants from contaminated sites) were only detected at three sites, and conversion of concentrations to toxic units (for *D. magna*) suggested that sites contaminated with these compounds were probably an unimportant stressor for the benthic fauna. This finding is in accordance with McKnight et al. (2012), who studied a single stream in a nearby catchment on Sjaelland.

Hydro-morphological conditions were generally severely degraded and dominated by low base-flow conditions and high proportions of soft substrate (often 100%). Several of the study streams were heavily channelised and managed, stream profiles were often > 1 m below the soil surface and the riparian vegetation was dominated by grass characterising frequent maintenance. These factors translated into very low physical habitat quality scores using the Danish Habitat Quality Index (Table 3), and are also likely to have a significant impact on the stream biota through e.g. increased sediment transport, loss of essential habitat types, increased habitat instability and the occurrence of low flow periods causing critically high temperatures and low oxygen concentrations.

4.2 Characterisation of macroinvertebrate communities

We found that all applied macroinvertebrate community measures indicated heavily impacted ecological status, and the SPEAR index most clearly described the variability within the macroinvertebrate communities which may be partly explained by the selection of biological traits that forms the basis of the SPEAR index. The frequent occurrence of anthropogenic disturbances favours species that have high tolerance for the respective types of disturbance and, furthermore, short life cycles and high migration abilities (enabling high recovery potential), and since SPEAR considers these biological traits (and other), this may be partly explain its higher explanatory power.

The %SPEAR$_{pesticides}$ abundance attained values below 21 for all sites (Fig. 4), which categorises them as having *poor* to *bad* ecological status according to Schletterer et al. (2010). For six of the sites, the %SPEAR$_{pesticides}$ abundance was even below 10 suggesting a very strong impact of pesticides on the macroinvertebrate communities. Among the previous studies reporting SPEAR values from 112 different streams from different locations in the world (primarily headwater streams), less than 10% of the sites were characterised by %SPEAR$_{pesticides}$ values below 10 (compare McKnight et al., 2012; Rasmussen et al., 2012; Schletterer et al., 2010; Schäfer et al., 2007; 2011; von der Ohe et al., 2007). Despite much weaker correlations with the DCA axis scores,

other qualitative indicators of ecological quality (EPT abundance, DSFI and LIFE scores) all obtained very low values indicating strong ecological impairment in the studied headwater streams. High dispersal abilities, polyvoltinism and low specificity of habitat requirements are all essential traits required if a species is to survive in a degraded and frequently disturbed site situated in a degraded landscape (Doledec et al., 1999; Gerisch et al., 2012; Statzner & Bêche, 2010). The SPEAR index considers the species physiological sensitivity to pesticide pollution, length of life cycles, migration ability and timing of terrestrial phases in its assessment of ecological quality. The DSFI index only uses species oxygen requirements in its characterisation of sensitive species, whereas the LIFE index is based on species preferences for flow and habitat types. Since the traits used in SPEAR (except the physiological sensitivity to pesticides) additionally are central for species' abilities to cope with high levels of recurring anthropogenic stress, this may partly explain why SPEAR performed best in the overall characterisation of the macroinvertebrate communities in multi-stressor environments. This may furthermore explain why there was no clear clustering of SPEAR species in the DCA (Fig. 3).

The control streams were characterised by a macroinvertebrate community structure indicative of higher ecological quality (Supplementary Material, Table C.4). Different measures of ecological quality provided similar information; indicating that the control streams were characterised by a fauna with higher ecological index scores (DSFI, LIFE and SPEAR) and abundance of EPT taxa. Moreover, several of the EPT taxa present in the control sites are characterised by semi- or univoltinism, such as *Isoperla grammatica* and all the caddis flies, indicating environmental conditions that are characterised by much lower levels of anthropogenic disturbance (Nilsson, 2005). The control sites therefore demonstrate that the regional species pool from which new colonisers can disperse from is relatively diverse – but may also be relatively restricted in space due to high land-use intensity in the catchment.
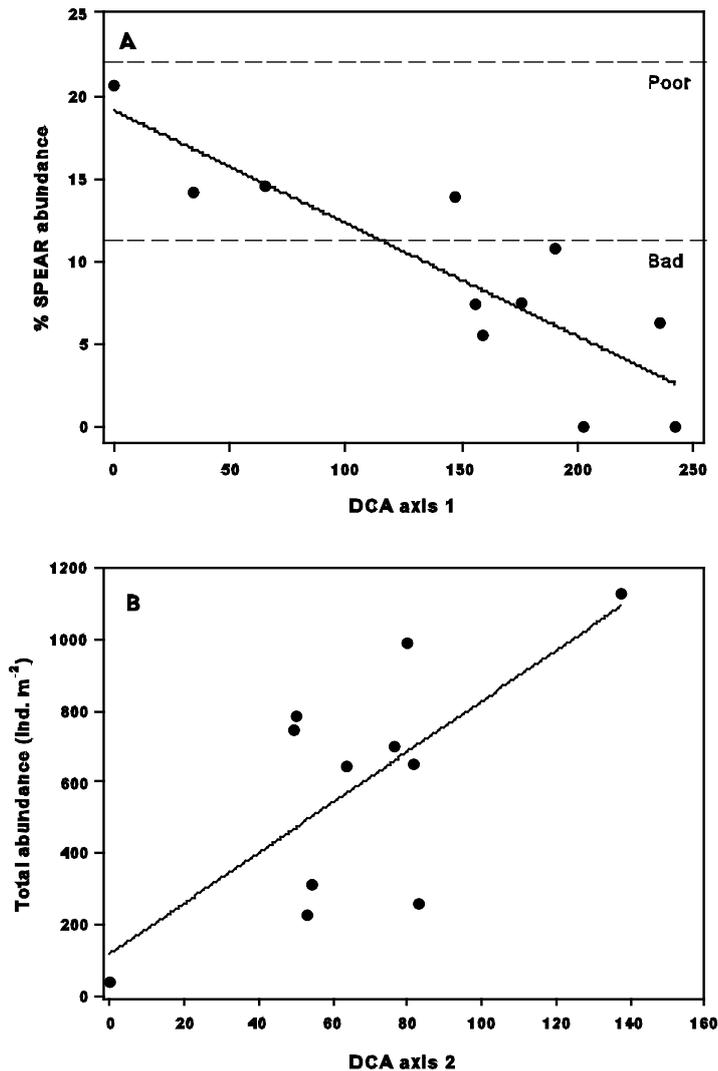
Figure 4: The abundance of species at risk for pesticide pollution (SPEAR) as a function of axis 1 sample scores from a community composition ordination (detrended correspondance analysis; DCA) (A) and total abundance of macroinvertebrates as a function of axis 2 sample scores from the community composition DCA (B). Dashed lines indicate water quality categories based on the SPEAR index (Schletterer et al., 2010).

4.3 Interactions between macroinvertebrate species and their environment

It was not possible to document the proportional structuring role of the measured environmental variables on the macroinvertebrate communities using a CCA or multi-correlation analyses which may be partly owed to the limited sampling size. Moreover, several of the measured stressors frequently exceeded thresholds for effects on the macroinvertebrate fauna entailing an increased

need for a larger sample size in order to disentangle the effects of specific stressors. Targeted mitigation efforts on single anthropogenic stressors in the catchment are therefore unlikely to have substantial effects on the ecological quality in these streams, and more holistic approaches should be preferred.

Another reason for the unsuccessful attempt to estimate the proportional importance of the dominating stressors in the catchment may be that the potential impact of pesticides was underestimated due to inadequate quality of the pesticide data, which for the 2010 field campaign only included herbicides that are rarely found to be acutely toxic to benthic macroinvertebrates in the field (Schulz, 2004). Since the very low % SPEAR$_{pesticides}$ abundances that characterised the study streams (Fig. 6) suggest that there was a significant impact of pesticides, we re-sampled a subset of the streams in 2011 in order to provide sufficient quality of pesticide data (including fungicides and insecticides) to further test the role of agricultural pesticides on the structure of macroinvertebrate communities. These results are discussed in the next section.

A multi-correlation evaluation of the relations between the eight macroinvertebrate community descriptors and the environmental variables highlighted several significant correlations, but only a limited number of the significant correlations were intuitively meaningful. For example, the LIFE score increased significantly with increasing toxicity of pesticides originating from groundwater inflow, total nitrogen concentration (primarily from nitrate-N) and chloride concentration. High LIFE scores are associated with communities that are adapted to higher current velocity and oxygen concentrations and increasing proportions of hard and clean substrate. In general, the LIFE scores were low compared to previous findings (e.g. Dunbar et al., 2010b), which is probably due to the fact that discharge was low in most of the study streams and several of the streams were characterised by low oxygen concentrations and dominated by soft substrate types, as confirmed in the multi-correlation analysis. Since the actual gradient in the hydromorphological parameters that

govern the LIFE score outputs was strongly skewed (sites with poor hydromorphological properties clearly dominating), we suggest that the LIFE score results should be interpreted with care, and we therefore also suggest that the positive correlations between the LIFE scores and pesticide toxicity (from groundwater inflow), nitrogen-N concentrations and chloride concentrations are Type II errors.

4.4 Effects of diffuse source pesticides on macroinvertebrate communities

Our findings indicate that (i) despite a potential reduction in bioavailability and ecotoxicity for sorbed pesticides, they may still significantly impact benthic macroinvertebrates, and (ii) suspended particle samples (SPS) add essential information concerning the antecedent occurrence of pesticides in the streams, which should be considered with the data from event-triggered water samples.

The second sampling campaign in 2011, conducted in a subset of the streams, revealed several fungicides in the stream water during storm-flow. The insecticide chlorpyrifos was only detected in the SPSs. These findings are perhaps not surprising, since chlorpyrifos has a high log K_{ow} (4.7) indicating that this compound is prone to be transported from fields to the adjacent streams as sorption complexes with organic microparticles (Liess et al., 1996). Chemical toxicity is thought to decrease when pesticides are sorbed to solid microparticles, as they become less bioavailable to non-target organisms (e.g. Yang et al., 2006a; You et al., 2008). In the calculations of toxicities of the SPS samples we therefore applied different adjustment factors (100 to 600) to compensate for the potentially reduced toxicities to non-target benthic macroinvertebrates, as suggested by the literature (see e.g. Maul et al., 2008; Maund et al., 2002).

Using the log $mTU_{D.magna}$ for the SPS samples (and 100 as adjustment factor), the relation between the toxic pressure from pesticides and %SPEAR$_{pesticides}$ abundance closely resembled previously published relations for German streams without upstream forested sections (Liess and von der Ohe, 2005). Notably, this relation was not seen when using the log $mTU_{D.magna}$ from event-

triggered water samples (Fig. 6), because chlorpyrifos (causing the high log $\text{mTU}_{D.magna}$) was only captured in the SPS samples. This clearly indicates the importance of the sampling technique used for the evaluation of pesticide effects in the field, which we recently highlighted in another study (McKnight et al., 2012).

The sorption of pesticides to particles often increases their half-lives, and the sedimentation and re-suspension of particle-bound pesticides are mechanisms that keep the pesticides in the stream ecosystems for a longer time. Moreover, organic microparticles constitute food resources and habitats for many benthic macroinvertebrates, and the sorption of pesticides to these ensures that the pesticides remain in close contact with the non-target benthic macroinvertebrates for longer periods of time. In fact, several pyrethroid insecticides have been detected in the bed sediment of streams in the United States at concentrations that are expected to impact sediment dwelling organisms (Kuivila et al., 2012). Similar comprehensive studies have not been conducted in Europe, but Feo et al. (2010) indicated that pyrethroid impacts may also occur in European streams.

The $\%\text{SPEAR}_{\text{pesticides}}$ abundance was again very low (< 10) for the five study streams that were re-sampled in 2011 (Fig. 6), and the detections of chlorpyrifos in SPSs seem to offer an explanation. Since the $\%\text{SPEAR}_{\text{pesticides}}$ abundance was low for all study streams, and since three out of three SPS samples confirmed the presence of insecticides, we cautiously conclude that pesticide pollution may be a major reason for the impaired ecological conditions in the headwater streams in the Hove catchment. Nonetheless, all data points were positioned below the previously published regression line for the $\%\text{SPEAR}_{\text{pesticides}}$ abundance as a function of log $\text{mTU}_{D.magna}$ (Fig. 6), indicating that the impact may be due to more than just the direct effects of pesticides on benthic macroinvertebrates. Rasmussen et al. (2012) showed that impaired physical habitat conditions (high proportions of soft substrate and low habitat heterogeneity) increases the effects of pesticides (decreasing $\%\text{SPEAR}_{\text{pesticide}}$ abundance). Rasmussen et al. (2012) argued that the majority of SPEAR species

had specific habitat preferences for hard substrate types and were characterised by long life cycles which are traits characteristics that strongly act against presence in a highly disturbed environment. Since soft substrate types were clearly dominating in the study streams, and in general were severely disturbed, the very low %SPEAR$_{pesticide}$ abundances may be partly due to the generally impoverished habitat conditions and frequent disturbance from dredging and sediment re-mobilisation. Our results infer that there may be a need for the development of an alternative pesticide index for physically impaired streams dominated by soft substrate types or an improved measure for the toxicity of pesticides adsorbed to microparticles. In contradiction to the conventional thought that sorption reduces pesticide toxicity by several orders of magnitude (Hill, 1989), Schulz and Liess (2001a; 2001b) showed that the adsorption of pyrethroids to suspended particles during acute exposure may not reduce the toxicity of pyrethroids to macroinvertebrates by more than a factor 10.

Conclusions

In this study, we showed that headwater streams in the Hove catchment are strongly impacted by numerous types of anthropogenic activities causing severe hydromorphological and chemical impairment as well as severe impairment of the macroinvertebrate communities. However, probably due to the strong overlap in the impairing side-effects of the many anthropogenic activities and due to limited sample size, we were not successfully able to distinguish between effects of specific anthropogenic stressors (i.e. contaminated sites, urban settlements and diffuse source agricultural pollution), and neither to rank-order their specific importance for the observed ecological impairment. This means, it is unlikely possible to gain significant positive effects of mitigation actions directed for single stressors only in this catchment, and more holistic mitigation approaches should be preferred. Many anthropogenic stressors at least partly forge a selection process for

similar trait characteristics within species (e.g. episodic pesticide pollution and physical impairment (large fractions of soft substrate types) both increases selection for short life cycles and high migration capacity). This forms an ambiguous problem in terms of disentangling the ecological effects of individual stressors that calls for further research.

Our results show that it is important that headwater streams be considered in River Basin Management Plans and other management and mitigation efforts conducted at the catchment level. This is needed, since many headwater streams are impacted by numerous anthropogenic activities causing the severe impairment documented in this study. Furthermore, headwater streams are essential to the dispersal of species between streams and river systems, and they have the potential to provide high biodiversity value to the systems due to their potentially unique habitat characteristics.

Acknowledgements

References

Allan JD. Landscapes and riverscapes: The influence of land use on stream ecosystems. Annu Rev Ecol Evol S 2004; 35: 257-284.

Beketov M, Liess M. Ecotoxicology and macroecology - Time for integration. Environ Pollut 2012; 162: 247-254.

Danish EPA. Pesticide statistics 2009 [In Danish]. Danish EPA, Ministry of Envioronment and Energy, 2010.

Ding Y, Weston DP, You J, Rothert AK, Lydy MJ. Toxicity of Sediment-Associated Pesticides to *Chironomus dilutus* and *Hyalella azteca*. Arc Environ Con Tox 2011; 61: 83-92.

Dodds WK, Oakes RM. Headwater influences on downstream water quality. Environ Manage 2008; 41: 367-377.

Doledec S, Statzner B, Bournard M. Species traits for future biomonitoring across ecoregions: patterns along a human-impacted river. Freshwater Biol 1999; 42: 737-758.

Dunbar MJ, Pedersen ML, Cadman D, Extence C, Waddingham J, Chadd R, et al. River discharge and local-scale physical habitat influence macroinvertebrate LIFE scores. Freshwater Biol 2010a; 55: 226-242.

Dunbar MJ, Warren M, Extence C, Baker L, Cadman D, Mould DJ, et al. Interaction between macroinvertebrates, discharge and physical habitat in upland rivers. Aquatic Conserv 2010b; 20: S31-S44.

Extence CA, Balbi DM, Chadd RP. River flow indexing using British benthic macroinvertebrates: A framework for setting hydroecological objectives. Regul River 1999; 15: 543-574.

Feo ML, Ginebreda A, Eljarrat E, Barcelo D. Presence of pyrethroid pesticides in water and sediments of Ebro River Delta. J Hydrol 2010; 393: 156-162.

Freeman MC, Pringle CM, Jackson CR. Hydrologic connectivity and the contribution of stream headwaters to ecological integrity at regional scales. J Am Water Resour As 2007; 43: 5-14.

Friberg N, Sandin L, Pedersen ML. Assessing the effects of hydromorphological degradation on macroinvertebrate indicators in rivers: Examples, constraints and outlook. Integr Environ Assess Manage 2009; 5: 86-96.

Friberg N, Skriver J, Larsen SE, Pedersen ML, Buffagni A. Stream macroninvertebrate occurrence along gradients in organic pollution and eutrophication. Freshwater Biol 2010; 55: 1405-1419.

Gerisch M, Agostinelli V, Henle K, Dziock F. More species, but all do the same: contrasting effects of flood disturbance on ground beetle functional and species diversity. Oikos 2012; 121: 508-515.

Hill IR. Aquatic organisms and pyrethroids. Pestic Sci 1989; 27: 429-465.

Kuivila KM, Hladik ML, Ingersoll CG, Kemble NE, Moran PW, Calhoun DL, et al. Occurrence and Potential Sources of Pyrethroid Insecticides in Stream Sediments from Seven U.S. Metropolitan Areas. Environ Sci Technol 2012; 46: 4297-4303.

Kürstein J, Andersen JA, Mahrt J. The Zealand model - an integrated model tool for the hydrological network (in Danish). Danish Hydrological Institute, 2009.

Larsen S, Ormerod SJ. Combined effects of habitat modification on trait composition and species nestedness in river invertebrates. Biol Conserv 2010; 143: 2638-2646.

Lassaletta L, Garcia-Gomez H, Gimeno BS, Rovira JV. Headwater streams: neglected ecosystems in the EU Water Framework Directive. Implications for nitrogen pollution control. Environ Sci Policy 2010; 13: 423-433.

Laubel A, Kronvang B, Fjordback C. Time-integrated suspended sediment sampling from a small lowland stream. Int Ver Theor Angew 2001; 28: 1420-1424.

Liess M, Schulz R, Neumann M. A method for monitoring pesticides bound to suspended particles in small streams. Chemosphere 1996; 32: 1963-1969.

Liess M, von der Ohe PC. Analyzing effects of pesticides on invertebrate communities in streams. Environ Toxicol Chem 2005; 24: 954-965.

Malaguerra F, Albrechtsen HJ, Thorling L, Binning PJ. Pesticides in water supply wells in Zealand, Denmark: A statistical analysis. Sci Tot Environ 2012; 414: 433-444.

Matson PA, Parton WJ, Power AG, Swift MJ. Agricultural intensification and ecosystem properties. Science 1997; 277: 504-509.

Matthaei CD, Piggott JJ, Townsend CR. Multiple stressors in agricultural streams: interactions among sediment addition, nutrient enrichment and water abstraction. J Appl Ecol 2010; 47: 639-649.

Maul JD, Brennan AA, Harwood AD, Lydy MJ. Effect of sediment-associated pyrethroids, fipronil, and metabolites on *Chironomus tentans* growthrate, body mass, condition index, immobilization and survival. Environ Toxicol Chem 2008; 27: 2582-2590.

Maund SJ, Hamer MJ, Lane MCG, Farrelly E, Rapley JH, Goggin UM, et al. Partitioning, bioavailability, and toxicity of the pyrethroid insecticide cypermethrin in sediments. Environ Toxicol Chem 2002; 21: 9-15.

McKnight US, Rasmussen JJ, Kronvang B, Bjerg PL, Binning PJ. Integrated assessment of the impact of chemical stressors on surface water ecosystems. Sci Tot Environ 2012; 427-428.

Meyer JL, Strayer DL, Wallace JB, Eggert SL, Helfman GS, Leonard NE. The contribution of headwater streams to biodiversity in river networks. J Am Water Resour As 2007; 43: 86-103.

Millenium Ecosystem Assessment. Ecosystems and human well-being: synthesis. Washington, DC: Island Press, 2005.

Milosevic N, Thomsen NI, Juhler RK, Alberechtsen H-J, Bjerg PL. Identification of discharge zones and quantification of contaminant mass discharges into a local stream from a landfill in a heterogeneous geologic setting. J Hydrol 2012; 446-447: 13-23.

Nilsson A. Aquatic insects of Northern Europe: A taxonomic handbook: Apollo Books, 2005.

Ormerod SJ, Dobson M, Hildrew AG, Townsend CR. Multiple stressors in freshwater ecosystems. Freshwater Biol 2010; 55: 1-4.

Pedersen ML. Effects of channelisation, riparian structure and catchment area on physical habitats in small lowland streams. Fund Appl Limnol 2009; 174: 89-99.

Pedersen ML, Friberg N. Influence of disturbance on habitats and biological communities in lowland streams. Fund Appl Limnol 2009; 174: 27-41.

Pedersen ML, Sode A, Kaarup P, Bundgaard P. Habitat quality in Danish streams. Testing of two indices and development of a National Physical Habitat Quality Index. Scientific Report. 590. National Environmental Research Institute, Silkeborg, Denmark, 2006.

Rasmussen JJ, Baattrup-Pedersen A, Larsen SE, Kronvang B. Local physical habitat quality cloud the effect of predicted pesticide runoff from agricultural land in Danish streams. J Environ Monitor 2011; 13: 943-950.

Rasmussen JJ, Wiberg-Larsen P, Baattrup-Pedersen A, Friberg N, Kronvang B. Physical structure of stream microhabitats influences the response of stream macroinvertebrate communities to pesticide stress. Environ Pollut 2012; 164: 142-149.

Schletterer M, Fureder L, Kuzovlev VV, Beketov MA. Testing the coherence of several macroinvertebrate indices and environmental factors in a large lowland river system (Volga River, Russia). Ecol Indic 2010; 10: 1083-1092.

Schulz R. Field studies on exposure, effects, and risk mitigation of aquatic nonpoint-source insecticide pollution: A review. J Environ Qual 2004; 33: 419-448.

Schulz R, Liess M. Acute and chronic effects of particle-associated fenvalerate on stream macroinvertebrates: A runoff simulation study using outdoor microcosms. Arch Environ Con Tox 2001a; 40: 481-488.

Schulz R, Liess M. Toxicity of aqueous-phase and suspended particle-associated fenvalerate: Chronic effects after pulse-dosed exposure of *Limnephilus lunatus* (Trichoptera). Environ Toxicol Chem 2001b; 20: 185-190.

Schäfer RB, Bundschuh M, Rouch DA, E. S, von der Ohe PC, Pettigrove V, et al. Effects of pesticide toxicity, salinity and other environmental variables on selected ecosystem functions in streams and the relevance for ecosystem services. Sci Tot Environ 2012a; 415: 69-78.

Schäfer RB, Caquet T, Siimes K, Mueller R, Lagadic L, Liess M. Effects of pesticides on community structure and ecosystem functions in agricultural streams of three biogeographical regions in Europe. Sci Tot Environ 2007; 382: 272-285.

Schäfer RB, Pettigrove V, Rose G, Allinson G, Wightwick A, von der Ohe PC, et al. Effects of pesticides monitored with three sampling methods in 24 sites on macroinvertebrates and microorganisms. Environ Sci Technol 2011; 45: 1665-1672.

Schäfer RB, von der Ohe PC, Rasmussen JJ, Kefford B, Beketov M, Schulz R, et al. Thresholds for the effects of pesticides on invertebrate communities and leaf breakdown in stream ecosystems. Environ Sci Technol 2012b; 46: 5134-5142.

Segner H. Moving beyond a descriptive aquatic toxicology: The value of biological process and trait information. Aquatic Toxicol 2011; 105: 50-55.

Skriver J, Friberg N, Kirkegaard J. Biological assessment of running waters in Denmark: introdution of the Danish Stream Fauna Index (DSFI). Ver Int Ver Limnol 2000; 27: 1822–1830.

Statzner B, Bêche LA. Can biological invertebrate traits resolve effects of multiple stressors on running water ecosystems? Freshwat Biol 2010; 55: 80-119.

Stoddard JL, Larsen DP, Hawkins CP, Johnson RK, Norris RH. Setting expectations for the ecological condition of streams: The concept of reference condition. Ecol Appl 2006; 16: 1267-1276.

ter Braak CJF, Prentice IC. A theory of gradient analysis. Adv Ecol Res 1988; 18: 271-317.

Tomlin CDS. The pesticide manual, a wold compendium. Farnham, Surrey, UK: Crop Protection Publications, 2000.

Townsend CR, Uhlmann SS, Matthaei CD. Individual and combined responses of stream ecosystems to multiple stressors. J Appl Ecol 2008; 45: 1810-1819.

von der Ohe PC, Prub A, Schafer RB, Liess M, de Deckere E, Brack W. Water quality indices across Europe - a comparison of the good ecological status of five river basins. J Environ Monitor 2007; 9: 970-978.

Warne MSJ, Hawker DW. The number of components in a mixture determines whether synergistic and antagonistic or additive toxicity predominate - the Funnel hypothesis. Ecotoxicol Environ Saf 1995; 31: 23-28.

Winslow SD, Martin JJ, Hallberg GR, Munch DJ, Freibis CP, al. e. Statistical procedures for the determination and verification of minimum reporting levels for drinking water methods. Environ Sci Technol 2006; 40: 281-288.

Yang W, Gan J, Hunter W, Spurlock F. Effect of suspended solids on bioavailability of pyrethroid insecticides. Environ Toxicol Chem 2006a; 25: 1585-1591.

Yang W, Spurlock F, Liu W, Gan J. Inhibition of aquatic toxicity of pyrethroid insecticides by suspended sediment. Environ Toxicol Chem 2006b; 25: 1913-1919.

You J, Pehkonen S, Weston DP, Lydy MJ. Chemical availability and sediment toxicity of pyrethroid insecticides to *Hyalella azteca*: Application to field sediment with unexpectedly low toxicity. Environ Toxicol Chem 2008; 27: 2124-2130.

Table 1. Percentages of different types of areal cover in the sub-catchments for each of the sampling sites. Natural area refers to forest and wet and dry meadows and other types of land use refer to grassland, uncultivated land and roads.

Parameter	Site 1	Site 2	Site3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	Site 11
Cultivated area in sub-catchment (%)	33	19	24	34	51	66	40	39	55	76	79
Urban area in sub-catchment (%)	26	22	8	2	3	3	5	3	4	3	1
Natural area in sub-catchment (%)	21	2	13	15	14	3	5	16	9	4	15
Other (%)	20	57	55	49	32	28	50	42	32	17	5

Table 2. List of pesticides included in the analysis packages for water samples in 2010 and 2011 and suspended particle samples in 2011. Capital letters indicate compound groups (H: herbicides, F: fungicides and I: insecticides).

Compounds screened	Water samples 2010	Water samples 2011	Suspended particles 2011
2,4-dichlorophenol (H)	X	X	
(2,4-D) 2,4-dichlorophenoxyacetic acid (H)	X	X	
2,6-dichlorophenol (H)	X	X	
4-chlor-2-methylphenol (H)	X	X	
4-chloro-2-methylphenol (H)	X	X	
(4-CPP) 2-(4-chlorphenoxy)propanoic acid (H)	X		
4-nitrophenol (H)	X	X	
Atrazine (H)	X	X	X
(BAM) 2,6-dichlorbenzamid (H)	X	X	
Bentazone (H)	X	X	X
Cyanazine (H)	X	X	
Desethylatrazine (H)	X	X	
Desisopropylatrazine (H)	X	X	
Dichlobenil (H)	X	X	
Dichlorprop (H)	X	X	
Diflufenican (H)	X	X	X
Dimethoat (H)	X	X	X
Dinoseb (H)	X	X	
Diuron (H)	X	X	X
(DNOC) 2-methyl-4,6-dinitrophenol (H)	X	X	
Hexazione (H)	X	X	
Hydroxyatrazine (H)	X	X	
Isoproturon (H)	X	X	X
(MCPA) (4-chloro-2-methylphenoxy)acetic acid (H)	X	X	
(MCPP) Mechlorprop (H)	X	X	
Metamitron (H)	X	X	
Pendimethalin (H)	X	X	X
Simazine (H)	X	X	
Terbutylazine (H)	X	X	X
(TCA) Trichloroacetic acid (H)	X		
Boscalid (F)		X	X
Epoxiconazole (F)		X	X
Fenpropimorf (F)			X
Hexachlorobenzene (F)			X
Propiconazole (F)		X	X
Pyraclostrobin (F)		X	X
Tebuconazole (F)		X	X
Chlorpyrifos (I)			X
Cypermethrin (I)		X	X
Deltamethrin (I)			X
(Lindane) Hexachlorocyclohexane(HCH)-gamma (I)			X

Lambda-cyhalothrin (I)		X
Pirimicarb (I)	X	X
Tau-fluvalinate (I)	X	X
Thiacloprid (I)	X	X

Table 3. Means, std. errors, minimum and maximum values of 27 environmental variables measured in the 11 study streams.

Variable	Mean	Std. Error	Minimum	Maximum
Water chemistry				
Ammonia-N (mg L^{-1})	0.155	0.038	0.054	0.438
Nitrate-N (mg L^{-1})	1.586	0.412	0.179	4.404
Total-N (mg L^{-1})	2.642	0.371	1.175	5.220
Phosphate-P (mg L^{-1})	0.079	0.019	0.005	0.195
Total-P (mg L^{-1})	0.355	0.131	0.047	1.359
Sulphate-S (mg L^{-1})	53.23	15.20	4.90	124.51
Cl (mg L^{-1})	87.97	17.33	29.35	207.61
Ca (mg L^{-1})	134.61	23.41	46.92	310.65
Fe (mg L^{-1})	0.965	0.775	0.062	8.691
Mn (mg L^{-1})	0.175	0.155	0.005	1.729
Mg (mg L^{-1})	11.21	2.07	3.35	24.15
Na (mg L^{-1})	69.34	14.45	25.42	187.87
K (mg L^{-1})	6.87	1.06	2.88	12.97
O_2 (mg L^{-1})	5.93	0.41	2.71	9.29
O_2 saturation (%)	58	4	27	91
BOD (mg L^{-1})	2.56	0.34	0.9	4.8
Physical parameters				
Conductivity (µS cm^{-})	735	60	467	1158
pH	7.32	0.08	6.99	8.05
Temperature (ºC)	15.3	0.5	11.6	16.7
Width (cm)	168	16	108	251
Depth (cm)	23	5	5	45
% boulder	5	3	0	38
% pebbles	7	4	0	42
% gravel	4	2	0	24
% sand	11	6	0	50
% silt	73	11	16	100
Discharge (L s^{-1})	19.5	8.7	0	83
Total plant cover (%)	34	10	1	100
Plant species richness	2.5	0.6	1	8
DHQI score	9.8	3.4	-3	29

Table 4. Overview of the herbicides (H), fungicides (F), insecticides (I) and xenobiotics detected in the Hove catchment streams, including minimum and maximum concentrations. Time of detection is indicated (August: grab samples during summer low flow; May and June: event triggered water samples; May-June (SP): Suspended particle samples). For suspended particle samples, the actual concentrations are given for sites 3, 6 and 7. EU priority pollutants are indicated with asterisks.

Compound (active ingredient)	Min concentration (μgL^{-1})	Max concentration (μgL^{-1})	Particle bound concentration (μg kg^{-1} dw)	Sampling event when detected
2,6-dichlorophenol (H)	0.011	0.064		Aug 2010
4-chlor-2-methylphenol (H)	0.085	0.085		May 2011
(4-CPP) 2-(4-chlorphenoxy)propanoic acid (H)	0.033	0.69		All water samples in 2010
Atrazine (H)*	0.015	0.015		Aug 2010
(BAM) 2,6-dichlorbenzamid (H)	0.023	1.7		All water samples
Bentazone (H)	0.012	0.012		Jun 2011
Desethylatrazine (H)	0.046	0.081		May & Aug 2010
Desisopropylatrazine (H)	0.022	0.022		May 2010
Dichlobenil (H)	0.010	0.051		Jun & Aug 2010; May & Jun 2011
Dichlorprop (H)	0.011	0.069		June & Aug 2010; Jun 2011
Diflufenican (H)			39; 0; 25	May-Jun 2011 (SP)
Diuron (H)*	0.012	0.053	0; 0; 140	Jun & Aug 2010; May & Jun 2011; May-Jun 2011 (SP)
(DNOC) 2-methyl-4,6-dinitrophenol (H)	0.011	0.31		All water samples
Hydroxyatrazine (H)	0.011	0.069		May, Jun & Aug 2010; Jun 2011
Isoproturon (H)*	0.011	0.23		All water samples in 2010
(MCPA) (4-chloro-2-methylphenoxy)acetic acid (H)	0.011	3.4		All water samples
(MCPP) Mechlorprop (H)	0.010	0.59		May, Jun & Aug 2010; Jun 2011
Metamitron (H)	0.063	0.92		May 2010
Simazine (H)*	0.010	0.11		Jun & Aug 2010
Terbutylazine (H)	0.010	0.15		Jun & Aug 2010; May & Jun 2011
(TCA) Trichloroacetic acid (H)	0.027	0.95		All water samples in 2010
Boscalid (F)	0.011	0.027		Jun 2011
Epoxiconazole (F)	0.055	0.055		Jun 2011
Fenpropimorf (F)			25; 0; 0	May-Jun 2011 (SP)
Hexachlorobenzene (F)*			20; 1; 2	May-Jun 2011 (SP)
Propiconazole (F)	0.036	0.082		Jun 2011
Chlorpyrifos (I)*			11; 0; 0	May-Jun 2011 (SP)
(Lindane) Hexachlorocyclohexane(HCH)-gamma (I)*			0; 3; 0	May-Jun 2011 (SP)
Toluene	1.0	5.1		Aug 2010
TCE	< 0.1	0.21		Aug 2010
Cis-1,2-DCE	<0.05	<0.05		Aug 2010

1   Table 5. Descriptive parameters for macroinvertebrate samples collected in May 2010.
2

Parameter	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	Site 11
Taxon richness	21	15	14	14	10	16	13	19	14	20	10
Tot. abund. (m^{-2})	1132	318	746	651	644	990	785	700	258	228	38
EPT taxon richness	1	2	1	2	0	2	0	2	2	1	1
EPT abund. (m^{-2})	51	13	7	11	0	20	0	2	5	25	5
Simpson diversity	0.26	0.31	0.32	0.44	0.62	0.42	0.31	0.54	0.31	0.13	0.23
DSFI	2	3	4	3	3	4	3	3	4	3	4
LIFE	5.64	7.09	6.36	5.7	5.75	6.36	5.8	6	5.88	5.73	5.71
SPEAR (%)	7.4	14.52	0	10.75	0	20.59	5.52	6.27	14.14	13.87	7.47

3 Table 6. Monte Carlo test results for eigenvalues and species-environment correlations based on
4 999 runs with randomised data. P-values are only given for the first CCA axis, since these are not
5 below the significance level ($\alpha = 0.05$).

Axis	Real data	Randomised data			P
		Mean	Minimum	Maximum	
Eigenvalue					
1	0.265	0.262	0.088	0.494	0.448
2	0.112	0.122	0.035	0.283	
3	0.108	0.051	0.014	0.126	
Spp-Envt Corr.					
1	0.808	0.812	0.576	0.980	0.531
2	0.800	0.728	0.465	0.960	
3	0.849	0.675	0.324	0.960	

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