Anionic Extraction for Efficient Recovery of Biobased 2,3-Butanediol-A Platform for Bulk and Fine Chemicals

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2,3-Butanediol (BDO) presents a promising platform molecule for the synthesis of basic and fine chemicals. Biotechnological production of BDO from renewable resources with living microbes enables high concentrations in the fermentation broth. The recovery of high-boiling BDO from an aqueous fermentation broth presents a subsequent challenge. A method is proposed for BDO isolation based on reversible complexation with phenylboronate in an anionic complex. BDO can be recovered by back-extraction into an acidic solution. The composition of the extracted species was determined by NMR spectroscopy, MS, and GC–MS methods. The conditions of extraction and back-extraction were optimized by using commercial BDO and finally applied to different fermentation broths. Up to 72–93% BDO can be extracted and up to 80–90% can be back-extracted under the optimized conditions. Purified bio-BDO was used in the presence of sulfuric acid for the synthesis of methyl ethyl ketone, an established organic solvent and discussed tailor-made biofuel.

**Introduction**

Biotechnologically manufactured 2,3-butanediol (BDO) was indicated as a promising platform chemical as long ago as the 1940s. Production of BDO was scaled up to pilot-plant volume by considering the diol as an intermediate for synthesis of 1,3-butadiene. Rapid development of refinery processes in the 1950s resulted in shutting down of biobased production of BDO owing to the availability of inexpensive fossil feedstocks. Later, with the aim of developing sustainable technologies, fermentative processes were revisited. Recent achievements in synthetic biology have enabled greatly improved efficiency of fermentation processes with the titer of BDO as high as 150 g L\(^{-1}\). Enzymatic synthesis of BDO has already been industrialized.

Along with the abovementioned utilization of BDO for the production of 1,3-butadiene, the diol can be applied to the production of other value-added chemicals. BDO can be converted into methyl ethyl ketone (MEK), which is currently used as a solvent, but can also be utilized as a fuel additive owing to its high heat capacity of 33.7 MJ kg\(^{-1}\). Moreover, BDO itself exhibits a high heat capacity of 27.2 MJ kg\(^{-1}\) and can be considered as a fuel blend. Alternatively, BDO can be converted into a dioxolane mixture, which can be potentially used as a gasoline blending component, diesel oxygenate, and industrial solvent. BDO is a valuable substrate for fine chemicals, for example, diacetyl applied as a flavoring agent in the food industry. Due to the presence of two stereocenters, three stereoisomers of BDO are known, namely, (2R,3S)-BDO, (2S,3R)-BDO, and the meso-form (2S,3R)-BDO (Figure 1S in the Supporting Information). Stereoselective production of BDO is of great interest, owing to its applicability in asymmetric synthesis with chiral auxiliaries possessing \(C_2\) symmetry. Nowadays, BDO is manufactured based on 2-butenes, which is available as a component of crack gases. Chlorohydrination of 2,3-butenes results in a mixture of cis- and trans-2,3-butenes oxides. The hydrolysis of this mixture gives rise to a commercial product composed of about 80% meso- and 20% rac-BDO. Regarding the stereoselectivity of BDO production, synthetic biology excels the refinery because tailoring of a microbial strain allows the production of the desired stereoisomer of biobased BDO.

The recovery of BDO from a fermentation broth still remains a tedious step in the BDO manufacturing process. A fermentation broth presents a complex mixture of organic substances and inorganic salts dissolved in water. The high boiling point of BDO of about 180 °C hampers its distillation from the solution: recovery first requires evaporation of water and all other...
components with lower boiling points. Other conventional separation methods, such as pervaporation,[12] steam stripping,[20] and reverse osmosis,[14] appeared to be energy intensive as well. Vacuum membrane distillation was proposed to enrich the fermentation broth with BDO upon vaporization of water.[12] Liquid–liquid extraction into an organic solvent attracted significant attention, although low partition coefficients of BDO were usually observed due to the high hydrophilicity of the diol. For example, biocompatible oleyl alcohol enables in situ recovery of BDO from a broth during fermentation.[13] 1-Butanol has been highlighted as a suitable solvent for BDO extraction because partitioning of BDO into 1-butanol (partition coefficient is ca. 1) is higher than that into other conventional organic solvents, although 1-butanol is partly miscible with water.[14] The addition of salts into the aqueous phase improves the partition coefficient to 5.2, owing to a salting-out effect.[14] The organic phase obtained after extraction can be concentrated by pervaporation with preferential permeation of water and 1-butanol.[14]

Aqueous two-phase extraction (ATPE) presents another method for postsynthetic[16] or even in situ[17] recovery of BDO. ATPE is based on the formation of two phases upon addition of two polymers (polyethylene glycol (PEG) and dextran),[16] or a mixture of a salt and an alcohol (17–25 wt% KH₂PO₄ + 21–24 wt% ethanol,[16] 20 wt% (NH₄)₂SO₄ + 34 wt% isopropanol,[16] 16 wt% (NH₄)₂SO₄ + 32 wt% ethanol[20]) to an aqueous solution of BDO. ATPE enables high partition coefficients of up to 28.34,[16] although utilization of highly concentrated solutions of salts requires corrosion-resistant equipment. Moreover, the recovery of salts from a highly polar medium can be challenging. Nevertheless, isolation of ammonium sulfate upon addition of methanol as an antisolvant was reported.[19,20]

Reactive extraction of BDO in the form of acetals by using butyraldehyde[21] or formaldehyde[22] was proposed. In the presence of acids, BDO reacts with aldehydes to form acetals that collect in the top oil phase, which can be separated.[22] Hydrolysis of acetals in the presence of an acid catalyst during reactive distillation releases BDO.[21] Efficient extraction of BDO in the form of acetals and recovery of the diol requires the addition of 0.5 M HCl or H₂SO₄ as catalysts, which results in the necessity for corrosion-resistant equipment and in the formation of acidic waste streams.[21] Reactive extraction of BDO from fermentation broths was performed after esterification with formic and acetic acid in the presence of H₂SO₄ as a catalyst. Pyrolysis of the obtained esters yields 1,3-butadiene.[23]

Herein, we propose a method for the recovery of BDO from fermentation broths by anionic extraction, utilizing reversible esterification with phenylboronates, as shown in Figure 1. Anionic extraction with phenylboronates has been investigated for the recovery and separation of monosaccharides[24] as well as for the isolation of 1,2-propanediol from aqueous solutions.[25] In the first step (extraction), BDO is extracted into an organic phase containing phenylboronic acid (PBA), Aliquat 336, and 1-octanol as a diluent. An anionic complex, BDO–PBA–OH, and a bulky cation of Aliquat 336 form an ion pair in the organic medium. BDO recovery is performed upon back-extraction with an acidified solution. The organic phase can be reused for the extraction. The recovered bio-BDO can be applied for the synthesis of MEK, whereas an original fermentation broth cannot. The results are discussed below in the context of challenges with water-based (bio)catalysis.

Results and Discussion
A commercially available solution of BDO and real fermentation broths were investigated. The concentrations of BDO and percentage of isomers are provided in Table 1. The commercial BDO comprised 75% meso-BDO and 25% of a mixture of (2R,3R)- and (2S,3S)-BDO, which is referred to as rac-BDO. Fermentation broths FB-1, FB-2, and FB-3 presented solutions of either meso- or rac-BDO, whereas FB-4 contained a mixture of them. In addition to BDO, the fermentation broths contained other organic compounds, such as glucose, acetoin, pyruvate, and ethanol. Detailed compositions of the fermentation broths are shown in Table 1 S in the Supporting Information.

![Image](image-url)

**Figure 1.** Isolation of BDO from fermentation broth through anionic extraction by using phenylboronates.

### Table 1. Composition of solutions of BDO applied in this study.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Abbreviation</th>
<th>Microorganism</th>
<th>BDO concentration [g L⁻¹]</th>
<th>meso-BDO [%]</th>
<th>rac-BDO [%]</th>
</tr>
</thead>
<tbody>
<tr>
<td>commercial BDO</td>
<td>-</td>
<td>-</td>
<td>pure</td>
<td>75</td>
<td>25</td>
</tr>
<tr>
<td>fermentation broths</td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>2</td>
<td>FB-1</td>
<td>L. lactis</td>
<td>15.8</td>
<td>100</td>
<td>0</td>
</tr>
<tr>
<td>3</td>
<td>FB-2</td>
<td>L. lactis</td>
<td>24.0</td>
<td>100</td>
<td>0</td>
</tr>
<tr>
<td>4</td>
<td>FB-3</td>
<td>P. polymyxa</td>
<td>26.9</td>
<td>0</td>
<td>100</td>
</tr>
<tr>
<td>5</td>
<td>FB-4</td>
<td>B. licheniformis</td>
<td>84.3</td>
<td>47</td>
<td>53</td>
</tr>
</tbody>
</table>

[a] Paenibacillus polymyxa. [b] Bacillus licheniformis.

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These are not the final page numbers!
complex designated as BDO–PBA–OH. According to data reported in the literature, anionic complexes of vic-diols with phenyl boronates are especially stable. 

Indeed, the addition of NaOH to an aqueous medium facilitates the extraction of BDO (Table 2, entries 1–3). Interestingly, a higher percentage of rac-BDO than that of meso-BDO is extracted upon addition of NaOH. This can be explained by the higher stability of complexes composed of cis-diols rather than those of trans-diols.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Additive</th>
<th>Additive concentration [mol L(^{-1})]</th>
<th>Extracted BDO [%]</th>
<th>BDO–BDO</th>
<th>rac-BDO</th>
<th>total</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>NaOH</td>
<td>0.50</td>
<td>72</td>
<td>93</td>
<td>77</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>NaOH</td>
<td>0.20</td>
<td>35</td>
<td>76</td>
<td>45</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>NaOH</td>
<td>0.06</td>
<td>18</td>
<td>55</td>
<td>27</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>H(_2)SO(_4)</td>
<td>–</td>
<td>18</td>
<td>18</td>
<td>18</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>H(_2)SO(_4)</td>
<td>1.05</td>
<td>30</td>
<td>67</td>
<td>39</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>H(_2)SO(_4)</td>
<td>2.05</td>
<td>33</td>
<td>71</td>
<td>42</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>H(_2)SO(_4)</td>
<td>5.15</td>
<td>44</td>
<td>80</td>
<td>52</td>
<td></td>
</tr>
</tbody>
</table>

[a] Extraction conditions: 0.5 m BDO (4 mL, 45 g L\(^{-1}\)) and additive (aqueous phase), 1 mL PBA; 1 mL Aliquat \(336\), and 1-octanol (4 mL, organic phase) under stirring at 750 rpm for 1 h at RT.

The composition of the extracted species was studied by NMR spectroscopy (Figure 2). First, the physical extraction of BDO into the organic phase was considered. An aqueous solution of BDO was allowed to stir with an organic phase comprised of CDCl\(_3\) and Aliquat \(336\). The NMR spectrum of the organic phase after extraction exhibited resonance signals corresponding to meso- and rac-BDO. Anionic extraction of BDO in the presence of NaOH resulted in the emergence of eight additional resonance signals, which were assigned to the complexes of meso- and rac-BDO with phenylboronate. Interestingly, introducing an asymmetrical center (a boron atom) results in lifting of the degeneracy in carbon chemical shifts corresponding to –CH\(_3\) and –CH groups. The \(^{11}\)B NMR spectrum of the organic phase after extraction from basic solutions exhibits a sole signal at \(\delta = 7\) ppm, which indicates sp\(^3\)-hybridization of boron (Figure 2S in the Supporting Information). Additionally, the presence of negatively charged complexes of BDO–PBA–OH was proven by ESI-MS. The mass spectrum of an organic phase after extraction exhibits a characteristic signal at \(m/z\) 193 (Figure 3S in the Supporting Information). Interestingly, signals with high \(m/z\) of up to 1100 were observed in the mass spectrum. The appearance of these signals can be explained by agglomeration of the amphiphilic Aliquat \(336\) ion pairs. Previously, Broekhuis et al. proposed the formation of such reverse micelles upon reactive extraction of 1,2-propanediol with phenylboronate. Thus, BDO is transferred into the organic phase as a mixture of complexes with phenylboronate and as physically extracted free BDO (Figure 2).

The addition of H\(_2\)SO\(_4\) to the aqueous phase upon extraction results in a changed NMR spectrum: signals corresponding to free meso- and rac-BDO vanish and new resonances at \(\delta = 16.6, 20.9, 75.9,\) and 80.7 ppm appear. Notably, already in the presence of H\(_2\)SO\(_4\) in low concentrations as 50 m\(\text{m}\), nearly no free BDO was detected in the organic medium (Figure 4S in the Supporting Information). The new resonances were assigned to the complexes of meso- and rac-BDO with PBA depicted in Figure 2. The \(^{11}\)B NMR spectrum of the organic phase after extraction from acidic solutions contains one broad signal at \(\delta = 31\) ppm, which corresponds to the presence of sp\(^2\)-hybridized boron (Figure 2S in the Supporting Information). Complexation between BDO and PBA upon extraction under acidic conditions was further studied by GC–MS. GC analysis of the organic phase after extraction from acidic solutions of BDO revealed the presence of two new peaks with similar retention times of 17.3 and 17.8 min (Figure 5S in the Supporting Information). These peaks were not present on the chromatogram of the organic phase before extraction. A GC–MS investigation showed that these two peaks contained a characteristic signal at \(m/z\) 176 that could be assigned to meso-BDO–PBA and rac-BDO–PBA complexes (Figure 6S in the Supporting Information). In summary, BDO is reactively extracted upon addition of H\(_2\)SO\(_4\) in the aqueous phase owing to the formation of PBA–BDO complexes (Eq. 2 and Figure 2). The formation of such complexes with sp\(^3\)-boron hybridization has been previously reported for glycosides upon transportation through a liquid membrane containing 3-(1-adamantylcarboxamido)phenylboronate and diols.
nylboronic acid. These complexes are very unstable in the presence of water and exist only in an organic phase.\(^{[26]}\)

Optimization of extraction and back-extraction conditions for fermentation broths

According to the results in Table 2, the presence of NaOH in amounts at least equimolar to the amount of BDO is required to achieve good extraction results. Excess NaOH indeed facilitates extraction by shifting the equilibrium of Equation (1) to the right (Table 3, entries 1–3). Excess NaOH is not desirable from a practical point of view because it leads to higher costs and the additional generation of waste. Moreover, the utilization of an equimolar amount of NaOH already results in a rather high percentage of extracted BDO of 77\% when using a model solution of a commercial diol. In addition, titration of an aqueous phase after extraction showed that the amount of extracted BDO equaled the amount of extracted OH\(_{-}/C_0\) species, which confirmed that the extraction mainly occurred due to complex formation according to Equation (1). Nevertheless, real fermentation broths usually contain other components that can influence extraction.\(^{[8,12]}\) Table 3 provides the results of BDO extraction from real fermentation broths. Fermentation broth FB-1 exhibited the lowest percentage of extracted BDO of 20–25\%. This can be explained by a high glucose content in FB-1: it comprised 30.7 and 19.8 g L\(^{-1}\) glucose and BDO, respectively (Table 15 in the Supporting Information). In the presence of alkali, glucose yields a complex mixture of acidic products as a result of isomerization, cleavage, and dehydra-

<table>
<thead>
<tr>
<th>Entry</th>
<th>BDO solution</th>
<th>NaOH/BDO(^{[b]})</th>
<th>Extracted BDO [%]</th>
<th>Neutralized OH(_{-}/C_0) (^{[c]})</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>commercial</td>
<td>2</td>
<td>84 93 86</td>
<td>0.9</td>
</tr>
<tr>
<td>2</td>
<td>commercial</td>
<td>1.5</td>
<td>82 93 85</td>
<td>1.0</td>
</tr>
<tr>
<td>3</td>
<td>commercial</td>
<td>1</td>
<td>72 93 77</td>
<td>0.9</td>
</tr>
<tr>
<td>4</td>
<td>FB-1</td>
<td>2</td>
<td>25</td>
<td>25 5.7</td>
</tr>
<tr>
<td>5</td>
<td>FB-1</td>
<td>1.5</td>
<td>22</td>
<td>22 4.5</td>
</tr>
<tr>
<td>6</td>
<td>FB-1</td>
<td>1</td>
<td>20</td>
<td>20 3.1</td>
</tr>
<tr>
<td>7</td>
<td>FB-2</td>
<td>2</td>
<td>73</td>
<td>73 1.5</td>
</tr>
<tr>
<td>8</td>
<td>FB-2</td>
<td>1.5</td>
<td>57</td>
<td>57 1.7</td>
</tr>
<tr>
<td>9</td>
<td>FB-2</td>
<td>1</td>
<td>34</td>
<td>34 1.8</td>
</tr>
<tr>
<td>10</td>
<td>FB-3</td>
<td>2</td>
<td>–</td>
<td>93 93 1.6</td>
</tr>
<tr>
<td>11</td>
<td>FB-3</td>
<td>1.5</td>
<td>–</td>
<td>84 84 1.5</td>
</tr>
<tr>
<td>12</td>
<td>FB-3</td>
<td>1</td>
<td>–</td>
<td>68 68 1.1</td>
</tr>
<tr>
<td>13</td>
<td>FB-4</td>
<td>2</td>
<td>63</td>
<td>84 74 1.1</td>
</tr>
<tr>
<td>14</td>
<td>FB-4</td>
<td>1.5</td>
<td>66</td>
<td>87 77 1.0</td>
</tr>
<tr>
<td>15</td>
<td>FB-4</td>
<td>1</td>
<td>47</td>
<td>89 68 0.9</td>
</tr>
</tbody>
</table>

\(^{[a]}\) Extraction conditions: a solution (4 mL) containing BDO and NaOH (aqueous phase); 1 mM PBA, 1 mM Aliquat\(^{\text{336}}\), and 1-octanol (4 mL, organic phase) under stirring at 750 rpm for 1 h at RT. \(^{[b]}\) Molar ratio of NaOH and BDO adjusted in aqueous phase prior to the extraction. \(^{[c]}\) Molar ratio of OH\(_{-}\) neutralized during the extraction [ideally due to the complex formation according to the Eq. (1)] to the amount of extracted BDO.

Figure 2. Assignment of NMR signals observed upon extraction of BDO (top), anionic extraction of BDO with phenylboronate (middle), and extraction of BDO due to complexation with PBA (bottom).
tion reactions.\textsuperscript{[27]} Formation of these organic acids result in partial neutralization of NaOH. In addition, glucose itself is a polyol, which is coextracted together with BDO. Extraction of BDO from FB-2–FB-4 appeared to be more efficient, compared with FB-1, although somewhat less efficient than the use of a commercial solution of BDO. In addition to BDO, glycerol was also coextracted from the fermentation broths (Table 2 S in the Supporting Information). Recovery of a polyol after anionic extraction with phenylboronates is typically performed upon back-extraction into an acidic medium. Addition of an acid shifts the equilibrium of Equation (1) to the left and a polyol is released. This approach appeared to be successful for the recovery of saccharides.\textsuperscript{[24]} However, as shown above, BDO also forms a complex with PBA under acidic conditions according to Equation (2). Therefore, excess acid was expected to facilitate the extraction of BDO due to the formation of a BDO–PBA complex.

The amount of back-extracted BDO indeed depends on the concentration of the acid used (Figure 3). Complete results of the extraction and back-extraction are provided in Table 3 S in the Supporting Information. The best recovery of BDO was observed under the applied conditions.

Recovery of a polyol after anionic extraction with phenylboronates is typically performed upon back-extraction into an acidic medium. Addition of an acid shifts the equilibrium of Equation (1) to the left and a polyol is released. This approach appeared to be successful for the recovery of saccharides.\textsuperscript{[24]} However, as shown above, BDO also forms a complex with PBA under acidic conditions according to Equation (2). Therefore, excess acid was expected to facilitate the extraction of BDO due to the formation of a BDO–PBA complex.

The amount of back-extracted BDO indeed depends on the concentration of the acid used (Figure 3). Complete results of the extraction and back-extraction are provided in Table 3 S in the Supporting Information. The best recovery of BDO was achieved by performing back-extraction in slightly acidic medium with [H\textsuperscript{+}] of less than 0.01 m. The percentage of back-extracted BDO depends on the stereochemistry: meso-BDO is recovered more readily compared with rac-BDO due to lower stability of a complex between phenylboronate and the former stereoisomer. In general, no difference in back-extraction of commercial BDO and bio-BDO was observed (Table 3 S in the Supporting Information). Notably, the amount of recovered BDO is limited to about 70–80% when using the same volumes of organic and aqueous phases. The recovery can nevertheless be improved up to 90% when using an excess of aqueous phase (Table 3 S in the Supporting Information). Figure 7 S in the Supporting Information shows HPLC traces of fermentation broths and refined solutions after extraction and back-extraction.

Recycling of the organic phase was performed by repeating extraction and back-extraction experiments. The organic phase was treated with a phosphate buffer after each recycling run. A somewhat declining BDO capacity of the organic phase was observed upon reuse. This can be explained by a partial leaching of boron upon extraction and back-extraction (Figure 4).

Conversion of BDO into MEK

Conversion of BDO into MEK takes place through a pinacol rearrangement in the presence of an acidic catalyst. The catalytic activity of molecular acids, such as H\textsubscript{2}SO\textsubscript{4} or H\textsubscript{3}PO\textsubscript{4} for this reaction has been known for a long time. A reduction of corrosion potential and a decrease of acidic waste streams can be achieved by application of solid catalysts, for example, sulfonated alumina and silica–alumina or boric acid modified HZSM-5 zeolites.\textsuperscript{[5]} Herein, we focused on the transformation of BDO into MEK in the presence of sulfuric acid. First, the possibility of using a fermentation broth directly for MEK synthesis was investigated. Commercial BDO could be readily converted into MEK (Figure 5), whereas a mixture of fermentation broth with sulfuric acid yielded no MEK, but a black precipitate of humins (Figure 8 S in the Supporting Information). This result contradicts previous publications reporting the successful direct application of fermentation broths for MEK synthesis.\textsuperscript{[28, 30]} The efficiency of BDO dehydration to MEK appears to depend significantly on the composition of the fermentation broth and probably on the composition of the catalyst. Herein, the other components of the fermentation broths gave rise to humins in the presence of acid and at elevated temperature (Table 1 S in the Supporting Information).

However, after the isolation of BDO through extraction with phenylboronate, bio-BDO can be converted into MEK as efficiently as commercial BDO (Figure 5). Under the reaction conditions, BDO was also converted into isobutyraldehyde in minor yields of 1–3\%.\textsuperscript{[24]} In addition, the formation of condensation products owing to the aldol reaction was observed.
Conclusions

Herein, we demonstrate the recovery of BDO from a complex fermentation broth by means of anionic extraction through complexation with phenylboronates. This method enables the selective recovery of BDO from the broth, although other components containing a 1,2-diol motif, such as glycerol or glucose, can be coextracted. Gaining an insight into processes of extraction and back-extraction allowed us to optimize their conditions. Up to 72–93 % BDO can be extracted and up to 80–90 % recovered under the optimized conditions. The organic phase was reused four times with only a minor decrease of the BDO capacity and moderate leaching of boron into the aqueous phase. Refined bio-BDO could be efficiently used for the synthesis of MEK, whereas the fermentation broths could not due to the formation of humins. The obtained results indicate that anionic extraction is a promising method for the recovery of BDO. Further research can be focused on reducing the amount of organic solvents and molecular acids/bases used as auxiliary chemicals to facilitate complexation. Moreover, the sustainability of MEK synthesis can be further improved through the utilization of a solid acidic catalyst. In situ removal of the product by reactive distillation presents another possibility for process intensification.

Experimental Section

Chemicals

2,3-Butanediol (98 %), Aliquat 336 (chloride form), and 1-octanol (> 99 %) were purchased from Sigma–Aldrich. Deuterium oxide (99.9 %) and CDCl 3 (99.9 %) were obtained from Deutero. [D 6 ]Benzene (99.9 %) and [D 6 ]acetone (99.8 %) were purchased from abcr. Microgranulate sodium hydroxide (98.8 %), methanol (99.8 %), sulfuric acid (> 95 %), and a standard solution of hydrochloric acid (0.1 m) were obtained from Chemsolute. PBA (98 %) was obtained from ChemPUR. Sodium dihydrogen phosphate monohydrate (Reag. Ph. Eur.) was provided by Merck. All solutions were prepared in distilled water.

Strain construction

*L. lactis* pNZ_BD1 was a derivative of the double lactate dehydrogenase-deficient strain NZ9000ΔldhDΔldhB that overexpressed the genes coding for α-acetolactate synthase (als, ilm_; 1309) and acetoin reductase (butA, ilm_; 1641) from *L. lactis* MG1363. [34] The als and butA genes were cloned into the nisin-inducible expression vector pNZ8048, [34] as previously described, [35] to provide plasmid pNZ_BD1. The plasmid was obtained and maintained in NZ9000. [34] The gene sequences of pNZ_BD1 were verified by sequencing and plasmid was electroporated [36] into NZ9000ΔldhDΔldhB originating strain NZ9000ΔldhDΔldhBpNZ_BD1 (herein denoted *L. lactis* pNZ_BD1).

Fermentation

*L. lactis* pNZ_BD1 was cultivated in New Brunswick BioFlo 110 bioreactors (Eppendorf, Hamburg, Germany) with a total volume of 1.3 L and a working volume of 0.5 L. The temperature was maintained at 30 °C and the pH value was set to 6.5 and controlled automatically with 4 M NaOH. The aeration rate was set to 0.5 L min⁻¹ (1vvm) and dissolved oxygen was kept at 40 % by automatically adjusting the stirring rate between 250 and 1200 rpm. The fed-batch fermentations were conducted by using a chemically defined medium (CDM) [32] and complex medium Difco™ M17 Broth (BD, USA) referred to as M17. In the cultivations, glucose (20 g L⁻¹) was added from the beginning. The media were furthermore complemented with CM and hemin. One hour after inoculation, nisin was added to induce the expression of the genes for BDO production. Three times during 45 h of fermentation, glucose (20 g L⁻¹) was added for process intensification.

Figure 5. Yield of MEK versus BDO conversion at 180 °C in the presence of H 2SO 4. Bio-BDO was applied after purification of FB-4 by anionic extraction with phenylboronate.
added (after 15, 17, and 23 h). Precultures were grown in 500 mL shake flasks filled with the respective medium (50 mL). They were inoculated with a cryoculture (250 mL) and incubated overnight at 30 °C. The starting optical density at λ = 600 nm (OD_600) was set to 0.25.

Cultivations of B. licheniformis DSM 8785 and P. polymyxa DSM 365 were conducted in a complex medium, according to a procedure reported by Nakashimada et al. The medium contained yeast extract (5 g L⁻¹), tryptone (5 g L⁻¹), K$_2$HPO$_4$ (7 g L⁻¹), KH$_2$PO$_4$ (5.5 g L⁻¹), (NH$_4$)$_2$SO$_4$ (1 g L⁻¹), MgSO$_4$·7H$_2$O (0.25 g L⁻¹), Na$_2$MoO$_4$·2H$_2$O (0.12 g L⁻¹), CaCl$_2$·2H$_2$O (0.021 g L⁻¹), Co(NO$_3$)$_2$·6H$_2$O (0.029 g L⁻¹), (NH$_4$)$_2$Fe(SO$_4$)$_2$·6H$_2$O (0.039 g L⁻¹), nicotinic acid (0.002 g L⁻¹), Na$_2$SeO$_3$ (0.0002 g L⁻¹), NiCl$_2$·6H$_2$O (0.00005 g L⁻¹), MnCl$_2$·4H$_2$O (0.005 g L⁻¹), H$_2$BO$_3$·(0.001 g L⁻¹), AlK(SO$_4$)$_2$·12H$_2$O (0.0002 g L⁻¹), CuCl$_2$·2H$_2$O (0.00001 g L⁻¹), and Na$_2$EDTA·2H$_2$O (0.0055 g L⁻¹). Glucose was added at concentrations of 180 g L⁻¹ for B. licheniformis and 70 g L⁻¹ for P. polymyxa. The medium was completed from different stock solutions that were sterilized separately. The cultivation was performed at 37 °C in 250 mL shake flasks that were placed in an orbital shaker with a shaking frequency of 100 rpm and a shaking diameter of 5 cm. As preculture, the medium (20 mL) was inoculated with a cryostock (200 mL). This culture was used to inoculate 40 mL per flask at an OD$_{600}$ of 0.1 as the main culture.

**Extraction**

The organic phase typically contained 1 mM PBA and 1 mM Aliquat 336 dissolved in 1-octanol. The organic phase was pretreated prior to extraction by stirring with 1 M Na$_2$HPO$_4$·Na$_2$HPO$_4$ phosphate buffer at pH 7.5 for 30 min at RT. In a typical extraction experiment, the organic phase (4 mL) and an aqueous phase (4 mL) were left stirred (750 rpm) for 1 h at RT. Thereafter, the phases were separated by centrifugation for 1 min at 7000 rpm. After splitting the phases, back-extraction was performed: the organic phase (4 mL) was back-extracted with monitoring of the final pH of the aqueous phase.

Concentrations of NaOH and H$_2$SO$_4$ in the solutions after extraction were determined by means of titration by means of NMR spectroscopy. 1H (400 MHz), 11C (101 MHz), and 13C (96 MHz) spectra were recorded on Bruker spectrometers. Chemical shifts are reported in δ (ppm) units by using internal and external standards. The chemical shifts are given in ppm. Structures of complexes of BDO with PBA or phenylboronate were proposed based on results of COSY, HSQC, HMBC, and APT NMR spectra. In addition, NMR spectroscopy investigations of BDO extracted from fermentation broths were performed.

Samples for analysis by ESI-MS were prepared by using a typical extraction procedure. After separation of the phases, the organic phase was diluted 1:50 with 1-octanol. The diluted samples were injected into an LCMS-2020 liquid chromatograph mass spectrometer (Shimadzu) operating in both positive and negative ionization modes. The mass spectra were collected in the range of m/z 50–2000 with a probe voltage of 4.5 kV and Q-array radiofrequency (RF) voltage of 60 V.

The samples for GC–MS were prepared according to a typical extraction procedure. The organic phase was diluted 1:10 with methanol and injected into a gas chromatograph equipped with a TRACE Ultra GC by Thermo Scientific with Rtx-5-Sil-MS column (30 m, inner diameter (ID) = 0.28 mm, d$_{f}$ = 0.25 μm, T = 50–280 °C, 5 iso, 8 °C/min). For GC–MS measurements, a Varian-Cp 3800 device with a 1200L Quadrupole MS/MS spectrometer was used.

**Conversion of BDO into MEK**

Experiments were carried out in a 45 mL Berghof autoclave equipped with a polytetrafluoroethylene (PTFE) inlet. Aqueous solutions (4 mL) containing BDO and H$_2$SO$_4$ were charged into an autoclave, sealed, and pressurized with about 30 bar (1 bar = 10$^5$ Pa) nitrogen. The reactions were performed under stirring at 750 rpm at 180 °C. The initial concentration of BDO was 0.25 or 0.5 M. The concentration of H$_2$SO$_4$ varied in the range of 0.05–0.18 M. The reaction was performed for 2–7 h. After the experiments, the reaction mixture was cooled in an ice bath and filtered through a polyamide syringe filter (Chromaphil, polyamide, pore size 0.2 μm).

Concentrations of BDO, other components of fermentation broths, and the products of dehydration to MEK were determined by HPLC on a Shimadzu Prominance LC-20 system. The separation was performed by using two successively connected organic acid resin columns (CS-Chromatographie, 100 mm×8.0 mm and 300 mm×8.0 mm) at 40 °C; the eluent (CF$_3$COOH (154 μL) in water (1 L)) was supplied at a flow rate of 1 mL min$^{-1}$. The system was equipped with a refractive index (RI) detector.

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Conflict of interest

The authors declare no conflict of interest.

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These are not the final page numbers!
**Complex separation:** Microbial production of 2,3-butanediol based on renewable resources enables high titers; however, the recovery of this hydrophilic high-boiling compound from aqueous fermentation broths remains challenging. Anionic extraction based on reversible esterification with phenylboronate is proposed for the isolation of 2,3-butanediol from aqueous solutions. The recovered bio-2,3-butanediol can be efficiently transformed into methyl ethyl ketone.

Anionic Extraction for Efficient Recovery of Biobased 2,3-Butanediol—a Platform for Bulk and Fine Chemicals