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Published in:
A C S Nano

Link to article, DOI:
10.1021/acsnano.8b00596

Publication date:
2018

Document Version
Publisher's PDF, also known as Version of record

Link back to DTU Orbit

Citation (APA):
Multifarious Biologic Loaded Liposomes that Stimulate the Mammalian Target of Rapamycin Signaling Pathway Show Retina Neuroprotection after Retina Damage

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Supporting Information

ABSTRACT: A common event in optic neuropathies is the loss of axons and death of retinal ganglion cells (RGCs) resulting in irreversible blindness. Mammalian target of rapamycin (mTOR) signaling pathway agonists have been shown to foster axon regeneration and RGC survival in animal models of optic nerve damage. However, many challenges remain in developing therapies that exploit cell growth and tissue remodeling including (i) activating/inhibiting cell pathways synergistically, (ii) avoiding tumorigenesis, and (iii) ensuring appropriate physiological tissue function. These challenges are further exacerbated by the need to overcome ocular physiological barriers and clearance mechanisms. Here we present liposomes loaded with multiple mTOR pathway stimulating biologics designed to enhance neuroprotection after retina damage. Liposomes were loaded with ciliary neurotrophic factor, insulin-like growth factor 1, a lipopeptide N-fragment osteopontin mimic, and lipopeptide phosphatase tension homologue inhibitors for enhancing axon regrowth and RGC survival after ON transection so far have been achieved by Bei et al. using a combination of adeno-associated virus (AAV) assisted overexpression of ciliary neurotrophic factor (CNTF), insulin-like growth factor 1 (IGF-1) and osteopontin (OPN), combined with multiple pharmacotherapies, pharmacotherapy, and cell therapy, etc. This is indicated by previous studies either involving the supplementation of different neurotrophic factors to protect RGCs in different animal models (induced hypertension, ON transection and ON crush) or the deletion of cell growth regulatory genes PTEN and SOCS3 to protect RGCs after axotomy.

Arguably, the most striking results in axon regrowth after ON transaction so far have been achieved by Bei et al. using a combination of adeno-associated virus (AAV) assisted overexpression of ciliary neurotrophic factor (CNTF), insulin-like growth factor 1 (IGF-1) and osteopontin (OPN), combined with multiple pharmacotherapies, pharmacotherapy, and cell therapy, etc. This is indicated by previous studies either involving the supplementation of different neurotrophic factors to protect RGCs in different animal models (induced hypertension, ON transection and ON crush) or the deletion of cell growth regulatory genes PTEN and SOCS3 to protect RGCs after axotomy.

Received: January 23, 2018
Accepted: July 13, 2018
Published: July 13, 2018

DOI: 10.1021/acsnano.8b00596
ACS Nano 2018, 12, 7497–7508

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with AAV codeletion of PTEN and SOCS3 genes. CNTF and IGF-1 have both been shown to promote axon regeneration by activation of the mTOR pathway. The cytokine CNTF is capable of activating several different signaling pathways such as JAK-signal transducers, MAPK, PI3K, and mTOR-p70S6K by binding to receptors at the cell surface. CNTF has been shown to stimulate mTOR activation of STAT3, hence implicating CNTF and mTOR as transcriptional regulators in neuroblastoma cells. CNTF supplementation has also been shown to prevent the decrease in mTOR activity in vitro in RGCs. IGF-1 is a protein hormone that binds to a receptor tyrosine kinases (IGF-1R) on the cell surface activating PI3K. IGF-1 and regulation of the MDM4/p53-IGF-1 pathway has been shown to be critical for axonal sprouting and neurological recovery after spinal cord injury.

OPN is a secreted phosphoprotein that binds to receptors recognizing the Arg-Gly-Asp protein motif. OPN is implicated in maintenance and reconfiguration of tissue integrity during inflammation by modulating the extracellular matrix (ECM). In a stroke model in Sprague–Dawley rat, OPN has been shown to be neuroprotective, and in combination with IGF-1, OPN has been shown to promote regeneration of adult RGCs after axotomy. Phosphatase tension homologue (PTEN) is a phosphatase that dephosphorylates PIP3 to PIP2 resulting in the Akt/mTOR pathway. PTEN deletion by virus mediated gene silencing has been shown to prevent RGC apoptosis after ON cut and enhance the regenerative potential of neurons in the corticospinal tract.

For clinical translation, combining mTOR pathway stimulating biologics for RGC neuroprotection requires modifications that would provide better control over cell pathway modulator (i.e., promoters, inhibitors, inducers, agonists and antagonists) delivery, release, clearance and residency time. Nanocarriers represent one of many possible solutions (e.g., microcarriers, implants, etc.) in delivering multiple cell pathway modulators while controlling modulator release. Nanocarriers protect sensitive cargos from degradation, can reduce the effect of clearance mechanisms and improve absorption of cargos across physiological barriers. Liposomes, phospholipid vesicles frequently containing cholesterol, are an attractive delivery system for retina diseases, offering loading of both hydrophobic and hydrophilic molecules, prolonged retention in the eye after intravitreal or subretinal injection, protection of cargo from degradation and have a history of clinical approval. The majority of liposomal research in central nervous system (CNS) aspects is associated with overcoming the blood brain barrier to improve the delivery of drugs to the brain. Liposomes and lipoplexes have been shown to effectively deliver the RPE65 gene, critical for vision, in RPE65 knockout blind mice leading to an improvement in vision. Limited work has been reported on neuroprotection and nerve regeneration. Cationic liposomes and corresponding lipoplexes have been used to transfer neurotrophic factor genes (e.g., glial derived neurotrophic factor, nerve growth factor etc.) in both spinal cord and brain injury models. These studies have shown neuroprotection and partial restoration of locomotor function. Immunomodulation, via the selective apoptosis of monocytes and phagocytic macrophages using clodronate loaded liposomes, has shown partial hindlimb recovery and neuron repair in a rat spinal cord injury model. To our knowledge, liposomes loaded with multiple cell pathway have not previously been reported for CNS neuroprotection and regeneration. Liposomes loaded with multiple cell pathway modulators have the advantage of delivering these modulators to the cell microenvironment at the same time, facilitating a synergistic effect on single or multiple cell pathways. Furthermore, signaling pathway modulators are often highly efficacious and only very low concentrations are required. This reduces the need for high drug loading and maximizing the space available to pack a range of therapeutic small molecules, peptides and proteins into the liposome.

In this study, we present two liposome formulations that promote neuroprotection through stimulating the mTOR pathway. Liposome aqueous cores were loaded with CNTF and IGF-1. Liposome membranes were loaded with lipid conjugated peptides OPP and either PAP2 or PAP4. OPP is a peptide analogue of the N-fragment of OPN (specifically the RGD and α3β1/α5β1 domains) that has been shown to mimic the function of recombinant human OPN and lower cytosolic Ca2+ in a way similar to OPN. PAPs are PTEN antagonist peptides with PAP2 targeting the ATP B type domain and PAP4 targeting the c-terminal tail of PTEN. We show that multifarious mTOR pathway stimulating biologic loaded liposomes significantly prevent RGC death and loss of retina electrophysiological function in a N-methyl-D-aspartic acid (NMDA) mouse model. To further explore combination therapy, we combine one neuroprotective liposome formulation with induced pluripotent stem cell (iPSC) derived RGC transplantation in the NMDA mouse model and show improved electrophysiological outcome of the transplantation.

RESULTS AND DISCUSSION

Liposomes. The palmitoyl (C16) conjugated peptides (i.e., lipopeptides) C16-OPP, C16-PAP2, and C16-PAP4 (referred to as OPP, PAP2, and PAP4 from now on), were all successfully synthesized and purified by semipreparative reverse phase HPLC to a purity of >95% confirmed by HPLC and MALDI-TOF MS (Supplementary Figures S1 and S2). Liposomes were produced as illustrated in Figure 1A. Liposomes showed low polydispersity index (PDI < 0.1) with diameters close to 100 nm in both DLS measurements (Figure 1B) and cryoTEM images (Figure 1C). CryoTEM images showed spherical liposomes that were predominantly unilamellar. All liposomes had a negative zeta potential, which is important for reducing cytotoxicity and in combination with surface PEylation has been shown to improve liposome diffusion throughout the vitreous. Lip A and Lip B showed a slightly more negative zeta potential than Lip C (~19 mV, ~18 mV, and ~13 mV, respectively). The zeta potential of all the liposomes is similar to previously reported values of unilamellar liposomes with similar phospholipid compositions.

The encapsulation efficacy (EE%) normalized to the lipid concentration was higher for Lip B than the EE% of Lip A, and both formulations showed a higher EE% of CNTF (~22 kDa) than IGF-1 (7.7 kDa). These EE% approximately corresponded to 700 nM IGF-1 and 763 nM CNTF concentrations for Lip A, while for Lip B the concentrations were 1.53 μM IGF-1 and 1.27 μM CNTF. These values indicate that IGF-1 and CNTF both loaded in approximately a 1:1 ratio in both formulations and that macromolecule size did not influence loading. These concentrations were well above the IC50 values of IGF-1 and CNTF (i.e., < 6 nM), while keeping the injected concentration of CNTF low enough to not induce suppression of retina electrophysiological function.

The difference in EE% between the two formulations is likely to be due to the different PTEN inhibitor lipopeptides
incorporated into the membrane. PAP2 has a cysteine with a free thiol group that can react with other thiol groups, either from other PAP2 lipopeptides in a liposome bilayer or on proteins (e.g., present in FBS, IGF-1, etc.), possibly reducing protein loading or increasing liposome leakiness during purification. This is supported by the observation that neither Lip A or Lip B leaked calcein (a small hydrophilic fluorescent dye) at 37°C in HBS. However, Lip A did leak calcein over time when 10% FBS was added to HBS at 37°C (Supplementary Figure S3), indicating an interaction that resulted in liposome bilayer instability. The low EE% ensured that the proteins were fully hydrated and did not aggregate inside the liposomes. This is supported by the absence of visible internal structures in the cryoTEM images in Figure 1C. The bioactivity of Lip A and Lip B was tested in vitro in HEK293T cells (Supplementary Figure S4). Phosphorylation of Akt and p70S6K was observed for both formulations, indicating that the cargos remained biologically active.

### Liposome Uptake in Retinal Organoids

The cellular uptake of liposomes was tested in vitro in retinal cell organoids differentiated from mouse embryonic stem (mES) cells (Figure 2A). This system has advantages over traditional 2D cultures, including more relevant pharmacokinetik results. Retinal organoids were cultured for 21 days and then exposed to liposomes labeled with Atto655 for 12 h. The level and cell specificity of uptake was quantified by flow cytometry (Supplementary Figure S5). The overall uptake of liposomes (Figure 2A) showed that Lip B had the lowest uptake with approximately 13% of cells showing uptake. The control liposome, Lip C, showed a slightly higher cellular uptake (18% of cells) than Lip B. The highest uptake was observed for Lip A with 23% of cells showing uptake. The variance between replicas of Lip A was significantly larger than that of either Lip B or Lip C. Statistical analysis between the liposome formulations was carried out, and none of the formulations showed significantly different uptake compared to Lip C. However, Lip A had a significantly higher uptake than Lip B. The higher uptake observed for Lip A might be explained by sulfur–sulfur interactions between the cysteine in PAP2 and thiol groups on the cell surface. Cell surface thiols interacting with thiols present on nanomaterials has been argued as a mechanism to enhance uptake into cells.

Retinal organoids were approximately 1 mm in diameter (Figure 2A). Retinal cells are between 9 and 12 μm in diameter and assuming organoids are densely packed perfect spheres then approximately 1% of the cells are located on the organoid surface, indicating that the liposomes were capable of moving (i.e., by active transport and/or passive diffusion) beyond the surface layer of cells. It has previously been reported that nanocarriers can penetrate beyond the surface layer of cells in 3D cell aggregates. To determine if liposome uptake was associated with specific cell types, organoids were dissociated and stained for five major retinal cell type markers. The markers used were antirecoverin-Rb-IgG (Chemicon) for photoreceptor cells,
Figure 2. Liposome uptake in retinal organoids. (A) Box and whiskers plot of the total liposome uptake (as % positive cells for Atto655) in retinal organoids (*P ≤ 0.05 between Lip A and Lip B, N = 4). Inset shows an example micrograph of a retinal organoid cultured to day 21 (scale bar, 1 mm). The box plot median values are 22.6%, 12.8%, and 18.4% for Lip A, Lip B, and Lip C, respectively. (B) Liposome uptake (as % positive cells for Atto655) in specific cell types found in retinal organoids using cell markers. The cell types shown are photoreceptors (recoverin), bipolar cells (PKC-σ), RGCs (RBPMs), horizontal cells (calbindin), and Müller glia cells (glutamine synthetase (GS)). Error bars show standard error of the mean (**P ≤ 0.0001 between Lip A and Lip B, no significant difference between cell types, N = 4).

Figure 3. Electroretinogram amplitude values from dark-adapted mice. (A) A-wave response for healthy and treated eyes. Box plot median values are −15.6 μV for healthy eye, −9.9 μV for Lip A, −15.5 μV for Lip B, and −29.1 μV for Lip C. (B) B-wave response for healthy and treated eyes (*P ≤ 0.05 between Lip A and Lip C). Box plot median values are 214.3 μV for healthy eye, 138.7 μV for Lip A, 104.7 μV for Lip B, and 83.5 μV for Lip C. (C) The change in scotopic threshold response, ΔSTR (pSTR − nSTR), between the healthy left (oculus sinister, OS) and liposome treated damaged right (oculus dexter, OD) eyes. Box plot median values are 27.6 μV for Lip A, 7.4 μV for Lip B, and 32.2 μV for Lip C. Highly negative values (greater than −10 μV) were excluded from ΔSTR assuming natural visual impairment in the healthy control eye. N = 8–10 mice per liposome treated group, N = 28 mice for healthy control eye.

in Vivo Preservation of Retinal Function by Loaded Liposomes. RC57BL/6J mice was induced in the right eye of C57BL/6J mice by a single intravitreal injection of N-methyl-D-aspartic acid (NMDA, 2 μL at 20 mM). Mice were divided into three treatment groups receiving 1 intravitreal injection of either Lip A, Lip B, or Lip C liposomes 2 h after the NMDA injection. Four weeks after the liposome treatment the retinal function of dark-adapted mice were evaluated by electroretinography (ERG) and eyes were enucleated for histological analysis (Supplementary Figure S6). NMDA treated mice receiving empty liposomes (Lip C) were used as controls. NMDA binds irreversibly to the NMDA receptor at the postsynaptic membrane leading to an excessive influx of positive ions (e.g., Ca²⁺), depolarizing the mitochondrial membrane and ultimately triggering apoptosis.⁷¹–⁷³ NMDA has shown retinal toxicity at low concentrations (e.g., 20 nM) with the degree of inner retina damage corresponding to NMDA concentration.⁷⁴ NMDA affects other retina cell types and intravitreal administered NMDA also induces partial optic nerve damage, likely linked to the damage of the inner retina. A 20 mM NMDA injection is an extreme model of retina damage. We observed no significant difference in a-wave response (Figure 3A), associated with photoreceptors, between healthy control eyes and NMDA-injected eyes. This confirmed that intravitreal NMDA injection at 20 mM concentration did not result in photoreceptor damage, which is consistent with previous observations.⁴⁻⁷⁵

The b-wave response, associated with the function of interneuron cells (e.g., amacrine and horizontal) and bipolar neuron cells,⁶⁷⁻⁷⁶ showed a significant decrease in all NMDA-injected groups (Figure 3B) compared to healthy controls. At low concentrations (<50 nM) NMDA has been shown to damage amacrine cells but bipolar cell damage has only been reported for higher concentrations (>200 nM).⁷⁸ At 20 mM NMDA concentrations, substantial bipolar and interneuron cell damage will have occurred. We observed a significant protective effect of Lip A compared to the empty liposome Lip C (P value < 0.02). Lip B showed no significant difference in b-wave response compared to either the Lip A or Lip C treated groups. The Lip B treated group did show a trend which indicated a minor preservation of b-wave response than the Lip C treated group. A-wave and b-wave amplitudes in healthy eyes between C57BL/6J mice were similar to previous reports.⁷⁹

Scotopic threshold response (STR, Figure 3C) has been related to the function of the inner retinal neurons more proximal than the bipolar cells (e.g., RGCs).²⁻⁷⁵,⁸⁰⁻⁸¹ To estimate the neuroprotective effect, the difference in STR amplitudes (pSTR − nSTR) between the healthy (left) and treated (right) eye for each animal was determined (i.e., ΔSTR_{OS-OD}; Figure 3C). A lower ΔSTR_{OS-OD} therefore means a return of function and...
an inner retinal neuronal response closer to the healthy eye, with full restoration giving a ΔSTR_{OS-OD} = 0. For ΔSTR_{OS-OD}, no significant difference was found between the treatment groups (ANOVA p > 0.05). However, both Lip A and Lip B treated groups showed clear trends toward better preservation of STR. For Lip A the spread of data points is divided into two groups suggesting a bimodal distribution (responsive and unresponsive), in which the responsive group has a ΔSTR_{OS-OD} of ~7 μV. The Lip B treated group showed a more unimodal distribution with more than 75% of the mice around the median ΔSTR_{OS-OD} of 7.4 μV (the median for Lip C treated group is 32.2 μV for comparison).

Extrapolation of the ERG data would indicate that Lip A had a significant protective effect on interneuron and bipolar neuron cells but a mixed effect on RGCs. Conversely, Lip B showed no significant protective effect on interneuron and bipolar cells but indicated a protective effect on RGCs. The reasons for this observation are likely manifold. PTEN inhibition may be a critical factor and peptides that inhibit regions of PTEN might induce different responses in different cell types. Equally, the concentrations of IGF-1 and CNTF needed for protection combined with appropriate release kinetics might be specific to specific cell types. Interestingly, NMDA activation of the p38 MAPK pathway has been shown to be pro-apoptotic for RGCs and pro-survival for photoreceptors.82 Given IGF-1 and CNTF MAPK pathway has been shown to be pro-apoptotic for RGCs (Figure 4A), while IGF-1 and CNTF will promote a number of signaling pathways (e.g., MAPK for RGCs, PTEN inhibition may be a critical factor and peptides that inhibit regions of PTEN might induce different responses in different cell types). Equally, the concentrations of IGF-1 and CNTF needed for protection combined with appropriate release kinetics might be specific to specific cell types. Interestingly, NMDA activation of the p38 MAPK pathway has been shown to be pro-apoptotic for RGCs and pro-survival for photoreceptors.82 Given IGF-1 and CNTF MAPK will promote a number of signaling pathways (e.g., MAPK for IGF-1) and that signaling pathway dynamics are likely different between specific cell types, loading multiple pathway regulators into liposomes may lead to specific effects (positive and negative) on specific cell types rather than promote positive effects in tissue as a whole.

**In Vivo Rescue of RGCs by Loaded Liposomes.** To evaluate the effect of a single intravitreal injection of Lip A and Lip B on host RGC survival after NMDA induced RGC death, retinal whole mounts were stained for RBPMs and imaged on a confocal microscope. Three images per retina were acquired, approximately 1 mm away from the optic nerve head. Example micrographs from a healthy control eye, an eye treated with Lip A, and an eye treated with Lip C are shown in Figure 4 panels A, B, and C, respectively. RGCs in each image were counted and the average number of surviving RGCs in the three images was calculated to enable quantitative comparison between the groups (Figure 4D). NMDA injection in combination with Lip C resulted in substantial RGC loss (~75% loss compared to the healthy controls). This observation is consistent with previous quantification of RGC loss in mice without empty liposomes.83 This result showed that Lip C did not have a neuroprotective effect. Treatment with either Lip A or Lip B was not able to completely prevent RGC loss.

**Statistical Analysis.** The number of surviving RGCs was compared between the treated groups and the control group using a two-tailed Student’s t-test (Figure 4D). NMDA injection in combination with Lip C resulted in substantial RGC loss (~75% loss compared to the healthy controls). This observation is consistent with previous quantification of RGC loss in mice without empty liposomes.83 This result showed that Lip C did not have a neuroprotective effect. Treatment with either Lip A or Lip B was not able to completely prevent RGC loss.

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Figure 4. RGC rescue by liposomes after NMDA induced damage of mouse retinas. Example retina micrographs from (A) a healthy control eye; (B) a Lip A treated eye, and (C) a Lip C treated eye. In all micrographs RGCs nuclei were labeled with RBPMs Alexa647 (red), white scale bars are 60 μm. (D) Box and whiskers plot showing the density of surviving RGCs in healthy and liposome treated damaged eyes (*P ≤ 0.05 between Lip A and Lip C). Each dot shows an average RGC density for an eye, calculated from three micrographs taken 1 mm away from the optic nerve head. The box plot median values are 1728 RGCs/mm² for the healthy eye, 553 RGCs/mm² for Lip A, 380 RGCs/mm² for Lip B, and 360 RGCs/mm² for Lip C. N = 6–8 mice per group.
Figure 5. Electretinogram amplitude values from dark-adapted mice undergoing treatment with liposomes and transplant RGCs (tRGCs). (A) A-wave response for healthy and treated eyes. (B) B-wave response for healthy and treated eyes (*P ≤ 0.05 between Lip A + tRGCs and Lip C + tRGCs. *P ≤ 0.05 between Lip A and Lip C). (C) ΔSTR\textsubscript{OD} for liposome treated and liposome plus tRGC treated damaged eyes compared to healthy controls. Highly negative values (greater than −10 μV) were excluded from the ΔSTR assuming natural visual impairment in the control eye. One outlier was identified in the Lip A + tRGCs group and excluded from the statistical analysis (*P ≤ 0.05 between Lip A and Lip A + tRGCs. *P ≤ 0.05 between Lip A and Lip C + tRGCs. **P ≤ 0.01 between Lip A + tRGCs and Lip C + tRGCs). N = 8–10 mice per treated group, N = 17–44 mice for healthy control (panels A and B). N = 7–9 mice per treated group (panel C).

Figure 6. Host and transplant RGC survival. (A) Box and whiskers plot showing host RGC density in healthy eyes (median = 1728 cells/mm²) and NMDA damaged host eyes (median values for Lip A + tRGCs = 528 RGCs/mm², Lip C = 360 RGCs/mm², and Lip C + tRGCs = 415 RGCs/mm²) 4 weeks after NMDA injection (*P ≤ 0.05 between Lip A and Lip C). (B) Box and whiskers plot of tRGC survival in combination with liposomes (median values of 219 and 409 for Lip A and Lip C, respectively). (C) Example micrographs from the Lip A + tRGCs group. Transplant RGCs are in green, host RGCs in red (white arrows indicate axons, scale bar = 60 μm). The insert figure shows an example tRGC with axon (scale bar = 60 μm). (D) Example whole retina tile scan from the Lip A + tRGCs group used for cell counting. In all microscopy images tRGCs are in green (GFP) and host RGCs are in red (RBPMS). White circles highlight example areas containing tRGCs (scale bar = 1 mm, see Supplementary Figure S9 for larger image). N = 6–8 mice per group (panels A and B).

There was no significant difference in b-wave response between Lip A treated and Lip A in combination with tRGCs treated mice. This result indicated that the transplant RGCs (tRGCs) did not contribute to the b-wave response and that Lip A alone had a protective effect. This is corroborated with the b-wave response for TRGs combined with Lip C, which resulted in similar b-wave amplitudes to empty liposomes (i.e., Lip C) alone. The ΔSTR\textsubscript{OD} data (Figure 5C) showed that a combination of Lip A and tRGCs resulted in a significant rescue of RGC associated electrophysiological function (median = 3.3 μV) compared to Lip A treatment alone (median = 27.6 μV). Lip C in combination with tRGCs showed no rescue of STR, while Lip A in combination with tRGCs resulted in a significant improvement in ΔSTR\textsubscript{OD} compared to all other treatments. The ERG data does indicate that the loaded liposomes (Lip A) support tRGCs in a manner that improved host STR. The observation that empty liposomes (Lip C) in combination with tRGCs showed no rescue of STR supports this argument. Interestingly, Lip A in combination with tRGCs significantly reduced ΔSTR\textsubscript{OD} variance and eliminated the bimodal distribution seen in Lip A alone. These results indicate that both activation of cell growth signal pathways and the protection of other retina cell types (e.g., amacrine, bipolar, etc.) may be critical to transplant RGCs restoring host tissue function.

The effect of the combined treatment on host RGCs survival was evaluated by RBPMS staining and confocal microscopy (Figure 6). The host RGC density of the healthy control eyes and the treated eyes are shown in Figure 6A. We were not able to identify a significant additive effect of tRGCs on host RGC survival (522 cells/mm² for Lip A and 527 cells/mm² for Lip A + tRGCs). This indicated that it was Lip A alone that promoted host RGC survival. Next, we investigated the effect of the liposomes on the survival of tRGCs. Retinal whole mounts were double-stained for GFP (marker for tRGCs) and RBPMS at higher levels than the tRGCs resulting in limited colocalization of red and green fluorescence for the tRGCs. It is worth emphasizing that the low RBPM expression in tRGCs is likely due to the tRGCs not yet being fully mature (Supplementary Figure S7). tRGC survival was observed in both Lip A and Lip C groups (Figure 6 and Supplementary Figure S8) 4 weeks after transplantation, showing a longer survival time for tRGCs compared to previous reports on transplanted cells. Axon sprouting was observed in both Lip A and Lip C groups when combined with tRGCs (Figure 6C). Surviving tRGC numbers after 1 month were very low and no significant difference in survival of the transplanted cells was observed between Lip A and Lip C groups. Transplant cell survival is a major challenge in cell transplantation. Additionally, the NMDA induced apoptotic environment in the host eye will
have also induced tRGC apoptosis as NDMA induced toxic effects to retina cells has been shown to have a prolonged duration (up to 14 days). The nonspecific (i.e., not associated with intact cells) green fluorescence observed in the whole retina tile scans is likely GFP debris from the tRGCs combined with tissue autofluorescence (Figure 6D). Transplant cell survival has been shown to be improved by injury and loss of host RGCs. The rescue of host RGCs by Lip A may explain the lack of an observed effect of Lip A on tRGC survival. Another possible explanation could be linked to the timing and concentration of the delivered mTOR pathway promoters. Delivery of growth factors at appropriate concentrations is important for RGC survival in the neonatal retina and likely to be critically important in inducing a significant positive effect on the survival of transplanted cells.

**CONCLUSION**

Liposomes loaded with multiple mTOR pathway stimulating biologics showed a significant improvement in retina electrophysiological function after a single injection in an NDMA mouse model with extensive retina damage. Multifarious mTOR pathway stimulating biologics showed a significant improvement in STRGC survival. The results show that multifarious mTOR pathway stimulating biologics can facilitate both neuroprotection across a number of specific cell types and RGC transplantation. Further research, particularly in dosing dynamics (e.g., modulator concentrations, modulator combinations, number of injections, time between injections, etc.), is required to develop clinically viable neuroprotective and transplant facilitating nanomedicines.

**METHODS**

**Lipopeptide Synthesis.** All peptides were synthesized using a Biotage Initiator Alstra peptide synthesizer (Biotage) on a TentaGel S RAM resin (Sigma-Aldrich) at the 0.5 mmol scale using established solid phase methods (Supplementary Figure S1). All chemicals (e.g., amino acids, solvents, coupling agents, etc.) were purchased from Sigma-Aldrich or Bachem. Briefly, couplings were 5 min at 75 °C using 4 equiv amino acid, 3.92 equiv O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethylyuronium hexafluorophosphate (HATU), and 8 equiv 2,4,6-collidine in DMF. Fmoc deprotection was done using 20% piperidine in DMF. Peptides were conjugated to palmitic acid (C16) using 4 equiv amino acid, 3.92 equiv 1,1,3,3-tetramethyluronium hexafluoroacetic acid, and triisopropylsilane (95:2.5:2.5). In the case of PAP2 cleavage was performed using trifluoroacetic acid, water, triisopropylsilane and ethanedithiol (92.5:2.5:2.5:2.5).

Lipopeptides were precipitated in cold diethyl ether and filtered off. Crude lipopeptide was dissolved in a acetonitrile:water (1:1) solution and purified using preparatory HPLC (Waters) on a C18 column (Xterra). A water (5% acetonitrile, 1% trifluoroacetic acid) acetonitrile (0.1% trifluoroacetic acid) gradient was used starting at 10% acetonitrile and increasing to 60% acetonitrile over 25 min. All lipopeptides had a purity ≥95%. Lipopeptide molecular weight was confirmed using a autoflex MALDI-ToF MS (DHB, 0.1% TFA matrix, Bruker) and purity by analytical HPLC (C8 column, Gilson). The lipopeptides had the following sequences: (OPP) C6−PVDVDPGRDGSLALGLRSK; (PAP2) C6−KHKNHKYKYNLCAE; and (PAP4) C6−TVEEPSNPEAS−SSTSVPDP. Lipopeptide HPLC chromatograms and MALDI-ToF MS spectra can be found in the Supporting Information (Supplementary Figure S2).

**Preparation of Liposomes and Loading of Proteins.** All lipids were purchased from Avanti Polar Lipids and had a purity of ≥98%. Liposomes were prepared by dissolving pure lipids in 9:1 tertiary butanol/water and then pipetting them together to give the relevant lipid mixture. The lipid mixtures were then lyophilized to dry lipid powders overnight using freeze-drying. The lipid powders were subsequently hydrated over 1 h by adding 10 mM HEPES buffered saline pH = 7.4 (HBS) at 60 °C and shaking every 10 min. The liposomes were extruded through a 100 nm pore size filter 21 times at 60 °C and stored at 4 °C until use. Liposomes were composed of 1,2-dipalmitoyl-sn-glycero-3-phosphatidylcholine (DPPC), cholesterol (Chol), 1,2-dipalmitoyl-sn-glycero-3-phosphoethanolamine-N-[methoxy(polyethylene glycol)-2000] ammonium salt (DPPE-PEG2000), OPP, and PAP2 or PAP4. Liposome A (Lip A) comprised DPPC/Chol/DPPE-PEG2000/OPP and PAP2 in a 53:40:5:1 mol % ratio. In liposome B (Lip B), PAP2 was replaced with PAP4 at the same mol %, all other lipid percentages were the same. A control liposome (Lip C) without any proteins or lipopeptides had a composition of DPPC/Chol/DPPE-PEG2000 (55:40:5 mol %).

The liposomes were loaded into preformed liposomes following established methods. Briefly, CNTF and IGF-1 (recombinant human CNTF and recombinant human IGF-1, Cell Guidance Systems) were dissolved in 0.25 mg/mL and mixed with liposomes. The mixture was snap frozen in liquid N2 and thawed in a water bath at 37 °C, this process was repeated once. Nonecapsulated proteins were removed by spin-filtration with a 100 kDa spin filter (Merck), using HBS as eluent. The concentration of encapsulated protein was determined by ELISA (R&D Systems) following the instructions of the manufacturer. Lipid concentrations were detected by ICP-MS (Thermo Scientific). Encapsulation efficiency (EE%) of the CNTF and IGF-1 was determined by normalizing the protein concentration before and after purification to the actual lipid concentration, as described in eq 1.

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EE\% = \frac{[\text{protein}]_{\text{final}} - [\text{protein}]_{\text{start}}}{[\text{lipid}]_{\text{start}}} \times 100
\] (1)

Normalizing protein concentration to lipid concentration, rather than describing EE% as a change in protein concentration only, takes into account changes in volume from purification steps.

**Liposome Characterization by DLS and Zeta-Potential.** Liposome hydrodynamic diameter and zeta-potential were measured on a Brookhaven ZetaPALS. For size measurements liposomes were diluted in HBS. Zeta-potential was measured on liposomes diluted in 10 mM HEPES, 5% glucose at pH 7.4.

**In Vitro Liposome Stability.** Liposome stability was evaluated at 4 °C by following changes in size and polydispersity index (PDI) of loaded liposomes over time. Sizes and PDI were measured by DLS as described above. The ability of the liposomes to retain the loaded content was assessed by encapsulation of a self-quenching concentration (20 mM) of the hydrophilic fluorophore, calcine (Sigma-Aldrich). Liposomes were formulated as described above, adding 20 mM calcine in HBS for hydration of the lipid powder. Nonencapsulated calcine was removed by size exclusion chromatography (SEC) on a Sephadex G50 column using HBS. The calcine loaded liposomes were then split into different vials, kept at 4 or 37 °C. A 100 μL aliquot of each sample at a concentration of 0.1 mM was subsequently transferred to a black flat bottom 96 well plate, and the fluorescent signal was measured in a plate reader (Wallac Victor 1420 Multilabel Counter) before and after lysing the liposomes with 2 μL of 10% Triton X (Sigma-Aldrich). The ratio of the fluorescent signal of lysed liposomes over intact liposomes was then plotted against time to generate a leakage profile.

**Bioactivity assay of liposomes by Western Bolt.** To test the ability of Lip A and Lip B to influence the mTOR/Akt pathway, HEK293T (Sigma-Aldrich) cultured in Dulbecco’s modified eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS) (Sigma) and 5% penicillin–streptomycin in a six well plate was
incubated for 12 h with 0.4 mM liposomes at 37 °C 5% CO2. Cells incubated with Lip C were used as a control. Semi-quantitative analysis for Akt activation was performed by Western blot (WB) following standard protocol. In brief, cells were washed with cold PBS and collected by mechanical scraping in 200 μL of cold PBS, with phosphatase inhibitor cocktail 2 (Sigma-Aldrich) and complete Mini EDTA free PBS containing Protease inhibitor cocktail (Roche). Cells were transferred to clean 1.5 mL tubes kept on ice. The cells were lysed by adding 200 μL of 0.6 M (pH ~9.6) 4X protein loading buffer (Li-Cor) with 10% mercaptoethanol (Sigma-Aldrich), and the mixture was boiled for 5 min before loading 15 μL of the whole cell lysate in a 1.5 mm Nu PAGE 4–12% Bis-Tris Gel (Invitrogen). Five microliters of chameleon 700 prestained protein ladder (Li-Cor) was loaded in the outermost wells. Electrophoresis was run at 100 V in Nu PAGE running buffer (Invitrogen) for 2 h.

The proteins were transferred to a nitrocellulose membrane (Invitrogen) in 25 mM Tris, 190 mM glycine, and 20% methanol buffer at pH 8.3. The nitrocellulose membrane was blocked with Odyssey blocking buffer (Invitrogen) in 25 mM Tris, 190 mM glycine, and 20% methanol for 2 h. The antibody was then stripped using Revitablot Western blot stripping buffer (Rockland) before rebinding and staining overnight with anti-Akt1/2/3-Rb-IgG (Abcam) at 4 °C. The membrane was washed with TBST and incubated for 1 h with secondary stain Gt-anti-Rabbit IgG DyLight 800 (Invitrogen) before imaging the membrane on an Odyssey FC imaging system (Li-Cor). The antibody was then stripped using Revitablot Western blot stripping buffer (Rockland) before rebinding and staining overnight with anti-Phospho-p70S6K (Thr421, Ser424)-Rb-IgG (Abcam) at 4 °C. Akt/1/2/3 was labeled with IRDye 680RD Gt-anti-Rb (Li-Cor) and imaged. The same methodology was used for p70S6 kinase, except anti-p70S6K-Rb-IgG (Invitrogen) and anti-Phospho-p70S6K (Thr421, Ser424)-Rb-IgG (Invitrogen) were used. Densitometry was made by drawing rectangles over appropriate bands, obtaining the intensity, and dividing the intensity by the control (e.g., Akt/1/2/3 or p70S6K).

**Culture of Retinal Organoids.** Murine embryonic stem cells (mESCs) were thawed and seeded in T75 flasks precoated with 1% Matrigel in DMEM for 20 min. Cells were cultured in ESC maintenance medium (ES medium, see Supporting Information for all media components), at 37 °C, 5% O2, 5% CO2, until 80% confluence. Cells were then collected washed with HBBS and dissociated with freshly activated papain for 3 min, vortexing thoroughly to mechanically disso-

**Liposome Uptake in Retinal Organoids.** For in vitro uptake studies, liposomes with the same membrane composition as Lip A, Lip B, and Lip C were prepared adding 0.5 mol % of the phospholipids to a well and incubating for 12 h with the mESC organoids. Liposomes with the same membrane composition as Lip A, Lip B, or empty Lip C, were collected washed with HBBS and incubated for 0.0001, 0.001, 0.01, 0.1, and 5 cd-s/m2 intensities with a 30 s recovery in between. The cell suspension in PBS. Four weeks after transplantation RGCs were intravitreally injected of 2 μL of cell suspension in PBS. Four weeks after transplantation ERGs were recorded, mice were euthanized, and eyes were collected.

**Electroretinography.** The function of retinal ganglion cells was assessed by electroretinography (ERG)-scotopic threshold response (STR) using a Diagnosys Espion 3 system with Ganzfeld bowl. C57BL/6j mice (Charles River Laboratories) were dark-adapted overnight prior to recordings. The mice were anesthetized by IP injection of 100–200 mg/kg ketamine and 20 mg/kg xylazine and placed on a heating pad to maintain temperature throughout the recordings. Pupils were dilated in both eyes by one drop of tropicamide. GelTal (hypromellose) was applied to the eyes to ensure conduction to the electrodes. The reference electrode (needle) was placed subcutaneously in the forehead and measuring electrodes (gold loops) were placed on the corneas. For the stimulation, a series of white flashes were used at 0.0001, 0.001, 0.01, 0.1, and 5 cd-s/m2 intensities with mice exposed to 5 flashes per intensity with a 30 s recovery in between each flash. Electroretinograms were recorded with positive STR, negative STR, for which a- and b-waves were analyzed. The STR data shown in this paper were obtained under 0.001 cd-s/m2 flash intensities while the a-wave and b-wave data were obtained for 0.1 cd-s/m2. These flash intensities produced electroretinograms with the best signal-to-noise ratio.

**Immunohistochemistry and Confocal Imaging.** Eyes from euthanized mice were collected in PBS and fixed in 4% PFA overnight.
Retinas were carefully dissected under a microscope. Retinas were blocked with 10% goat serum overnight at room temperature. Retinas were then washed with 0.1% triton × 0.1% tween 20 in PBS three times before incubation with anti-RBPSM-Rb-IgG (Abcam) and anti-GFP-m-iG (Abcam) overnight at 4 °C. After they were washed three times as described above, the retinas were incubated with antimouse-Alexa488 and antibiot-Alexa488 1 h at room temperature. Lastly the retinas were washed, incubated with 0.5 μg/mL DAPI in PBS for 1 min at room temperature, and washed again before mounting on glass slides using 9% poly vinyl-alcohol, 22% glycerol, 2% 1,4-diazabicyclo[2.2.2]octane in 88 mM Tris-HCl, pH 8.5.

Slides were imaged on a Leica TCS SP5 confocal microscope using a 40× oil emission objective. The predefined settings for the dyes used were chosen in the Leica software. Sequential scanning was used to avoid spillover between the DAPI channel and the Alexa488 channel. Retinas were imaged approximately 1 mm from the optic nerve head. To quantify surviving host RGS, z-stacks of approximately 16 μm with a step size of 0.5 μm were collected. Stacks were z-projected to form one image in ImageJ. 2–3 images were obtained per retina. Surviving RGCs were counted manually. To evaluate survival of transplanted RGCs that might be heterogeneous distributed across the retina, the whole retina was imaged using a Zeiss Axio Scan Z.1 slide scanner (20× objective). Signals were recorded in the 488, 560, and 647 nm channels. To eliminate some of the green autofluorescence from the retina, the signal from the 560 nm channel was subtracted from the 488 nm channel before counting the green cells in Zeiss Zen Blue and Black lite software. Cells were counted manually.

**Statistical Analysis.** Statistical analysis was performed using GraphPad Prism 7 software. The following statistical methods were used to analyze the biological data: (Figure 2A) one-way ANOVA post hoc Tukey HSD test; (Figure 2B) two-way ANOVA; (Figure 3A) one-way ANOVA, no significance; (Figure 3B) one-way ANOVA post hoc Tukey HSD test; (Figure 3C) Kruskal–Wallis, no significance; (Figure 4D) one-way ANOVA post hoc Tukey HSD; (Figure 5A) one-way ANOVA, no significance; (Figure 5B) one-way ANOVA post hoc Tukey HSD test; (Figure 5C) Kruskal–Wallis post hoc Dunn’s test; (Figure 6A) One-way ANOVA post hoc Tukey HSD; (Figure 6B) Unpaired t test, no significance. The threshold of statistical significance (alpha) for all analyses was 0.05. Distribution curves and quantile–quantile (Q–Q) plots were used to determine whether data sets were parametric or nonparametric. Statistical significance between healthy control eyes and treated eyes was significant and not shown for clarity.

**ASSOCIATED CONTENT**

Supporting Information
The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsnano.8b00596.

Lipopeptide mass spectra and chromatography traces; calcine leakage from liposomes; liposome bioactivity Western blots; flow cytometry scatter plots; electrophoretograms; retina micrograms, and cell culture media compositions (PDF)

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P.B., M.Y., A.Z.E., and A.J.U. conceived the concept and designed the experiments. A.Z.E., P.K., T.L.A., and A.J.U. manufactured liposomes and performed all liposome related analysis. R.E. and A.J.U. synthesized and analyzed lipopeptides. A.Z.E., J.O., F.M., P.B., and M.Y. performed cell culture experiments and cell analysis. A.Z.E., J.O., P.B., and M.Y. performed in vivo studies, immunohistochemistry and ERG analysis. A.Z.E. and A.J.U. wrote the manuscript with contributions of all authors. All authors have given approval to the final version of this manuscript.

**Notes**

The authors declare no competing financial interest.

**ACKNOWLEDGMENTS**

We would like to thank the Velux Foundation (A.Z.E. and A.J.U.) and Shore Fellowship (P.B.) for financial support. The Animal core and Confocal Imaging facility at the Schepps Eye Research Institute was supported by NIH National Eye Institute Core Grant P30EYE003790. P.B. and M.Y. would like to thank the Laboratory of Joshua Sanes (Harvard University) for kindly providing the Thy1-GFP iPSCs.

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DOI: 10.1021/acsnano.8b00596
ACS Nano 2018, 12, 7497–7508


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