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Abundance and diversity of the fecal resistome in slaughter pigs and broilers in nine European countries

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1 **Abstract**

2 Antimicrobial resistance (AMR) in bacteria and associated human morbidity and mortality is increasing. Use
3 of antimicrobials in livestock selects for AMR that can subsequently be transferred to humans. This flow of
4 AMR between reservoirs demands surveillance in livestock as well as in humans. As part of the EFFORT
5 project (www.effort-against-amr.eu), we have quantified and characterized the acquired resistance gene
6 pools (resistomes) of 181 pig and 178 poultry farms from nine European countries, generating more than
7 5,000 gigabases of DNA sequence, using shotgun metagenomics. We quantified acquired AMR using the
8 ResFinder database and a database constructed for this study, consisting of AMR genes identified through
9 screening environmental DNA. The pig and poultry resistomes were very different in abundance and
10 composition. There was a significant country effect on the resistomes, more so in pigs than poultry. We
11 found higher AMR loads in pigs, while poultry resistomes were more diverse. We detected several recently
12 described, critical AMR genes, including *mcr-1* and *optrA*, the abundance of which differed both between
13 host species and countries. We found that the total acquired AMR level, was associated with the overall
14 country-specific antimicrobial usage in livestock and that countries with comparable usage patterns had
15 similar resistomes. Novel, functionally-determined AMR genes were, however, not associated with total
16 drug use.

17

18 **Introduction**

19 Antimicrobial resistance (AMR) is considered one of the largest threats to human health.¹ In addition to the
20 use of antimicrobial agents for humans, livestock is considered an important source of AMR, potentially
21 compromising human health.² Besides AMR in zoonotic pathogens, AMR in commensal bacteria is
22 worrisome because of its ability to spread horizontally to pathogens.

23 Multiple studies have shown that use of antimicrobials in livestock will lead to increased occurrence of AMR
24 and that reduction of usage will eventually lead to reduced resistance.³⁻⁸ A number of national surveillance
25 programs have been implemented to monitor the occurrence of AMR in different reservoirs and follow
26 trends over time.^{1,9-11} There are major differences in antimicrobial consumption patterns between different
27 countries globally and also within Europe.¹² Major differences in the occurrence of AMR have also been
28 observed among indicator organisms (e.g. *E. coli*) isolated from different European countries.^{3,13} Current
29 monitoring efforts are mainly based on culturing indicator bacteria followed by phenotypic AMR
30 determination.^{13,14} This procedure only targets a limited number of species present in the gut microbiota

31 and thus likely represents only a fraction of its resistome (the collective pool of AMR genes). Such
32 metagenomic approaches have been used in a number of recent studies and it has been shown that
33 metagenomic read mapping describes AMR abundance in bacterial communities more accurately than
34 commonly used technologies on selected indicator organisms.¹⁵⁻¹⁷ A recent study focused on sampling a
35 diverse group of individual pigs from eleven farms in three countries showed that genetics, age, diet and
36 country all likely influence the pig microbiota, but little information is available for poultry.¹⁶

37 As part of the EU-funded EFFORT project (www.effort-against-amr.eu), we sampled over 9000 animals in
38 181 pig and 178 poultry herds in nine European countries, generating herd-level composite samples as
39 previously described.¹⁷ This gives us a unique insight into the diversity and structure of the acquired pig and
40 broiler resistomes across Europe. We sampled animals as close as possible to slaughter to elucidate the
41 potential consumer exposure to AMR associated with meat production in Europe. Association between
42 AMR gene abundance and country level antimicrobial usage was analyzed. We hereby provide an overview
43 of AMR in the two most intensively raised European livestock species. To our knowledge, this study
44 represents the single largest metagenomic AMR monitoring effort of livestock: both in terms of countries
45 (9), herds included (359), individual animals sampled (over 9,000) and sequencing effort (5,000+ gigabases).

46

47 **Methods**

48 **Farm selection and sampling**

49 The sampling protocol for pig and broiler farms that has been agreed on by the EFFORT consortium is
50 described below. Selection of farms and sampling procedure followed these guidelines to the extent
51 possible, but some deviations from the protocol were occasionally necessary. A detailed description of the
52 sampling conducted in the individual countries is provided in supplementary material.

53 **Selection of pig and poultry farms**

54 In each participating country, 20 conventional integrated pig farrow-to-finisher non-mixed farms were
55 selected. The farms needed to have a minimum of 150 sows and 600 fatteners and employ batch
56 production to ensure that the majority of the animals of the sampled group originated from the same birth
57 cohort. All-in all-out production at compartment level was preferred and all fatteners sampled were
58 required to have been on the same site during their entire life. Selected farms should have no contact
59 through livestock trade, and have a random regional distribution.

60 In each country, 20 conventional broiler farms (no breeders) were selected. The farms should have all-in all-
61 out production, with a thinning procedure from day 30 onwards allowed. All selected farms should have no
62 intended slaughter age higher than 50 days, no slow growing breeds (intended growth rate less than 55
63 gram/day) and no stocking density lower than 10 birds/m². Only one flock/house per holding should be
64 sampled and the flock should be between 20,000 and 40,000 birds. If possible, selected farms should have
65 a random regional distribution.

66 **Procedure for sampling**

67 We sampled pig farms between May 2014 and December 2015, and tried to minimize seasonal influences.
68 The sampled fatteners should be as close to slaughter as possible (i.e. within the last week). A total of 25
69 fresh, still warm and undisturbed fecal droppings were sampled from pen floors (a minimum of 10 g of
70 feces per sample) randomly divided over all eligible compartments/stables of fatteners close to slaughter.

71 Broilers were sampled between May 2014 and June 2016, and we tried to minimize seasonal influences. On
72 each farm, 25 undisturbed, fresh main bowel droppings were collected from the floor of the house (a
73 minimum of 3 g feces per sample). The flocks should be sampled as close to slaughter as possible (last week
74 before the final depopulation).

75 All samples were collected aseptically in plastic containers and were stored at 4°C and transported to the
76 laboratory within 24 hours after sampling.

77 **Pooling and handling of samples**

78 Upon arrival in the laboratory, individual fecal samples were homogenized by stirring thoroughly with a
79 sterile tongue depressor/ spoon for a few minutes. From each pig sample, two 2 ml cryotubes were filled
80 and frozen immediately at -80°C (alternatively at -20°C for maximum 4 days, before transferring to -80°C).
81 For broiler samples, at least 0.5 g feces was added to two cryotubes. Sample pooling was either done
82 immediately or the frozen tubes were shipped to the Technical University of Denmark (DTU) on dry ice for
83 pooling. Individual samples from the same herd were defrosted and placed on ice briefly before weighing.
84 Following weighing, they were pooled with 0.5 g of feces from each sample and stirred for a few min with a
85 sterile device (e.g. disposable wooden tong depressor). All samples were only thawed once shortly before
86 DNA extraction.

87 After removing two mis-labeled samples, we ended up with composite fecal samples from 178 broiler
88 flocks and 181 pig herds.

89 **Sampling to estimate the effect of random sampling**

90 To study the potential effect of sampling randomness and the reproducibility of our sampling protocol, two
91 of the pig herds were chosen for triplicate sampling. These two herds were sampled three times on the
92 same day (25 samples x 3 sampling rounds), resulting in six pooled samples (2 herds x 3 sampling rounds),
93 from which the within-farm variation was assessed. A table with all samples and their metadata is included
94 as Supplementary Table 1.

95 **DNA extraction and sequencing**

96 From each of the pooled, herd-level fecal samples, DNA was extracted using a modified QIAamp Fast DNA
97 stool mini kit protocol (Qiagen, cat. no. 51604), as previously described.¹⁸ One major modification is the
98 addition of a bead beating step in the beginning of DNA extraction. The protocol can be found at
99 https://figshare.com/articles/SOP_-_DNA_Isolation_QIAamp_Fast_DNA_Stool_Modified/3475406. DNA-
100 purification of all pooled samples was processed centrally at Technical University of Denmark (DTU), and
101 the DNA was stored in duplicates at -20°C until further use.

102 DNA was shipped on dry ice for library preparation and sequencing at the Oklahoma Medical Research
103 Foundation (OMRF). There, DNA from all samples was mechanically sheared to a targeted fragment size of
104 300bp using ultrasonication (Covaris E220evolution). For pooled pig samples, library preparation was
105 performed with the NEXTflex PCR-free library preparation kit (Bioo Scientific). For poultry samples, due to a
106 lower DNA availability, the minimal amplification-based KAPA Hyper kit (Kapa Biosystems) was used. For all
107 samples, the Bioo NEXTflex-96 adapter set (Bioo Scientific) was used. In batches of roughly sixty samples,
108 the libraries were multiplexed and sequenced on the HiSeq3000 platform (Illumina), using 2x150bp paired-
109 end sequencing per flow cell. A total of 17 Belgian, Danish and Dutch pig fecal samples were sequenced on
110 the HiSeq2500 platform (Illumina), using 2x100bp paired-end sequencing, before converting to HiSeq3000
111 for the remaining samples (See Supplementary Table 1).

112 **Bioinformatics processing**

113 The FASTQ files with sequencing data for each sample were analyzed following the principles from the
114 previously described MGmapper tool.¹⁵ The reads were first cleaned by cleaning out adaptors using BBduk
115 (BBMap v39.92 - Bushnell B. -<https://sourceforge.net/projects/bbmap/>), and by removing reads that
116 aligned to the internal sequencing control phi-X174 as determined by the BWA-MEM algorithm.¹⁹ Trimmed
117 read pairs were aligned using the BWA-MEM algorithm Prokaryotic RefSeq genomes from NCBI GenBank
118 (downloaded on Nov. 18, 2016) with the “reference” or “representative” genome tags. The BWA-MEM
119 algorithm (v0.7.15) was slightly modified such that the insert size was estimated for the entire sample
120 unaffected by the number of CPU cores used. This estimate was then used for re-analyzing the cleaned

121 read pairs using BWA-MEM, ensuring that the thresholds for a read pair to map as a 'proper pair' were the
122 same for all read pairs, and avoiding bad insert size estimates when few read pairs aligned.

123 The read pairs were aligned to the bacterial NCBI genomes again, and to the AMR genes present in the
124 ResFinder database (accessed Nov. 17, 2016).²⁰ ResFinder is a manually curated database of horizontally
125 acquired AMR genes and thus does not include intrinsic AMR genes and mutated housekeeping genes
126 providing AMR by changing the drug target. Technical duplicate read pairs were then removed using
127 'MarkDuplicates' from the Picard command line tools (v2.8.3; <http://broadinstitute.github.io/picard/>).

128 For each ResFinder reference sequence, we counted the number of read pairs that properly aligned with at
129 least 50 bp aligning from both the forward and reverse reads. Each read pair matching a ResFinder
130 reference was assigned to the first highest-scoring reference, as done in MGmapper. The same was done
131 for NCBI microbial genomes in order to quantify the bacteriome and get a measure of the microbial
132 proportion within each sample. This total was used to normalize the ResFinder counts, by computing an
133 FPKM-value (fragments per kilobase reference per million bacterial fragments) for each ResFinder
134 reference sequence. The FPKM values were computed by dividing the mapping count on each reference
135 with its gene length and the total number of bacterial read pairs for the samples and multiplying by 10^9 .²¹
136 Raw mapping count data and their associated FPKM values can be found in Supplementary Table 2 and 3.

137 Because ResFinder contains many representatives of certain gene families, a high degree of homology
138 exists, with long stretches of the references being identical. This causes unspecific mapping between high-
139 identity sequences. To eliminate this random noise, we chose to aggregate read counts and relative
140 abundances post-mapping at higher levels based on sequence identity. We clustered all the ResFinder
141 genes using CD-HIT-EST (v4.6.6) at a 90% identity level and otherwise default settings.²² The resulting gene
142 clusters were manually inspected and named to reflect their contents while avoiding conflicts with other
143 clusters (Supplementary Table 4). Abundances were aggregated according to these clusters of high-identity
144 genes and the resistance-class-level as annotated in ResFinder. These two levels, "gene" and "class", were
145 used for all downstream analysis.

146 **Functional resistance database**

147 Previous studies have identified a wide array of novel AMR genes in various reservoirs using functional
148 metagenomics, referred to as functional AMR genes.^{23–26} By cloning random DNA fragments from complex
149 microbiomes into an expression vector expressed in a host (typically *E. coli*) and selecting for growth in the
150 presence of certain antibiotics, they have been found to provide AMR to many antibiotics.^{23–26} We

151 constructed a functional resistance database (FRD) from 3,416 AMR gene variants identified in four major
152 studies, using 23 different antimicrobials for selection.^{23–26}

153 Briefly, in each of these studies, DNA was extracted from environmental and human fecal samples,
154 fragmented and cloned into a plasmid vector and screened for AMR functionality in *E. coli* cultured with
155 one of multiple antimicrobials. AMR-granting plasmid inserts were then sequenced and the responsible
156 open reading frame was identified. The protocol for the database construction can be found at
157 <https://cge.cbs.dtu.dk/services/ResFinderFG/>. Genes were quantified using MGmapper as was done for
158 ResFinder. Genes with more than 90% identity to ResFinder genes were removed post mapping to obtain
159 the new AMR genes without overlap with ResFinder. The resulting data was aggregated to 90% gene
160 clusters, using CD-HIT-EST, as was done for ResFinder.²² The most frequent gene clusters remaining were
161 derived from genes selected using: trimethoprim, chloramphenicol, co-trimoxazole, cycloserine, amoxicillin,
162 gentamicin, penicillin and tetracycline.

163 **Principal coordinate analysis and resistome clustering**

164 For principal coordinate analysis (PCoA), the gene-cluster level FPKM matrix was Hellinger-transformed and
165 the Bray-Curtis (BC) dissimilarities between all samples were calculated using the R package *vegan*.²⁷ PCoA
166 was carried out for both pigs and poultry; combined and separately, using the *vegan* function ‘betadisper’.
167 The same analysis was used to test whether host animal and country were significant predictors of within-
168 group beta diversity dispersion. The effects of country on sample dissimilarities was determined using
169 ‘permutational multivariate analysis of variance using distance matrices’ (‘adonis2’ function in *vegan*
170 package), separately for pig and poultry.

171 **Antimicrobial use in livestock**

172 Data for national livestock antimicrobial usage (AMU) was obtained from the European Medicines Agency’s
173 2014 European Surveillance of Veterinary Antimicrobial Consumption (ESVAC) report and was stratified by
174 major drug family.²⁸ The mass of active compound sold for use in animals in 2014 was divided by the
175 Population Correction Unit (PCU) in 10⁶ kg - approximating the biomass. The PCU is a unit that allows inter-
176 species integration by adjusting for import/export and differences in average weight between species when
177 they are most likely to receive antimicrobial treatment. The estimate was multiplied by 1000 to obtain drug
178 mg/kg PCU livestock. The country-specific veterinary drug use can be found in Supplementary Table 5.

179 **Procrustes analyses**

180 In order to test the association between country-specific AMU patterns and the resistomes, we performed
181 Procrustes analysis using the *vegan* R package as follows. A PCoA was generated from Euclidean distances

182 between the samples in the PCU-corrected AMU (Supplementary Table 5, Supplementary Figure 1). The
183 AMU PCoA was tested against the previously mentioned Hellinger-transformed, resistome BC dissimilarity
184 PCoA using the 'protest' function with the default 999 permutations, again separately for pigs and poultry.
185 The gene-cluster FPKM ResFinder matrix and the genus-level FPKM taxonomy matrix were Hellinger
186 transformed and BC dissimilarities were calculated. They were ordinated using non-metric
187 multidimensional scaling (NMDS) with the 'metaMDS' *vegan* function (999 permutations) for pig and
188 poultry samples separately. The symmetric Procrustes correlation coefficients between the bacteriome and
189 resistome ordinations, p-values and plots were obtained using the 'protest' and 'procrustes' functions in
190 *vegan*.²⁹

191 **Alpha diversity**

192 For all samples, we computed the within-herd resistome diversity using Simpson diversity index (1-D),
193 Chao1 richness estimate and Pielou's evenness.³⁰ The raw read count matrix was rarified to 10,000 hits for
194 all samples for alpha diversity estimation, leading to the exclusion of 10 samples.

195 **Visualization**

196 Heatmaps were produced using the *heatmap* R package. For heatmaps showing individual gene
197 abundances, the BC dissimilarities between samples were used. For all other dendrograms, the Pearson
198 product-moment correlation coefficients (PPMCC) were used. Complete-linkage clustering was used for all
199 dendrogram clustering. For sample similarities, BC dissimilarity was converted to a similarity percentage,
200 i.e., $100*(1- BC)$.

201 The circular BC sample dendrogram was exported in Newick format using the *ape* package and further
202 annotated with the Interactive Tree of Life tool.^{31,32} Bar-, box- and scatter plots were produced using the
203 *ggplot2* R library.³³ The R library *RcolorBrewer* was used to generate the color palettes used. The library is
204 based on work by Cynthia A. Brewer (www.ColorBrewer.org).

205 **Statistical analyses**

206 All statistics was done in Microsoft R Open (MRO) 3.3.2, using the libraries and procedures detailed below.
207 Exact package versions can be found here: <https://mran.revolutionanalytics.com/snapshot/2016-11-01/bin/windows/contrib/3.3/>. For statistical tests, only the first sampling from triple-sampled herds was
208 included (see Supplementary Table 1). Unless otherwise mentioned, all statistical analyses were performed
209 on pigs and poultry separately.
210

211 **Effect of AMU on total AMR**

212 For testing the effect of total AMU on total metagenomic AMR abundance (sum of all genes), we used the
213 *lme4* 1.1-12 package to make linear mixed effects regression models with total livestock drug usage as the
214 independent variable, total AMR abundance (FPKM) as the dependent variable and country as a mixed
215 effect intercept, adjusting for the fact that AMR abundance observed in farms from the same country is
216 correlated due to factors that are not included in this study.³⁴ The effect of AMU was modelled on a
217 logarithmic scale, which resulted in lower Akaike's information criteria compared with modelling AMU on a
218 linear scale. Country- and sample-level residuals were plotted and inspected for normality and
219 homoscedasticity. Pig sample residuals and country residuals were normal and so were poultry country
220 residuals. Poultry sample residuals had a longer right tail. Square-root transforming the poultry AMR data,
221 gave more normal residuals and a similar conclusion ($p < 0.05$). The effect and significance of drug usage was
222 assessed using likelihood-ratio tests, comparing the random effect models with and without the AMU
223 effect.

224 **Differential abundance analysis**

225 To identify differentially abundant AMR genes per country, we analyzed the read pair mapping count
226 matrix using the *DESeq2* package as previously recommended for metagenomic read count data.^{35,36} This
227 was done on the raw read pair matrix, following recommendations that rarefying is not warranted in
228 metagenomic studies.³⁶ The read-pair count matrices for pigs and poultry were analyzed separately. The
229 number of mapped bacterial pairs was divided by the minimum number of mapped bacterial pairs and was
230 used as the size factor. For each gene, we used a two-sided Wald test to determine whether the fold
231 change between countries differed from 0 and extracted all the country-versus-country results. P-values
232 were adjusted for the false discovery rate (FDR) using the Benjamini-Hochberg approach and we used a
233 significance threshold of $\alpha: 0.05$.³⁷

234 **Core resistome**

235 The core resistomes determined here, were the set of AMR gene clusters with mapping read pairs in at
236 least 95% of the samples. The core resistomes were determined separately for the pig and poultry
237 reservoirs.

238

239 **Results**

240 In total, DNA from 365 pooled samples was extracted and shotgun sequenced, resulting in more than 36
241 billion sequences (18 billion paired-end (PE) reads), comprising more than 5 terabases of DNA. This yielded

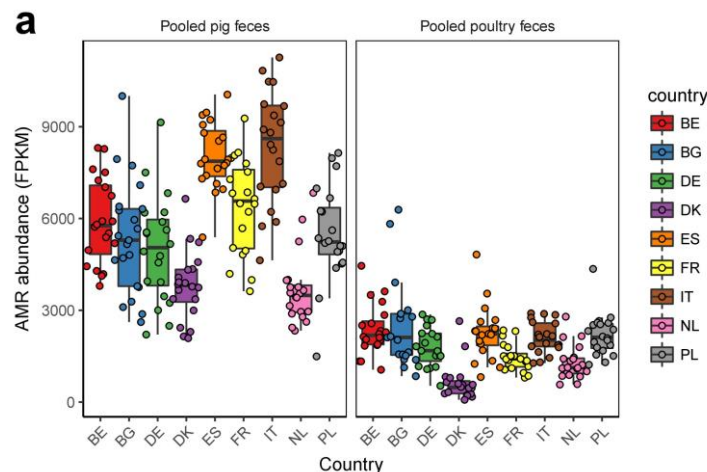
242 an average of 50 million (SD: 18×10^6) PE reads per pooled sample. This was similar for pig and poultry
243 samples, though the former varied more than the latter.

244

245 **Acquired resistome characterization**

246 The total AMR gene level varied significantly across samples, both depending on host animal and country of
247 origin. In general, pigs had a higher AMR level than poultry (Figure 1a). The highest AMR levels were found
248 in Italian pigs from where the top four resistance-scoring samples originated, all in excess of 10,000 FPKM
249 AMR. At the lower end of the spectrum were Danish poultry samples that occupied the eleven samples
250 with least AMR, all below 500 FPKM.

251



252



253

254 **Figure 1. Overview of AMR abundance and composition.** From the read mapping results to the ResFinder database, AMR
255 abundance was calculated for each reference gene in each sample. (a) Boxplots showing the total AMR level per sample,
256 stratified by host species and country. Each herd is also represented by a dot with sideways jitter to minimize overplotting.
257 Horizontal box lines represent Q1, median and Q3. Whiskers denote range of points within $Q1-1.5*IQR$ and $Q3+1.5*IQR$. (b)
258 Stacked bar chart of AMR abundance per type (colors) per sample (x-axis), proportional to total AMR within each sample.

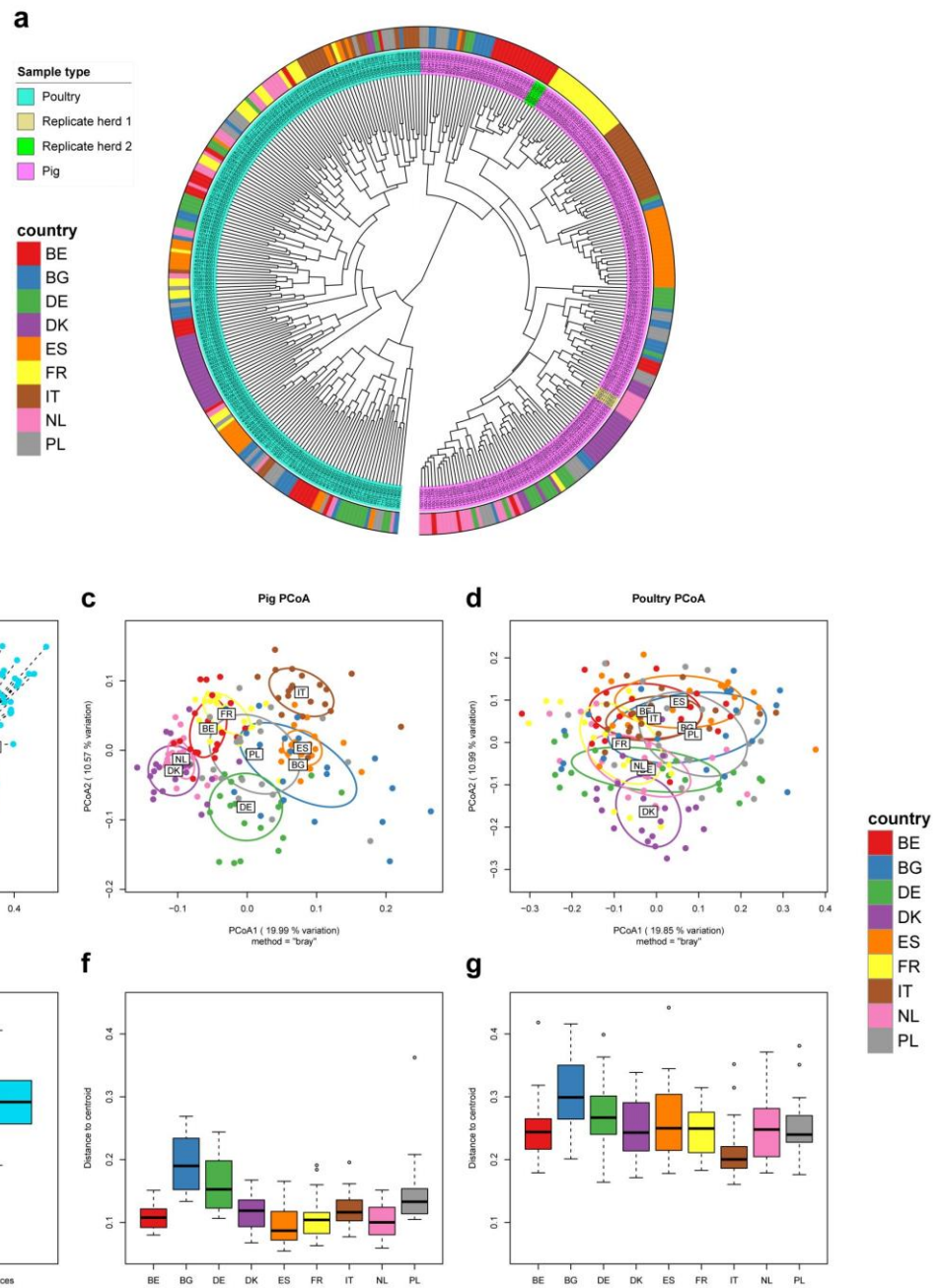
259 We summed the relative abundance of AMR to the corresponding drug class level for each sample to look
260 for major trends across species and countries (Figure 1b). When considering the proportion of the total
261 resistome that each AMR type takes up, the pig samples were relatively homogenous: tetracycline AMR
262 was by far the most common, followed by macrolide AMR. Beta-lactam and aminoglycoside AMR genes
263 followed with other kinds of AMR being rare. Italian pigs had a notably larger proportion of phenicol AMR
264 compared with other countries and it seemed to be consistent across all Italian farms. A subset of Bulgarian
265 pig farms had a similar proportion of phenicol AMR.

266 Among the poultry farms, there was less consistency: both within and between countries, the relative
267 proportions of AMR per drug class varied more. Tetracycline, macrolide, beta-lactam and aminoglycoside
268 AMR made up the majority of AMR, as in pig samples, but the two latter classes had very minimal
269 contributions in a subset of herds. Sulfonamide and trimethoprim AMR were more abundant in poultry
270 samples compared with pig samples, across all countries. In many Polish poultry herds, quinolone AMR
271 made up a sizeable fraction of the combined resistome. This was also true for a few non-Polish herds,
272 notably in Bulgaria. For non-proportional graphical representations of the AMR load stratified by sample
273 and drug class, see the supplementary material for an unscaled, stacked bar chart (Supplementary Figure 2)
274 and a heatmap (Supplementary Figure 3). Class level AMR relative abundances can be found in
275 Supplementary Table 6.

276 To characterize the individual components of the resistome, we summed relative abundance to the gene-
277 cluster level as we had done at drug class level. We found evidence for 407 different gene clusters across all
278 pig and poultry samples (Supplementary Table 2).

279 We calculated the BC dissimilarities between all samples' gene-level resistomes and visualized it in a
280 dendrogram (Figure 2a). There was a perfect host separation, with all pig samples clustering separately
281 from all poultry, suggesting pig and poultry resistomes are very different. Within the pig cluster, the
282 country separation was more pronounced than for poultry. An exception was Danish poultry, where 18/20
283 samples clustered.

284 To assess the reproducibility of our protocol, from sampling through sequencing, we evaluated the
285 similarities between resistomes of two triple-sampled swine herds. Dutch triple-sampled herd pools had
286 the highest similarities of all samples, ranging from 93.6% - 93.7% BC similarity. The Belgian triple-sampled
287 herd pools had values ranging from 91.5% - 93.3% similarity. No replicated sample pool had higher
288 similarity to other herds than to its own replicates and the two sets of three samples can thus be seen
289 clustering as expected (Figure 2a). A sample similarity heatmap is found in Supplementary Figure 4.



290

291

292 **Figure 2. Resistome clustering is influenced by both host animal and country. (a) Dendrogram showing complete linkage**
 293 **clustering of BC dissimilarities between all pig and poultry resistomes. Triple-sampled pig herds are highlighted in separate**
 294 **colors. (b-d) PCoA plots for pig and poultry samples combined (b), pig samples (c) and poultry samples (d), respectively. Ellipses**
 295 **denote standard deviation for distance of each member to its group centroid (labeled). (e-g) Boxplots of distances for each**
 296 **group's samples to its centroid for pig and poultry samples combined (e), pig samples (f) and poultry samples (g), respectively.**
 297 **Horizontal box lines represent Q1, median and Q3. Whiskers denote range of points within $Q1-1.5 \cdot IQR$ and $Q3+1.5 \cdot IQR$.**

298

299 We ordinated the gene-level resistomes for all samples (Figure 2b) and pig and poultry samples separately
300 (Figure 2cd). As with hierarchical clustering, there was a clear separation of pig and poultry samples, along
301 the first principal coordinate, which explained 48% of the variation across all resistomes.

302 When analyzing the two reservoirs separately, we observed clustering according to country of origin in pigs
303 (Figure 2c), while clustering was more diffuse for poultry (Figure 2d). We tested for the country effect and
304 found it to be significant in both pigs (adonis2 $p < 0.001$) and poultry (adonis2 $p < 0.001$). In poultry however,
305 the country effect only explained roughly a quarter of the variation, while country explained roughly half of
306 the variation in pigs (data not shown). In the pig resistome ordination, the Danish and Dutch samples
307 clustered closely together. The same could be seen for French and Belgian resistomes and to a lesser
308 degree, Italian and Spanish samples. Bulgaria, Germany and Poland showed larger dispersions than the
309 other countries. Poultry samples had a higher dispersion than pig samples (Figure 2e, beta-dispersion
310 $p < 0.001$). Beta-dispersion levels varied significantly between countries in both pigs (Figure 2f, beta-
311 dispersion $p < 0.001$) and poultry (Figure 2g, beta-dispersion $p < 0.001$).

312 We visualized the AMR gene abundances in a heatmap to look at the overall structure and composition of
313 the resistomes and the co-occurrence of AMR genes (Supplementary Figure 5). Some AMR genes were
314 more abundant in one species, while others, including *tet(W)* and *erm(B)* were ubiquitous in all samples, for
315 both species. Among the pig samples, the Italian samples stood out: several chloramphenicol AMR genes,
316 including *cat(pC194)*, *catP*, and *cat_2*, were much more abundant in Italy, compared to the other countries,
317 consistent with our inspection of AMR at class level (Figure 1). Several AMR genes known to be co-located
318 indeed co-occurred across samples. The genes in the vancomycin AMR *VanA* cassette were co-located in a
319 number of poultry samples. This was also true for the *VanB* cassette members, clustering together, but
320 separately from *VanA*, showing ability to distinguish variants of homologous genes. As earlier indicated, the
321 poultry samples showed less country-based clustering than pigs. An exception were the Danish poultry
322 samples. These had noticeably lower abundance of many AMR genes that were widespread in other
323 countries.

324 **Core resistome**

325 To determine whether specific genes were unique to each of the host animals, we examined the set of AMR
326 genes that was consistently observed within each animal species (evidence for it in 95% of samples). We
327 identified 33 core AMR genes in pigs and 49 core AMR genes in poultry, with 24 being shared between the
328 two hosts (Supplementary Figure 6). Hence, only nine AMR genes were pig-core genes without also being
329 poultry-core genes. These included the genes making up the Van-G vancomycin cassette, *tet(C)*, *bla_{ACI}* and

330 *cfxA*. Twenty- five AMR genes were poultry-core genes without also being pig-core genes and include the
331 Enterobacteriaceae-associated *strAB*, *sul2*, *blaTEM* and *tet(A)* genes.

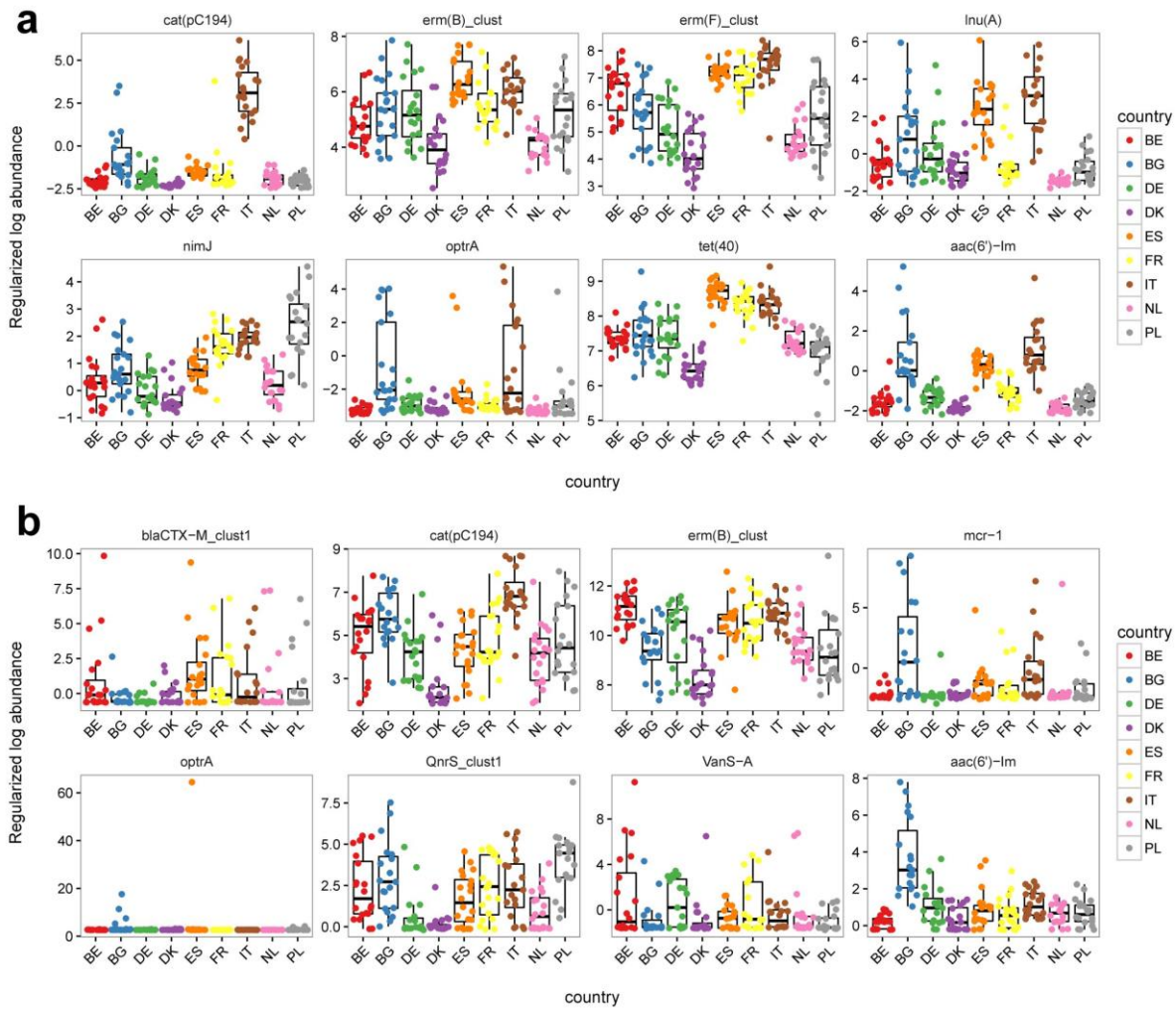
332

333 **Differential abundance analysis**

334 In order to test which specific genes differed in abundance between countries, we carried out a differential
335 abundance analysis for ResFinder gene read pair counts. Heavy overrepresentation of low unadjusted p-
336 values indicated a large effect of country in both in the pig and poultry datasets (Supplementary Figure 7).
337 Of special interest was the newly characterized *Enterococcus*-associated linezolid-resistance gene *optrA*,
338 that had a significantly higher abundance in Bulgarian poultry farms, compared with poultry farms in all
339 other countries (FDR < 0.05) (Figure 3b). A single Spanish farm did, however, have even higher *optrA*
340 abundance than any other farm. Among the pig herds, the *optrA* gene was more abundant in Bulgarian and
341 Italian herds than anywhere else (except for two farms in Spain) (FDR<0.05).

342 The newly identified colistin-resistance gene *mcr-1* was significantly more abundant in Bulgarian and Italian
343 poultry farms, compared with most other countries (FDR<0.05). France, Poland and Spain had intermediate
344 levels, while Denmark, the Netherlands and Germany had the lowest levels (Figure 3b).

345 As previously noted from visual inspection of heatmaps, multiple chloramphenicol AMR genes including
346 *cat(pC194)* were much more abundant in Italian pigs than other pigs. The ESBL *bla_{CTX-M}* gene cluster 1 also
347 showed country dependency, being significantly more abundant in poultry samples from Spain, Poland,
348 Italy, France and Belgium than Germany (FDR<0.05). Differential abundance analysis results can be found in
349 Supplementary Table 7 and 8 for pig and poultry respectively.



350

351 **Figure 3. AMR genes differ in abundance between countries. A handpicked subset of genes that differed significantly in**
352 **abundance between at least two countries' pig farms (a) or poultry farms (b). The regularized log abundance (rlog) is shown on**
353 **the y-axis in boxplots and points. Points were sideways jittered to reduce overplotting. Horizontal box lines represent Q1,**
354 **median and Q3. Whiskers denote range of points within $Q1-1.5*IQR$ and $Q3+1.5*IQR$.**

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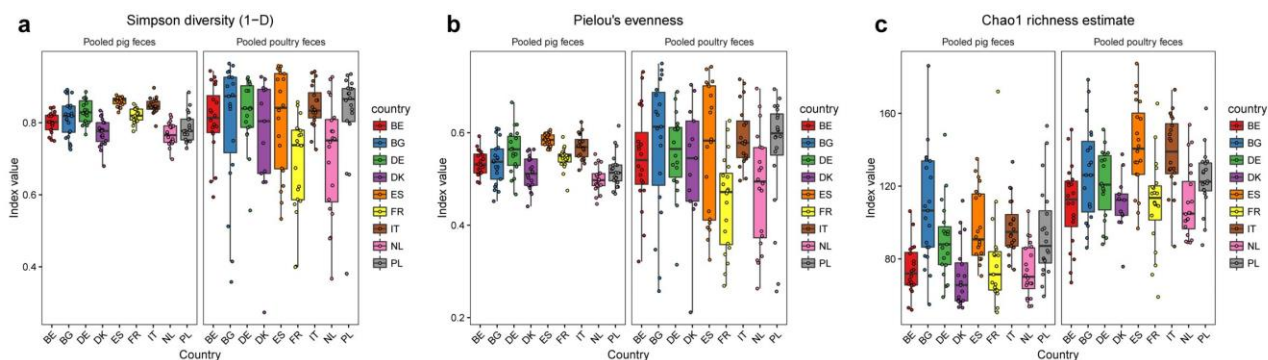
361

362 Alpha diversity and richness

363 We calculated several alpha diversity indexes for each farm resistome (Figure 4) The range of AMR diversity
364 was generally much larger for poultry samples, having both lower and higher diversity than pig samples,
365 which had a tighter spread of diversity. The poultry samples had a higher estimated richness than pigs (i.e.
366 a higher number of unique AMR genes per sample). Alpha diversity indexes can be found in Supplementary
367 Table 9.

368 Interestingly, countries with high AMR richness in pigs also tended to have high AMR richness in poultry.
369 Spain had the highest median richness in both reservoirs, followed by Italy. Poland and Bulgaria together
370 had the third and fourth highest AMR richness in pigs and poultry. Overall, the median estimated Chao1
371 richness per country correlated significantly between the reservoirs (Spearman's rho: 0.88, $p < 0.01$). This
372 was neither true for evenness nor diversity ($p > 0.05$). Rarefaction curves for pig and poultry resistomes can
373 be found in Supplementary Figure 8.

374



375

376 **Figure 4. Resistome alpha diversity and richness differs between animal host and countries. From the read count pair matrix,**
377 **several indexes were calculated. (a) Simpson diversity index, (b) Pielou's evenness, and (c) the Chao1 richness estimate.**
378 **Horizontal box lines represent Q1, median and Q3. Whiskers denote range of points within $Q1-1.5*IQR$ and $Q3+1.5*IQR$.**

379

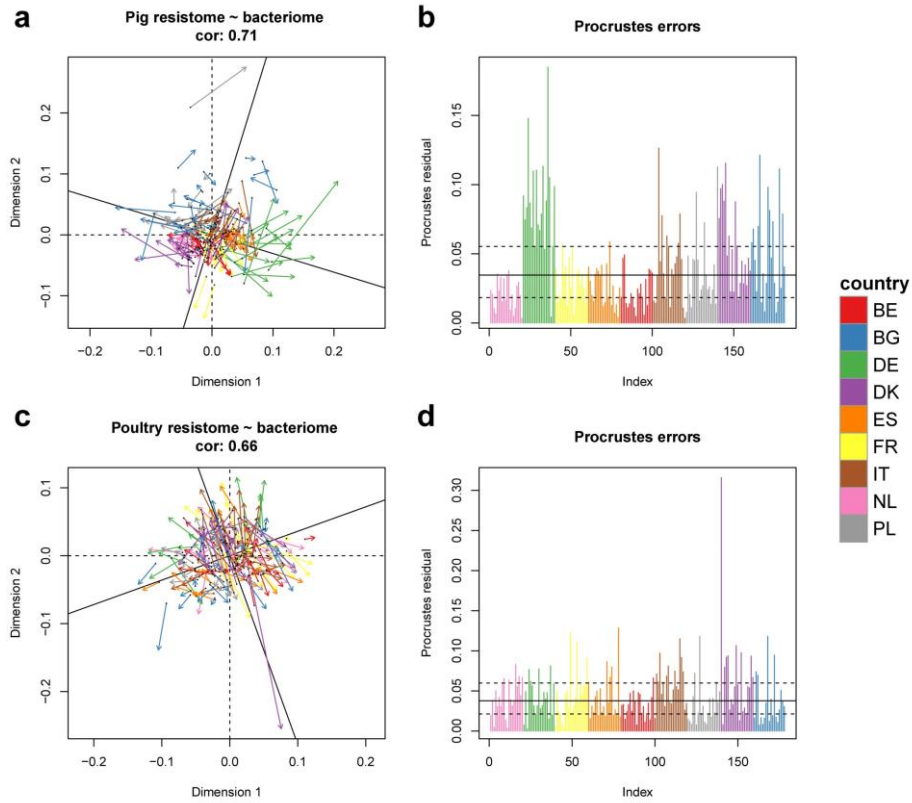
380

381 Association between bacteriome and resistome

382 To test the degree to which bacterial genus composition of the microbiota dictates the resistomes,
383 Procrustes analyses were performed. We compared ordinations of the pig microbiome with the pig
384 resistome and the poultry microbiome with the poultry resistome (Figure 5). We found that for both pig

385 and poultry, the bacterial composition correlated significantly with the resistome ($p < 0.001$). Samples with
386 similar taxonomic compositions thus tended to have similar resistome compositions.

387



388

389 **Figure 5. Microbial composition dictates the resistome. Procrustes analysis plots for pig (a/b) and poultry (c/d). Procrustes**
390 **rotations of bacterial composition onto their corresponding resistome composition (a/c). Bar charts of Procrustes residuals**
391 **(b/d).**

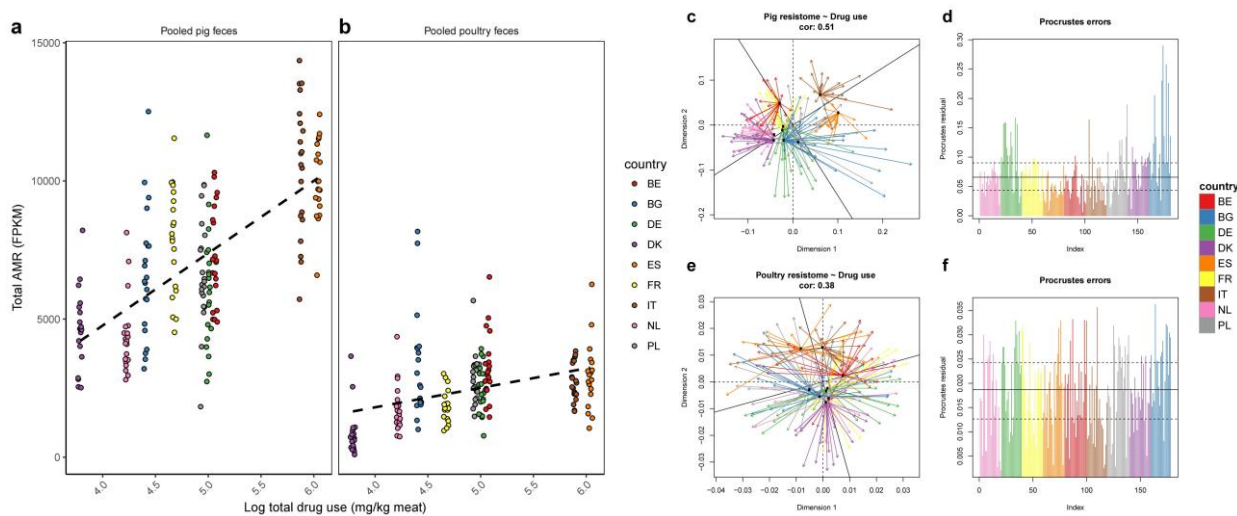
392

393 The correspondence between the two datasets was slightly stronger in pigs (Procrustes symmetric
394 correlation: 0.71) than in poultry (Procrustes symmetric correlation: 0.66). Interestingly, in pig samples we
395 saw a country effect on the strength of association between the bacteriome and the resistome. In Dutch
396 and Belgian pig herds, ordinations based on bacterial genera and AMR genes gave similar results (Figure
397 5b). For samples from Bulgaria, Denmark, Italy and especially Germany however, the Procrustes residuals
398 were larger. This was less evident for poultry, though a single Danish poultry herd had a very unusual
399 resistome, considering its taxonomic composition (Figure 5d). Stress plots for NMDS can be found in
400 Supplementary Figure 9.

401

402 AMR and drug use association

403 We found that the total country-level veterinary AMU was positively associated with AMR in both pigs and
404 poultry. The AMR abundance increased by 1736-3507 (95% CI, $\beta=2621$) FPKM in pigs when the AMU
405 increased by 1 \log_e unit (36.8% increase in AMU) (Figure 6a) and to a lesser degree in poultry, where the
406 AMR abundance increase by 68 - 1330 FPKM (95% CI, $\beta=700$) when the AMU increase 1 \log_e unit (Figure
407 6b). For pigs, the variance between farms within countries was 7 times larger than the variance between
408 countries in general, whereas in poultry the variance was 4 times larger within-country than between
409 countries.



410

411 **Figure 6. National veterinary AMU affects total metagenomic AMR. (a-b) Scatter plots of country AMU and pooled sample total**
412 **AMR. A slight sideways jitter was added to the points to minimize overplotting. Trend lines are shown for (a) pigs and (b)**
413 **poultry. (c-f) Procrustes errors from rotation of resistome principal coordinate analysis (PCoA) onto country AMU PCoA (c-d) and**
414 **poultry (e-f). (c/e) Procrustes rotations with arrows starting in AMU and ending in corresponding AMR ordination. (d/f) Line**
415 **plots showing length of each residual arrow in Procrustes rotation.**

416

417 To test if the AMU pattern was associated with AMR gene profiles, we compared the AMR abundance
418 matrix with AMU matrix, comprised of the 15 recorded AM classes (Supplementary Table 5). We found an
419 association between the veterinary AMU pattern and the pig resistomes (Procrustes correlation: 0.51,
420 $p<0.001$) (Figure 6cd). The poultry resistomes were also significantly associated with AMU, albeit with a
421 lower correlation, likely due to their larger beta-diversity and lower degree of country clustering
422 (Procrustes correlation: 0.38, $p<0.001$) (Figure 6ef).

423

424

425 **Functional AMR genes**

426 In addition to using ResFinder, we also ran most analyses with the FRD database, to elucidate whether the
427 functionally determined AMR genes behave similarly to the acquired AMR genes in ResFinder. If FRD genes
428 serve similar AMR functionality as the acquired ResFinder genes, we would expect similar results.

429 Using the FRD database, we found both similar and different patterns, compared with using ResFinder.
430 There was still a perfect separation between pig and poultry samples, but the country separation in pigs
431 was less distinct than when using ResFinder (Supplementary Figure 10). Though less variation could be
432 explained by two axes, the PCoA plot of pig samples now clustered German and Spanish samples, with the
433 remaining countries being more similar. The resistome richness showed similar patterns to ResFinder:
434 Spanish, Italian, Polish and Bulgarian samples had a higher estimated richness in both pig and poultry,
435 compared to the other countries. The Procrustes correlation between the resistome and drug usage was
436 lower (0.40 for pig and 0.25 for poultry). This result was echoed by the lack of association between total
437 AMR and total AMU, for both pig and poultry ($p>0.05$, Supplementary Figure 11).

438

439 **Discussion**

440 Using a metagenomic shotgun sequencing strategy, we were able to detect and quantify more than 400
441 AMR genes across 181 pig and 178 poultry herds in 9 European countries.

442 A recent study including Chinese, Danish and French pigs showed the Chinese pig resistomes clustered
443 separately, while Danish and French overlapped.¹⁶ Here we demonstrate that even among European
444 countries, the livestock resistomes differ in a country-specific manner that might be explained by
445 differential AMU so that countries with similarly high and diverse AMU (Spain, Italy) have similar
446 resistomes, the same way as countries with similarly low AMU (Denmark, Netherlands) also have similar pig
447 resistomes.

448 We found that the beta diversity dispersion seems to be country dependent, particularly in pigs, with
449 Bulgarian, German and Polish pig herds having more dispersed AMR. While we cannot currently explain
450 this, we consider possible causes as differences in trade and management, among others.

451 We found the recently discovered plasmid-borne colistin resistance gene *mcr-1* in a number of poultry
452 herds, especially in Bulgaria, Spain and Italy. Spain and Italy had the highest reported veterinary colistin
453 usage among the surveyed countries, whereas Bulgaria has a low reported usage, uncharacteristic for the

454 high *mcr-1* level found here.¹³ This gene was recently discovered in China and identified throughout the
455 world and has been identified in pigs, poultry and human clinical infections alike.³⁸

456 A newly characterized enterococcal linezolid-resistance gene, *optrA* was detected in a subset of pig
457 samples, with Bulgaria, Italy and Spain having the highest abundances. The *optrA* gene provides AMR to
458 both oxazolidinone and amphenicols, including the veterinary-used florfenicol.^{13,39} The high abundance of
459 this gene in these countries can likely be explained by the fact that they have the highest veterinary
460 amphenicol usage (together with Croatia, which we did not sample) among the 26 countries surveyed by
461 ESVAC.¹² This explanation fits well with the fact that Bulgaria, Italy and Spain also had the highest
462 abundances of chloramphenicol AMR genes such as *cat(pC194)* in poultry.

463 Another AMR gene of special interest, the *bla*_{CTX-M}, was also observed in the poultry herds. The higher
464 abundance of *bla*_{CTX-M} cluster 1 in Spain, Italy, Poland and Belgium, could possibly be explained by co-
465 selection by fluoroquinolones, which is used more in Spain, Poland, Italy and Belgium than other sampled
466 countries. *qnr* and *bla*_{CTX-M} genes are frequently co-located on large ESBL plasmids. Veterinary
467 cephalosporin usage did not seem to explain the observed levels.

468 Poland and Spain use far more fluoroquinolones veterinarily than other countries included in this study. We
469 found that plasmid mediated quinolone AMR (*qnr*-genes) was frequently abundant in Polish, but not in
470 Spanish, poultry. In Bulgaria, quinolone AMR was also frequently observed, although their reported AMU
471 did not follow the same trend.

472 Interestingly, we observed that country-wise estimated AMR richness significantly correlated between pig
473 and poultry. Also, the countries with a high AMR richness were also the ones with higher AMR abundance
474 (Italy, Spain, Bulgaria and Poland). While having higher abundance of the ubiquitous core genes, these
475 countries had a higher number of unique genes in both pig and poultry production. The fact that countries'
476 AMR abundance and gene richness in pig and poultry tend to follow each other, could perhaps be explained
477 with policy: if a country has strict AMU regulations in one livestock species, chances are that similar
478 regulations are in place for other livestock species. Better host-separated, preferably herd-level, AMU data
479 is needed to further explore this.

480

481 It has previously been reported that soil community composition structures the soil resistome.²⁶ Using a
482 wide array of environmental matrices, including fecal and waste water samples, this has also been shown
483 for human habitats.⁴⁰ We found the same to be true for pig and poultry resistomes, and additionally, we
484 showed that the taxa-AMR association strength differs between countries. Horizontal gene transfer (HGT)

485 could explain this phenomenon, if a larger proportion of certain countries' resistome is mobile and AMR
486 genes are more frequently introduced and re-introduced to genera. On the other hand, vertical AMR
487 transmission can also play a role; if e.g. one country's livestock is more isolated from trade. This would
488 mean more livestock generations for the microbiome and resistome to diverge from those in other
489 countries.

490 The determination of a set of core AMR genes for pig and poultry likely represents an underestimate of the
491 true core resistomes. Requiring a certain percentage of samples to show evidence for a gene does not
492 account for differential sequencing depth. The core genes discovered here thus likely represent a
493 conservative subset of the true core resistomes.

494 In contrast to ResFinder, when using FRD we found no relationship between total drug use and total
495 functional AMR abundance. This suggests that while many genes can provide AMR when cloned into e.g. *E.*
496 *coli* in functional metagenomic assays, they might not provide AMR functionality in their natural hosts with
497 natural expression levels. If most of them did, we could expect to see AM-based selection and an
498 association to drug usage, like it is observed for the AMR genes in ResFinder - a database of genes known to
499 provide AMR to their natural hosts. This finding echoes previous sentiments that one should carefully
500 consider the risk to human health imposed by individual AMR genes.⁴¹ Some FRD genes might represent
501 high risk, but we currently do not know what subset that is. Creating the FRD is a first step in trying to
502 catalog the many AMR genes found in functional metagenomic studies. Screening sequenced pathogenic
503 isolates and metagenomic assemblies for FRD genes, would be a good start for assessing their host range
504 and risk potential.

505 The AMU data used in this study is not optimal. There is variation in drug use within each country's farms
506 that we did not account for by using a national average and we are unable to accurately distinguish
507 between drugs used in different livestock species. Moreover the PCU denominator used by ESVAC may vary
508 greatly between countries with differences in livestock populations and imports/exports that contribute to
509 the biomass as calculated. Furthermore, the integrated herds enrolled in this study might represent only a
510 limited subset of the overall livestock production in some countries. However, even with our crude AMU
511 estimates, we found significant associations with total AMR abundance. The similar conclusion when
512 considering the specific drug usage profile of each country indicates that the resistome is responding to
513 AMU. The AMR-AMU association is well-documented for specific cultured indicator species and certain AM
514 drugs, but relatively unknown when considering the whole microbiota and resistome and the newer
515 approach of metagenomic shotgun sequencing.^{3,8} We do not know why the pig samples had a large within-

516 country spread of total AMR, but perhaps the more heterogeneous production system and production
517 management is responsible.

518 DNA extractions from the pooled poultry samples resulted in relatively low DNA yields. The protocol used
519 was optimized for pig feces, human feces and sewage, but not poultry feces.¹⁸ The lower yields
520 necessitated the use of a PCR-based library preparation kit, that can influence downstream analysis of
521 shotgun sequencing.⁴² While the large difference between pig and poultry resistomes in our study is likely
522 real, we caution the use of sensitive, quantitative analyses when comparing between samples prepared
523 using different library preparation kits. For this reason, we have mostly tested within each reservoir. For
524 future studies, one might consider using larger volumes of bird feces or otherwise optimize the protocol to
525 ensure PCR-free library preparation is possible for all samples.

526 The sensitivity of metagenomic approaches does not yet rival phenotypic alternatives such as selective
527 enrichment. We relied on read-mapping as opposed to a metagenomic assembly-based strategy for greater
528 sensitivity. Still, there are AMR genes in important pathogens that we know are likely present but are below
529 our detection limit. For example, we only found evidence for *bla*_{CTX-M} in three pig herds, whereas in
530 phenotypic studies, the prevalence is high even among farms with no cephalosporin usage.⁴³ Another
531 tradeoff with the use of reference-based read mapping is our inability to identify mutated housekeeping
532 genes granting AMR or distinguish between close homologs of the same AMR gene, e.g., the non-ESBL
533 *bla*_{TEM} genes from those encoding ESBL variants.

534 The primary concern with read-mapping techniques, the lack of genomic context, can be solved using
535 metagenomic assembly and binning approaches.^{16,44,45} In this way, AMR alleles in full length, their genomic
536 context and their associated taxa have been identified in both pig, poultry and human fecal samples.⁴⁶ As
537 shown previously, the association between AMR and AMU is similar for metagenomics and traditional
538 phenotypic methods, but several aspects make metagenomics an intriguing monitoring tool.¹⁷ The fact that
539 both types of analyses (quantitative, sensitive read mapping and qualitative, context-giving binning) use the
540 same raw data, makes metagenomics an attractive tool. In addition, the digital nature of shotgun data
541 would also allow future re-use and form the basis of an invaluable historical archive, potentially usable for
542 both AMR and pathogen tracking worldwide. While it is possible to identify thousands of AMR genes from
543 environmental samples using functional metagenomics, and then track them using shotgun sequencing,
544 their association to drug use and potential risk to human and animal health requires much further work to
545 inform effective drug policies.

546 We found that the metagenomic resistome varied significantly in livestock, with large differences between
547 the pig and poultry reservoir, but also within each livestock species, in a country-dependent manner.
548 Within each country, we found different levels of variation, with some countries having more homogenous
549 herds than others. Differences were seen both in total AMR abundance, but also abundances of AMR types
550 and specific genes, including clinically relevant AMR genes. Some of this variation, we attributed to
551 differential drug usage between the countries. We also identified the microbiome background as an
552 important factor in determining the resistome in livestock, but found the strength of the association was
553 country-dependent, at least in pigs. Interestingly, we found that AMR richness in one livestock species in a
554 country is linked to the abundance in another livestock species. Finally, we observed some indications that
555 newly described AMR genes from functionally metagenomic studies, might not provide AMR functionality
556 when expressed in their natural host, even though they have the potential at the right expression levels in
557 the right organism.

558 **Data availability**

559 The DNA sequences (reads) from the 363 metagenomic samples from the 359 herds are deposited in the
560 European Nucleotide Archive (ENA) under the project accession number PRJEB22062.

561

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576 **Author contributions**

577 FMA, DH, JAW, TH, DM and the EFFORT group designed the study. FMA and BEK detailed the sampling and
578 sequencing. ASRD and the EFFORT group carried out sampling. BEK and SJP conducted the DNA purification
579 and organized with PM the sequencing. RBH, OL and TNP created the read-mapping pipeline. ER created
580 the functional resistome database. PM, OL, RECL, LC, LAMS, HS, AB and HV carried out the bioinformatics
581 and statistical analysis. PM created the figures and drafted the manuscript. All authors helped review, edit
582 and complete the manuscript.

583

584 **Competing interests**

585 The authors declare no competing financial interests.

586 **References**

587

- 588 1. Who. ANTIMICROBIAL RESISTANCE Global Report on Surveillance 2014. *World Heal. Organ.* (2014).
- 589 2. Aarestrup, F. M. The livestock reservoir for antimicrobial resistance: a personal view on changing
590 patterns of risks, effects of interventions and the way forward. *Philos. Trans. R. Soc. Lond. B. Biol.*
591 *Sci.* **370**, 20140085 (2015).
- 592 3. Chantziaras, I., Boyen, F., Callens, B. & Dewulf, J. Correlation between veterinary antimicrobial use
593 and antimicrobial resistance in food-producing animals: A report on seven countries. *J. Antimicrob.*
594 *Chemother.* **69**, 827–834 (2014).
- 595 4. Dunlop, R. H. *et al.* Associations among antimicrobial drug treatments and antimicrobial resistance
596 of fecal *Escherichia coli* of swine on 34 farrow-to-finish farms in Ontario, Canada. *Prev. Vet. Med.* **34**,
597 283–305 (1998).
- 598 5. Aarestrup, F. M., Seyfarth, A. M., Emborg, H., Pedersen, K. & Bager, F. Effect of Abolishment of the
599 Use of Antimicrobial Agents for Growth Promotion on Occurrence of Antimicrobial Resistance in
600 Fecal Enterococci from Food Animals in Denmark. *Antimicrob. Agents Chemother* **45**, 2054–2059
601 (2001).
- 602 6. Agersø, Y. & Aarestrup, F. M. Voluntary ban on cephalosporin use in Danish pig production has
603 effectively reduced extended-spectrum cephalosporinase-producing *Escherichia coli* in slaughter
604 pigs. *J. Antimicrob. Chemother.* **68**, 569–72 (2013).

- 605 7. Dutil, L. *et al.* Ceftiofur resistance in *Salmonella enterica* serovar Heidelberg from chicken meat and
606 humans, Canada. *Emerg. Infect. Dis.* **16**, 48–54 (2010).
- 607 8. Dorado-García, A. *et al.* Quantitative assessment of antimicrobial resistance in livestock during the
608 course of a nationwide antimicrobial use reduction in the Netherlands. *J. Antimicrob. Chemother.*
609 dkw308 (2016). doi:10.1093/jac/dkw308
- 610 9. Aarestrup, F. M. *et al.* Resistance to antimicrobial agents used for animal therapy in pathogenic-,
611 zoonotic- and indicator bacteria isolated from different food animals in Denmark: a baseline study
612 for the Danish Integrated Antimicrobial Resistance Monitoring Programme (DANMAP). *APMIS* **106**,
613 745–770 (1998).
- 614 10. Bronzwaer, S. Harmonised monitoring of antimicrobial resistance in *Salmonella* and *Campylobacter*
615 isolates from food animals in the European Union. *Clin. Microbiol. Infect.* **14**, 522–533 (2008).
- 616 11. McEwan, S. A., Aarestrup, F. M. & Jordan, D. in *Antimicrobial resistance in bacteria of animal origin*
617 397–413 (ASM Press, 2006).
- 618 12. Veterinary Medicines Division. European Medicines Agency, European Surveillance of Veterinary
619 Antimicrobial Consumption, 2014. (2016).
- 620 13. ECDC. EFSA. ENA. ECDC/EFSA/EMA first joint report on the integrated analysis of the consumption of
621 antimicrobial agents and occurrence of antimicrobial resistance in bacteria from humans and food-
622 producing animals. *EFSA J.* **13**, 4006 (2015).
- 623 14. EFSA. Technical specifications on the harmonised monitoring and reporting of antimicrobial
624 resistance in *Salmonella*, *Campylobacter* and indicator *Escherichia coli* and *Enterococcus* spp .
625 bacteria transmitted through food 1. *EFSA J.* **10**, 64 (2012).
- 626 15. Nordahl Petersen, T. *et al.* Meta-genomic analysis of toilet waste from long distance flights; a step
627 towards global surveillance of infectious diseases and antimicrobial resistance. *Sci. Rep.* **5**, 11444
628 (2015).
- 629 16. Xiao, L. *et al.* A reference gene catalogue of the pig gut microbiome. *Nat. Microbiol.* **1**, 16161 (2016).
- 630 17. Munk, P. *et al.* A sampling and metagenomic sequencing-based methodology for monitoring
631 antimicrobial resistance in swine herds. *J. Antimicrob. Chemother.* dkw415 (2016).
632 doi:10.1093/jac/dkw415

- 633 18. Knudsen, B. E. *et al.* Impact of Sample Type and DNA Isolation Procedure on Genomic Inference of
634 Microbiome Composition. *mSystems* **1**, 64394 (2016).
- 635 19. Li, H. & Durbin, R. Fast and accurate short read alignment with Burrows-Wheeler transform.
636 *Bioinformatics* **25**, 1754–1760 (2009).
- 637 20. Zankari, E. *et al.* Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.*
638 **67**, 2640–2644 (2012).
- 639 21. Trapnell, C. *et al.* Differential gene and transcript expression analysis of RNA-seq experiments with
640 TopHat and Cufflinks. *Nat. Protoc.* **7**, 562–78 (2012).
- 641 22. Fu, L., Niu, B., Zhu, Z., Wu, S. & Li, W. CD-HIT: Accelerated for clustering the next-generation
642 sequencing data. *Bioinformatics* **28**, 3150–3152 (2012).
- 643 23. Pehrsson, E. C., Forsberg, K. J., Gibson, M. K., Ahmadi, S. & Dantas, G. Novel resistance functions
644 uncovered using functional metagenomic investigations of resistance reservoirs. *Front. Microbiol.* **4**,
645 1–11 (2013).
- 646 24. Moore, A. M. *et al.* Pediatric fecal microbiota harbor diverse and novel antibiotic resistance genes.
647 *PLoS One* **8**, (2013).
- 648 25. Sommer, M. O. a, Dantas, G. & Church, G. M. Functional characterization of the antibiotic resistance
649 reservoir in the human microflora. *Science* **325**, 1128–31 (2009).
- 650 26. Forsberg, K. J. *et al.* Bacterial phylogeny structures soil resistomes across habitats. *Nature* **509**, 612–
651 6 (2014).
- 652 27. Legendre, P. & Gallagher, E. D. Ecologically meaningful transformations for ordination of species
653 data. *Oecologia* **129**, 271–280 (2001).
- 654 28. Veterinary Medicines Division. European Medicines Agency, European Surveillance of Veterinary
655 Antimicrobial Consumption, 2016. (2016).
- 656 29. Jari Oksanen, F. Guillaume Blanchet, Michael Friendly, Roeland Kindt, Pierre Legendre, D., McGlinn,
657 Peter R. Minchin, R. B. O’Hara, Gavin L. Simpson, Peter Solymos, M. H. H. & Stevens. vegan:
658 Community Ecology Package. (2016).
- 659 30. Pielou, E. C. The measurement of diversity in different types of biological collections. *J. Theor. Biol.*

- 660 **13**, 131–144 (1966).
- 661 31. Paradis, E., Claude, J. & Strimmer, K. APE: Analyses of phylogenetics and evolution in R language.
662 *Bioinformatics* **20**, 289–290 (2004).
- 663 32. Letunic, I. & Bork, P. Interactive Tree Of Life (iTOL): An online tool for phylogenetic tree display and
664 annotation. *Bioinformatics* **23**, 127–128 (2007).
- 665 33. Wickham, H. *ggplot2: Elegant Graphics for Data Analysis*. (Springer-Verlag New York, 2009).
- 666 34. Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting Linear Mixed-Effects Models using lme4. *eprint*
667 *arXiv:1406.5823* **67**, 51 (2014).
- 668 35. Love, M. I., Huber, W. & Anders, S. Moderated estimation of fold change and dispersion for RNA-seq
669 data with DESeq2. *Genome Biol.* **15**, 550 (2014).
- 670 36. McMurdie, P. J. & Holmes, S. Waste not, want not: why rarefying microbiome data is inadmissible.
671 *PLoS Comput. Biol.* **10**, e1003531 (2014).
- 672 37. Hochberg, Y. & Benjamini, Y. Controlling the False Discovery Rate : a Practical and Powerful
673 Approach to Multiple Testing. **57**, 289–300 (1995).
- 674 38. H Hasman, AM Hammerum , F Hansen , RS Hendriksen, B Olesen, Y Agersø, E Zankari, P
675 Leekitcharoenphon, M Stegger , RS Kaas, LM Cavaco, DS Hansen, FM Aarestrup, R. S. Detection of
676 Mcr-1 Encoding Plasmid-Mediated Colistin-Resistant Escherichia Coli Isolates From Human
677 Bloodstream Infection and Imported Chicken Meat, Denmark 2015. *Euro surveill* **20**, pii=30085
678 (2015).
- 679 39. Wang, Y. *et al.* A novel gene, *optrA*, that confers transferable resistance to oxazolidinones and
680 phenicols and its presence in *Enterococcus faecalis* and *Enterococcus faecium* of human and animal
681 origin. *J. Antimicrob. Chemother.* **70**, 2182–2190 (2015).
- 682 40. Sun, M. *et al.* Interconnected microbiomes and resistomes in low-income human habitats SI. *Nat. SI*
683 **61**, 5985–5991 (2016).
- 684 41. Martínez, J. L., Coque, T. M. & Baquero, F. What is a resistance gene ? Ranking risk in resistomes.
685 *Nat. Publ. Gr.* **13**, 116–123 (2014).
- 686 42. Bowers, R. M. *et al.* Impact of library preparation protocols and template quantity on the

- 687 metagenomic reconstruction of a mock microbial community. *BMC Genomics* **16**, 856 (2015).
- 688 43. Hammerum, A. M. *et al.* Characterization of extended-spectrum β -lactamase (ESBL)-producing
689 *Escherichia coli* obtained from Danish pigs, pig farmers and their families from farms with high or no
690 consumption of third- or fourth-generation cephalosporins. *J. Antimicrob. Chemother.* 1–8 (2014).
691 doi:10.1093/jac/dku180
- 692 44. Nielsen, H. B. *et al.* Identification and assembly of genomes and genetic elements in complex
693 metagenomic samples without using reference genomes. *Nat. Biotechnol.* **32**, (2014).
- 694 45. Anantharaman, K. *et al.* Thousands of microbial genomes shed light on interconnected
695 biogeochemical processes in an aquifer system. *Nat. Commun.* **7**, 1–11 (2016).
- 696 46. Ma, L. *et al.* Metagenomic Assembly Reveals Hosts of Antibiotic Resistance Genes and the Shared
697 Resistome in Pig, Chicken, and Human Feces. *Environ. Sci. Technol.* **50**, 420–427 (2016).
- 698