



IEDB-AR: immune epitope database-analysis resource in 2019

Dhanda, Sandeep Kumar; Mahajan, Swapnil; Paul, Sinu; Yan, Zhen; Kim, Haeuk; Jespersen, Martin Closter; Jurtz, Vanessa ; Andreatta, Massimo; Greenbaum, Jason A.; Marcatili, Paolo

Total number of authors:
13

Published in:
Nucleic Acids Research

Link to article, DOI:
[10.1093/nar/gkz452](https://doi.org/10.1093/nar/gkz452)

Publication date:
2019

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):
Dhanda, S. K., Mahajan, S., Paul, S., Yan, Z., Kim, H., Jespersen, M. C., Jurtz, V., Andreatta, M., Greenbaum, J. A., Marcatili, P., Sette, A., Nielsen, M., & Peters, B. (2019). IEDB-AR: immune epitope database-analysis resource in 2019. *Nucleic Acids Research*, 47(1), W502-W506. <https://doi.org/10.1093/nar/gkz452>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

IEDB-AR: immune epitope database—analysis resource in 2019

Sandeep Kumar Dhanda^{1,†}, Swapnil Mahajan^{1,†}, Sinu Paul^{1,†}, Zhen Yan¹, Haeuk Kim¹, Martin Closter Jespersen², Vanessa Jurtz², Massimo Andreata^{2,3}, Jason A. Greenbaum¹, Paolo Marcatili², Alessandro Sette^{1,4}, Morten Nielsen^{2,3} and Bjoern Peters^{1,4,*}

¹Division of Vaccine Discovery, La Jolla Institute for Allergy and Immunology, La Jolla, CA 92037, USA, ²Department of Health Technology, Technical University of Denmark, Kgs. Lyngby, Denmark, ³Instituto de Investigaciones Biotecnológicas, Universidad Nacional de San Martín, Argentina and ⁴Department of Medicine, University of California, San Diego, CA 92122, USA

Received February 12, 2019; Revised May 01, 2019; Editorial Decision May 09, 2019; Accepted May 10, 2019

ABSTRACT

The Immune Epitope Database Analysis Resource (IEDB-AR, <http://tools.iedb.org/>) is a companion website to the IEDB that provides computational tools focused on the prediction and analysis of B and T cell epitopes. All of the tools are freely available through the public website and many are also available through a REST API and/or a downloadable command-line tool. A virtual machine image of the entire site is also freely available for non-commercial use and contains most of the tools on the public site. Here, we describe the tools and functionalities that are available in the IEDB-AR, focusing on the 10 new tools that have been added since the last report in the 2012 NAR webserver edition. In addition, many of the tools that were already hosted on the site in 2012 have received updates to newest versions, including NetMHC, NetMHCpan, BepiPred and DiscoTope. Overall, this IEDB-AR update provides a substantial set of updated and novel features for epitope prediction and analysis.

INTRODUCTION

The adaptive immune system in vertebrates can recognize a large repertoire of antigens from a broad spectrum of pathogens. B and T cell receptors are responsible for recognizing these diverse set of antigens and triggering immune responses. The specific regions recognized on these antigens by B and T cell receptors are termed as epitopes. Thus, understanding the mechanism of immune receptor:epitope interactions is important in developing diagnostics, therapeutics, and vaccines against infectious and autoimmune diseases, cancers and allergies.

The Immune Epitope Database (IEDB) captures experiments that identify and characterize epitopes and epitope specific immune receptors along with various other details such as host organism, immune exposures, and induced immune responses (1). A companion site, IEDB-Analysis Resource (IEDB-AR), hosts various B and T cell epitope prediction tools based on algorithms trained and validated on the IEDB data along with epitope analysis tools. Since the last update, the number of monthly users visiting the IEDB-AR has more than tripled from under 1,500 in 2012 to over 4,500 in 2018 (Supplementary Figure S1). New epitope prediction and analysis tools are regularly added in the IEDB-AR with features to advance epitope-based therapeutics and vaccine development (2). For example, a tool to reduce undesired immunogenicity of therapeutic proteins was implemented recently (3). Here, we describe the newly implemented tools (Table 1), updates to the previously existing tools, and novel functionalities that have been added since the last report in the 2012 NAR webserver edition (4).

T CELL EPITOPE PREDICTION TOOLS

A total of 6 new tools were added in the category of T cell epitope prediction. These include TepiTool, a T cell peptide:MHC binding prediction tool with a new user-friendly interface, tools for prediction of naturally processed MHC class I and class II ligands, deimmunization of therapeutic proteins and prediction of T cell immunogenicity beyond MHC binding affinity. In addition to the newly added tools, many of the previously existing tools have been re-trained and updated as more data were made available. The latest versions of the prediction methods in T cell epitope prediction tools are listed in Table 2. While the latest versions are provided as the default methods, many of the tools allow the user to select previous versions where available. The newly added tools are described briefly in the following sections.

*To whom correspondence should be addressed. Tel: +1 858 752 6914; Fax: +1 858 752 6987; Email: bpeters@lji.org

†The authors wish it to be known that, in their opinion, the first three authors should be regarded as Joint First Authors.

Table 1. New and updated tools in the IEDB-AR

Category	Name	Update type	Key features	Purpose
T cell	TepiTool	New tool	Interactive and easy to use tool for immunologists	Prediction of T cell epitopes.
	MHC-NP	New tool	Uses binding and ligand elution data to train the model.	Prediction of naturally processed ligands for MHC class I.
	MHCII-NP	New tool	Uses motif informations in the ligand elution dataset from IEDB	Prediction of naturally processed ligands for MHC class II.
	Immunogenicity	New tool	Uses properties and position of amino acids to predict immunogenicity	Predicting immunogenicity for MHC-class I epitopes.
	CD4EpiScore	New tool	Combines the prediction from immunogenicity and MHC binding algorithms	Predicting CD4 T cell reactivity in human population.
	Deimmunization	New tool	Predicts non-immunogenic regions based on reduced binding to a set of reference MHC II alleles	Identification of immunogenic regions and suggested amino acid substitutions to reduce immunogenicity.
B cell / T cell	LYRA	New tool	Easy to use and fast antibody and TCR structure prediction.	Template-based 3D structure modeling of B- and T-cell receptors.
B cell	BepiPred2.0	New version	Training on conformational epitope dataset using random forest algorithm	Prediction of linear B-cell epitopes.
	DiscoTope2.0	New version	Novel spatial neighborhood and surface exposure definitions.	Prediction of discontinuous B-cell epitopes.
Analysis tools	RATE	New tool	Infers HLA restriction by generating a matrix of subjects and given immune response	Inferring allele restriction for epitopes based on immune response data from HLA-typed subjects.
	ImmuneBrowser	New tool	User specified epitopes and source proteins.	Aggregating and mapping the immune response from heterogeneous epitope data to source proteins.
	Cluster2.0	Re-engineered	Multiple clustering methods and visualization.	Grouping and visualizing peptides similar in sequence.

Table 2. Methods and versions available in the IEDB T cell epitope prediction tools

MHC class	Prediction method	Versions available	Reference
MHC class I	IEDB consensus (Recommended ^a)	2.18 (default)	Moutaftsi <i>et al.</i> (22)
	NetMHCpan	4.0 (default), 3.0, 2.8	Jurtz <i>et al.</i> (23)
	NetMHC (also called ANN)	4.0 (default), 3.4	Andreata and Nielsen (24)
	SMPMBEC	1.0	Kim <i>et al.</i> (25)
	SMM	1.0	Peters and Sette (26)
	Complib_sidney2008	1.0	Sidney <i>et al.</i> (27)
	PickPocket	1.1	Zhang <i>et al.</i> (28)
	NetMHCcons	1.1	Karosiene <i>et al.</i> (29)
	netMHCstabpan	1.0	Rasmussen <i>et al.</i> (30)
	IEDB consensus (Recommended ^a)	2.17	Wang <i>et al.</i> (31)
	MHC II	NetMHCIIpan	3.1
NN-align		2.2	Nielsen and Lund (33)
SMM-align		1.1	Nielsen <i>et al.</i> (34)
Combinatorial Library		1.0	Sidney <i>et al.</i> (27)
Sturniolo		1.0	Sturniolo <i>et al.</i> (35)

^aRecommended methods can change based on regular benchmarking evaluations.

TepiTool

TepiTool (<http://tools.iedb.org/tepitool>) (5) is a new interface for IEDB T cell epitope predictions and is designed as a step-by-step wizard combining both MHC class I and class II prediction methods. The tool provides recommended default values at each step for the prediction and selection of an optimal set of peptides for a given application. TepiTool also offers additional functionalities that go beyond binding predictions. For example, conservancy analysis of peptides among the input sequences and different options for selecting the top-predicted peptides. Once the prediction task is finished, the user is provided with a concise set of top-predicted peptides and links to download the complete prediction results and conservancy estimates. The prediction results and links are also emailed to the user, if an email address is provided.

Prediction of naturally processed ligands for MHC class I and class II

MHC-NP (<http://tools.iedb.org/mhcnp>) (6) is a tool for predicting peptides that are naturally processed by the MHC

class I pathway and bind to MHC class I molecules. The tool can predict MHC I ligands for six human and two mouse MHC alleles. Similarly, MHCII-NP (<http://tools.iedb.org/mhciinp>) (7) is a tool for predicting naturally processed MHC II ligands. These tools were developed by training on the naturally processed peptides eluted from MHC molecules.

Immunogenicity

This new tool (<http://tools.iedb.org/immunogenicity>) is intended to classify peptides that bind to MHC class I (pMHC) into two categories: epitopes and non-epitopes (8). It is based on an analysis of amino acid composition of the peptide at non-anchor positions, where the side chains of amino acids are likely to be in contact with the TCR.

CD4EpiScore

CD4EpiScore (<http://tools.iedb.org/cd4episcore>) is a new tool for predicting the immunogenicity of CD4-restricted peptides in human populations that utilizes a neural network to identify patterns associated with immunogenicity

(9). It has been validated on a series of independent datasets reported in the literature for different ethnicities and diverse antigens using a variety of experimental approaches.

Deimmunization

The Deimmunization tool (<http://tools.iedb.org/deimmunization>) was added to IEDB-AR to address the issue of undesired immune reactivity to therapeutically important proteins. In a stepwise wizard, this tool makes use of the class II peptide:MHC binding prediction tools to predict potentially immunogenic regions in protein sequences and suggest amino acid substitutions to reduce their immunogenicity (3). As a proof-of-concept, the tool has been validated experimentally on recombinant factor VIIa (Vatreptacog alpha), which was discontinued from clinical trials due to immunogenicity issues (10).

B CELL EPI TOPE PREDICTION TOOLS

The IEDB-AR hosts linear B cell epitope prediction tools, such as BepiPred (11), various amino acid physicochemical property based scales (<http://tools.iedb.org/bcell/>), and discontinuous B cell epitope prediction tools, such as DiscoTope (<http://tools.iedb.org/discotope/>) (12) and ElliPro (<http://tools.iedb.org/ellipro/>) (13). Since the last update of IEDB-AR, the recommended B cell epitope prediction methods, BepiPred (14) and DiscoTope (15), were updated to their 2.0 versions.

LYRA

A new tool named LYRA (Lymphocyte Receptor Automated Modelling) was added to model the 3D structures of B and T cell receptors (16). The LYRA tool (<http://tools.iedb.org/lyra/>) predicts the structure of B- and T-cell receptors from their amino acid sequence. Using homology modelling, it selects the best framework templates and, if necessary, models the complementary determining regions (CDRs) based on the predicted canonical structure (17) of each loop, which are then grafted onto the framework templates. The results page shows the aligned sequence and a visualization of the structure allowing for quick inspection of the CDRs in both sequence and structure.

ANALYSIS TOOLS

The analysis section of IEDB-AR contains tools that automate common tasks when working with sets of epitopes or epitope-candidates. Updates to this section include a revised version of epitope clustering along with a new tool to map epitopes to source proteins, and a tool to infer allele restrictions of epitopes from immune response data on HLA-typed subjects.

Cluster2.0

The epitope clustering tool (<http://tools.iedb.org/cluster2>) has been completely re-engineered to group peptidic epitopes based on their sequence similarity. In addition to providing three different clustering approaches, this new version also supports interactive graphical visualizations of the clusters to show connectivity among peptides (18).

ImmunomeBrowser

The ImmunomeBrowser tool in the IEDB website maps epitopes to their source antigen and provides a visualization of the observed immune responses across all tested regions of the protein. In a new customizable version of this tool (<http://tools.iedb.org/immunomebrowser/>), we have extended this application to perform a similar analysis for user-provided datasets of epitopes and source antigens (19).

Restrictor Analysis Tool for Epitopes (RATE)

RATE (<http://tools.iedb.org/rate>) (20) is an automated method that can computationally infer the HLA restrictions of epitopes, given large datasets of T cell responses in HLA typed subjects. The tool takes two inputs, the alleles expressed by the subjects and the immune response of the peptides in the subjects. It then calculates the odds ratios for each allele being the restricting allele for a specific peptide and estimates significance using Fisher's exact test. The tool was developed with a focus on class II alleles but can also be applied to class I alleles.

NEW FEATURES IN THE IEDB-AR

In addition to implementing new tools, IEDB-AR development since 2012 has also targeted improvements that address how users want to interact with the tools. Two of the most readily apparent and prevalent new features are the ability to submit prediction jobs for processing in the background and the architectural changes made at the hardware and software levels to improve stability and support parallelization.

Background batch job processing

With the release of version 2.17 of the IEDB-AR in June 2017, users were given the ability to provide an email address upon submission of a class I or class II peptide binding prediction job. This enhancement has allowed users to run larger prediction jobs (in terms of the number of input sequences and predicted alleles) than would be possible directly through the web interface. Upon completion of the job, results are sent as an email attachment to the user. Since the initial implementation, this feature has been added to TepiTool and the Deimmunization tool - both of which are computationally intensive and could timeout with a reasonable-sized request through the web interface. For tools that support batch processing, it is available through the web interface as well as the API.

Hardware & software architectural changes to improve stability and support parallelization

Several architectural changes have been made to the hardware and software in order to decouple the front-end from the back-end, improve stability, and support parallelization of several tools. At the hardware level, a separate job-processing cluster was created to run all CPU-intensive tasks, such as binding predictions. These machines are physically separated from the web server so that heavy processing has little effect on web site performance. To make use

of the redesigned server architecture, the backend software was completely reengineered in Python and Django with special attention to make use of a message queuing system (RabbitMQ) and task manager (Celery). With this integrated system in place, it has allowed parallelization of jobs for several of the resource tools with speedups as great as 15-fold over the single-threaded version. It has also enabled efficient use of resources and prioritization of jobs based upon their origins. As an example, a separate resource queue has been configured in collaboration with the Griffith lab to support the CPU-intensive requests of their pVAC-Seq pipeline (21) while keeping the IEDB systems responsive to requests from other users.

AVAILABILITY OF THE TOOLS

IEDB application programming interface (IEDB-API)

In addition to the main web interface, public-facing APIs are made available for several of the tools hosted at the IEDB-AR. Included among these tools are the MHC class I and class II binding and processing predictions, MHC-NP, and the B cell linear epitope predictor. To provide a consistent experience, each of the APIs adhere to a similar interface, with parameter names shared among them where possible. All of the APIs work via HTTP POST requests and return responses in plain text. The MHC class I and II binding prediction APIs are very heavily utilized, accounting for over 300,000 predictions each month and upward of 90% of the jobs run through the IEDB-AR.

Software distribution packages

While the goal of the public IEDB-AR server is to accommodate as many prediction jobs as reasonably possible, resources can be limiting for extremely large requests. This is one of the many reasons that the IEDB-AR team provides downloadable packages to run the predictions locally on the user hardware. Currently, 8 different standalone packages are available, covering the most widely used tools, and new packages are developed based upon user demand and available resources. Additionally, a complete virtual machine image of the IEDB-AR is made available to external entities through license agreements. These two modes of distribution cover a broad range of use cases, enabling users to run many of the IEDB-AR tools on their own hardware and in complete privacy.

SUPPLEMENTARY DATA

[Supplementary Data](#) are available at NAR Online.

FUNDING

National Institutes of Health [HHSN272201200010C, 75N93019C00001]. Funding for open access charge: National Institutes of Health [HHSN272201200010C, 75N93019C00001].

Conflict of interest statement. None declared.

REFERENCES

- Vita, R., Mahajan, S., Overton, J.A., Dhanda, S.K., Martini, S., Cantrell, J.R., Wheeler, D.K., Sette, A. and Peters, B. (2019) The Immune Epitope Database (IEDB): 2018 update. *Nucleic Acids Res.*, **47**, D339–D343.
- Fleri, W., Paul, S., Dhanda, S.K., Mahajan, S., Xu, X., Peters, B. and Sette, A. (2017) The immune epitope database and analysis resource in epitope discovery and synthetic vaccine design. *Front. Immunol.*, **8**, 278.
- Dhanda, S.K., Grifoni, A., Pham, J., Vaughan, K., Sidney, J., Peters, B. and Sette, A. (2018) Development of a strategy and computational application to select candidate protein analogues with reduced HLA binding and immunogenicity. *Immunology*, **153**, 118–132.
- Kim, Y., Ponomarenko, J., Zhu, Z., Tamang, D., Wang, P., Greenbaum, J., Lundegaard, C., Sette, A., Lund, O., Bourne, P.E. *et al.* (2012) Immune epitope database analysis resource. *Nucleic Acids Res.*, **40**, W525–W530.
- Paul, S., Sidney, J., Sette, A. and Peters, B. (2016) TepiTool: a pipeline for computational prediction of T cell epitope candidates. *Curr. Protoc. Immunol.*, **114**, 18.19.11–18.19.24.
- Giguere, S., Drouin, A., Lacoste, A., Marchand, M., Corbeil, J. and Laviolette, F. (2013) MHC-NP: predicting peptides naturally processed by the MHC. *J. Immunol. Methods*, **400–401**, 30–36.
- Paul, S., Karosiene, E., Dhanda, S.K., Jurtz, V., Edwards, L., Nielsen, M., Sette, A. and Peters, B. (2018) Determination of a predictive cleavage motif for eluted major histocompatibility complex Class II ligands. *Front. Immunol.*, **9**, 1795.
- Calis, J.J., Maybeno, M., Greenbaum, J.A., Weiskopf, D., De Silva, A.D., Sette, A., Kesmir, C. and Peters, B. (2013) Properties of MHC class I presented peptides that enhance immunogenicity. *PLoS Comput. Biol.*, **9**, e1003266.
- Dhanda, S.K., Karosiene, E., Edwards, L., Grifoni, A., Paul, S., Andreatta, M., Weiskopf, D., Sidney, J., Nielsen, M., Peters, B. *et al.* (2018) Predicting HLA CD4 immunogenicity in human populations. *Front. Immunol.*, **9**, 1369.
- Lentz, S.R., Ehrenforth, S., Karim, F.A., Matsushita, T., Weldingh, K.N., Windyga, J., Mahlangu, J.N. and Adept, I. (2014) Recombinant factor VIIa analog in the management of hemophilia with inhibitors: results from a multicenter, randomized, controlled trial of vatreptacog alfa. *J. Thromb. Haemost.*, **12**, 1244–1253.
- Larsen, J.E., Lund, O. and Nielsen, M. (2006) Improved method for predicting linear B-cell epitopes. *Immunome Res.*, **2**, 2.
- Haste Andersen, P., Nielsen, M. and Lund, O. (2006) Prediction of residues in discontinuous B-cell epitopes using protein 3D structures. *Protein Sci.*, **15**, 2558–2567.
- Ponomarenko, J., Bui, H.H., Li, W., Fusseder, N., Bourne, P.E., Sette, A. and Peters, B. (2008) ElliPro: a new structure-based tool for the prediction of antibody epitopes. *BMC Bioinformatics*, **9**, 514.
- Jespersen, M.C., Peters, B., Nielsen, M. and Marcatili, P. (2017) BepiPred-2.0: improving sequence-based B-cell epitope prediction using conformational epitopes. *Nucleic Acids Res.*, **45**, W24–W29.
- Kringelum, J.V., Lundegaard, C., Lund, O. and Nielsen, M. (2012) Reliable B cell epitope predictions: impacts of method development and improved benchmarking. *PLoS Comput. Biol.*, **8**, e1002829.
- Klausen, M.S., Anderson, M.V., Jespersen, M.C., Nielsen, M. and Marcatili, P. (2015) LYRA, a webserver for lymphocyte receptor structural modeling. *Nucleic Acids Res.*, **43**, W349–W355.
- Chothia, C. and Lesk, A.M. (1987) Canonical structures for the hypervariable regions of immunoglobulins. *J. Mol. Biol.*, **196**, 901–917.
- Dhanda, S.K., Vaughan, K., Schulten, V., Grifoni, A., Weiskopf, D., Sidney, J., Peters, B. and Sette, A. (2018) Development of a novel clustering tool for linear peptide sequences. *Immunology*, **155**, 331–345.
- Dhanda, S.K., Vita, R., Ha, B., Grifoni, A., Peters, B. and Sette, A. (2018) ImmunomeBrowser: a tool to aggregate and visualize complex and heterogeneous epitopes in reference proteins. *Bioinformatics*, **34**, 3931–3933.
- Paul, S., Arlehamn, C.S.L., Schulten, V., Westernberg, L., Sidney, J., Peters, B. and Sette, A. (2017) Experimental validation of the RATE tool for inferring HLA restrictions of T cell epitopes. *BMC Immunol.*, **18**, 20.

21. Hundal,J., Carreno,B.M., Petti,A.A., Linette,G.P., Griffith,O.L., Mardis,E.R. and Griffith,M. (2016) pVAC-Seq: a genome-guided in silico approach to identifying tumor neoantigens. *Genome Med*, **8**, 11.
22. Moutafsi,M., Peters,B., Pasquetto,V., Tschärke,D.C., Sidney,J., Bui,H.H., Grey,H. and Sette,A. (2006) A consensus epitope prediction approach identifies the breadth of murine T(CD8+)-cell responses to vaccinia virus. *Nat. Biotechnol.*, **24**, 817–819.
23. Jurtz,V., Paul,S., Andreatta,M., Marcatili,P., Peters,B. and Nielsen,M. (2017) NetMHCpan-4.0: Improved Peptide-MHC Class I interaction predictions integrating eluted ligand and peptide binding affinity data. *J. Immunol.*, **199**, 3360–3368.
24. Andreatta,M. and Nielsen,M. (2016) Gapped sequence alignment using artificial neural networks: application to the MHC class I system. *Bioinformatics*, **32**, 511–517.
25. Kim,Y., Sidney,J., Pinilla,C., Sette,A. and Peters,B. (2009) Derivation of an amino acid similarity matrix for peptide: MHC binding and its application as a Bayesian prior. *BMC Bioinformatics*, **10**, 394.
26. Peters,B. and Sette,A. (2005) Generating quantitative models describing the sequence specificity of biological processes with the stabilized matrix method. *BMC Bioinformatics*, **6**, 132.
27. Sidney,J., Assarsson,E., Moore,C., Ngo,S., Pinilla,C., Sette,A. and Peters,B. (2008) Quantitative peptide binding motifs for 19 human and mouse MHC class I molecules derived using positional scanning combinatorial peptide libraries. *Immunome Res.*, **4**, 2.
28. Zhang,H., Lund,O. and Nielsen,M. (2009) The PickPocket method for predicting binding specificities for receptors based on receptor pocket similarities: application to MHC-peptide binding. *Bioinformatics*, **25**, 1293–1299.
29. Karosiene,E., Lundegaard,C., Lund,O. and Nielsen,M. (2012) NetMHCcons: a consensus method for the major histocompatibility complex class I predictions. *Immunogenetics*, **64**, 177–186.
30. Rasmussen,M., Fenoy,E., Harndahl,M., Kristensen,A.B., Nielsen,I.K., Nielsen,M. and Buus,S. (2016) Pan-specific prediction of peptide-MHC class I complex stability, a correlate of T cell immunogenicity. *J. Immunol.*, **197**, 1517–1524.
31. Wang,P., Sidney,J., Kim,Y., Sette,A., Lund,O., Nielsen,M. and Peters,B. (2010) Peptide binding predictions for HLA DR, DP and DQ molecules. *BMC Bioinformatics*, **11**, 568.
32. Andreatta,M., Karosiene,E., Rasmussen,M., Stryhn,A., Buus,S. and Nielsen,M. (2015) Accurate pan-specific prediction of peptide-MHC class II binding affinity with improved binding core identification. *Immunogenetics*, **67**, 641–650.
33. Nielsen,M. and Lund,O. (2009) NN-align. An artificial neural network-based alignment algorithm for MHC class II peptide binding prediction. *BMC Bioinformatics*, **10**, 296.
34. Nielsen,M., Lundegaard,C. and Lund,O. (2007) Prediction of MHC class II binding affinity using SMM-align, a novel stabilization matrix alignment method. *BMC Bioinformatics*, **8**, 238.
35. Sturniolo,T., Bono,E., Ding,J., Radrizzani,L., Tuereci,O., Sahin,U., Braxenthaler,M., Gallazzi,F., Protti,M.P., Sinigaglia,F. *et al.* (1999) Generation of tissue-specific and promiscuous HLA ligand databases using DNA microarrays and virtual HLA class II matrices. *Nat. Biotechnol.*, **17**, 555–561.