



Miniaturising supercritical angle fluorescence and total internal reflection fluorescence structures for a microfluidic system using in diagnostic applications

Nguyen, Trieu; Wolff, Anders

Publication date:
2019

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):

Nguyen, T., & Wolff, A. (2019). *Miniaturising supercritical angle fluorescence and total internal reflection fluorescence structures for a microfluidic system using in diagnostic applications*. Abstract from The 23rd International Conference on Miniaturized Systems for Chemistry and Life Sciences (μ TAS 2019/microtas 2019), Basel, Switzerland.

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

MINIATURISING SUPERCRITICAL ANGLE FLUORESCENCE AND TOTAL INTERNAL REFLECTION FLUORESCENCE STRUCTURES FOR A MICROFLUIDIC SYSTEM USING IN DIAGNOSTIC APPLICATIONS

Trieu Nguyen*, and Anders Wolff

*Department of Biotechnology and Biomedicine, Technical University of Denmark, Lyngby, Denmark
(*tring@dtu.dk, bios.trieu@gmail.com)*

ABSTRACT

This paper presents, for the first time, an update of a microfluidic system inspiring from the work of Hill et al. [1] in which they reported a macroscopic (millimetre scale) disposal biochip produced for diagnostic applications. The employing principal of the chips in the previous work was based on the combination of supercritical angle fluorescence (SAF) and total internal reflection fluorescence (TIRF) for a fluorescent signal measurement (Fig. 1 & 2).

KEYWORDS: polymer, microfluidics, supercritical angle fluorescence, total internal reflection fluorescence.

INTRODUCTION

When a fluorophore sits at the interface between two different media, a large portion of the fluorescence is emitted into the medium with higher refractive index (the denser material) at the angle larger than the critical angle of the two media. The fluorescence observed in this phenomenon is hence called super-critical-angle fluorescence. One can use total internal reflection (TIR) phenomenon to direct this fluorescence to the observation points where a camera can be placed to capture the fluorescent signal. Inversely, excitation light can be directed from the light source using TIR to the interface where they hit and excite the fluorophores. By doing so, we solely excite the fluorophores which are located in the vicinity of the interface (within a distance less than 100 nm from the surface, Fig. 3), hence prevent the excitation of the fluorescent molecules in the bulk (far from the surface) which are the cause of signal noise. Therefore, SAF and TIRF are combined in this detection technique. We have successfully fabricated microarrays of SAF structures, presented in MicroTAS 2018 [2].

We in this work miniaturise the fluidic system, which has the combination of SAF and TIRF structures down to microscale with a robust and low-cost fabrication procedure. Going down from millimetre to micrometre scale enhances not only the numbers of reaction chambers and structures for multiple detections but also the capability of using in a portable device (due to lightweight) for point of care detection.

EXPERIMENTS AND RESULTS

The structures that we miniaturised, including two separated parts, i.e. the SAF part, and TIRF light directing part, (the annular mask). These two parts are then attached using a chip holder (made from aluminium by micro-milling) (Fig. 4).

For the TIRF light directing part, we use the cutting-edge maskless aligner (MLA150 from HEIDELBERG Instrument (Germany)) and e-beam deposition for creating the annular mask structures after finishing by a lift-off step (Fig. 5). We created the annular masks both on glass slides and on the disposable cyclic olefin copolymer (COC) chips for the sake of comparison. The results showed that we successfully fabricated the annular masks on both glass slide and COC polymer (Fig. 6). The lift-off time for the later is much longer (20 hours versus 5 mins), assumably due to the higher adhesion between COC polymer and the photoresist. For the SAF part, micro-milling is used [2].

This work has opened up a possibility for using a combination of SAF and TIRF in a microfluidic chip for point of care testing which may conduct either solid-phase polymer chain reaction or loop-mediated isothermal amplification.

REFERENCES

[1] D. Hill *et al.* *Biomed. Microdevices*, vol. 13, no. 4, pp. 759–767, 2011; [2] T. Nguyen et al., MicroTAS2018, pages 2284–2285

ACKNOWLEDGEMENTS VIVALDI project, project number: 773422

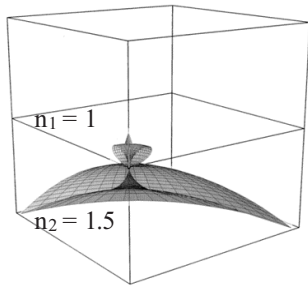


Figure 1: Large amount of the radiation emitted into the higher refractive index medium [2]. See also Fig. 1 of ref. [1]

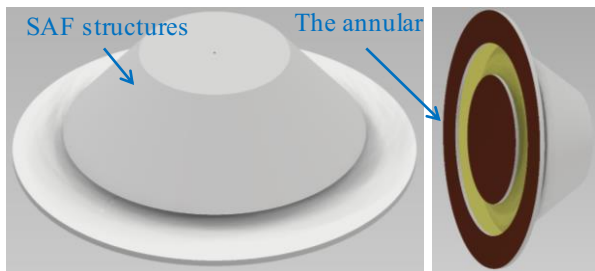


Figure 2: The CAD illustration of SAF and TIRF light directing annular ([Left] top view and [Right] side view).

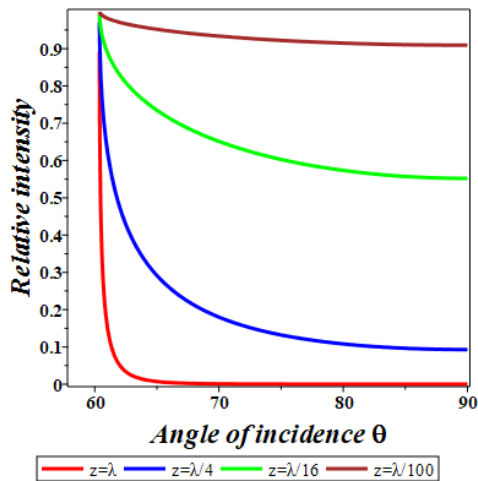


Figure 3: Our theoretical modelling of the relative fluorescent intensity from the interface versus the incidence angle with different z (z is the distance of the fluorophore from the interface). The two media are water and glass, and the critical angle is 61 degree. λ is the excitation wavelength

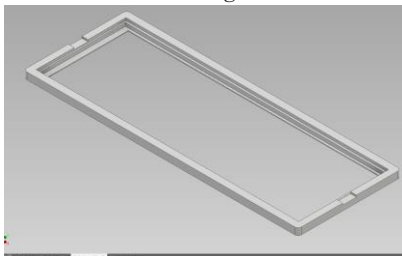


Figure 4: The CAD design of the chip holder.

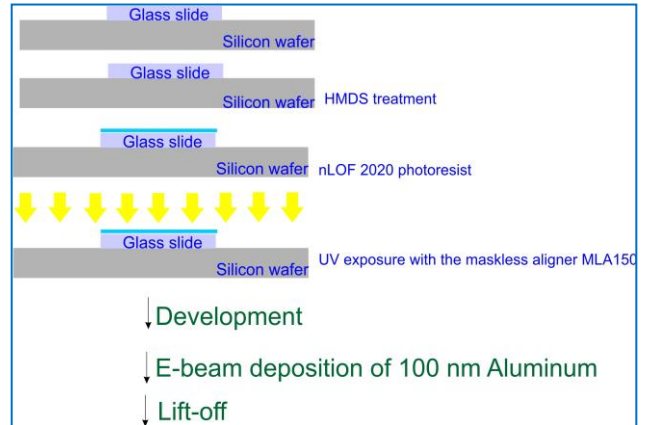


Figure 5: The lift-off process flow for fabrication of the annular structures used as TIRF light directing. Our work is done in NanoLab of Denmark Technical University.

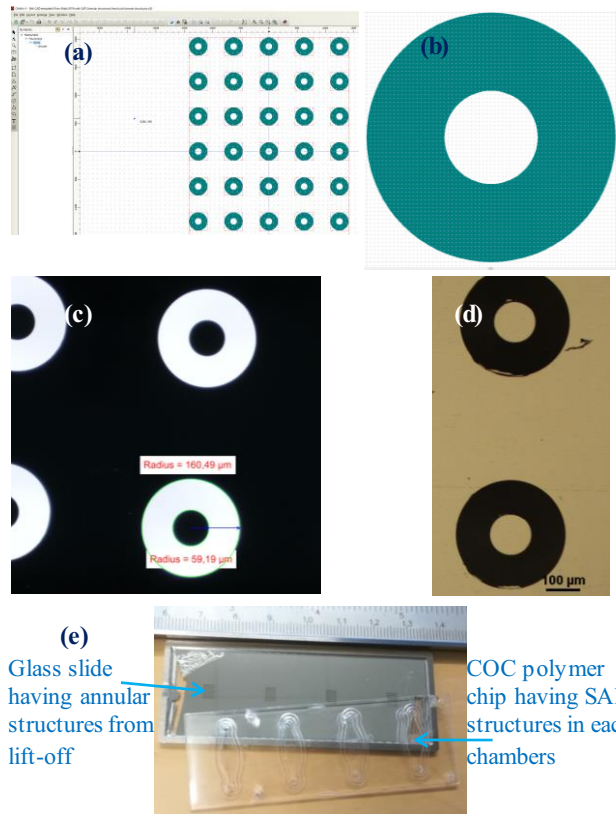


Figure 6: (a) The design of the mask. This design is then used with the maskless aligner for UV lithography. (b) Large view of the individual annular mask from the design. (c) The microscopic images of the fabricated annular on the glass slide surface after the lift-off step. The dimension of the annular agreed with the design. (d) The microscopic images of the fabricated annular on the COC polymer surface. The results show that we can obtain a clean (without a trace of metal), and quicker lift-off time for the case of using glass slide as the substrate compared to the COC polymer. (e) Digital picture of the chips and the glass slide having annular structures assembled in the chip holder.