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EFFECTIVENESS OF DENITRIFYING BIOREACTORS ON WATER POLLUTANT REDUCTION FROM AGRICULTURAL AREAS

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HIGHLIGHTS

- Denitrifying 'woodchip' bioreactors treat nitrate-N in a variety of applications and geographies.
- This review focuses on subsurface drainage bioreactors and bed-style designs (including in-ditch).
- Monitoring and reporting recommendations are provided to advance bioreactor science and engineering.

ABSTRACT.

Denitrifying bioreactors enhance the natural process of denitrification in a practical way to treat nitrate-nitrogen (N) in a variety of N-laden water matrices. The design and construction of bioreactors for treatment of subsurface drainage in the United States is guided by the United States Department of Agriculture Natural Resources Conservation Service Conservation Practice Standard 605. This review consolidates the state of the science for denitrifying bioreactors using case-studies from across the globe with an emphasis on full-size bioreactor nitrate-N removal and cost effectiveness. The focus is on bed-style bioreactors (including in-ditch modifications), although there is mention of denitrifying walls which broaden the applicability of bioreactor technology in some areas. Subsurface drainage denitrifying bioreactors have been assessed as removing between 20 to 40% of annual nitrate-N loss in the Midwest, and an evaluation across peer-reviewed literature published over the past three years showed bioreactors around the world have been generally consistent with that (N load reduction median: 46%; mean \pm st dev: 40 \pm 26%; n = 15). Reported N removal rates were on the order of 5.1 g N/m³-d (median; mean \pm st dev: 7.2 \pm 9.6 g N/m³-d; n = 27). Subsurface drainage bioreactor installation costs have ranged from less than \$5,000 to \$27,000 with estimated cost efficiencies ranging from less than \$2.50/kg N-y to roughly \$20/kg N-y (although they can be as high as \$48/kg N-y). A suggested monitoring set-up is described primarily for the context of conservation practitioners and watershed groups for assessing annual nitrate-N load removal performance of subsurface drainage denitrifying bioreactors. Recommended

minimum reporting measures for assessing and comparing annual N-removal performance include: bioreactor dimensions and installation date; fill media size, porosity, and type; nitrate-N concentrations and water temperatures; bioreactor flow treatment details; basic drainage system and bioreactor design characteristics; and N removal rate and efficiency.

Keywords.

Groundwater; Nitrate; Non-point source pollution; Subsurface drainage; Tile;

INTRODUCTION

This article is part of a collection that provides a comprehensive review and evaluation of the performance and cost effectiveness of selected agricultural conservation practices on nutrient and sediment reduction. The focus of this article is on denitrifying bioreactors for treatment of nitrate-N from agricultural water discharge.

The term denitrifying bioreactor most generally refers to a trench filled with carbonaceous media through which nitrate-laden water is routed (Christianson et al., 2012a; Schipper et al., 2010a). Wood is most commonly used as the carbon source inside these bioreactors. Denitrifying bacteria are fueled by the wood-sourced carbon to convert the nitrate-N in the water to dinitrogen gas. In more scientific terms, the woodchips supply dissolved organic carbon (DOC) to serve as the terminal electron acceptor in the dissimilatory stepwise reduction of nitrate-N to dinitrogen performed by chemo-heterotrophic denitrifying bacteria under anoxic conditions. This enhancement of the biological process of denitrification lends the name *bio*-reactor.

Schipper et al. (2010a) proposed definitions of three types of denitrifying bioreactors – beds, walls, and layers – to distinguish between designs based on their hydrological connections. The bed designs have generally been the most common with recent interest leading to in-ditch variations (fig. 1a and b). The second most common type, denitrifying bioreactor walls, require less complicated design and construction (fig. 1c). Denitrifying layers are not discussed here.

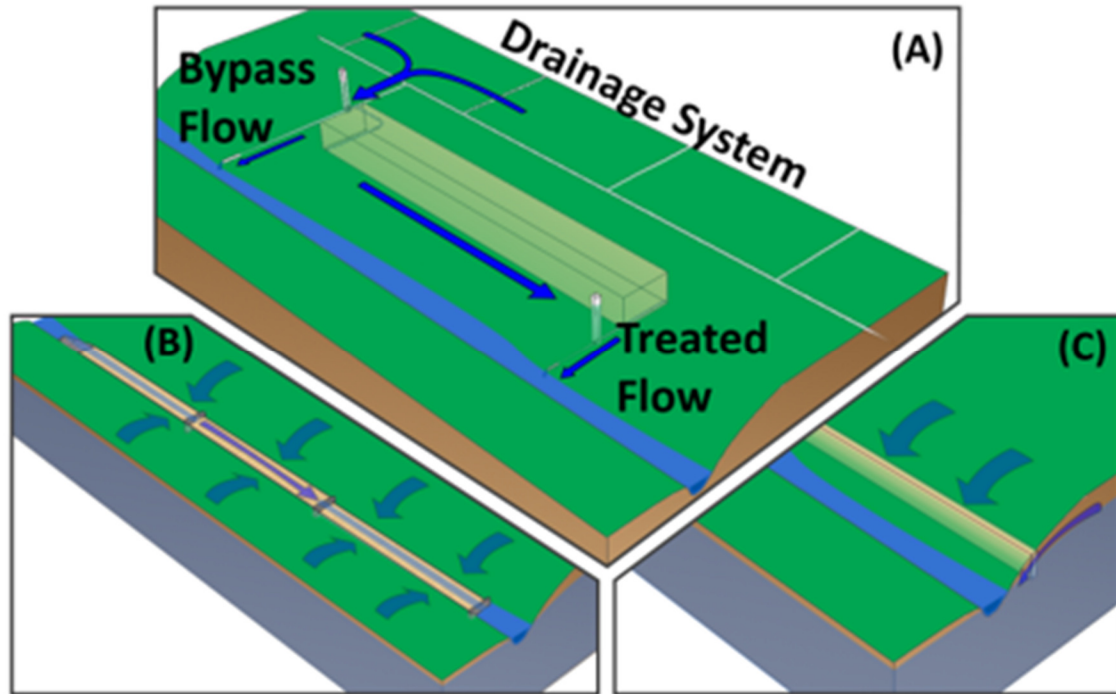


Figure 1. Three types of wood-based denitrifying bioreactors discussed in this article: (a) bed-style with internal plumbing; (b) in-ditch beds with protective gravel or mesh; and (c) wood-based denitrifying wall installed parallel to stream or ditch.

Denitrification beds are woodchip-filled pits or trenches below the soil surface that receive water through a pipe either connected to a subsurface tile drain or a wastewater outlet (fig. 1a). Beds are the most common type of woodchip bioreactor in the US, and their design and construction for the treatment of nitrate in subsurface drainage water is guided by the United States Department of Agriculture Natural Resources Conservation Service Conservation Practice Standard (Conservation Practice Standard 605: Denitrifying Bioreactor; USDA NRCS, 2015). Subsurface drainage flow is driven by the head gradient created across the bioreactor following principles of flow through porous media; pumps are typically not used for this application (e.g., Christianson et al., 2012b; Rosen and Christianson, 2017; Woli et al., 2010). Internal plumbing manifolds and control structures route and control the saturated depth and flow. Top-down and other flow configurations have been used in other bed designs (e.g., Bruun et al., 2016; Schipper et al., 2010b; von Ahnen et al., 2018).

The NRCS Conservation Practice Standard recommends a design hydraulic retention time of 3 h at a design flow rate that is at least 15% of the peak estimated flow rate from the drainage system (USDA NRCS, 2015). While this “15% criterion” is the most commonly used, designers can also choose to design a bioreactor based on treating the peak flow from a 10-year, 24-hour drain flow event or treating at least 60 percent of the long-term average annual flow from the drainage system (USDA NRCS, 2015). Development of this Conservation Practice Standard in the early 2010s lent important credibility to the practice

of denitrifying bioreactors and critically allowed federal incentive payments to assist with bioreactor construction. The standard was based on the few published field-scale studies available at the time, but because practice standards undergo periodic review and revision, future considerations should include refining the standard (e.g., Should the standard be load reduction outcomes-based rather than capacity-based? Are there sufficient data to support a load reduction-based standard?) and streamlining the design process (e.g., Are standardized plug-and-play bioreactor designs possible?).

Retrofitting bed-style bioreactors to be situated in streams or drainage ditches is a practical idea supported by farmers to minimize the risk of land removed from production (fig. 1b). Consideration must be given to minimizing flow restriction with in-ditch bioreactors as flow conveyance is the primary responsibility of a ditch network. These designs have included wooden berms, gravel, and/or wire or plastic mesh to create woodchip “bags” or “mattresses” (Chase et al., 2019; Christianson et al., 2017; Dhaese et al., 2019; Pfannerstill et al., 2016; Robertson and Merkley, 2009). Design of these in-ditch systems has borrowed from the NRCS Conservation Practice Standard, but these systems have unique design, construction, and maintenance concerns (e.g., by-pass flow goes over the top of the bioreactor, sedimentation is a significant issue).

Denitrification walls are installed across the flow path of groundwater often parallel and adjacent to an affected stream or drainage ditch (fig. 1c; Schipper and Vojvodic-Vukovic, 1998; Schmidt and Clark, 2012). The wall can be constructed using 100% carbon material or as a mixture of carbon material and native soil. There is no lining, and where possible, installing the bottom of the porous wall into a soil layer of low permeability is recommended to minimize flow short-circuiting under the reactive barrier. A major distinction between denitrifying beds and walls is that flow rates through beds tend to be much greater than that of walls. It is worth noting that while the NRCS CPS 605 was written with denitrifying beds in mind, it applies to sites where there is a need to reduce the nitrate-nitrogen concentration in subsurface drainage flow and does not require a certain type of implementation.

The overall objective of this paper is to consolidate the state of the science for full-size denitrifying bioreactors across the globe to supplement the existing reviews and syntheses on this topic (Addy et al., 2016; Christianson et al., 2012a; Schipper et al., 2010a). “Field-scale” and “full-size” bioreactors were loosely defined as meeting most of these factors: a bioreactor that was built on an operational farm (including some research farms); a given bioreactor (or bioreactor system, i.e., paired bioreactors in parallel) that was sized to receive drainage from a given drainage main (or other outflow system, e.g., agricultural wastewater); a bioreactor with a drainage treatment area of greater than 2 ha; no on-site replication was possible given the magnitude of the treatment system. An additional distinction between “pilot-scale” and “field-scale” is that the former is generally mainly intended for comparative assessments between treatments as compared to providing a direct assessment of

bioreactor performance under “real-world” conditions at full scale. Bioreactors treating subsurface drainage are emphasized since this special collection focuses on USDA NRCS Conservation Practice Standards, but a variety of designs, applications, and water chemistries are included to more comprehensively reflect the current state of knowledge.

PERFORMANCE EFFECTIVENESS

SEDIMENT

Denitrifying woodchip bioreactors are capable of removing sediment via the physical process of filtration, but this capability can eventually hinder N-removal performance. There are little field-scale data on sediment removal, likely because one of the main applications of denitrifying bioreactors in the US is subsurface drainage water which tends to have low sediment and suspended solids. When woodchip bioreactors are paired downstream of wetlands or sedimentation basins, total suspended solids (TSS) removal is generally excellent across the entire system (i.e., >90%; Choudhury et al., 2016; Tanner et al., 2012). Christianson et al. (2016) achieved >90% TSS removal from aquaculture wastewater in pilot-scale bioreactors operated for more than 250 days, and in a similar wastewater study, Lepine et al. (2020) reported TSS removal decreased from >90% to at least 64% after 637 days of operation when a total of 26 kg of TSS had been loaded into each bioreactor. Choudhury et al. (2016) estimated only 7% of the woodchip pore space had been filled by sediment after a settling basin-woodchip bioreactor treatment system had been operated for seven months for high TSS wash water (5800 ± 2700 mg TSS/L). Woodchip clogging and siltation will eventually reduce bioreactor performance via flow restriction and development of preferential flow paths (Christianson et al., 2020). Using a woodchip bioreactor for sediment removal will shorten the design life for denitrification purposes but the extent and timeframe of this will likely depend on factors including the water matrix, loading rates, and bioreactor media.

NITRATE-NITROGEN

This description of full-size bioreactor nitrate-N removal performance is organized as a series of case studies contributed by the wide geography of co-authors. A range of applications and designs are presented to highlight local contextual insights and practical on-site knowledge. However, these case studies do not serve as an exhaustive list of every full-size bioreactor across the globe. A number of additional studies (e.g., Canadian studies: Gottschall et al. (2019), Husk et al. (2017); Danish studies: Bruun et al. (2016); Swedish treatment of mine drainage: Nordstrom and Herbert, 2019) are mentioned more appropriately in other sections per the focus of the given study.

In general, subsurface drainage denitrifying bioreactors remove from 20 to 40% of the annual nitrate-N loss according to science reviews in Midwestern state nutrient strategies (IDALS, 2014; IDOA, 2015). This typically ranges from 0.5 to 20 kg N/ha removed from fields with N losses of 1.0 to >60 kg N/ha (Christianson et al., 2012b; Woli et al., 2010). A 2016 meta-analysis showed N removal rates most commonly range from 0.5 to 10 g N removed per cubic meter bioreactor per day (g N/m³-d) for bed-style designs (Addy et al., 2016). A more recent compilation of studies is presented in Appendix A which shows that the majority of bioreactor beds in the most current pool of literature (since 2016) are treating inflow nitrate-N concentrations in the range of 5.20-9.75 mg NO₃-N/L (25th and 75th percentiles; Figure 2a) with hydraulic retention times of generally less than 2 d (75th percentile: 44 h; Figure 2b). Nitrogen load reductions across the most current literature (fig. 2c; median: 46%; mean ± st dev: 40±26%) were consistent with Midwestern state strategy documents, although these bioreactors were from a variety of locations worldwide and the inclusion of untreated bypass flow in these load reduction efficiencies was not reported consistently across literature (i.e., a mix is represented here). Nitrogen removal rates aligned with the earlier meta-analysis by Addy et al. (2016) with a median of 5.1 g N/m³-d (fig. 2c; mean ± st dev: 7.2±9.6 g N/m³-d;).

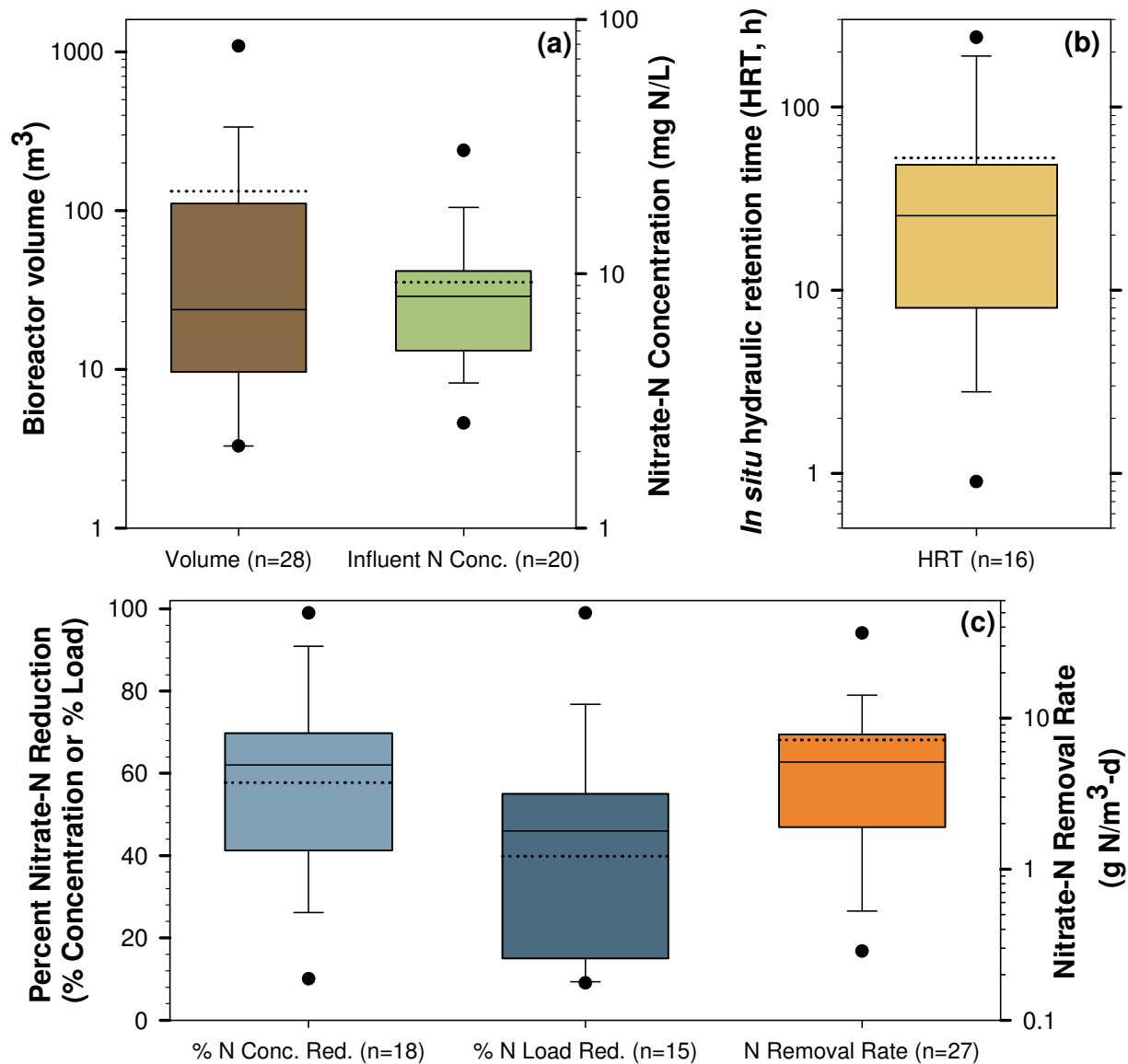


Figure 2. Survey of details compiled from full-size peer-reviewed bioreactor studies published since the meta-analysis by Addy et al. (2016). See Appendix A for data. Boxes, stems, and dots represent the 25th and 75th, 10th and 90th, and 5th and 95th percentiles, respectively; the solid line is the median and the dotted line is the mean.

Subsurface drainage denitrifying bioreactors in the U.S. Midwest

Illinois

Some of the earliest documented subsurface drainage denitrifying bioreactor studies in the United States were performed in Illinois (Cooke et al., 2001). By 2019, Illinois had 37 field-scale bioreactors treating approximately 1,345 drained acres which was an increase from 20 reported bioreactors (611 treated acres) in 2017 (IDOA and IEPA, 2019). Many of these are research

and demonstration sites and are not designed following the USDA NRCS Conservation Practice Standard (USDA NRCS, 2015). The bioreactors currently being monitored that were designed to the NRCS standard have treated drainage areas of 2.8-20 ha and tend to be 9.8-23 m long and 1.8-5.8 m wide with the depth set by the depth of the existing drainage system. Roughly seven site-years from four NRCS-designed bioreactors generally show 15-98% of the water from the field has been treated (2,900-26,200 m³ treated annually) with 12-98% of the nitrate routed into the bioreactor removed (5.4-168 kg N removed annually; 0.8-8.3 kg N/ha removed annually; removal rates: 2.1-6.6 g N/m³-d). Nitrate-N loss reductions at the edge of the field considering the un-treated bypass flow tend to be around 20% for these NRCS-designed bioreactors (L. Christianson, unpublished). The Illinois Nutrient Loss Reduction Strategy currently assesses bioreactors at a 25% edge-of-field N loss reduction (IDOA, 2015).

Full-size bioreactor research in Illinois currently focuses on refining design criteria and processes and advancing unique designs. A bioreactor designed with baffles to more efficiently route flow has produced edge-of-field N load reductions of 22-24% over three years by treating 40-43% of the total flow from the field (0.71-1.3 g N/m³-d; 2.70-4.03 kg N/ha removed annually; Christianson, unpublished; Dougherty, 2018). Pairing bioreactors: (1) to operate in parallel under high flow conditions, (2) to treat in-ditch drainage water and water that has been diverted out of the ditch, and (3) with phosphorus removal structures are being trialed. Across the state, some of the most significant observed challenges include side wall-slumping as the woodchips degrade and plugging of the inflow pipe likely due to excessive microbial growth and/or siltation near the inlet (David et al., 2016). Side-wall slumping can be mitigated by refilling with woodchips (open-top bioreactors, at least), but consistent underlying causes of early woodchip subsidence have yet to be determined. Modeling efforts to better predict bioreactor performance, with the ultimate aim of improving design procedures, are also a major emphasis of the work in Illinois (Cooke and Bell, 2014; Jang et al., 2018).

Iowa

One of the first studies of multiple field-scale bioreactors monitored four bioreactors in Iowa (Christianson et al., 2012b). Over 14 site-years, annual nitrate removal rates ranged from 0.38 to 7.76 g N/m³-d. Nitrate load reductions ranged from 12% to 76% (mean 45%) for flow through the bioreactor and 12% to 57% (mean 32%) for total flow (including bypass flow), which translated to 0.5 to 15.5 kg N/ha removed. Jones and Kult (2016) monitored five bioreactors in north central Iowa treating 10 to 20 ha of drainage area. All five bioreactors were designed to treat 20% of the estimated peak flow rate at a 4-h design hydraulic retention time. Average nitrate load reductions ranged from 50% to 80% for flow through the bioreactor, and for days

when flow was measured, average nitrate removal rates ranged from 1.67 to 6.68 g N/m³. The science assessment in the Iowa Nutrient Reduction Strategy assigns an average nitrate load reduction of 43% ± 21% (mean ± standard deviation) to bioreactors (IDALS, 2014), and there was a conservative estimate of 25 bioreactors in the state at the end of 2017 (IDALS, 2019).

The Greene County bioreactor, constructed in 2008 and monitored in both the Christianson et al. (2012b) and Jones and Kult (2016) studies, was recharged with new woodchips in 2017. A series of lab tests paired with field observations indicated that hydraulic changes from sedimentation and woodchip degradation created potential flow restrictions and preferential pathways. This, along with degradation in wood carbon content and quality, resulted in the need to recharge the bioreactor after 9 y (Christianson et al., 2020).

Minnesota

Woodchip bioreactors were introduced in Minnesota in 2009. The first two monitored field-scale bioreactors were located at Dundas and Claremont, MN and were of a narrow, long design (Ranaivoson et al., 2012). Nitrate-N and total P load removal at the Dundas site (LWD: 27 x 0.90 x 1.2 m, woodchip depth) were 47 and 78%, respectively, over a test period of three six-day cycles at a 21-h hydraulic retention time (Ranaivoson et al., 2019). There are sporadic reports of unexpectedly early woodchip subsidence (e.g., within a few years of installation) at some Minnesota sites. Minnesota's northern climate creates cold weather challenges for bioreactor performance. Design and bioreactor fill modifications may be needed to improve performance under such conditions (Feyereisen et al., 2016). Current field research includes hydrologic and performance analysis of a unique multi-bed, cascading system at a community scale (250-ha watershed).

South Dakota

South Dakota State University has monitored four bioreactors as part of its ongoing research into management practices to reduce nutrient transport to surface waters. Two bioreactors were installed in 2012 (Baltic and Montrose, SD), one in 2013 (Arlington, SD), and one in 2014 (Hartford, SD) (Partheeban, 2014). These bioreactors were designed with hydraulic retention times ranging from 4.9 to 6.3 h at design flow rates that were 18 to 25% of the estimated peak flow rate from the drainage system. Dimensions of the initial three ranged from lengths of 35-40 m and widths of 3.7-6.4 m (Partheeban, 2014). Installation costs ranged from nearly \$7,900 to \$10,400. The bioreactors receive subsurface drainage from predominantly corn (*Zea mays*) and soybean (*Glycine max*) row crop systems, with the Hartford bioreactor inflow likely also receiving some manure influence from the nearby cattle confinement area.

During the monitoring periods (late spring to early fall), the bioreactors received flow for a majority of the time. Nine of the

11 site-years had flow into the bioreactor for over 71% of the monitoring period with three site-years receiving flow 100% of the monitoring period. Average flow rates were 5.6, 24.5, 9.7, and 0.62 L/s for the Montrose, Arlington, Baltic, and Hartford bioreactors, respectively, demonstrating a variety of flow regimes (≈ 0.7 to 31 mm/d). The bioreactors generally removed the majority of the entering N load (i.e., >50% concentration reductions). Mean nitrate concentration reductions for the individual sites were 54% (Baltic, three years monitored, $n = 41$), 58% (Montrose, three years monitored, $n = 39$), 60% (Hartford, two years monitored, $n = 19$), and 80% (Arlington, three years monitored, $n = 39$), which were relatively consistent with Fig. 2c.

Drainage denitrifying bioreactors in the Eastern U.S.

Maryland and the Delmarva Peninsula

Ditches and tile drainage have benefitted production within this region's coastal plain but have created an efficient pathway for nutrients to enter the Chesapeake Bay. There have been three field-scale tile drainage bioreactors monitored in Maryland and four field-scale ditch bioreactors monitored in Maryland and one in Delaware. Rosen and Christianson (2017) reported the three tile drainage bioreactors were able to reduce nitrate-N loads between 9 and 62% and had an overall average load reduction efficiency of 24% (removal rates of 0.40-5.36 g N/m³-d). Beyond tile drainage, both bed-style and wall-style bioreactors have been adapted for the extensive ditch network found throughout Maryland and Delaware to improve the relevance of this practice to achieve Chesapeake Bay water quality goals. Three design modifications have been trialed: ditch-diversion bioreactors designed according to the USDA NRCS Conservation Practice Standard, in-ditch bioreactors, and sawdust denitrification walls. The two ditch-diversion bioreactors have achieved 2 and 25% N load reduction efficiencies (Rosen, unpublished; Christianson et al., 2017). Load reduction efficiencies could not be calculated for the in-ditch bioreactors or sawdust denitrification walls due to difficulties accurately assessing flow volumes; however, both designs achieved N concentration reductions >65%.

From a practical application perspective, the in-ditch bioreactor will need annual maintenance to reduce sedimentation. The sawdust wall, although inexpensive to install, will need hydrogeologic investigations to determine groundwater flow direction and to ensure groundwater does not bypass under or around the wall. The ditch diversion bioreactor follows the USDA NRCS practice standard making design and implementation relatively straightforward, but, as with the wall, site selection is critical because ditch networks can intercept various sources and volumes of water that have variable concentrations of nitrate-N. The ditch-diversion bioreactor also proved relatively expensive to install (>\$20,000). Bioreactor beds have recently been recommended to receive a 20% Total Nitrogen (TN) loss reduction credit in the Chesapeake Bay total maximum daily load model (Bryant et al., 2019).

New York State

Ten denitrifying bioreactors have been constructed on five farms throughout central and western New York for research and monitoring. These were generally located on dairy farms treating effluent from between 2 and 12 ha of tile drained fields. Bioreactors were sized based on estimates of field drainage and a design hydraulic retention time of 20 h (LWD: 6 to 9 m x 3 to 4.5 m x 1 to 1.5 m). At some sites, paired bioreactors were used, with one of each pair amended with biochar (10:1 woodchip to biochar ratio by volume) although the impacts of these additions are dependent upon the specific biochar's properties. The N removal rate across all bioreactors averaged approximately 5 g N/m³-d (annual range: 3 to 15 g N/m³-d). Temperature had a significant control on N removal rate, with temperatures below 5°C resulting in lower rates (Hassanpour et al., 2017). Low nitrate inflow concentration and low DOC in effluent also resulted in limited removal rates, though only low DOC resulted in low removal efficiency. Both removal rate and efficiency dropped significantly after storm events; however, the highest instantaneous removal rates were measured during peak flows (Pluer et al., 2019).

International bioreactor applications

New Zealand

In New Zealand, the first denitrification wall (40 m long) was constructed in early 1996 to determine whether this approach could be used to remove nitrate from shallow groundwater of a grazed dairy farm. A series of studies demonstrated that nitrate removal exceeded 95% (Schipper and Vojvodic-Vukovic, 1998), denitrification was the mechanism of removal (Schipper et al., 2005; Schipper and Vojvodic-Vukovic, 2000) and that removal occurred for at least 15 years (Long et al., 2011). Nitrate-N removal rates (~1 g N/m³-d) were lower than reported for denitrification beds because there were lower carbon contents (roughly 5% C by weight) as sawdust was mixed with soil on-site and because nitrate inputs were low due to slow movement of groundwater (Addy et al., 2016). A second wall (120 m long) installed nearby into sandy soils experienced bypass flow due to a large reduction in hydraulic conductivity (Schipper et al., 2004) due to repacking of sand (Barkle et al., 2008) demonstrating that care was needed with denitrifying wall installation in aquifers with high conductivity. From the mid-2000s, large denitrification beds (LWD ranging from 40 to 150 m x 4 to 7 m x generally 1.5 m) were constructed for treatment of N-laden water from subsurface drains, domestic wastes, dairy farm effluent and hydroponic glasshouse runoff production. High rates of nitrate removal were demonstrated (e.g., 5-10 g N/m³-d; Schipper et al., 2010b; Warneke et al., 2011b) that were shown to be supported by denitrification (Warneke et al., 2011a).

Southeast Queensland, Australia

The Pumicestone Passage in southeast Queensland is a valuable ecological and recreational waterway located 70 km north of Brisbane between Bribie Island and the Australian mainland. Land use adjacent to catchment waterways is dominated by intensive horticulture comprised predominantly of strawberry, pineapple, and macadamia production. Annual water quality assessments for the Passage and its freshwater catchment have shown a decline since 2006, with significant loads of nitrate contributed by the agricultural area in the northern region of the catchment. Manca et al. (2020) described the design and performance of two denitrification walls (LWD: 20 x 1.0 x 1.4 m; $\approx 27 \text{ m}^3$ each; 3 m apart) which were constructed in June 2017 and filled with either a softwood (*Pinus caribaea*) or hardwood mix (*Eucalyptus* spp.). Design parameters primarily accounted for depth to the aquitard, available land, surface drainage ditches, size of the excavator and attachments (i.e., the wall width was set to the width of the excavator bucket), required volume of woodchips, and sampling needs. The installation cost of the bioreactors was approximately \$50AUD/m³ (\approx \$35USD/m³) including materials and excavation.

The denitrification walls removed all the nitrate-N transported in groundwater (inflow range 0.0-22.7 mg NO₃-N/L) based on weekly sampling over two years. Such high removal was facilitated by the development of anaerobic conditions (average <1.1 mg/L DO) and high temperatures (21-27 °C) with persistent nitrate limitation and higher methane production during transition to unsaturated conditions. The average removal rates for the softwood and hardwood walls were 2.0 and 1.6 g N/m³-d, respectively, consistent with previous denitrification walls (Fahrner, 2002; Jaynes et al., 2008; Robertson et al., 2000). Results were similar to values reported by Schipper et al. (2005) due to occurrence of nitrate limitation in the walls, and such limitations necessarily caused uncertainty in the quantification of maximum removal rates..

In 2015, the Australian and Queensland Governments released the Great Barrier Reef 2050 Long-Term Sustainability Plan to provide an overarching framework for managing water quality in Great Barrier Reef lagoon (Commonwealth of Australia, 2018). Denitrifying bioreactors have been identified as a potential treatment system option for reducing point source and diffuse environmental nitrate loading from agriculture into the lagoon. Bioreactors are currently being trialed in a number of catchments throughout Queensland to test their N removal efficacy across differing climates, soil types, and agricultural production systems. As of November 2019, it is estimated 30 comparably monitored bioreactors are operating in Australia with more than 20 of the systems in Northern Queensland within close proximity to the Great Barrier Reef (i.e., the “Australian Bioreactor Network” (ABC Rural News, 2020)).

Belgium

A woodchip bioreactor (LWD: 65 x 2 x 1 m; 127 m³ of hardwood species) was installed in July 2018 in a ditch in Peer, Belgium (Flanders), that transports nitrate-rich drainage water from sandy fields. The relatively high nitrate concentrations (e.g., > 30 mg NO₃-N/L) cannot be linked directly to current in-field fertilizer management but are likely rather due to historic nitrate-rich groundwater reaching the surface. From June 2018 to July 2019, the mean bioreactor inflow and outflow NO₃-N concentrations were 28.6 and 8.9 mg NO₃-N/L, respectively (mean of 36 measurements), which corresponded to an average concentration reduction of 69%. The maximum concentration reduction for a given sample event was 97%. There was an anecdotal increase in N-removal during higher temperatures, which has been well documented by others (Addy et al., 2016).

The oak (genus *Quercus*) wood used in the bioreactor leached high amounts of color (even blackish color) for several weeks after installation. Very low dissolved oxygen content was measured in the ditch for at least 9 d following installation. One and a half years after installing the bioreactor, an additional 15 m³ of woodchips was added to prevent too much bypass flow over the top of the in-ditch design.

As an EU-member state, Flanders is held to a threshold value of 11.4 mg NO₃-N/L (or, 50 mg NO₃-/L) in surface and groundwater (Council of the European Union, 1991). Despite more than twenty years of work toward these goals, many monitoring locations in Flanders still exceed these values. In 2019, the Flemish Government imposed new measures to accelerate progress in areas where the water quality threshold value has not been achieved including: (1) reduction of fertilizer rates; (2) increase in cover crop area; and (3) liquid animal manure can only be transported to arable fields by a recognized contractor. From 2020 onward, equivalent measures that have been approved by a special commission can be used by farmers to replace these three, given the N mitigation effectiveness of the equivalent measure has been proven. Denitrifying woodchip bioreactors could possibly be a useful end-of-pipe “equivalent measure” technique in catchments where high nitrate concentrations in surface water cannot be linked to current fertilization practices.

Denmark

In Denmark, full-scale woodchip bioreactors in the size range of 350 to 6,000 m³ have been installed to treat the effluents from commercial recirculating aquaculture systems (RAS) rearing rainbow trout (*Oncorhynchus mykiss*) in freshwater. Aquacultural effluents typically contain higher loads of particulate organic matter compared to subsurface agricultural drainage, which poses a higher risk for bioreactor clogging. To reduce the risk for bioreactor clogging and associated head loss, woodchip bioreactors were built as vertical down-flow filters with depths of 1.0 to 1.5 m with several outlet drainage pipes in the bottom

to facilitate even distribution of water within the bioreactors. The bioreactors were installed within constructed wetlands to reduce the incoming particulate load and dissolved oxygen into the woodchips and alleviate the environmental impact of leached nutrients and organic matter on receiving water bodies during bioreactor start-up. In a one-year monitoring study, three full-scale woodchip bioreactors, which were operated at empty bed contact times of 15-21 h (i.e., volume of the empty reactor divided by the flow rate; doesn't consider woodchip porosity), achieved average nitrate-nitrogen removal rates of 4.5 – 7.8 g N/m³-d at average water temperatures of 10°C (von Ahnen et al., 2018). The monitoring study confirmed that woodchip bioreactors can be applied as a technologically simple, low-maintenance method to remove nitrogen from aquaculture effluents, where their operation benefits from stable flow rates and nitrate loads. The costs for installation of woodchip bioreactors at Danish recirculated trout farms were 400-500 DKK/m³ bioreactor volume (\$60 - \$75USD/m³). The cost per kg N removed for this relatively high-solids application will be highly dependent on the required frequency of woodchip replacement (Lepine et al., 2018). The oldest woodchip bioreactor at a Danish RAS farm has only been operating for 2.5 years (at the time of writing) and still removes nitrate without any observations indicating replacement of woodchips is necessary.

TOTAL NITROGEN

Much of the total nitrogen (TN) load in subsurface drainage in the US, the current most common application for woodchip bioreactors, is comprised of nitrate-nitrogen. The recent expert panel recommendation for the Chesapeake Bay Total Maximum Daily Load model credits denitrifying bioreactors designed to the NRCS standard with a 20% TN reduction credit based on this assumption (Bryant et al., 2019). However, this will be water matrix specific as nitrogen speciation will vary in wastewater or subsurface drainage in other locations (e.g., Irish drainage waters can contain relatively more ammonium-N than nitrate-N; Clagnan et al., 2018).

DISSOLVED AND TOTAL PHOSPHORUS

Phosphorus (P) is a macronutrient required for plant growth, and thus, woodchips contain and will leach dissolved P. There are few full-scale studies documenting P dynamics in denitrifying woodchip bioreactors treating subsurface drainage water; the results show a mix of P leaching and removal although most generally document P leaching upon bioreactor start-up. Flushing of P beyond the first year was observed by David et al. (2016) at a bioreactor in Illinois (LWD: 15 x 6.0 x 1.3 m; 20 ha drainage area; installed 2012) where dissolved reactive phosphorus (DRP) loads entering and leaving the bioreactor were 0.1, 1.0, and 0.3 kg DRP versus 1.3, 4.7, and 1.0 kg DRP, respectively, across a three year period (2012-2014). The two years where total phosphorus (TP) was monitored showed a similar trend (inflow: 1.9 and 0.5 kg TP; outflow: 6.4 and 1.4 kg TP). Other

bioreactors in Illinois have confirmed initial P flushing with Bell et al. (2015) showing outflow DRP concentrations can be an order of magnitude greater than the inflow within the first month of operation (≈ 0.1 - 4.5 and <0.1 mg DRP/L, respectively). Over the next six months of monitoring, the outflow concentrations were consistently elevated above the inflow, but these differences were not statistically significant. Phosphorus removal in woodchip bioreactors is also possible, however, as Dougherty (2018) reported consistent P removal at a third bioreactor in Illinois (LWD: $16.8 \times 10.7 \times 0.91$ m; 14 ha drainage area; installed 2016). The bioreactor removed 72 and 74% of the inflow dissolved P load in its first and second year, respectively, which equated to 0.01 and 0.08 g P removed/ m^3 -d for the two periods. These trends of initial P flushing with the possibility of removal generally hold across a variety of bioreactor applications such as agricultural wash water (Choudhury et al., 2016), greenhouse effluent (Warneke et al., 2011b), and aquacultural wastewater (Sharrer et al., 2016: removal rates of 0.51 to 0.74 g TP/ m^3 -d and -0.74 to 0.23 g DRP/ m^3 -d; von Ahnen et al., 2018: -0.2 to 0.1 g TP/ m^3 -d).

Much work at the intersection of P and denitrifying woodchip bioreactors has been around the integration of P-sorbing media with the woodchips. Husk et al. (2018) compared one mixed media bioreactor containing a gabion filled with activated alumina and gravel upstream of woodchips with three woodchip-only controls (range of LWDs: 12 - 27×0.9 - 1.8×0.9 ; drainage areas: 0.6-1.3 ha). They reported that all four bioreactors released P during the first year of operation, but overall, the bioreactor with the P-sorption media was more effective for P removal than the woodchip-only bioreactors, with removal rates of 0.36 and 0.02 g TP removed/ m^3 -d, respectively. Neither the mixed media bioreactor nor the woodchip-only bioreactors were able to treat the drainage water to below the threshold for eutrophication of freshwaters of 0.03 mg TP/L. Nevertheless, this serves as one of the most current comprehensive studies of woodchip bioreactor P removal particularly given the variety of P forms evaluated. The authors noted: *“This three-year field-scale study demonstrates that an agricultural subsurface drainage system with a woodchips-only bioreactor had lower P load in its outflow and a significant reduction in the bioavailable, soluble P fractions”* (Husk et al., 2018).

There will be variability in the P-sorbing media used as Bock et al. (2018) reported differences between inlet and outlet TP concentrations at a bioreactor (LWD: $5.8 \times 5.3 \times 0.8$ m; 6.5 ha drainage area) filled with volumetric ratio of 90/10 woodchips/biochar were not significant (mean inflow and outflow: 0.13 and 0.03 mg TP/L, respectively). Several pilot- and lab-scale studies investigating the combination of woodchips and P-sorbing media have documented some dissolved P removal (generally 5-15%) with woodchip-only treatments or column sections (Goodwin et al., 2015; Zoski et al., 2013; Li et al., 2018). Gottschall et al. (2016) reported small replicated bioreactors (LWD: $2.4 \times 1.5 \times 0.9$ m) packed with only woodchips had TP and DRP removal efficiencies of 28 and 35%, respectively. Bioreactors amended with alum-based drinking water treatment residuals

provided more P removal (64% TP and 89% DRP; Gottschall et al., 2016). Hua et al. (2016) reported as much as 75% phosphate removal and removal rates as high as 0.88 g P/m³-d by woodchips themselves in two phases of a small experiment pairing woodchips and steel slag.

In summary, there was variability in denitrifying bioreactor P leaching/removal across literature. The mechanisms and consistency of this removal are unclear (Warneke et al., 2011b). The importance of P (and its variety of forms, e.g., Husk et al., 2018) for freshwater eutrophication make this an important area of continued research for woodchip bioreactors.

REMOVAL OF ADDITIONAL POLLUTANTS

The removal of pollutants beyond those mentioned above is a ripe topic for bioreactor research. Much early work in this area has necessarily been done at the lab-scale (e.g., Ilhan et al. (2010): atrazine, enrofloxacin, and sulfamethazine; Krause-Camilo et al. (2013): atrazine; Zoski et al. (2013): *E. coli*; Liu et al. (2014): perchlorate; Bell (2019): plant pathogens of the *Phytophthora* species) with a handful of field-scale studies reported here. For example, herbicide dissipation was evaluated at one of the Minnesota field-scale bioreactors described above (Dundas, MN). After three six-day cycles at a 21-h hydraulic retention time, load reductions of acetochlor and atrazine were 70 and 53%, respectively. However, adsorption was identified as the removal mechanism, and modeling indicated herbicide breakthrough would occur within 18 days (Ranaivoson et al., 2019). Other recent work by Hassanpour et al. (2019) corroborated woodchip bioreactors' ability to remove atrazine from drainage water in the field. Gottschall et al. (2016) reported woodchip-only bioreactors receiving drainage from plots applied with liquid swine manure removed more than 70% of the tylosin, chlortetracycline, and isochlortetracycline in the water. They reiterated the removal mechanism for veterinary antibiotic compounds appeared to be sorption; however, the ultimate fate of these chemicals is not known. Removal of microorganisms including bacteria and viruses has been documented with field-scale bioreactors (Rambags et al., 2016; Tanner et al., 2012). Most recently, mesocosm work by Rambags et al. (2019) demonstrated ANAMMOX, where ammonium oxidation is combined with nitrate reduction to produce the final product of nitrogen gas, was equally important to denitrification as a removal mechanism. The controls on this process were unclear but it will only occur when there is sufficient dissolved ammonium and nitrate present.

LIMITATIONS AND TRADEOFFS

No given conservation practice is a “silver bullet” for every situation or pollutant, thus bioreactors have their limitations and tradeoffs, some of which were mentioned in the above sections. For example, practical maintenance considerations observed in

the field include sporadic occurrences of woodchip slumping (IL and MN sections above) and the need to remove sediment from in-ditch bioreactors (MD section above). Ditch-diversion bioreactors with a structure for water control to back up water within a ditch can also create stagnant water conditions facilitating an anaerobic environment conducive to phosphorus release (Sharpley et al., 2007). This is an important consideration for areas in Maryland and Delaware that have phosphorus-saturated soils. Such potential pollution swapping (i.e., release of one pollutant while trying to treat another) is an important consideration (Healy et al., 2012).

Many bioreactor pollution swapping studies have explored losses of dissolved organics and nutrients (e.g., DOC, chemical/biological oxygen demand, organic nitrogen, ammonium, phosphorus) at the bioreactor outlet especially during bioreactor start-up and emissions of greenhouse gases (nitrous oxide (N_2O), methane, and carbon dioxide). Many small-scale bioreactor fill media comparisons have reported these start-up nutrient flushing issues (e.g., Gibert et al., 2008; Cameron and Schipper, 2010; Healy et al., 2012) and colored bioreactor outflow water has been observed during start-up at the field-scale (e.g., Belgium section above). Additional woodchip leaching concerns such as phenols and tannic acid are starting to be investigated (Wickramarathne et al., 2020; Lepine et al., Under Review). These issues subside given sufficient flushing which has ranged on the order of 15 to roughly 300 cumulative pore volumes or a week to approximately six months depending on bioreactor size, fill media, and initial flow rate (Healy et al., 2012; Christianson et al., 2016; von Ahnen et al., 2016; Wickramarathne et al., 2020).

When considering greenhouse gas emissions, it is critical to not only measure emissions from bioreactors but to compare these to emissions that would have occurred in the absence of the bioreactor. For example, woodchips used for constructing a bioreactor would likely degrade with time in any case so that carbon dioxide emissions from a bioreactor do not represent a net carbon dioxide contribution to the atmosphere. Similarly, in the absence of a bioreactor, a proportion of the leached nitrate could also be converted to N_2O in receiving waters (e.g., Moorman et al., 2010). In one of the first full-scale bioreactor studies to evaluate N_2O losses, Elgood et al. (2010) reported only 0.6% of the consumed $\text{NO}_3\text{-N}$ was released as N_2O in an in-stream bioreactor. Goeller et al. (2019) recently reported greenhouse gas fluxes from a subsurface drainage bed-style bioreactor in New Zealand were similar to emissions from the surrounding pasture, and Bock et al. (2018) reported greenhouse gas emissions from a bioreactor in Virginia, USA were similar to other nitrate treatment systems and were not “environmentally concerning”. Similarly, Woli et al. (2010) reported surface N_2O fluxes at a bioreactor in Illinois were “of little concern”. However, current work does make clear that dissolved gasses in addition to surface fluxes should be considered. Warneke et al. (2011b) and Bruun et al. (2017) both reported the majority of N_2O losses were in the dissolved form rather than emitted via the surface

although total N₂O emissions were still only 4.3% and between 2.2-5.5%, respectively, of removed nitrate. Bruun et al. (2017) and Elgood et al. (2010) both observed that bioreactors could, at times, be N₂O sinks from the atmosphere. However, methane emissions, which are another greenhouse gas consideration, have been observed under conditions leading to full consumption of nitrate including low hydraulic loading and high temperatures (Elgood et al., 2010; Bruun et al., 2017).

Highly anaerobic conditions leading to methanogenesis can also be conducive to sulfate reduction. On one hand, sulfate reduction in bioreactors, which has been relatively widely documented (Warneke et al., 2011b; Shih et al., 2011; Christianson et al., 2012b; Corbett et al., 2020), could be considered treatment of an additional pollutant. On the other hand, sulfate reduction is associated with mercury methylation in natural systems. Mercury methylation in woodchip bioreactors can be avoided by maintaining bioreactor outflow nitrate concentrations of at least 0.5 mg NO₃-N/L (Shih et al., 2011). Natarajan (2015) found no evidence of methylmercury generation when water with low nitrate-N concentrations was pumped into four field-scale bioreactors in Minnesota operating at hydraulic retention times ≥ 24 h. Hudson and Cooke (2015) also measured the potential for methyl mercury production but in bioreactor designs where the bioreactor bottom was 30 cm below the tile invert to provide longer residence times. They found negligible methyl mercury values during winter, but high values during summer, particularly during times of low or no flow likely due to highly reducing conditions in the stagnant pool of water below the tile invert. These findings led to new design recommendations in which the bottom of the bioreactor is at the same level as the tile invert. This recommendation was incorporated into the NRCS Conservation Practice Standard which states: *“If reducing conditions may result in the production of methyl mercury, make additional provisions to ensure that stagnant conditions do not develop in the media chamber”* (USDA NRCS, 2015).

In terms of practice limitations, nitrate removal in bioreactors has been reported to be limited by cold climates as noted in Minnesota studies or low DOC during storm flows as noted in the New York State section. Analysis of nitrate removal rates across regional or seasonal temperature gradients would provide added insight into bioreactor functioning and performance. Jang et al. (2018) and Nordstrom and Herbert (2019) described recent advances to bioreactor models to incorporate temperature sensitivity. Heating water to improve nitrate removal has been trialed with limited success (Rendall, 2015; Cameron and Schipper, 2011).

There are also monitoring limitations and challenges for quantifying N removal performance of a variety of bioreactor designs (MD section above). For example, sawdust denitrifying walls may require hydrogeological investigations to determine N loading and treatment effectiveness. Christianson et al. (2017) suggested it may be more practical to create groundwater flow rate estimates for regions within the Chesapeake Bay watershed which could be used for denitrifying wall N loading estimates.

Overall, more long term studies, which face many challenges including funding cycles/limitations, staff turnover, and the need for long term commitments by field site partners (e.g., landowners), are needed to advance bioreactor technologies.

COST-EFFECTIVENESS

Christianson et al. (2012b) reported subsurface drainage bioreactor installation costs generally ranged from \$6,940 to \$11,820 in the relatively early days of field-scale bioreactor research in the Midwest. Scaled to drainage treatment area, these costs were \$190/ha to \$590/ha, and for the one site where the volume was reported along with cost, the resulting cost efficiency was \$77/m³. In this review, bioreactor bed installation costs have ranged from \$7,900 to \$10,400 in South Dakota to more than \$20,000 in the Chesapeake Bay watershed. Work in the Chesapeake Bay illustrates beds are more expensive than denitrifying walls (e.g., \$18,000 and \$27,000 vs. <\$3,000, respectively; Christianson et al., 2017), although costs will be site and design specific. Schmidt and Clark (2012) reported a relatively early denitrifying wall cost of approximately \$20,000 which equated to \$168/m³ (LWD: 55 x 1.7 x 1.8 m). The Australian denitrifying walls described here were more cost efficient at approximately \$50AUD/m³ (\approx \$35USD/m³).

Initial evaluations of bioreactor N-removal cost efficiency ranged from \$2.40/kg N to \$15.20/kg N (Schipper et al., 2010a; assumed 20-y life; 4% annual interest) which have been corroborated with more recent estimations. For example, Sarris and Burbery (2018) assumed a 10-y life in a numerical simulation and determined the average treatment cost would be \$9.70 \pm \$3.10NZD/kg N (mean \pm stdev: \$6.60 \pm \$2.10USD/kg N) for an in-stream bioreactor in New Zealand. Lepine et al. (2018) detailed the full itemization for initial bioreactor installation (\$47,840; \$140/m³) and subsequent woodchip replacement (\$19,470/replacement) assuming a variety of clogging scenarios with aquaculture wastewater. They estimated bioreactors had conservative cost efficiencies of \$5.50/kg N to \$13.40/kg N with improved cost efficiencies occurring at reduced frequency of woodchip replacement (10-y planning horizon; 10.6% discount rate). Easton et al. (2019) reported cost efficiencies of \$0.54 to \$7.60/kg N-y for bioreactors treating legacy N in spring water with an economy of scale noted for treatment of larger springs (> 500 m³/d flows) and greater cost efficiency for springs with higher nitrate-N concentrations (7.3 versus 3.8 mg NO₃-N/L). Bioreactor cost efficiencies by DeBoe et al. (2017) ranged higher at \$15/kg N-y to \$48/kg N-y using a case study installation cost of \$11,060 (\$181/m³). This equated to a total annualized cost of \$994/y assuming a 15-y planning horizon and 4% interest rate. Importantly, they concluded that while bioreactors are a cost-effective practice, adoption may be slow due to the relatively high up-front cost and low incentives. Note, the NRCS Conservation Practice Standard establishes 10 years as the practice life for denitrifying bioreactors treating subsurface drainage (USDA NRCS, 2015).

Christianson et al. (2013) developed cost efficiencies for seven subsurface drainage conservation practices and reported constructed practices of bioreactors and wetlands had competitive cost efficiencies of less than \$3.00/kg N-y. These calculations assumed very small design service costs of only \$40/h for two days of bioreactor design work. Where bioreactors require engineering plans signed and sealed by a professional engineer to receive public funding, design costs may add 50% to 100% of the installation cost (average design cost of \$7,500) for bioreactors designed by private design firms. Additionally, the small project size and distributed nature (both spatially and temporally) of typical subsurface drainage bioreactor projects limits the appeal for many engineering consultants to provide this service. Design services provided by USDA NRCS, while free to the farmer or landowner, still come with a transaction cost in terms of design times that can be lengthier due to agency staffing constraints and competing program priorities.

In summary, reported subsurface drainage bioreactor installation costs have been less than \$5,000 (e.g., if contractor and design fees are waived) to \$27,000, volumetric costs have ranged from \$77/m³ to nearly \$200/m³, and cost efficiencies have been estimated at less than \$2.50/kg N-y ranging generally to about \$20/kg N-y (although can be as high as \$48/kg N-y). There are too few woodchip bioreactor data to estimate sediment or phosphorus removal cost efficiencies.

MONITORING RECOMMENDATIONS

One common subsurface drainage denitrifying bioreactor monitoring configuration uses pressure transducers with associated data loggers in the control structures, weir equations, and water quality sampling to estimate nitrate-N removal. Monitoring methods described below are suggested for the context of conservation practitioners, watershed groups, and others interested in documenting bioreactor annual nitrate-N load removal performance of bed designs. Beyond this guidance, specific methods for research studies should be guided by study objectives. Additionally, while the recommended guidance below is for assessment of N load reductions, water quality sampling alone without flow monitoring can provide a simple practitioner-oriented assessment of if nitrate concentrations are being reduced across the bioreactor (i.e., assessment of if a bioreactor is working).

FLOW MONITORING

Control structures using stop logs that act as weirs to direct and control flow provide convenient locations to monitor bioreactor flow and water chemistry (Figure 3; USDA NRCS, 2012). Water in the control structures cascades over the top stop log, and the water depth is recorded with a pressure transducer placed upstream of the stop logs. The pressure transducer either

has an internal datalogger or is connected to an external datalogger. That recorded water height is then entered into a calibrated weir equation to calculate flow rate. While the estimation of flow through control structures in this way violates many of the conditions for standard weir equations, the use of empirically developed weir equations for control structures has proven effective in practice. In some bioreactor installations, a single combined control structure is used to manage inflow, bypass, and outflow. However, it is difficult to measure multiple flows within the small volume of the control structure, so if monitoring flow is an objective, separate inflow and outflow control structures are strongly considered.

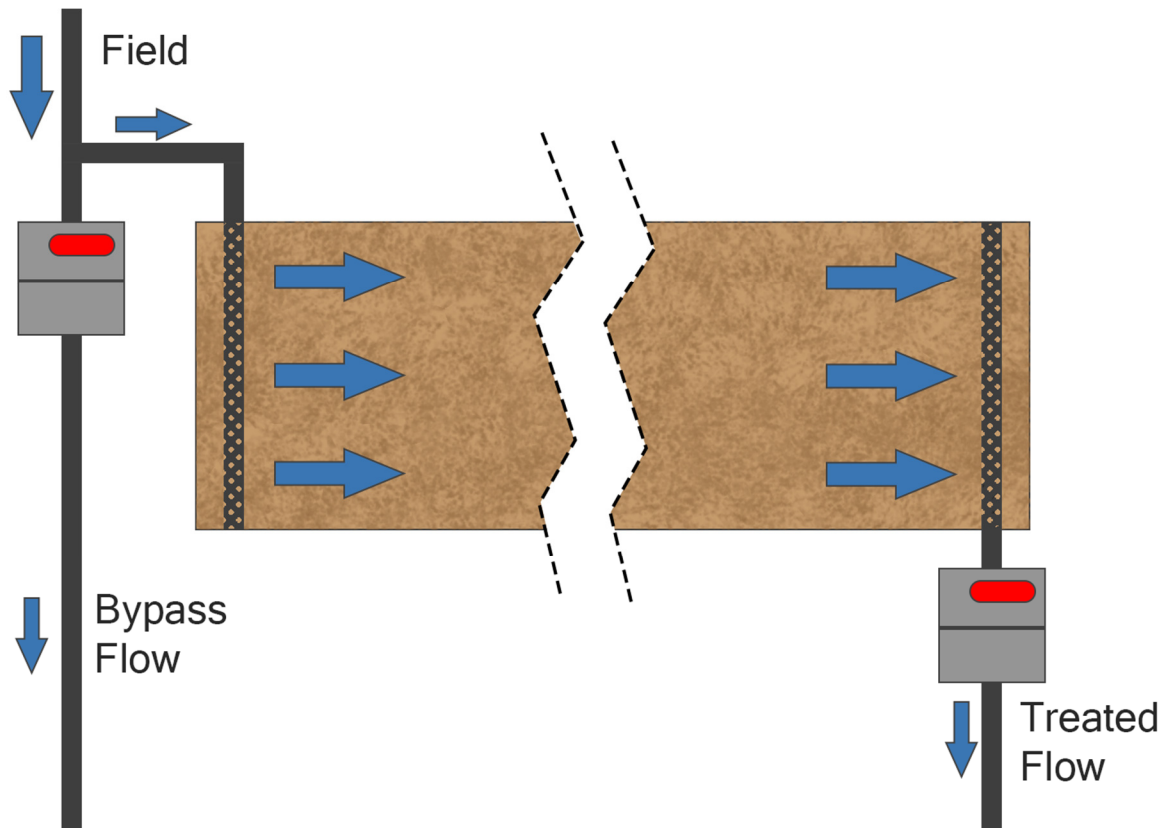


Figure 3. Example pressure transducer (red oval) placement in the inflow and outflow control structures for monitoring bypass flow and bioreactor outflow, respectively. Total flow from the field is often calculated as bypass flow volume + bioreactor outflow volume.

In common subsurface drainage bioreactor monitoring configurations where two-chamber control structures are used at both the inlet and outlet, neither the bioreactor inflow volume nor the total flow volume from the field are measured (e.g., Figure 3). It is a common misconception that the pressure transducer placed in the inflow control structure yields water depths that can be used to calculate the bioreactor inflow. The pressure transducer placed in the inflow structure is used to estimate the bypass flow and the pressure transducer placed in the outflow control structure is used to estimate the outflow. This is because the pressure transducer data must be paired with a weir equation to convert the logged pressures (that is, the water depths) into

flow rates. Total drainage flow from the field is estimated as the bypass flow plus the bioreactor outflow because those two individual flows are generally the only two monitored. In other words, the bypass flow and the bioreactor outflow are the only two streams flowing over a stop log weir in a given control structure. The bioreactor inflow is then usually assumed to be equal to the bioreactor outflow. This assumes there are no gains or losses of water within the bioreactor itself, so for monitoring flow, a lined bioreactor design is preferred to make this assumption more valid. Individually monitoring the total flow from the field and/or the bioreactor inflow would require more expensive instrumentation as well as more elaborate plumbing (e.g., additional control structures). Placing the pressure transducers as shown in Figure 3 is also a convenient way to estimate the saturated volume of the bioreactor.

It is recommended to calibrate and validate the pressure transducers in house (e.g., using a flat bottom bucket) prior to deployment. If the pressure transducers are not vented, a barometric logger needs to be used to correct the logged pressures for changes in barometric pressure; some anecdotal evidence indicates pressure transducers that internally account for barometric pressure may be simpler. Each pressure transducer should be secured firmly within the control structure, so it does not move. The calculation of water depth flowing over the stop log weir is a relative calculation based on the logged depth of water minus the height of the stop logs. Any potential change in elevation of the pressure transducer due to disturbance can result in an inappropriately logged water depth, which would give a false resulting depth of water flowing over the stop log. The pressure transducer can be attached (e.g., zip-tied) to the bottom of a tall “L” made of PVC pipe. The top of the “L” should be screwed or otherwise secured to the inside of the control structure to minimize movement.

It is also recommended to manually measure and record the depth of water on the upstream side of the control structures during each site visit (see example field log sheet in Fig. A1). Use of continuously logging pressure transducers provides relatively high-frequency water depth data for use in weir equations, but this should be verified with manual readings especially if the pressure transducers have moved or if there is accumulated sediment in the structure. Control structure water depths can be manually estimated by applying KolorKut™ paste to a tape measure or measuring rod which is then inserted to the bottom of the control structure. A more expensive but more precise option is to use an electronic tape to measure the depth of the water table to the top edge of the control structure.

It is helpful to record the height of the stop logs in the control structures as often as possible and account for all management activities altering these (Fig. A1) because the calculation of water depth flowing over the stop log weir is a relative calculation involving the total height of the stop logs. A pressure transducer will log the entire depth of water (i.e., pressure) in the control structure, but it is only the depth of water flowing over the weir that is used in the weir equation. Thus, having an exact and

current account of the (1) number of stop logs and (2) the height of each is critical for this weir-based flow estimation method.

Selection of the appropriate weir equation can vary based on the type and size of the control structure, the type of stop logs used, and the amount of flow. Chun and Cooke (2008) developed rectangular weir equations for use with standard rectangular stop logs. Their set of equations includes a low- and a high-flow equation for use with 15.2 cm (6-in) control structures and a low- and a high-flow equation for structures that are 20.3 to 61.0 cm (8- to 24-in). V-notch weir stop logs with a sharp-crested, stainless-steel edge and the associated flow equation have recently been developed (Christianson et al., 2019). A compound weir equation should be used for when the water height exceeds the top of the V-notch weir plate. In-house calibration of weirs is always recommended. It is not unusual for poorly drained areas where bioreactors are implemented to become submerged. In those cases, it can be difficult to accurately estimate flow and an orifice approach is recommended (e.g., Cooke et al., 2004). Given many bioreactors are placed at lower landscape locations prone to flooding, it is suggested researchers describe how flooding events are handled in data processing.

Other flow monitoring methods include use of area velocity meters with a propeller or ultrasonic doppler water velocity measures. The former has moving parts and, in the author's experience, can be subject to erroneous readings if flow is hampered (e.g., sediment interference). Other anecdotal experience has indicated ultrasonic velocity meters may have challenges with subsurface drainage water due to the characteristic low turbidity. In locations where the bioreactor is pumped, using a mechanical flow meter of appropriate size may be the most reliable option. In cases where the bioreactor has a clear and accessible outlet, a calibrated bucket and stopwatch can be used for a low-tech monitoring option. If monitoring funding only allows for grab sample collection, performing replicated bucket/stopwatch flow measurements at the time water samples are collected will give a snapshot flow estimate that, while not a complete picture of flow and load reduction performance, can be useful to better assess bioreactor performance than just water quality sampling alone.

WATER CHEMISTRY SAMPLING

Water chemistry sampling and analysis should be performed on bioreactor inflow and outflow water to determine nutrient concentrations, which combined with flow data, allows calculation of loading reductions. Nutrient concentrations sampled in the inflow structure are generally applied to calculate both inflow loading and bypass loading. In other words, it is assumed the nutrient concentration in the bypass flow water doesn't change between the inflow control structure and the receiving water body. There has been some evidence nitrate concentrations in the control structures can stratify (Rendall, 2015) meaning it is important to collect samples in the same way during each sample event.

When the objective is to monitor annual N loading in subsurface drainage (or in this case, annual N loading reduction due to a bioreactor), weekly samples for nitrate analysis have generally been recommended. Wang et al. (2003) found there was a 92% probability of estimating the annual nitrate-N mass loss within $\pm 15\%$ of the “true” value with weekly sampling versus a 68% probability for monthly sampling. Williams et al. (2015) recently refined this by recommending nitrate-N samples be collected every 2.7-6.0 d for a $\pm 10\%$ desired uncertainty for tile drainage nitrate loading (or, every 1.6-1.8 d for a $\pm 5\%$ desired uncertainty). Sampling to capture phosphorus dynamics requires increased frequency (e.g., every 13-26 h; Williams et al, 2015). Capturing more finite changes in water chemistry and variations in bioreactor performance due to fluctuating environmental parameters would require higher resolution sampling to meet study objectives (even beyond auto-sampler capability). Such enhanced frequency is now possible with more real-time and/or continuous sensors (e.g., Maxwell et al., 2019), although these methods are not yet widely used. An emerging alternative approach is diffuse gradients through thin films (DGT) that are relatively simple devices operating on Fick’s law of diffusion. Corbett et al. (2020) demonstrated that DGT accurately captured average nitrate concentrations over a 24 h period along the length of two bioreactors and allowed calculation of removal rates.

A combined inflow-outflow control structure is sometimes used to minimize the cost of the purchasing two control structures (e.g., David et al., 2016). This configuration may not be ideal for monitoring nutrient removal performance as the single control structure necessarily mixes bypass and bioreactor outflow during higher flow events making it very difficult to sample those flows separately during those times. As mentioned above, separate inflow and outflow control structures are recommended for bioreactors intended to be monitored.

DATA REPORTING

Collection and reporting of a minimum set of consistent criteria at field sites would advance denitrifying bioreactor science and engineering (Table 1). Several years ago, Christianson and Schipper (2016) suggested the research community would benefit from publication of bioreactor performance data in more original forms to allow testing of models and for development of future hypotheses. They suggested, at minimum, inclusion of bioreactor location, installation date, and nitrate concentration, flow, and temperature data. Those suggestions are reiterated here along with additional measures depending on funding and staffing capabilities. Recent bioreactor monitoring guidance from Queensland, Australia recommends “core” and “satellite” bioreactor sites with appropriate levels of monitoring at each (State of Queensland, 2018). Table 1 follows a similar idea with three levels of monitoring, again, depending on funding and capacity. For a practitioner audience, the “minimum suggested to report” is particularly important; it may be better to have as many bioreactors as possible consistently reporting basic

information compared to having fewer sites where data may be more in-depth but more case-study in nature.

Table 1. Three levels of suggested reporting criteria for field-scale bioreactors treating agricultural drainage. This could be easily modified to report data from other types of full-size bioreactors (e.g., aquacultural wastewater) that are influenced similarly by field conditions. HRT is hydraulic retention time; ORP is oxidation-reduction potential.

Reporting type	Minimum suggested to report	Suggested additional reporting	Supplemental suggestions for future modeling or meta-analytical work
In-field characteristics	Known or estimated drainage area Drainage criteria for bioreactor design capacity per the NRCS standard Presence of surface inlets Cropping system/rotation/land use	Additional drainage details: pattern or targeted drainage system; drain spacing and depth; drainage coefficient; drainage management zones Soil types in drainage area	Field management information: nutrient management such as source, timing, rate, and placement; tillage; planting/harvesting dates; crop yield; cover cropping information Additional drainage details: drainage intensity (e.g., Skaggs, 2017)
Bioreactor design	Bioreactor dimensions (LWD, where D is the depth of woodchips) and volume Installation date Design HRT and design flow rate (or associated criteria) Presence/absence of a soil cover	Typical saturation depth <i>In situ</i> HRT Manifold orientation and flow direction (e.g., up flow, down flow, horizontal flow) Stop log management	As-built survey indicating elevations of all bioreactor components Conservative tracer testing metrics like hydraulic efficiency
Bioreactor fill	Size range within which the majority of the media falls Estimated drainable porosity General type (hardwood/softwood)	Particle D ₁₀ and D ₅₀ Particle uniformity coefficient Bulk density Total porosity Wood species General woodchip shape (e.g., irregular, rectangular, flat, squared)	Full particle size distribution Saturated hydraulic conductivity Estimate of carbon source longevity Type of wood-chipping equipment
Water chemistry and quality (generally for the inflow and outflow; additional internal measurements are desirable supplemental information)	Nitrate-N concentrations Water temperature	Dissolved oxygen pH	ORP Total nitrogen Dissolved organic carbon Sulfate Total and/or dissolved phosphorus Methyl mercury Additional pollutants
Flow criteria	Bioreactor flow rates Volume and/or percentage of water treated	Precipitation during monitoring period	Long-term average annual precipitation
Performance criteria	N removal rate N removal efficiency Undesirable effects over time (e.g., early slumping, flow restriction)	Mass load/loss from the field Mass load/loss removed Mass load/loss bypassing	Load reduction of phosphorus or other contaminants

The 2016 recommendations urged tandem reporting of N removal rate (g nitrate-nitrogen removed per m³ of bioreactor per d) and N removal efficiency (% reduction; Christianson and Schipper, 2016), and the latter is now been expanded to include explicit masses of N removed (kg N; kg N/ha) and flow volumes treated (m³; mm equivalent depth). The method of calculating the removal rate and hydraulic retention time should be made explicit, including use of the entire reactor volume or the saturated reactor volume (when the entire bed is not saturated) and use of the total or effective/drainable porosity. Additional monitoring

and reporting guidance is needed for denitrifying walls and in-ditch bioreactors. While the recommendations in Table 1 generally apply, specific nuances such as flow estimates for shallow ground water at denitrifying walls require special attention.

CONCLUSION

Denitrifying bioreactors are an effective and cost-efficient technology for treatment of nitrate in subsurface drainage and other waters. This review documents a variety of successes with and some limitations of full-scale bioreactors for a variety of applications around the world. An evaluation across peer-reviewed literature published over the past three years showed bed-style bioreactors provided $40 \pm 26\%$ N load reduction (mean \pm st dev; median: 46%; $n = 15$; N removal rate mean \pm st dev: 7.2 ± 9.6 g N/m³-d; $n = 27$). Bioreactors designed to any given standard may not achieve this mean value consistently as this analysis included a variety of bioreactors treating a range of water matrices. Subsurface drainage bioreactor installation costs have been less than \$5,000 to \$27,000 with cost efficiencies estimated at less than \$2.50/kg N ranging generally to about \$20/kg N. Bioreactors in high-solids applications (i.e., not subsurface drainage) provide effective solids and sediment removal, but there are little field-scale data on this. Denitrifying bioreactors treating subsurface drainage show a mix of phosphorus leaching and removal although most generally experience phosphorus leaching at bioreactor start-up. The mechanisms and consistency of these phosphorus dynamics are unclear.

Despite N-removal successes, there is much room to advance the science and engineering of bioreactor technology. Key research areas include: improving understanding of removal of additional pollutants, compounds, and biologicals; reducing pollution swapping especially at start-up; advances to simplify and streamline the design process; innovative design modifications; development of management strategies to address observed challenges such as clogging and slumping. Moving this field forward requires more strategic and coordinated monitoring efforts as new bioreactors continue to be designed and constructed. A suggested monitoring set-up is described for assessing annual nitrate-N load removal performance of subsurface drainage denitrifying bioreactors. Recommended minimum reporting measures include: bioreactor dimensions and installation date; fill media size, porosity, and type; nitrate-N concentrations and water temperature; bioreactor flow treatment details; basic in-field cropping and drainage system characteristics; and N removal rate and efficiency.

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APPENDIX A

Table A1. Compilation of field studies reported after the meta-analysis performed by Addy et al. (2016). Abbreviations as follows: V is bioreactor volume; HRT is reported hydraulic retention time; NRE is N removal efficiency (noted as concentration or load); NRR is N removal rate; WC is woodchips; WTR is waste treatment plant residuals; CF is constant pumped flow. Special notes as follows: * = Concentration reduction; † = Load reduction; ‡ = These bioreactors have high dissolved organic carbon (DOC) in the influent likely resulting in higher NRE and NRR; ** = Additional per. comm. with the author.

Source	Location	Type	Installed	Volume	Area treated	HRT	Influent N Concentration	N removal efficiency	N removal rate	Reporting period	Notes
				m ³	ha	h or d	mg N/L	%	g N m ⁻³ d ⁻¹		
Bock et al., 2018	Middlesex Co, VA	bed	Aug. 2014	25.3	6.5	3-20h	3.7	9.5†	0.56	Sept. 2015-Sept. 2016	90% WC10% biochar (v:v)
Christianson et al., 2017	Caroline Co, MD	stream bed	Nov. 2015	248.9	35	N/A	3.98	75*, 25†	0.97	295 d	Treated ditch water from rotating crops
Christianson et al., 2017	Somerset Co, MD	stream bed	Dec. 2015	20.8-29.3	6.6	~24h	2.53	65*	N/A	4 months of 1st year	In-ditch bioreactor – built in 3 segments
Christianson et al., 2017	Somerset Co, MD	stream bed	Feb. 2015	19.8	0.08	N/A	5.5 & 4.9	>90*	1.9 & 2.9	~1 year	Sawdust wall
Goeller et al., 2019	Canterbury Plains, NZ	bed	Oct. 2015	25.0	160	0.9h	N/A	10*	50.9	Dec. 2015-Oct. 2017	Closed pit – tile drainage from dairy pasture‡
Goeller et al., 2019	Canterbury Plains, NZ	walls	Oct. 2015	20.0, 24.0	160	2.7h, 5.5h	N/A	57*, 99*	N/A	Dec. 2015-Oct. 2017	2 sawdust walls, riparian seep from dairy pasture

Gottschall et al., 2016	Eastern Ontario, Canada	bed	Oct. 2011	3.3, 3.3, 3.3	0.14, 0.14, 0.14	N/A	N/A	39*, 62*, 63*	6.58, 7.80, 6.36	Sept. 2013-May 2014	6 bioreactors - 2 replicates WC, WC and 10% WTR, WC and 20% WTR
Hassanpour et al., 2017	Tompkins Co, NY	beds	Oct. 2012	9.5	4	2.2d, 2.1d	9.3	42*, 55*	3.8, 4.7	2013-2015	1 WC and 1 WC+10% Biochar - closed pit
Hassanpour et al., 2017	Chemung Co, NY	beds	Jun. 2013	9.5, 17.1	5	0.5d, 0.3d	6.2	68*, 66*	13.5, 15.1	2013-2015	1 WC and 1 WC+10% biochar – closed pit
Hassanpour et al., 2017	Steuben Co, NY	beds	Jul. 2013	12.1, 17.1	6, 9	2.8d, 2.3d	18.4, 16.6	58*, 62*	4.7, 6.7	2013-2015	1 WC and 1 WC+2% biochar – closed pit
Husk et al., 2017	South Central Quebec, Canada	beds	Jul. 2012	33.0, 10.1, 22.7, 33.0	0.61, 0.69, 0.93, 1.28	14.1h, 5.3h, 27h, 14.h	5.3	99†	6.84 (median)	2012-2015	4 bioreactors monitored 3 years for 12 site years of data cumulative. All closed pit
Hoffmann et al., 2019	Jutland, Denmark	beds	Fall 2012	100	78	N/A	14-May	55†, 56†, 43†, 46†, 50†, 55†	1.67-2.22	2013-2014	6 bioreactors with mixed media (seashells/willow WC) at 50:50 and 25:75 ratios. Closed pit with vegetation
Pfannerstill et al., 2016	North German lowlands	stream bed	2010	N/A	N/A	N/A	8.9	28*, 15†	N/A	Aug. 2010-April 2012	drainage reactive ditch with tile from a farm - 12 m3 WC
Pluer et al., 2018	Sarasota, FL		Nov. 2013	4.32	16.4	CF	N/A	N/A	7.9	Nov. 2013-Nov. 2014	open pit, Submerged bios in retention ponds‡
Pluer et al., 2018	Sarasota, FL		Oct. 2013	4.32	22.9	CF	N/A	N/A	3.4	Nov. 2013-Nov. 2014	open pit, Submerged bios in retention ponds‡
Rambags et al., 2016	Newstead, NZ	bed	May-13	114	CF	7-10d	avg 31.2	>99*	~14	Aug. 2013-June 2015	closed pit trapezoidal, treats human waste‡
Rosen and Christianson, 2017	Caroline, Co, MD	bed	Nov. 2013	169.3	34.7	>10d	4.7, 7.7	9†, 16†	0.4, 0.21	Aug. 2014-Aug. 2015, Aug. 2015-May 2016	closed pit treats dairy farm
Rosen and Christianson, 2017	Queen Anne's Co, MD	bed	Dec. 2013	90.9	25.2	> 1 week	9.2, 8.6	62†, 47†	5.36, 5.12	Aug. 2014-Aug. 2015, Aug. 2015-April 2016	closed pit treats organic grain farm
Rosen and Christianson, 2017	Caroline, Co, MD	bed	Nov. 2014	252.6	40.1	42 ± 56 h	13.5	10†	1.53	Dec. 2014 – July 2015	closed pit treats conventional row crops
von Ahnen et al., 2016	North Jutland Region, Denmark	bed	Oct. 2015	12.5	CF	3.3-6.5 h**	5.6	N/A	7.06	first 147 d	treats aquaculture - start-up - rainbow trout farm‡
von Ahnen et al., 2018	Denmark	bed	Mar-17	300	CF	17.7 h**	5.3	80*	4.8	52 wk from install	Horizontal flow – rainbow trout aquaculture‡
von Ahnen et al., 2018	Denmark	bed	Jul-17	660	CF	11.0 h**	10.5	29*	4.5	28 wk starting 6 wk after install	vertical/down flow - rainbow trout aquaculture‡
von Ahnen et al., 2018	Denmark	bed	Jan. 2017	1440	CF	10.4 h**	9.5	48*	7.8	52 wk starting 7 wk after install	vertical/down flow - rainbow trout aquaculture‡

Field site: Christian County Bioreactor **Date/Time:** 16 June 2019 14:30
Personnel: Laura Christianson with help from Ronnie Chacon

Weather and site conditions: Party cloudy, hot (high of 85 F), standing water in some access road ruts
(state if anything is surprising or unusual)

Recent relevant weather: Rained yesterday (state the amount if known)

Bioreactor observations: Bioreactor is getting a little weedy; no sulfide smell present near either structure;

Stop log measurement and management

	Inflow Structure	Outflow Structure
Number of plates when arriving on-site	4	3
Plate heights	7"+7"+7"+V plate	5"+5"+V plate
Were the plates changed today?	No	No
New number of plates	none	none
New plate height	none	none
Was flow observed over the stop logs?	No	Yes
Manual water depth recorded	Kolor Kut value of 20 7/8ths inches	Kolor Kut value of 16 1/4ths inches
Additional notes	V-notch plate present	Slime observed; V-notch plate present

Sample collection:	Inflow Structure	Outflow Structure
Manual grab sample	(number of samples; notes)	(number of samples; notes)
Auto-sampled	(number of samples; notes)	(number of samples; notes)
Samples from on-site personnel	(number of samples; notes)	(number of samples; notes)

Sampling notes: The outflow autosampler malfunctioned for the outflow samples from day 09-14 June;
only partial samples were collected; The inflow sampler was flooded, all samples lost.

Flow data:	Inflow Structure	Outflow Structure
Pressure transducer downloaded	(file name or notes)	(file name or notes)

Barometric pressure transducer: (file name or notes)

Flow notes: Launched a new pressure transducer for the inflow structure (which flooded);
bringing the old one back to the lab for testing

Additional testing notes: We can likely start a tracer test here soon (bypass flow has ceased);
Did not collect well samples this time; need to do this next week

Future action items: Replaced inflow pressure transducer; need to replace one of the spare ones in the work truck.
Saw Farmer John on-site, he reminded me about the upcoming field day on June 28
Bring weedeater next time

Figure A1. Example field log sheet for personnel collecting data at denitrifying bioreactors treating subsurface drainage with example entries in blue italics.

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