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Continuous Cultivation of Photosynthetic Bacteria for Fatty

Acids Production

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Abstract

In the present work, we introduced a novel approach for microbial fatty acids (FA) production. Photosynthetic bacteria, *Rhodobacter sphaeroides* KD131, were cultivated in a

1 continuous-flow, stirred-tank reactor (CFSTR) at various substrate (lactate) concentrations.
2
3 At hydraulic retention time (HRT) 4 d, cell concentration continuously increased from 0.97 g
4
5 dcw/L to 2.05 g dcw/L as lactate concentration increased from 30 mM to 60 mM. At 70 mM,
6
7 however, cell concentration fluctuated with incomplete substrate degradation. By installing a
8
9 membrane unit to CFSTR, a stable performance was observed under much higher substrate
10
11 loading (lactate 100 mM and HRT 1.5 d). A maximum cell concentration of 16.2 g dcw/L,
12
13 cell productivity of 1.9 g dcw/L/d, and FA productivity of 665 mg FA/L/d were attained, and
14
15 these values were comparable with those achieved using microalgae. The FA content of *R.*
16
17 *sphaeroides* was around 35% of dry cell weight, mainly composed of vaccenic acid (C18:1,
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19 omega-7).
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28 **Keywords:** photosynthetic bacteria; fatty acids; membrane-coupled bioreactor; lactate; cell
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30 productivity
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35 1. Introduction

36
37 Environmental concerns, energy shortage, and consequent increasing energy costs have
38
39 emphasized the need to produce sustainable and renewable fuels (Steen et al., 2010). To this
40
41 end, huge effort is now being focused on the production of lipids using microalgae (Chen et
42
43 al., 2011). Under unfavourable culture conditions, they are able to store neutral lipids, 20-
44
45 70% of their body in heterotrophic or autotrophic ways, mainly in the form of triacylglycerol
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47 (TAG). TAG is convenient storage compound for carbon and energy, possessing high
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49 calorific value, and can be used as industrial chemical and bioenergy feedstock (Alvarez et al.,
50
51 2002).
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1 On the other hand, although photosynthetic bacteria do not accumulate neutral lipids, they are
2
3 able to synthesize fatty acids, principally for glycerol-based membrane lipids (Carlozzi et al.,
4
5 2010). The oil content (20-40% of dry biomass weight) of photosynthetic bacteria is
6
7 generally lower than that of microalgae, and therefore, the research on this subject has been
8
9 scarce. However, they are simpler to cultivate than microalgae that they do not require
10
11 stressful environments for lipids production. In addition, bacteria are much more genetically
12
13 manipulatable than algae. The expression of specific genes could result in the overproduction
14
15 of fatty acids and their secretion to the broth (Steen et al., 2010).
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20 Until now, the use of purple non-sulfur phototrophic bacteria has been proposed mainly for
21
22 the production of hydrogen and polyhydroxybutyrate (PHB), as well as for wastewater
23
24 treatment (Khatipov et al., 1998; Kim et al., 2006; Wu et al., 2012). Electrons contained in
25
26 organic materials can be released as hydrogen by nitrogenase with the help of light energy,
27
28 and PHB is formed when PNS bacteria are faced with a suboptimal environment. Recently,
29
30 the co-production of hydrogen and lipids under anaerobic conditions was suggested (Carlozzi
31
32 et al., 2010). During the cultivation of *Rhodospseudomonas palustris* under anaerobic light
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34 conditions, lipid content of 22-39% of dry biomass weight was observed. However, this study
35
36 was limited to batch operation: it is important to maximize the lipid productivity by
37
38 continuous operation. In addition, the effective use and design of a continuous culture are
39
40 known to lower the production cost (Chen et al., 2011).
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45 During the production of photosynthetic bacteria in a continuous culture system, the ability to
46
47 maintain a balanced concentration of chemical substances and cells is critical to obtain higher
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49 productivity than in a batch culture system. However, there are many difficulties in
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51 maintaining a continuous culture for long periods without mutation and microbial
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1 contamination. Population changes due to mutation or microbial contamination in the pure
2 culture of a microbial strain have frequently been reported in continuous culture systems
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4 (Toda, 2003).
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7
8 The use of a membrane-coupled bioreactor has recently gained recognition as a solution to
9 the above problems by decreasing contamination risks and cleaning frequencies, and by
10 higher cell concentration compared to a standard stirred tank bioreactor (Glazyrina et al.,
11 2010). In addition, it is possible to reuse the water and nutrients in the permeate from the
12 membrane filter by retaining cells that are larger than the diameter of membrane pore. Recent
13 works employing various microorganisms immobilized in membrane-based culture systems
14 have shown that extremely high, sludge-like densities of ca. 10^{12} cells per ml were possible in
15 such systems and that the productivity of membrane reactors continues at high levels for
16 more than two weeks (Lee et al., 2008; Ríos et al., 2012).
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19
20 In this study, we suggested a novel approach for microbial fatty acids production, which is
21 competitive with a traditional way of using microalgae. Photosynthetic bacteria, *Rhodobacter*
22 *sphaeroides* KD131, were cultivated for fatty acids production using lactate as a substrate.
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24

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26 Organic acids that mostly consisted of lactate can be easily obtained from fermentation
27 products in agricultural and food waste (Kim et al., 2009). To maximize the cell productivity,
28 a continuous-flow, stirred-tank reactor (CFSTR) was operated at various substrate
29 concentrations and hydraulic retention times (HRT). A membrane unit was installed in the
30 CFSTR for further increase of cell concentration and productivity. To our knowledge, this is
31 the first attempt to apply the concept of a membrane-coupled bioreactor aiming at continuous
32 cell harvesting. In addition, the fatty acids profile of the bacteria was assayed and the
33 performance obtained in this study was compared with that using microalgae.
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2. Methods

2.1 Inoculum preparation

The phototropic bacterium *R. sphaeroides* KD131 isolated from mudflats along the coast of Daebu Island in the West Sea of South Korea was cultivated for fatty acids production (Kim and Kim, 2012). The KD131 strain was pre-cultured in a modified Siström's broth (Kim et al., 2012) containing 4 mM $(\text{NH}_4)_2\text{SO}_4$, 0.3 mM L-aspartic acid, and 20 mM lactate at 30°C for 24 hr under 54 W/m² irradiance using a halogen lamp (12V, 50W).

2.2 Reactor operation

A 1.2 L glass reactor (effective volume of 1.0 L, 200 mm high by 80 mm diameter) installed with a membrane unit was designed for the continuous cultivation of *R. sphaeroides* (Fig. 1). Three halogen lamps were properly located to adjust the light intensity at 54 W/m² on the reactor wall. The membranes are made of high density polyethylene (HDPE) with a normal pore size of 0.4 μm and an effective filtration area of 0.006 m². A certain amount of centrifuged microorganism was inoculated to reach an initial cell concentration of 0.5 g dcw/L. After purging with Ar gas for 1 hr, the reactor was operated for 48 hr by batch mode, and then switched to a continuous mode. It was agitated using a magnetic stirrer at 150 rpm. Feedstock contained 30 mM of lactate with nutrient medium as mentioned above. All experiments were conducted in a constant temperature room at 30±1°C.

First, the reactor was operated by a CFSTR mode (w/o membrane unit operation). Lactate concentration was gradually increased from 30 mM to 70 mM at a fixed HRT 4 d. As the performance failure was observed at 70 mM, lactate concentration was decreased to 40 mM, and the reactor was operated as membrane-coupled bioreactor mode. Once a day, 50 mL of

1 cell broth, which was harvested for fatty acids production, was pulled out corresponding to a
2
3 cell retention time (CRT) of 20 d, while HRT was kept at 4 d. In order to increase substrate
4
5 loading, first, substrate concentration was gradually increased up to 120 mM. Afterwards,
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7 HRT was gradually shortened to 1 d at a fixed lactate concentration of 100 mM. The
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9 CRT/HRT ratio was kept at 5. Permeation of membranes was continuously maintained by
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11 using a peristaltic pump according to the HRT. One cycle of membrane filtration consisted of
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13 45 min of filtration and 15 min of releasing. In order to reach the steady-state and to obtain
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15 average performance values, the reactor was operated for more than five times of HRT at
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17 each operating condition.
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25 **2.3 Analysis**

26 Residual lactate was analyzed by a high performance liquid chromatograph (HPLC)
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28 (Finnigan Spectra SYSTEM LC, Thermo Electron Co.) with an ultraviolet (210 nm) detector
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30 (UV1000, Thermo Electron) and an 100 mm × 7.8 mm Fast Acid Analysis column (Bio-Rad
31
32 Lab.) using 0.005 M H₂SO₄ as mobile phase. The liquid samples were pretreated with a 0.45
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34 μm membrane filter before injection to both HPLCs. Cell concentration and fatty acids were
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36 measured according to the methods described in Kim et al. (2012) and Carlozzi et al. (2010),
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38 respectively.
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48 **3. Results and Discussion**

49 **3.1 CFSTR performance**

50 High substrate concentration allows energy-efficient operation and results in concentrated
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52 biomass unless substrate inhibition occurs. In microbial oil production, in particular, a high
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1 level of biomass concentration is essential for the ease of downstream processing such as
2
3 harvesting, dewatering, and extracting (Chen et al., 2010).

4
5 Fig. 2 shows the daily performance of CFSTR in terms of cell concentration and lactate
6 degradation at HRT 4 d. As lactate concentration increased from 30 mM to 60 mM, cell
7
8 concentration continuously increased from 0.97 g dcw/L to 2.05 g dcw/L. The experimental
9
10 period used to obtain average performance values are shown in Table 1. At 70 mM, however,
11
12 cell concentration started to fluctuate with incomplete substrate degradation, suggesting
13
14 substrate inhibition. At similar substrate strength, the inhibition has been observed in the
15
16 continuous culture of heterotrophic microalgae and photosynthetic bacteria (Chen and Johns
17
18 1996; Xie et al., 2012). In the CFSTR operation of *R. sphaeroides* fed with lactate at HRT 4 d,
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20 the highest cell concentration of 2.05 g dcw/L and cell productivity of 0.54 g dcw/L/d were
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22 obtained at 60 mM. Cell productivity was calculated by dividing cell concentration to the
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24 corresponding CRT.
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33 It appears that cell yield, the conversion ratio of input substrate to cell growth, was constant
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35 at around 50%, although the substrate concentration changed. Cell yield was calculated by
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37 assuming the composition of $C_5H_7O_2N$, resulting in a chemical oxygen demand (COD) value
38
39 of 1.42 g COD/g dcw (Kim et al., 2012). The reported cell yield of photosynthetic bacteria
40
41 varied as 20-70% depending on the carbon sources, nutrient composition, and operational
42
43 parameters such as pH and HRT (Kim et al., 2012; Yilmaz et al., 2010). 50% cell yield meant
44
45 that half of the electrons contained in the substrate were used for cell growth while remaining
46
47 electrons were converted to other metabolites such as soluble microbial products (SMPs).
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52 SMPs are defined as soluble organic compounds produced during substrate metabolism and
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54 biomass decay and they have been important research topics in biological wastewater
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1 treatment for more than two decades (Laspidou and Rittmann, 2002). However, it was only
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3 recently recognized in the field of using photosynthetic bacteria that SMPs have a sizable
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5 impact on the electron distribution (Kim et al., 2012; Yilmaz et al., 2010). Further researches
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7 on SMP issue such as formation mechanisms, characteristics, and metabolic and genetic
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9 control strategies are needed to increase the cell yield.
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16 **3.2 Membrane-coupled bioreactor performance - Effect of substrate concentration**

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18 In the concept of a membrane-coupled bioreactor, a complete retention of biomass by
19
20 membrane process makes it possible to maintain a high biomass concentration, and thus, a
21
22 stable performance under high substrate loading can be expected (Judd, 2008).
23
24

25 In this study, first, lactate concentration gradually increased from 40 to 100 mM while the
26
27 HRT and CRT were fixed at 4 d and 20 d, respectively. The cell concentration gradually
28
29 increased and reached 14.38 g dcw/L at 100 mM (Fig. 3). Compared to the cell concentration
30
31 in the CFSTR, this value was seven times higher, which could significantly reduce the cost in
32
33 the downstreaming process (Table 2). To our knowledge, this is the highest cell concentration
34
35 achieved to date in various application fields of photosynthetic bacteria. In photo-
36
37 fermentative H₂ production, cell concentration was maintained at a low level (generally <3 g
38
39 dcw/L) since light penetration is essential for the activation of nitrogenase, the central
40
41 enzyme producing hydrogen (Pattanamanee et al., 2012). In wastewater treatment using
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43 photosynthetic bacteria, several attempts have been made to increase cell concentration, but
44
45 the result was less than 5 g dcw/L (Kaewsuk et al., 2010).
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51
52 The reactor performance significantly dropped immediately after increasing lactate
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54 concentration to 120 mM with incomplete lactate degradation. It seemed that *R. sphaeroides*
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1 could not be adapted at this high substrate concentration and the cell concentration decreased
2
3 by the pullout of biomass for CRT control. To recover the performance, lactate concentration
4
5 decreased to 100 mM at further operation.
6
7

10 **3.3 Membrane-coupled bioreactor performance – HRT and CRT control**

11 Although high cell concentration was achieved by installing a membrane unit, cell
12
13 productivity was only increased by 30% compared to the reactor performance without
14
15 membrane, due to long CRT of 20 d (4 d for reactor without membrane). The way to increase
16
17 cell productivity is to reduce CRT while increasing the substrate loading. As larger amount of
18
19 biomass is removed from the reactor at short CRT, more substrate should be supplied to
20
21 maintain the biomass concentration in the reactor. From the 193th day, HRT and CRT were
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23 gradually shortened to 1 d and 5 d, respectively, while keeping the CRT/HRT ratio at 5,
24
25 meaning that the same amount of substrate was supplied to keep the biomass concentration
26
27 unless the cell yield changed.
28
29

30 As HRT and CRT gradually shortened to 2 d and 10 d, respectively, both cell concentration
31
32 and cell productivity increased up to 16.20 g dcw/L and 1.62 g dcw/L/d, respectively (Fig. 2,
33
34 Table 2). As the CRT/HRT ratio was kept at the same substrate concentration, the increased
35
36 cell concentration indicates the increased cell yield. In the application of membrane-coupled
37
38 bioreactor, high cell yield has generally been observed under short CRT (Duan et al., 2009;
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40 Huang et al., 2001). At HRT 1.5 d and CRT 7.5 d, the cell concentration decreased to 14.29 g
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42 dcw/L with a small amount of lactate remaining. However, the overall mass of cultivated
43
44 biomass was enhanced to 1.90 g dcw/L/d, which was 3.5 times higher than that in the CFSTR
45
46 operation. Further decrease of HRT and CRT to 1 d and 5 d, respectively, caused the failure
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1 of reactor operation. Moreover, the trans membrane pressure (TMP), which was maintained
2 less than 20 kPa during the previous operation, increased up to 30 kPa. Because of membrane
3 fouling, it was impossible to continue the reactor operation. The flux could be calculated
4 based on the effective surface area of membrane module, 0.006 m². The calculated flux at
5 HRT 4, 2, 1.5, and 1 d are 1.74, 3.47, 4.65, and 6.94 L/m²/h, respectively. The significant
6 increase of TMP was not observed during the operation periods under the HRT 4 to 1.5 days,
7 but the significant membrane fouling was occurred at right after reactor operation under the
8 HRT 1 d.
9

10 Using the stepwise increase of flux, the critical flux could be obtained. Critical flux is a
11 criterion for the transition between concentration polarization and fouling. The critical flux is
12 reached when irreversible fouling occurs locally on the membrane (Bacchin, 2004). The
13 reactor was operated under an intermittent filtration regime (45 min filtration/15 min pause),
14 and no irreversible fouling was observed under the flux of 1.74 - 4.65 L/m²/h. On the other
15 hand, a significant increase of TMP was observed under the flux of 6.94 L/m²/h. Therefore,
16 the critical flux for this membrane module and reactor system can be determined as 4.65
17 L/m²/h.
18

19 3.4 Fatty acids profile and the performance comparison

20 The fatty acids profile of *R. sphaeroides* is shown in Fig. 4. During the membrane-coupled
21 bioreactor operation, the microbial samples for fatty acid analysis were taken five times after
22 reaching a steady-state at each substrate concentration, and the average values were obtained.
23 The fatty acids content of *R. sphaeroides* investigated here was around 35% of the dry cell
24 weight, and did not vary according to the substrate concentration. Although some microalgae
25

1 species can accumulate fatty acids over 50% of dry cell weight, it is obvious that *R.*
2
3 *sphaeroides* are also suitable biomass resources for fatty acids generation. Ratledge and
4
5 Wynn (2002) defined the microorganisms that can accumulate fatty acids at more than 20%
6
7 of their body as oleaginous species. In addition, unlike phototrophic microalgae, dense cell
8
9 cultivation was possible in the case of *R. sphaeroides* which was attributed their ability to
10
11 capture every possible photon with high efficiency under light low intensity (Adams and
12
13 Hunter, 2012). It is known that intracytoplasmic membranes (ICMs) containing structural
14
15 components necessary for photosynthetic growth are developed when the light resource is
16
17 limited.
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22 The main fatty acid compositions of *R. sphaeroides* were C16-C18, which are appropriate for
23
24 biodiesel production. In addition, the fatty acids from *R. sphaeroides* have special features in
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26 that they are mainly composed of unsaturated fatty acids whereas saturated forms are
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28 dominant in other microorganisms including microalgae, cyanobacteria, and *Escherichia-coli*
29
30 (Chen et al., 2010; Lu, 2010; Steen et al., 2010). Saturated fatty acids have good oxidative
31
32 stability, but poor fuel properties at low temperatures, which is a disadvantage in winter
33
34 operation (Park et al., 2008). Also, unsaturated fatty acids are the raw materials for dietary
35
36 supplements such as omega-3, omega-6, and omega-7 fatty acids. Vaccenic acid, comprising
37
38 60% of fatty acids of *R. sphaeroides*, is a main source of omega-7 fatty acids, which has a
39
40 crucial role in maintaining the health of skin and mucous membranes.
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47 From the continuous cultivation of *R. sphaeroides* using the membrane-coupled bioreactor,
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49 maximum cell concentration of 16.2 g dcw/L, cell productivity of 1.9 g dcw/L/d, and fatty
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51 acid productivity of 665 mg FA/L/d were attained, and these values were comparable with
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53 those achieved using microalgae (Table 3). Fatty acids productivity, representing the
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1 combined effects of fatty acid content and cell productivity, is a suitable index to indicate the
2 effectiveness of a process (Chen et al., 2011). In the case of fatty acids production by
3 photoautotrophic microalgae, continuous and pilot-scale operation has frequently been
4 reported. The fatty acids productivity was in a range of 30-180 mg FA/L/d, which was
5 comparably lower than that achieved in this study. On the other hand, higher cell productivity
6 and fatty acids productivity have been reported in the heterotrophic microalgae cultivation.
7 However, most of them were obtained from the batch operation.

8 In microalgae cultivation, the membrane has only been applied for the harvesting of cultured
9 microalgae as an extracting process even though it has potential to integrate the cultivation
10 and harvesting process together. In this study, the membrane-coupled bioreactor for
11 cultivation and harvesting of photosynthetic bacteria was successfully operated continuously.
12 The productivity of cells and fatty acids was higher than photoautotrophic microalgae and it
13 was comparable with the microalgae production under heterotrophic conditions. It is
14 suggested that the novel method developed here that is cultivating photosynthetic bacteria
15 using membrane-coupled bioreactor could be an alternative way of microbial fatty acids
16 production.

42 **4. Conclusions**

43 Photosynthetic bacteria, *R. sphaeroides*, were continuously cultivated for fatty acids
44 production. By adopting a membrane-coupled bioreactor operation mode, a maximum cell
45 concentration of 16.2 g dcw/L, cell productivity of 1.9 g dcw/L/d, and fatty acids productivity
46 of 665 mg FA/L/d were attained, which were comparable with those achieved using
47 microalgae. The fatty acids content of *R. sphaeroides* was around 35% of the dry cell weight

1 with mainly composed of unsaturated fatty acids, in particular, vaccenic acid (C18:1, omega-
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3
4 7). This work has high importance in the perspective of introducing a novel approach for
5
6 microbial fatty acids production.
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13
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15
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Table 1 Average cell production performances at various substrate concentrations in the CFSTR

Substrate concentration (mM)	Data acquired (th day)	Cell concentration (g dcw/L)	Cell yield (%)	Cell productivity (g dcw/L/d)	Lactate degradation (%)
30	6-27	0.97±0.06	47.6±1.9	0.24±0.02	99.7
40	33-54	1.39±0.04	51.5±1.4	0.35±0.01	99.9
50	58-79	1.70±0.05	50.4±1.5	0.43±0.01	99.7
60	85-106	2.05±0.10	50.5±2.5	0.51±0.01	99.9

Table 2 Average cell production performances at various operation conditions in the membrane-coupled bioreactor

Substrate concentration (mM)	HRT, SRT (day)	Data acquired (th day)	Cell concentration (g dcw/L)	Cell productivity (g dcw/L/d)	Lactate degradation (%)
40	4, 20	19-41	5.77±0.17	0.29±0.01	99.9
60	4, 20	49-81	8.74±0.36	0.44±0.22	99.9
80	4, 20	91-122	11.27±0.37	0.57±0.01	99.9
	4, 20	134-162	14.38±0.33	0.72±0.02	99.9
100	3, 15	210-232	15.14±0.26	1.01±0.02	99.9
	2, 10	248-272	16.20±0.12	1.62±0.01	99.9
	1.5, 7.5	284-309	14.29±0.29	1.90±0.04	97.2

Table 3 Performance comparison of fatty acids productivity obtained in this study with microalgae

Cultivation conditions	Microbial species	Cell productivity (g dcw/L/d)	Fatty acids content (% of dcw)	Fatty acid productivity (mg FA/L/d)	Reactor and operation	Reference
Photo-heterotroph	<i>Rhodobacter sphaeroides</i>	1.9	35	665	Continuous (1 L) (HRT 1.5 d, CRT 7.5 d)	This study
	<i>Chlorella</i> sp.	0.53	34	178.8	Fed-batch (0.8 L)	Chiu et al., 2008
	<i>Chlorella</i> sp.	3.8	N/A ^a	N/A ^a	Fed-batch (400 L) Flat plate type	Doucha et al., 2005
Photo-autotroph	<i>Chlorella vulgaris</i> , <i>Scenedesmus</i> sp, etc.	0.17 - 0.28	16.1 - 21.1	30.4 - 53.9	Batch (0.25 L)	Rodolfi et al., 2009
	<i>Nannochloropsis</i> sp.	0.36	32.3	117	Continuous (110 L) (HRT 2.5 d)	Rodolfi et al., 2009
	<i>Chlorella minutissima</i>	0.137	36 - 44	60.2	Continuous (3 L) (HRT 3 d)	Tang et al., 2012
	<i>Synechocystis aquatilis</i>	0.932	18.58	177.1	Continuous (8 L) (HRT 8.5 d)	Kaiwan-arporn et al., 2012
	<i>Chlorella protothecoides</i>	1.7 - 7.4	43.0 - 57.8	732.7 - 3701.1	Batch (5 L)	Xiong et al., 2008
Heterotroph	<i>Chlorella vulgaris</i>	0.08 - 0.15	23.0 - 36.0	27.0 - 35.0	Batch (1 L)	Liang et al., 2009
	<i>Galdieria sulphuraria</i>	14.6 - 50	N/A ^a	N/A ^a	Continuous (3 L, HRT 2 d)	Graverholt and Eriksen, 2007

^aN/A = Not attained

Fig. 1 Schematic of membrane-coupled bioreactor system for mass cultivation of photosynthetic bacteria

Fig. 2 Daily variation of cell concentration and lactate degradation in the CFSTR at various substrate concentrations

Fig. 3 Daily variation of cell concentration and lactate degradation in the membrane-coupled bioreactor at various operating conditions

Fig. 4 Fatty acids profile of *Rhodobacter sphaeroides*

Fig. 1 Schematic of membrane-coupled bioreactor system for mass cultivation of photosynthetic bacteria

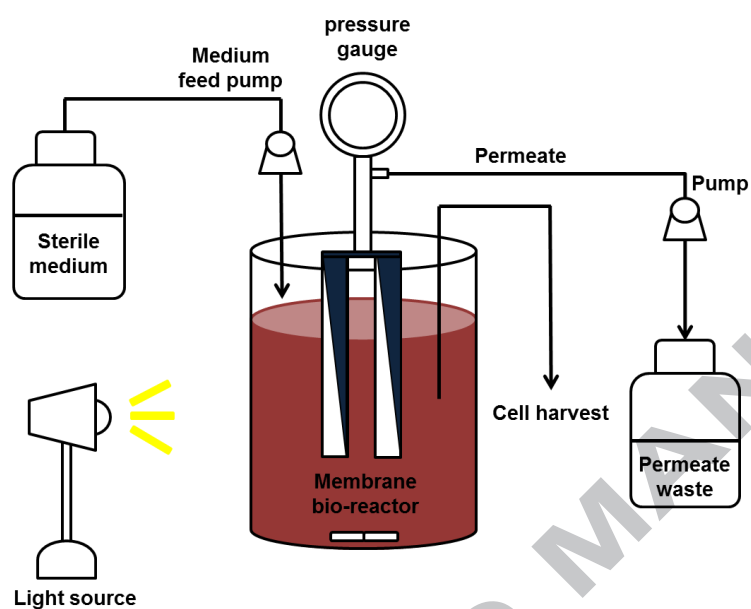


Fig. 2 Daily variation of cell concentration and lactate degradation in the CFSTR at various substrate concentrations

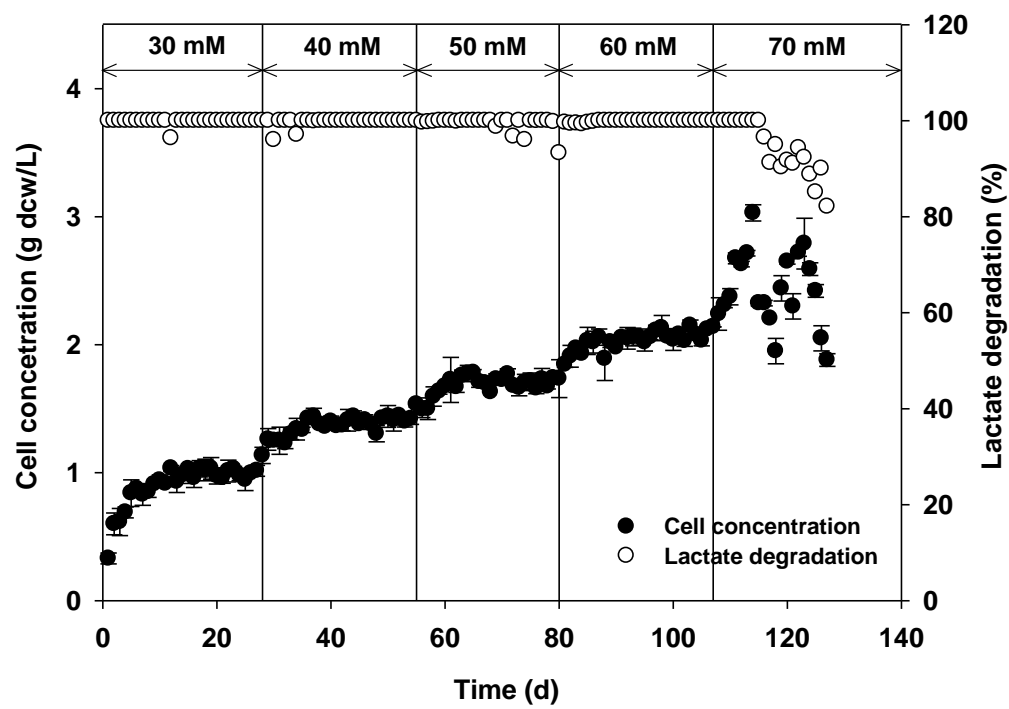


Fig. 3 Daily variation of cell concentration and lactate degradation in the membrane-coupled bioreactor at various operating conditions

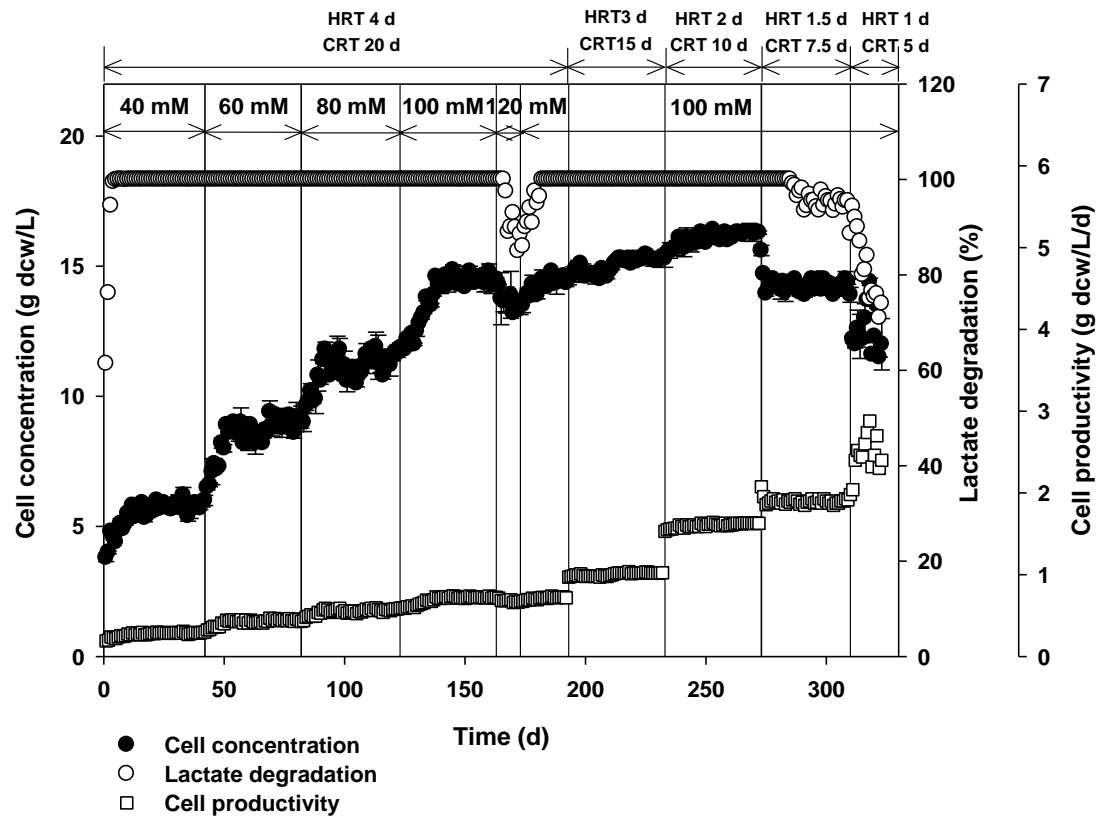
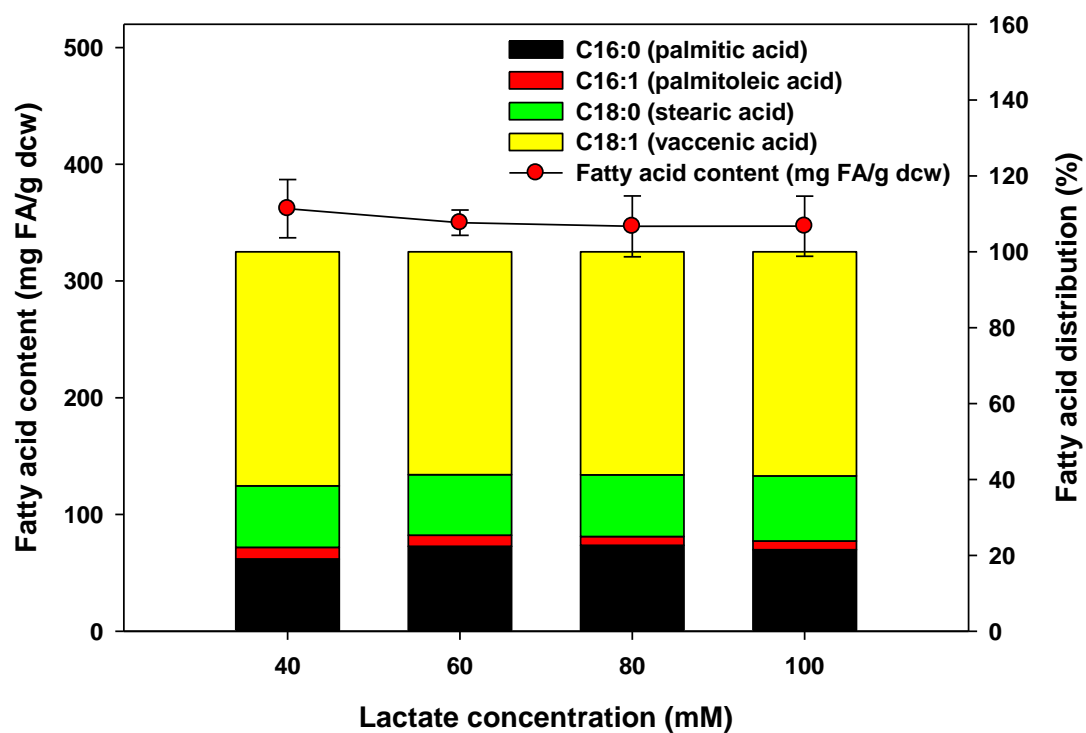
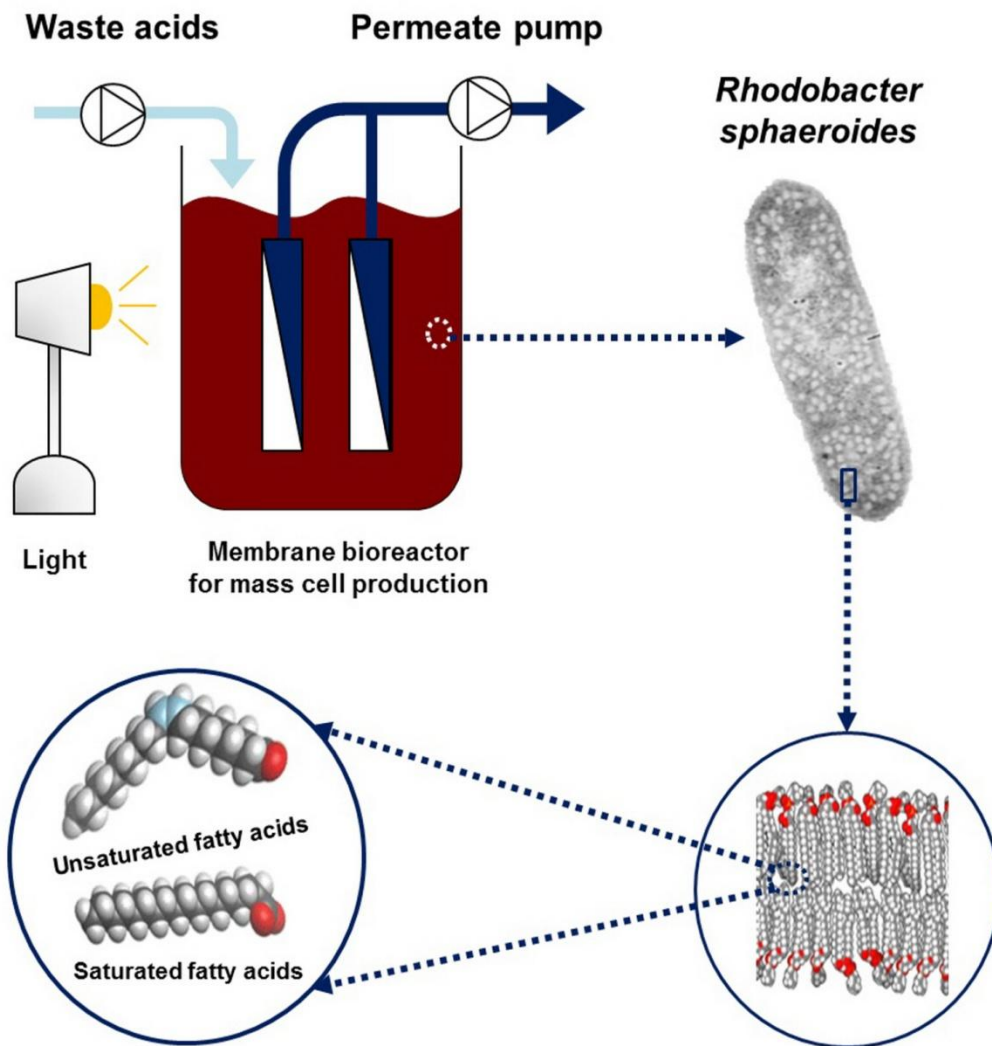


Fig. 4 Fatty acids profile of *Rhodobacter sphaeroides*



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Highlights

- Photosynthetic bacteria cultivation for fatty acids (FA) production from lactate
- Continuous-flow, stirred-tank reactor and membrane-coupled bioreactor
- Maximum cell productivity of 1.9 g dcw/L/d and FA productivity of 665 mg FA/L/d