



## Protein network reconstruction of CHO cell secretory pathway

Lund, Anne Mathilde; Kaas, Christian Schrøder; Kildegaard, Helene Fastrup; Kristensen, Claus; Andersen, Mikael Rørdam

*Publication date:*  
2014

*Document Version*  
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

*Citation (APA):*  
Lund, A. M., Kaas, C. S., Kildegaard, H. F., Kristensen, C., & Andersen, M. R. (2014). *Protein network reconstruction of CHO cell secretory pathway*. Poster session presented at CHOgenome workshop, Vienna, Austria.

---

### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# Protein network reconstruction of CHO cell secretory pathway

Anne Mathilde Lund<sup>1,\*</sup>, Christian Schrøder Kaas<sup>2</sup>, Helene Faustrup Kildegaard<sup>3</sup>, Claus Kristensen<sup>2</sup>, Mikael Rørdam Andersen<sup>1</sup>

(1) Department of Systems Biology, Technical University of Denmark, Kgs. Lyngby, Denmark; (2) Current address: Novo Nordisk, Maaloev, Denmark; (3) Current address: Novo Nordisk Foundation Center for Biosustainability, Technical University of Denmark, Hoersholm, Denmark  
(\* ) amalu@bio.dtu.dk

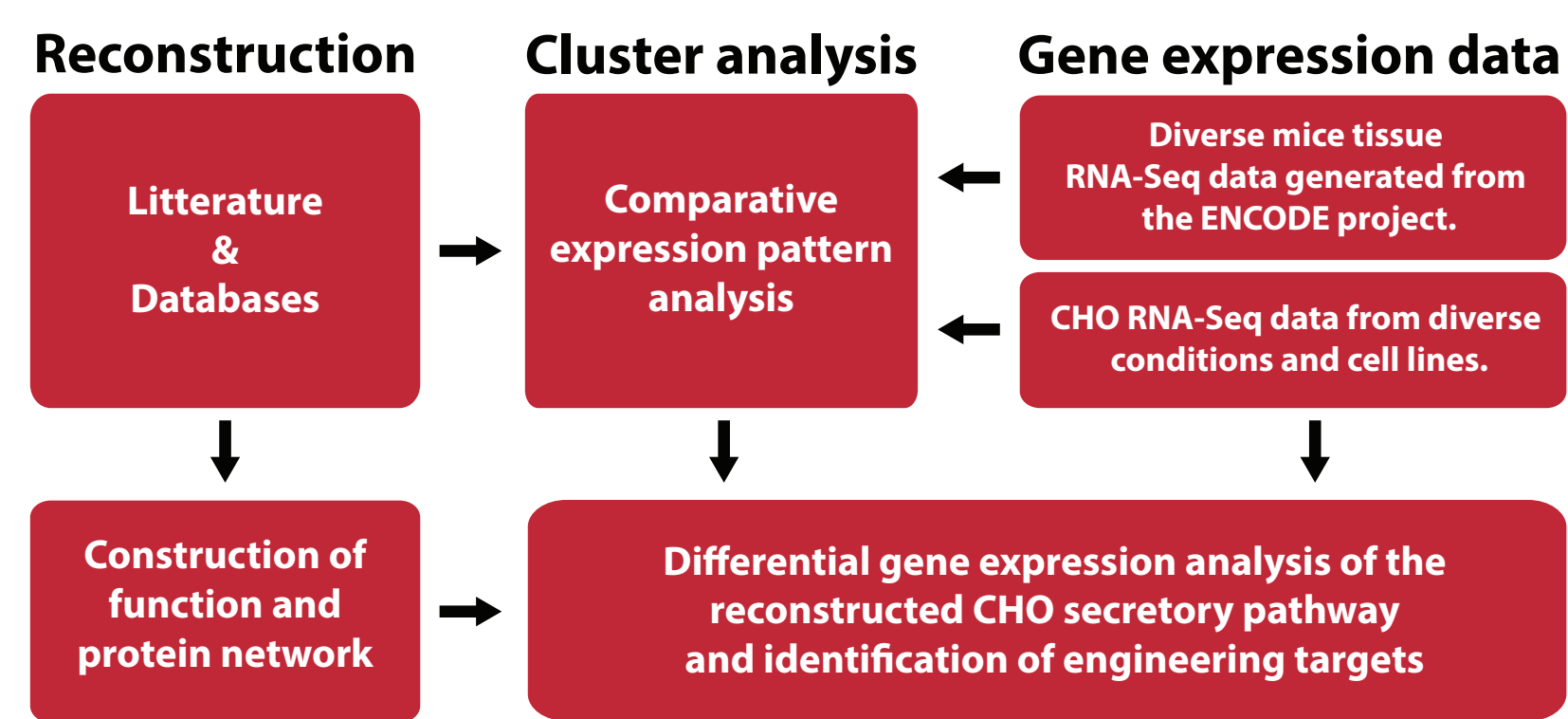
## Introduction

Protein secretion is one of the major bottlenecks in the productivity of recombinant protein in mammalian cells. So far, there have been limited studies of the cell biology of the CHO cell and the potential of cell line engineering. To elucidate the poorly understood cellular processes that control and limit recombinant protein production and secretion, a system-wide study was initiated to identify possibly engineering targets relevant for therapeutic protein production.

### Objective and Strategy

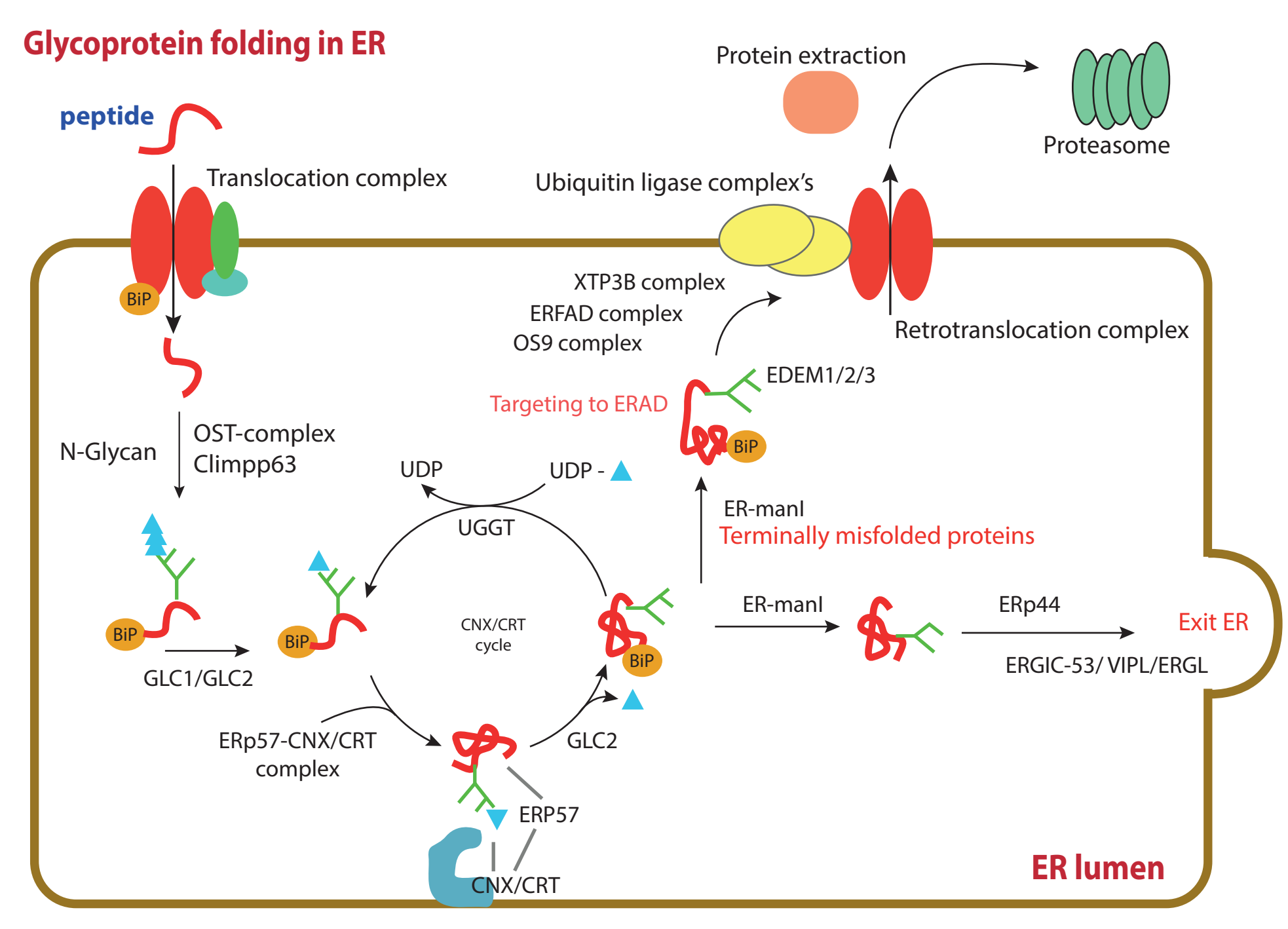
- Introduce a more systematic approach in improving the protein production in CHO cell lines
- Employ a guided approach that integrates protein function interaction network, gene expression and comparative studies of mouse and CHO cells.
- Identify functional gene targets within the secretory pathway for modification in order to increase protein production.

Workflow for reconstruction of CHO cell pathways:



### Pathway reconstruction

Proteins associated or linked to early secretion pathway were identified by manually curate available literature on mouse models and cell lines. The proteins found were used to identify CHO-K1 genes of the ERAD and protein folding pathway.



### RNA-Seq transcriptome data

RNA was extracted with TRIzol reagent under different growth conditions and treatments from diverse sets of CHO cell lines.

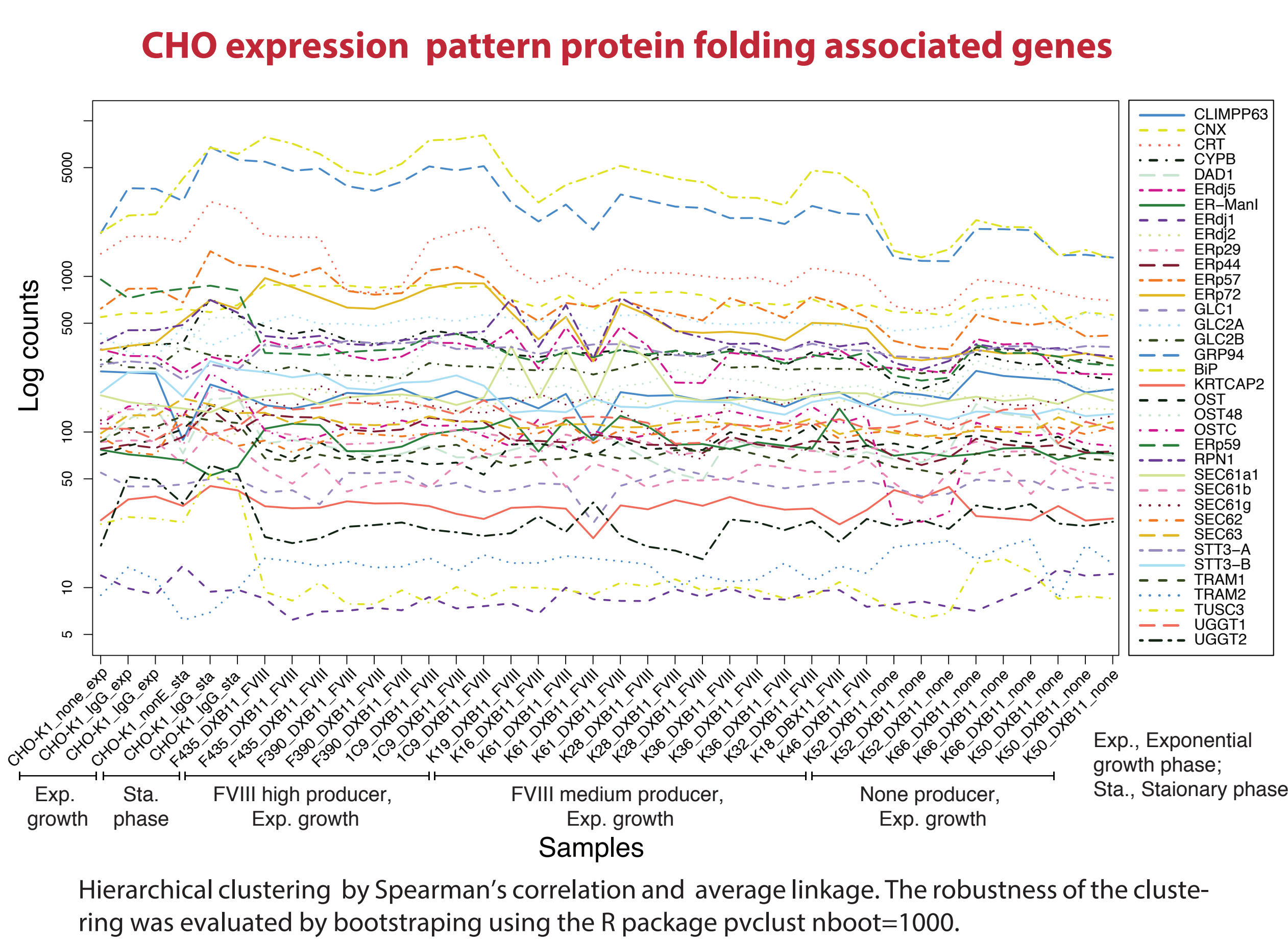
Samples:

CHO-K1		CHO-DXB11	
exp. growth	stationary	NaBu	exp. growth
IgG producer		FVIII producer	
None producer		high	medium low

Library and paired-end RNA sequencing were performed by AROS a/s on Illumina HiSeq 2000 platform with a sequencing depth of min. 35 mio reads. Pre-processing and mapping of RNA-seq reads performed with Bowtie [1] and TopHat [2] using the CHO-K1 genome [3] as reference.

### Gene expression cluster analysis

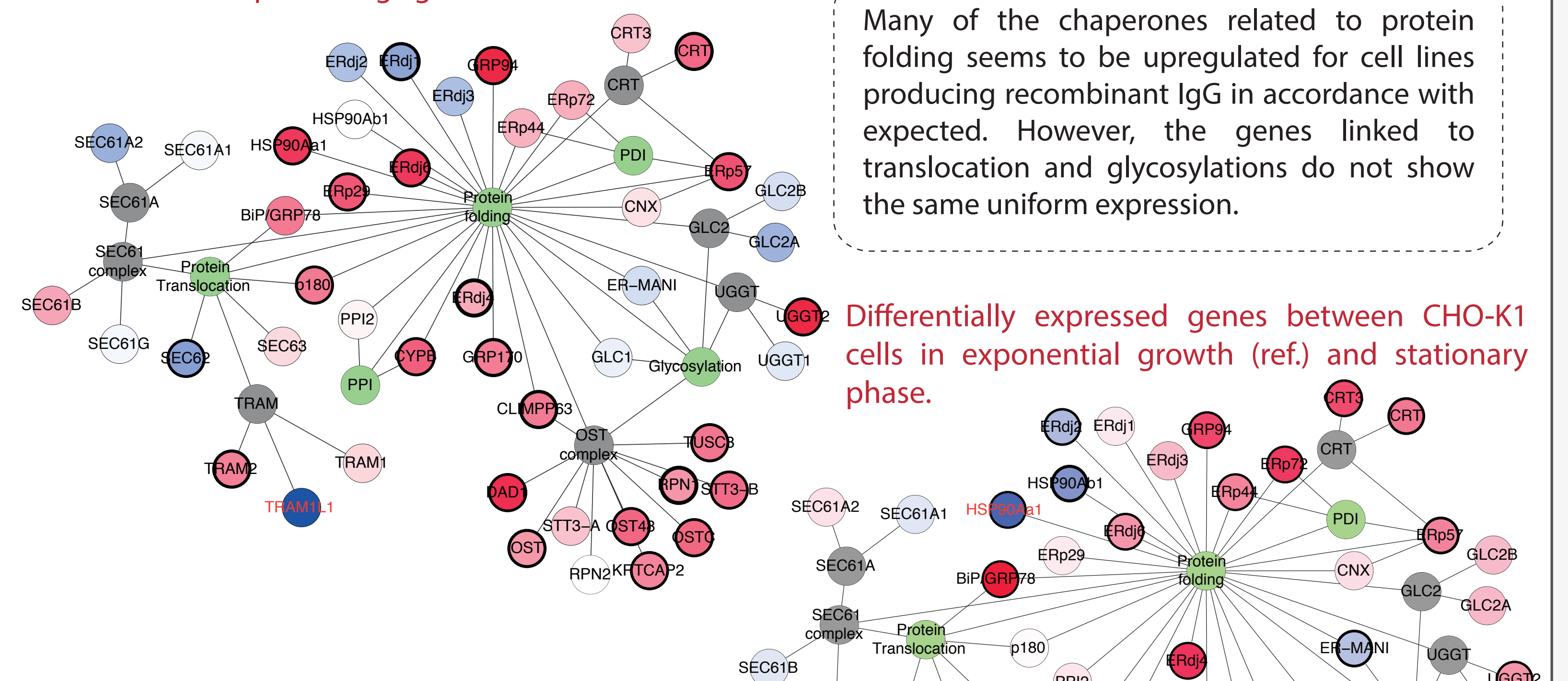
A comparative expression analysis of CHO cells and mice allowed to evaluate CHO cell genes expression patterns and for identification of specific proteins and association with changed network arrangement in CHO cells.



### Integrated gene expression network

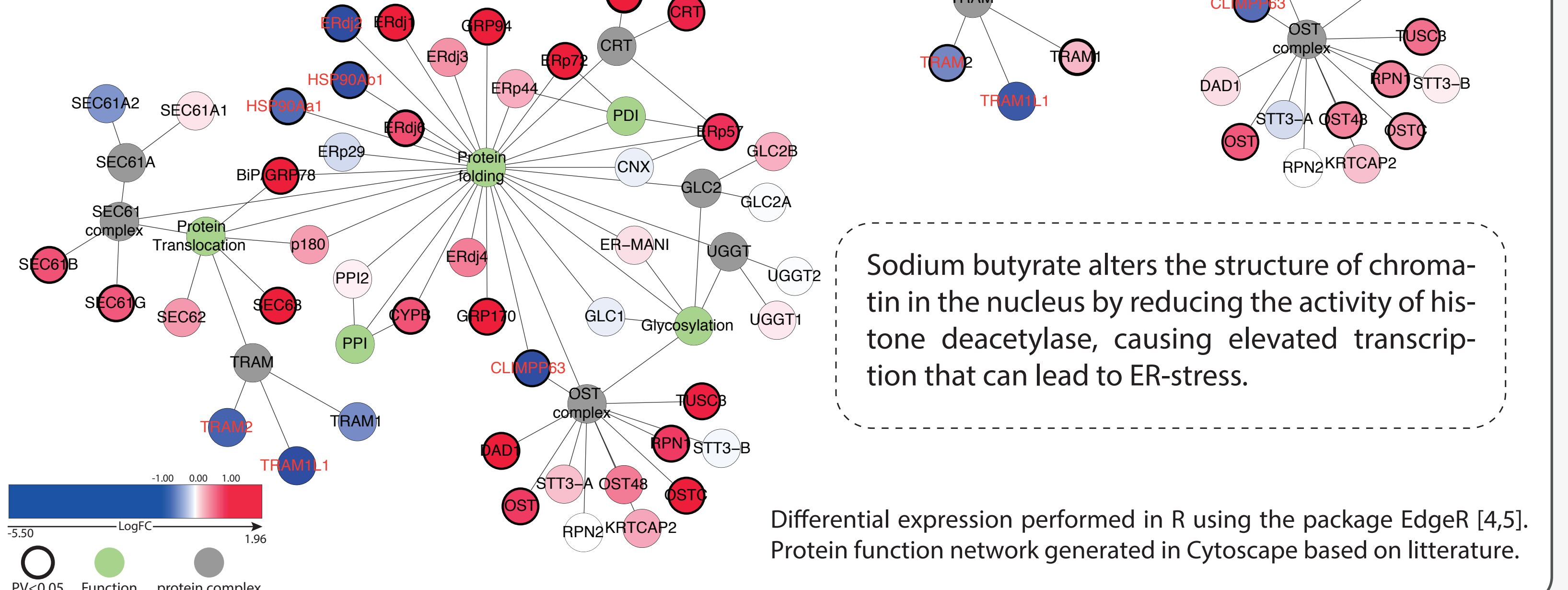
The sub-network of genes associated with translocation and protein folding.

Differentially expressed genes between CHO-K1 cells (ref.) and CHO-K1 cells producing IgG.



Many of the chaperones related to protein folding seems to be upregulated for cell lines producing recombinant IgG in accordance with expected. However, the genes linked to translocation and glycosylations do not show the same uniform expression.

Differentially expressed genes between CHO-K1 cells and CHO-K1 cells treated with NaBu.



Sodium butyrate alters the structure of chromatin in the nucleus by reducing the activity of histone deacetylase, causing elevated transcription that can lead to ER-stress.

Differential expression performed in R using the package EdgeR [4,5]. Protein function network generated in Cytoscape based on literature.

### Conclusion

- This case study showed how a gene function interaction network and gene expression clusters may give insight to biological gene clusters and expression behaviour.
- Possible genetic targets can be evaluated in relation to the systemic network as well as identification of new targets.
- Identify changed regulation in functions or pathways within the secretory pathways.

### References

1. B. Langmead, C. Trapnell, M. Pop, and S. Salzberg, Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biology* 2009, 10:R25.
2. C. Trapnell, L. Pachter, and S.L. Salzberg, TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 2009, 25(9):1105-1111.
3. Xu et al., The genomic sequence of the Chinese hamster ovary (CHO)-K1 cell line. *Nat Biotechnol.* 2011, 29(8):735-741.
4. M.D. Robinson, D.J. McCarthy, and G.K. Smyth, edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* 2010, 26:139-140
5. D.J. McCarthy, Y. Chen, and G.K. Smyth, Differential expression analysis of multifactor RNA-Seq experiments with respect to biological variation. *Nucleic Acids Res.* 2012, 40(10):4288-4297

