



Transfer of Lactic-Acid Bacterial Strains from the Feed to the Sow, the Environment, and the Piglets

Pedersen, Karl; Christensen, G. W.; Steffensen, M.; Schyum, P.; Johansen, A. K.

Published in:
ACTA Veterinaria Scandinavia

Link to article, DOI:
[10.1186/BF03547295](https://doi.org/10.1186/BF03547295)

Publication date:
1992

Document Version
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

Citation (APA):
Pedersen, K., Christensen, G. W., Steffensen, M., Schyum, P., & Johansen, A. K. (1992). Transfer of Lactic-Acid Bacterial Strains from the Feed to the Sow, the Environment, and the Piglets. *ACTA Veterinaria Scandinavia*, 33(4), 297-303. <https://doi.org/10.1186/BF03547295>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Transfer of Lactic Acid Bacterial Strains from the Feed to the Sow, the Environment, and the Piglets

By K. Pedersen, G. W. Christensen, M. Steffensen, P. Schyum and A. K. Johansen

Department of Veterinary Microbiology, Royal Veterinary and Agricultural University, Frederiksberg, Denmark.

Pedersen, K., G. W. Christensen, M. Steffensen, P. Schyum and A. K. Johansen: Transfer of lactic acid bacteria strains from the feed to the sow, the environment and the piglets. Acta vet. scand. 1992, 33, 297-303. – The spread of lactic acid bacterial strains to the environment and to newborn piglets was investigated after feeding of such strains to sows. Rifampicin resistant bacterial strains were fed to sows, 10^{10} c.f.u. per day, during the period from 1 week before expected farrowing until 1 week after farrowing. Fecal samples from the sows and samples of litter were collected for bacteriological examination together with swabs from the pens, the skin of the sows, and from the rectum of the piglets. The test strains were only excreted in relatively low amounts in the feces of the sows, approximately 10^3 - 10^6 c.f.u. per gram. They were not able to displace the normal lactic acid bacterial flora in the sows nor were they transmitted to the intestinal tract of the piglets to any significant extent. After the last administration the test strains disappeared from both feces, skin, and environment, indicating that no permanent colonization had taken place, although considerable differences in duration of persistence were noticed between test strains.

lactobacilli; Enterococcus faecium; intestinal tract.

Introduction

Lactic acid bacteria begin to colonize the gastrointestinal tract of piglets and a number of other animal species shortly after birth (Savage 1977), and during the following days a succession of lactic acid bacterial strains takes place (Tannock *et al.* 1990a). Experiments based on plasmid profiling indicate that the strains colonizing the intestine of piglets mainly are intestinal strains excreted by the sow (Tannock *et al.* 1990a). Even in humans evidence for the transfer of *Enterobacteriaceae* and bifidobacteria from feces of the mother to the infant has been obtained (Tannock *et al.* 1990b). Lactic acid bacteria are often claimed to play an important part for the well-being of the host (Fuller 1989). Experiments spreading some light on the

ecology of lactic acid bacteria with respect to the transfer from mother animal to piglets may be important for the understanding of the acquisition of lactic acid bacteria by the newborn. Furthermore, such experiments may be useful in evaluating the possibilities of administration of potentially beneficial strains to piglets by feeding them to the mother animal. This paper describes an attempt to transfer lactic acid bacterial strains from sows to their newborn piglets by feeding strains to the sows. The excretion of the test strains in the feces of the sows, their contamination of the environment, the bedding, and the skin of the sows, together with the transmission to the digestive tract of the piglets was followed.

Materials and methods

Experimental animals

Four crossbred sows and their piglets were used in the experiments, and housed under standard Danish farming conditions. The sows were kept in separate pens with straw bedding. After farrowing they had access to only approximately one half of their pen, while the piglets had free access to the whole area. The sows received a standard feed mainly based on grained barley with a supplement of protein while the piglets only received maternal milk. Piglets were earmarked and had their tail, deciduous corner incisors, and canine teeth trimmed shortly after birth. Neither sows nor piglets received any medication before or during the experiments.

Bacterial cultures

One of the sows received a culture of *Lactobacillus acidophilus* SS28, while a second was given a culture of *Enterococcus faecium* Cernelle 68 (SF68). The last 2 sows received a mixture of 4 *Lactobacillus* strains: *L. salivarius* SS129 and SS258, *L. gasseri* SS131 and *L. crispatus* SS151. The *Lactobacillus* strains were all isolated from pigs and were all able to adhere to porcine stratified squamous epithelial cells in vitro as previously described (Pedersen & Tannock 1989). *E. faecium* SF68 originated from the probiotic product Cernivet (Bioferment, Lugano, Switzerland) (Lewenstein et al. 1979, Carbone et al. 1980). All strains were rifampicin resistant spontaneous mutants of the parent strains collected on gradient plates of MRS agar (Oxoid) or Mitis-Salivarius agar (Difco) containing rifampicin with a maximum concentration 100 µg/ml. The *L. acidophilus* SS28 strain was propagated in MRS broth (Oxoid) while *E. faecium* SF68 was propagated in brain-heart infusion broth (Difco) at 37°C.

Overnight cultures were centrifuged for 5 min at 6000 rpm. Pellets were resuspended in fresh broth, added 20% sterile glycerol and kept at -80°C in ampoules containing 1 c.f.u., 1 ampoule for each day. The mixture of the 4 *Lactobacillus* strains was freeze dried in a carrier medium of glucose and electrolytes and packed in air and light tight metal ampoules each containing 10¹⁰ viable cells (Lactobacillus Bio Systems, Hørsholm, Denmark). The stability of the rifampicin resistance was examined by repeated subculture in MRS broth or brain-heart infusion broth with and without rifampicin. After incubation at 37°C for 24 h, subcultures were made on MRS or Mitis-Salivarius agar plates with and without rifampicin.

Administration of cultures and collection of samples

From 1 week before expected farrowing to 1 week after farrowing the sows received a daily dose of 10¹⁰ c.f.u. of 1 of the test cultures by adding cultures to the feed. Usually all feed was eaten within 30 min after farrowing. The administration periods ranged between 15 and 18 days. Until farrowing fecal samples and samples of the bedding were collected every 2 or 3 days. Feces were collected from rectum. Furthermore, swabs were taken from the environment. With a metal template a 10 cm² surface area was swabbed with a sterile cotton swab at 5 different sites in the pens, from both floor and equipment. Similarly, 10 cm² of skin was swabbed at 5 different sites on the ventral part of the sow, including the udder. After farrowing the sampling also included rectal swabs from 5 piglets from each litter, randomly selected and marked with numbers from 1 to 5. During the first 2 weeks samples were collected every 2 or 3 days. Thereafter the intervals were prolonged, but samp-

continued until the test strains were no longer detectable. All swabs were immediately transferred to tubes with Stuart Transport Medium (Difco). All samples were kept at 0-5°C until laboratory treatment (less than 2 h).

Culture from samples

Feces and litter samples were suspended in sterile saline as 10-fold dilutions (wt/vol) from which further 10-fold dilutions (vol/vol) were made. The 5 swabs from the environment were pooled in saline and mixed on a whirlmixer, and likewise the 5 swabs from the skin. Ten-fold dilutions from saline (vol/vol) were made. Rectal swabs from the piglets were treated separately. In experiments with *Lactobacillus* strains inoculation from relevant dilutions took place on Rogosa SL agar (Difco) and Rogosa SL agar added rifampicin 50 µg/ml for the enumeration of total lactobacilli and test strains, respectively. Plates were incubated anaerobically in an

atmosphere of N₂, H₂, and CO₂ (GasPak jars, BBL Microbiology Systems, Cockeysville, Md.) for 2 days at 37°C. In the experiment with *E. faecium* SF68 relevant dilutions were plated on Mitis-Salivarius agar (Difco) with and without rifampicin, 100 µg/ml, for the enumeration of total streptococci/enterococci and *E. faecium* SF68, respectively. Mitis Salivarius agar was supplied with nalidixic acid, 15 µg/ml, instead of Chapman tellurite (Barrow *et al.* 1977). Plates were incubated aerobically for 2 days at 37°C. Results were expressed as c.f.u. per gram (feces and litter), per 5 swabs (50 cm² from skin and environment), or per rectal swab.

Results

In all experiments using *Lactobacillus* cultures the total fecal excretion of lactobacilli by the sows was very stable, approximately 10⁸ - 10⁹ viable counts per gram. Counts from 1 of the sows given the mixture of 4 *Lactobacillus* strains are shown in Fig. 1. In the

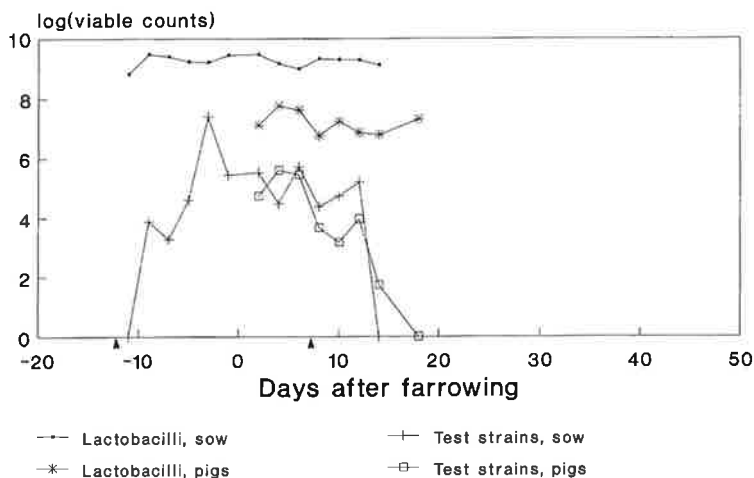


Figure 1. Fecal counts of total lactobacilli and *Lactobacillus* test strains in the sow and the piglets (average of 5 piglets). Counts for the sow are expressed as viable counts per gram feces and for the pigs as viable counts per rectal swab. Test strains were a mixture of 4 *Lactobacillus* strains. First and last day of administration of test strains to the sow is indicated with arrows.

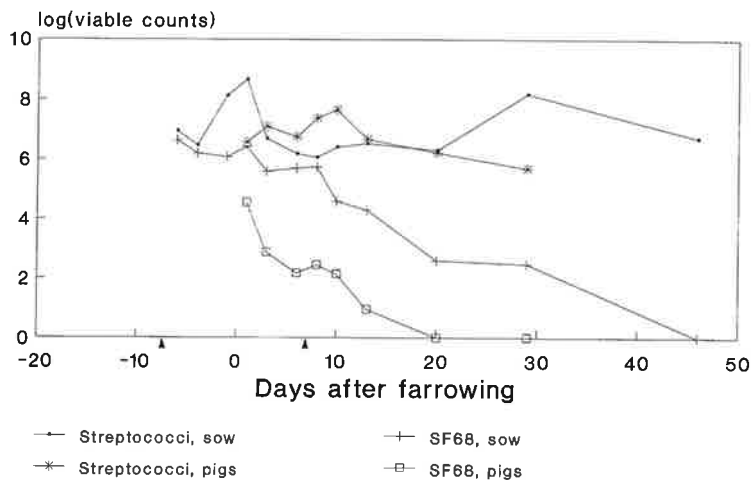


Figure 2. Fecal counts of total streptococci/enterococci and *Enterococcus faecium* SF68 in the sow and piglets (average of 5 piglets). Counts for the sow are expressed as viable counts per gram feces and for piglets as viable counts per rectal swab. First and last day of administration of SF68 to the sow is indicated by arrows.

experiment with *E. faecium* SF68 the total counts of streptococci/enterococci were more fluctuating, but generally ranged in the interval 10^7 - 10^8 c.f.u. per gram (Fig. 2). The counts of the test strains in fecal samples increased during the first few days and reached a maximum 5 - 8 days after the first administration. Thereafter the counts dropped significantly.

None of the *Lactobacillus* test strains reached a higher proportion than about 2.5% of the total *Lactobacillus* counts in feces of the sows (data not shown). In contrast the *E. faecium* SF68 strain was able to reach a much higher proportion of the total Streptococcus/Enterococcus flora, up to 50%. Already 3 days after the final administration, however, the fraction had fallen below 1%. In litter samples and swabs from the environment the total counts of *Lactobacilli* and Streptococci/Enterococci fluctuated more than in fecal samples but ranged around 10^7 viable cells

per gram and per 50 cm², respectively (not shown). Counts in swabs from the piglets fluctuated considerably.

After the last administration, the *Lactobacillus* strains maintained populations about 10^5 - 10^6 c.f.u. per gram of feces for a short period. The *Lactobacillus* mixture strains maintained these counts for 5 - 7 days (Fig. 1) the SS28 strain only for 3 days (data not shown). Thereafter the excretion of the test strains abruptly dropped. This drop was reflected in the counts from skin and in the environment. Four days after the last administration the SS28 strain could no longer be detected. The *E. faecium* strain disappeared more gradually (Fig. 2) and could be detected longer than any of the *Lactobacillus* strains. Twentytwo days after the last administration it could still be detected in both feces, litter, skin, and environment but at 39 days it had disappeared.

Counts of test strains and total *Lactobacillus*

in piglets from one litter receiving the four *Lactobacillus* strains are shown in Fig. 1, while counts of *E. faecium* SF68 and total *Enterococcus*/*Streptococcus* are shown in Fig. 2. The excretion of the test strains by the piglets reached a maximum within a few days and thereafter dropped, although the strains were still fed to the sows. The highest counts of the test strains were observed for almost all piglets at day 1 - 3, but even then they only constituted a very small fraction of the total counts of lactobacilli and streptococci/enterococci, respectively, usually below 1%. However, there was a considerable variation between the piglets in each litter. In 1 of the 2 litters from sows receiving the mixture of the 4 *Lactobacillus* strains the test strains constituted 22.5 % of the lactobacilli in 1 piglet 6 days after birth - a sudden and remarkable rise from 0.08 % at day 4 (Table 1).

Discussion

None of the test cultures were transmitted to the piglets to any significant extent, and none of them were able to colonize permanently inside or outside the animals, although considerable differences in persistence were noticed between strains. They disappeared

from both animals and environment within 4 - 39 days after the last administration. These results are in accordance with those from other experiments (Jonsson 1985, Jonsson *et al.* 1985, Pedersen & Tannock 1987, Pedersen & Tannock 1989). Jonsson (1985) showed that lactic acid bacterial strains fed to newborn piglets could be detected in the feces after 7 days, but had disappeared at 19 days. No effect on performance was observed in her study. Results presented in the literature suggest that a succession of lactic acid bacterial strains takes place in neonatal piglets (Tannock *et al.* 1990a). This may indicate that strains are usually only able to colonize for a certain period of time and thereafter will be supplanted by other strains. However, the capacity of 1 particular strain to colonize more permanently in humans has been suggested (Goldin & Gorbach 1987). The mechanisms behind the succession of lactic acid bacteria are poorly understood. The *Lactobacillus* strains in the present study all originated from the digestive tract of piglets and were able to adhere to porcine squamous epithelial cells *in vitro*. The strains are therefore considered indigenous porcine *Lactobacillus* strains, although they have been shown to be unable to colonize permanently in a previous inoculation study in piglets where they only persisted for a certain period and with considerable differences between strains (Pedersen & Tannock 1989). The *E. faecium* SF68 strain was tested because it is probably the most popular probiotic strain in the world, both for animals and humans.

In most samples collected from the piglets the test strains constituted less than 1% of the total *Lactobacillus* or *Streptococcus*/*Enterococcus* flora. In 1 piglet, however, from 1 of the litters receiving the *Lactobacillus* mixture, the figure suddenly rose from 0.08% at day 4 to 22.5% at day 6 followed by

Table 1. Counts of test strains in rectal swabs of piglets from 1 sow receiving a mixture of 4 *Lactobacillus* strains expressed as % of the total *Lactobacillus* counts.

Days after birth	Piglet 1	Piglet 2	Piglet 3	Piglet 4	Piglet 5
2	1,78	0,34	0,01	0,02	0,08
4	0,08	0,43	0	2,3	0,001
6	22,5	0,003	0,004	3,1	0,02
8	3,4	0,001	0	0,06	0
10	0,49	0	0	0,004	0
12	0,21	0,006	0	0,005	0
14	0,013	0,006	0	0	0
17	0	0	0	0	0

a decline to 3.4% and 0.49% at day 8 and 10, respectively. In this piglet the test strains appeared to establish and multiply for a few days.

The lack of ability to survive in the environment for all test strains, together with the close parallelity between the excretion of the strains in the feces and the counts retrieved in the environment, indicates that the main source of these strains was the mother animal and that essentially no multiplication took place outside the digestive tract. The results therefore suggest that the mother animal was the primary source of the piglets' flora of lactic acid bacteria. This is in accordance with recently published results (Tannock et al. 1990a).

The maximum fecal excretion of the test strains from the sows occurred after 5 - 8 days, i.e. more than 1 week before the last administration. The reason for this is unknown. It was observed in all 4 sows and also in the piglets from which the maximal excretion was reached 1 - 3 days after birth.

For a period *E. faecium* SF68 constituted more than half the total Enterococcus/Streptococcus flora in feces from the sow and it disappeared more slowly than did the Lactobacillus test strains. This may be explained by a better ability to survive in the environment or capacity to persist in the intestine.

The highly fluctuating counts for all test cultures obtained in swabs from the skin may be considered to merely reflect the degree of fecal contamination rather than any colonization of the skin by these microorganisms.

It was not possible from the present data to detect any differences in colonization patterns between freeze dried cultures and frozen broth cultures. Further experiments should be carried out to examine this question.

The rifampicin resistance was found to be a

stable trait as was described by other authors (Compeau et al. 1988). Naturally occurring resistance to rifampicin among Danish lactobacilli and enterococci is negligible (Pedersen unpublished observation).

It was concluded that the intestinal Lactobacillus flora of the sows was too stable to be affected to any significant degree by the given doses of any of the test cultures, whereas the Streptococcus/Enterococcus flora was markedly influenced by the *E. faecium* strain.

References

- Barrow PA, Fuller R, Newport MJ: Changes in the microflora and physiology of the anterior intestinal tract of pigs weaned at 2 days, with special reference to the pathogenesis of diarrhoea. *Infect. Immun.* 1977, 18, 586-595.
- Carbone M, Bonina L, Fera MT: Microbiological properties of *Streptococcus faecium* SF 68 strain and its relationships with other microorganisms. *Boll. Ist. sieroter. milan.* 1980, 59, 591-598.
- Compeau G, Al-Achi BJ, Platsouka E, Levy J: Survival of rifampicin-resistant mutants of *Pseudomonas fluorescens* and *Pseudomonas putida* in soil systems. *Appl. environ. Microbiol.* 1988, 54, 2432-2438.
- Fuller R: A review. Probiotics in man and animals. *J. appl. Bact.* 1989, 66, 365-378.
- Goldin BR, Gorbach SL: *Lactobacillus GG* - a new strain with properties favourable for survival, adhesion and antimicrobial activity in the gastrointestinal tract. *FEMS Microbiol. Lett.* 1987, 46, P72, F1.
- Jonsson E: Persistence in the gut of suckling piglets of a *Lactobacillus* strain and its influence on performance and health. In: Just A, Jørgensen H, Fernandez JA (eds): Proceedings of the International Seminar on Digestive Physiology in the Pig, Copenhagen 16th - 18th May 1985. Beretning fra Statens Husdyrbrugsforsknings Landhusholdningsselskabets Forlag, Frederiksberg, 1985, 288-291.
- Jonsson E, Björck L, Claesson CO: Survival of orally administered *Lactobacillus* strains in the gut of cannulated pigs. *Livest. Prod. Sci.* 1985, 14, 279-285.
- Lewenstein A, Frigerio G, Moroni M: Biological properties of SF 68, a new approach for

treatment of diarrheal diseases. *Curr. ther. Res.* 1979, 26, 967-981.

Pedersen K, Tannock GW: Colonization and persistence of two *Lactobacillus* strains in the digestive tract of piglets. *FEMS Microbiol. Rev.* 1987, 46, P47, F5.

Pedersen K, Tannock GW: Colonization of the porcine gastrointestinal tract by lactobacilli. *Appl. environ. Microbiol.* 1989, 55, 279-283.

Savage DC: Microbial ecology of the gastrointestinal tract. *Ann. Rev. Microbiol.* 1977, 31, 107-133.

Tannock GW, Fuller R, Pedersen K: *Lactobacillus* succession in the piglet digestive tract demonstrated by plasmid profiling. *Appl. environ. Microbiol.* 1990a, 56, 1310-1316.

Tannock GW, Fuller R, Smith SL, Hall MA: Plasmid profiling of the family *Enterobacteriaceae*, lactobacilli, and bifidobacteria to study the transmission of bacteria from mother to infant. *J. clin. Microbiol.* 1990b, 28, 1225-1228.

Sammendrag

Spredning af mælkesyrebakterier fra foderet til soen, miljøet og grisene.

Spredningen af stammer af mælkesyrebakterier til miljøet og pattegrisene blev undersøgt efter tildeling af stammerne til søerne via foderet. Rifampicinresistente teststammer blev i mængder af 10^{10} c.f.u. pr. dag tildelt søerne fra en uge før forventet faring til en uge efter faring. Fæcesprøver fra søerne og prøver af strøelsen blev udtaget til bakteriologisk undersøgelse sammen med svaberprøver fra stierne, fra søernes hud og fra rectum af grisene. Teststammerne blev kun udskilt i fæces fra søerne i relativt lave antal, omkring $10^3 - 10^6$ c.f.u. pr. g. De var således ikke i stand til at fortrænge den naturlige flora af mælkesyrebakterier i tarmen, ligesom de heller ikke i nævneværdig grad blev overført til grisenes tarmkanal. Efter sidste tildeling af teststammerne forsvandt disse fra såvel fæces som fra huden og miljøet. Der forekom således ingen permanent kolonisering, men der var dog betydelige forskelle i persistens mellem de enkelte testkulturer.

(Received March 23, 1992; accepted July 2, 1992).

Reprints may be requested from: K. Pedersen, Department of Veterinary Microbiology, Royal Veterinary and Agricultural University, 13 Bülowvej, DK-1870 Frederiksberg C, Denmark.

